Supporting Information

Sulfonylation-Induced *N*- to *O*-Acetyl Migration in 2-Acetamidoethanol Derivatives

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Experimental Procedures

Benzyl 2-acetamido-4,6-O-benzylidene-2-deoxy-α-D-glucopyranoside (1a)

The compound **1a** was prepared from D-(+)-glucosamine by a variation of the known procedures.¹ Sodium methoxide (32.6 g, 0.60 mol) was added to a suspension of D-(+)-glucosamine hydrochloride (100 g, 0.46 mol) in MeOH (1 L), and the mixture was stirred for 30 min at 40 °C. After addition of acetic anhydride (79 mL, 0.83 mol), the resulting suspension was stirred vigorously for 22 h at 40 °C, and then cooled to 0 °C. After filtration of the reaction mixture, the filtered white solid was washed with cold MeOH and dried to afford *N*-acetyl D-glucosamine (100 g, 97%) as a white powder.

Acetyl chloride (27.4 mL, 0.39 mol) was slowly added to a suspension of *N*-acetyl D-glucosamine (85 g, 0.39 mol) in anhydrous benzyl alcohol (300 mL) under nitrogen atmosphere. The suspension was stirred at room temperature for 2 h, heated to 50 °C for 4 h, and then cooled to room temperature. The resulting yellow solution was slowly poured onto Et_2O (3 L) in ice-water bath, and the mixture was vigorously stirred for 2 h at 0 °C. The precipitate was recovered by filtration and dried under vacuum to afford benzyl 2-acetamido-2-deoxy- α -D-glucopyranoside (113 g, 94%) as a white solid.

Benzaldehyde dimethylacetal (58 mL, 0.39 mol) and *p*-toluenesulfonic acid monohydrate (1.8 g, 9.6 mmol) were added to a solution of benzyl 2-acetamido-2-deoxy- α -D-glucopyranoside (60 g, 0.19 mol) in anhydrous DMF (500 mL), and the mixture was stirred at 70 °C for 24 h. The resulting mixture was cooled to 0 °C and then triethylamine (8.1 mL, 56 mmol) was added. After stirring for 30 min at room temperature, the solvent and extra reagent were removed under reduced pressure to give a white solid. MeOH was added to the solid and the resulting suspension was stirred vigorously for 5 min. After filtration, filtered white solid was washed well with MeOH and dried under vacuum to afford **1a** (67.7 g, 88%) as a white powder. **1a**: ¹H NMR (500 HMz, DMSO-*d*₆) δ 1.85 (s, 3H, *CH*₃CO), 3.51 (dd, *J* = 8.5, 8.5 Hz, 1H, H-4), 3.63–3.80 (m, 3H, H-3, H-5, H-6a), 3.85 (ddd, *J* = 3.6, 8.2, 8.4 Hz, 1H, H-2), 4.14 (dd, *J* = 2.8, 8.6 Hz, 1H, H-6b), 4.48 and 4.70 (AB, *J* = 12.7 Hz, 2H, OCH₂Ph), 4.82 (d, *J* = 2.4 Hz, 1H, H-1), 5.19 (d, *J* = 5.8 Hz, 1H, OH), 5.61 (s, 1H, CHPh), 7.26–7.49 (m, 10H), 8.00 (d, *J* = 8.0 Hz, 1H, NH); ¹³C NMR (126 MHz, DMSO-*d*₆) δ 22.5, 54.2, 62.8, 67.2, 68.5, 82.1, 96.9, 100.9, 126.4, 127.6, 127.6, 128.0, 128.3, 137.7, 137.7, 169.4; IR 3399, 3302, 1651, 1552, 1452, 1374, 1129, 1087, 1059 cm⁻¹; HRMS (FAB) calcd for C₂₂H₂₅NO₆ (M + H⁺), 400.1760, found 400.1763.

General procedures for the tosylation of 2-acetamidoethanol derivatives (Condition A or B)

Condition A: *p*-Toluenesulfonyl chloride (1.05 equiv) and pyridine (3.0 equiv) were added to a solution of substrate in CH_2Cl_2 at 0 °C, and the mixture was stirred at room temperature for 24 h. The solvent was removed under reduced pressure, and the crude product was purified by silica gel column chromatography. **Condition B:** *p*-Toluenesulfonyl chloride (2.0 equiv) was added to a solution of substrate in pyridine, and the mixture was refluxed for 30 h. The mixture was cooled to room temperature, and the solvent was removed under reduced pressure. The crude product was purified by silica gel column temperature, and the solvent was removed under reduced pressure.

Products of the tosylation of the compound 1a

Benzyl 2-acetamido-4,6-*O*-benzylidene-2-deoxy-3-*O*-*p*-toluenesulfonyl-α-D-glucopyranoside (2a) Benzyl 3-*O*-acetyl-4,6-*O*-benzylidene-2-deoxy-2-*p*-toluenesulfoneamido-α-D-glucopyranoside (3a)

The condition B was used for compound **1a** (100 mg, 0.25 mmol), and the crude product was purified by silica gel column chromatography (AcOEt/hexane, 1:3) to afford the product **2a** (46 mg, 33%) as a white solid and the product **3a** (74 mg, 53%) as a white solid. **2a**: ¹H NMR (500 HMz, CDCl₃) δ 2.02 (s, 3H, CH₃CO), 2.28 (s, 3H, CH₃Ph), 3.68 (dd, J = 9.6, 9.6 Hz, 1H, H-4), 3.72 (dd, J = 10.7, 10.7 Hz, 1H, H-6a), 3.88 (ddd, J = 4.8, 9.8, 9.8 Hz, 1H, H-5), 4.21 (dd, J = 5.0, 10.4 Hz, 1H, H-6b), 4.46 (ddd, J = 3.6, 9.2, 10.4 Hz, 1H, H-2), 4.53 and 4.73 (AB, J = 11.9 Hz, 2H, OCH₂Ph), 4.92 (dd,

¹ (a) Berger, I.; Nazarov, A. A.; Hartinger, C. G.; Groessl, M.; Valiahdi, S-M.; Jakupec, M. A.; Keppler, B. K. *ChemMedChem* **2007**, *2*, 505–514. (b) Babic, A.; Pecar, S. *Tetrahedron: Asymmetry* **2008**, *19*, 2265–2271.

J = 10.0, 10.0 Hz, 1H, H-3), 5.02 (d, *J* = 3.8 Hz, 1H, H-1), 5.37 (s, 1H, *CHP*h), 6.00 (d, *J* = 9.0 Hz, 1H, N*H*), 6.97 (d, *J* = 8.0 Hz, 2H, Ts), 7.19–7.44 (m, 10H), 7.67 (d, *J* = 8.4 Hz, 2H, Ts); ¹³C NMR (126 MHz, CDCl₃) δ 21.9 (*C*H₃Ph), 23.4 (*C*H₃CO), 52.5 (C-2), 63.6 (C-5), 68.9 (C-6), 70.6 (OCH₂Ph), 78.4 (C-3), 79.2 (C-4), 98.0 (C-1), 101.8 (CHPh), 126.4, 128.1, 128.3, 128.4, 128.6, 128.9, 129.2, 129.5, 133.9, 136.7, 136.8, 144.6, 170.6 (*C*=O); IR 3324, 1656, 1547, 1362, 1182, 1123, 1097 cm⁻¹; HRMS (FAB) calcd for C₂₉H₃₂NO₈S (M + H⁺), 554.1849, found 554.1848. **3a**: ¹H NMR (500 HMz, CDCl₃) δ 1.72 (s, 3H, *CH*₃CO), 2.37 (s, 3H, *CH*₃Ph), 3.53 (ddd, *J* = 3.8, 10.2, 10.2 Hz, 1H, H-2), 3.56 (dd, *J* = 9.7, 9.7 Hz, 1H, H-4), 3.67 (dd, *J* = 10.3, 10.3 Hz, 1H, H-6a), 3.83 (ddd, *J* = 4.8, 9.8, 9.8 Hz, 1H, H-5), 4.15 (dd, *J* = 4.8, 10.4 Hz, 1H, H-6b), 4.37 and 4.62 (AB, *J* = 11.6 Hz, 2H, OCH₂Ph), 4.71 (d, *J* = 3.8 Hz, 1H, H-1), 5.00 (d, *J* = 10.2 Hz, 1H, N*H*), 5.24 (dd, *J* = 10.1, 10.1 Hz, 1H, H-3), 5.42 (s, 1H, *CHP*h), 7.21 (d, *J* = 8.4 Hz, 2H, Ts), 7.26–7.39 (m, 10H), 7.62 (d, *J* = 8.4 Hz, 2H, Ts); ¹³C NMR (126 MHz, CDCl₃) δ 20.8 (*C*H₃CO), 21.7 (*C*H₃Ph), 56.8 (C-2), 63.1 (C-5), 68.9 (C-6), 69.4 (C-3), 70.5 (OCH₂Ph), 79.4 (C-4), 98.0 (C-1), 101.7 (CHPh), 126.3, 127.1, 128.4, 128.6, 128.6, 128.9, 129.3, 129.9, 136.4, 137.0, 138.4, 143.7, 170.8 (*C*=O); IR 3356, 1734, 1498, 1337, 1234, 1163, 1090 cm⁻¹; HRMS (ESI) calcd for C₂₉H₃₁NNaO₈S (M + Na⁺), 576.1659, found 576.1662.

N-(4-Acetamidobenzyl)acetamide (14)

Acetic anhydride (3.4 mL, 36 mmol) and pyridine (6.6 mL, 82 mmol) were added to a solution of 4aminobenzylamine (2.0 g, 16 mmol) in CH₂Cl₂ (100 mL), and the mixture was stirred for 40 h. After removal of the solvent and extra reagent under reduced pressure, the resulting yellow solid was washed with Et₂O to afford the product **14** (3.3 g, 99%) as a light yellow solid. **14**: ¹H NMR (500 HMz, CD₃OD) δ 1.97 (s, 3H, CH₃CO), 2.11 (s, 3H, CH₃CO), 4.30 (s, 2H, CH₂), 7.22 (d, *J* = 8.6 Hz, 2H), 7.49 (d, *J* = 8.6 Hz, 2H); ¹³C NMR (126 MHz, CD₃OD) δ 22.7 (CH₃CO), 23.9 (CH₃CO), 43.9 (CH₂), 121.4, 129.3, 135.8, 139.2, 171.8 (*C*=O), 173.2 (*C*=O); HRMS (ESI) calcd for C₁₁H₁₄N₂NaO₂ (M + Na⁺), 229.0947, found 229.0966.

trans-2-(Acetamido)cyclohexanol (1b)

Sodium methoxide (356 mg, 6.6 mmol) was added to a solution of *trans*-2-aminocyclohexanol hydrochloride (1.0 g, 6.6 mmol) in MeOH (30 mL), and the mixture was stirred for 15 min. After addition of acetic anhydride (0.69 mL, 7.3 mmol), the resulting mixture was stirred at room temperature for 40 h, and then the solvent and extra reagent were removed under reduced pressure. CHCl₃ was added to the residue and filtrated. The combined filtrate was concentrated to dryness, and the resulting colorless paste was added hexane. The resulting suspension was filtrated to afford the product **1b** (1.03 g, quant.) as a white solid. **1b**: ¹H NMR (500 HMz, CDCl₃) δ 1.11–1.38 (m, 4H), 1.64–1.77 (m, 2H), 1.94 (m, 1H), 2.01 (s, 3H, CH₃CO), 2.05 (m, 1H), 3.31 (ddd, *J* = 4.4, 10.0, 10.0, 1H, H-2), 3.60 (m, 1H, H-1), 5.64 (br s, 1H, OH), 6.16 (br s, 1H, NH); ¹³C NMR (126 MHz, CDCl₃) δ 23.5 (CH₃CO), 24.3 and 24.8 (C-4, C-5), 31.7 (C-3), 34.6 (C-6), 55.9 (C-2), 75.0 (C-1), 172.3 (*C*=O); IR 3275, 1646, 1562, 1450, 1376, 1140, 1069 cm⁻¹; HRMS (ESI) calcd for C₈H₁₆NO₂ (M + H⁺), 158.1176, found 158.1197.

Products of the tosylation of the compound **1b** *trans*-**2**-**Acetamido**-**1**-**O**-**tosyl-cyclohexanol** (**2b**) *trans*-**1**-**O**-**Acetyl-2**-(tosylamido)cyclohexanol (**3b**) *trans*-**2**-(Tosylamido)cyclohexanol (**15b**) *trans*-**2**-**Acetamido**-**1**-**O**-acetyl-cyclohexanol (**16b**)

The condition A was used for compound **1b** (150 mg, 0.93 mmol), and the crude product was purified by silica gel column chromatography (AcOEt/hexane, 1:3 to 1:0) to afford the product **2b** (27 mg, 9%) as a white solid, the product **3b** (107 mg, 36%) as a white solid, the product **15b** (64 mg, 25%) as a white solid and the product **16b** (36 mg, 19%) as a white solid. The structures of the products **3b**,

15b and **16b** were confirmed by comparison to the reported data.² **2b**: ¹H NMR (500 HMz, CDCl₃) δ 1.10–1.37 (m, 3H), 1.54 (m, 1H), 1.64 (m, 1H), 1.73 (m, 1H), 1.88 (m, 1H), 1.94 (s, 3H, CH₃CO), 2.19 (m, 1H), 2.46 (s, 3H, CH_3Ph), 3.85 (m, 1H, H-2), 4.34 (ddd, J = 4.6, 10.8, 10.8 Hz, 1H, H-1), 5.78 (d, J = 7.2 Hz, 1H, NH), 7.35 (d, J = 8.6 Hz, 2H, Ts), 7.78 (d, J = 8.0 Hz, 2H, Ts); ¹³C NMR (126 MHz, CDCl₃) § 21.9 (CH₃Ph), 23.7 (CH₃CO), 24.0 (C-4), 24.4 (C-5), 32.1 (C-6), 32.2 (C-3), 52.5 (C-2), 82.5 (C-1), 127.8, 130.1, 134.2, 145.1, 170.2 (C=O); IR 3436, 1638, 1561, 1370, 1188, 1175 cm⁻¹; HRMS (ESI) calcd for $C_{15}H_{21}NNaO_4S$ (M + Na⁺), 334.1083, found 334.1075. **3b**: ¹H NMR (500 HMz, CDCl₃) δ 1.14–1.38 (m, 4H), 1.64 (m, 1H), 1.69 (m, 1H), 1.76 (s, 3H, CH₃CO), 1.92 (m, 1H), 2.02 (m, 1H), 2.42 (s, 3H, CH₃Ph), 3.19 (m, 1H, H-2), 4.55 (ddd, J = 4.5, 10.2, 10.2 Hz, 1H, H-1), 4.95 (d, J = 7.6 Hz, 1H, NH), 7.29 (d, J = 8.0 Hz, 2H, Ts), 7.74 (d, J = 8.2 Hz, 2H, Ts); ¹³C NMR (126 MHz, CDCl₃) δ 21.1 (CH₃CO), 21.7 (CH₃Ph), 23.9 and 24.4 (C-4, C-5), 31.3 (C-6), 33.7 (C-3), 57.1 (C-2), 74.3 (C-1), 127.1, 129.8, 138.8, 143.2, 171.6 (C=O); IR 3261, 1715, 1599, 1453, 1335, 1257, 1163, 1094, 1046 cm⁻¹ HRMS (ESI) calcd for $C_{15}H_{22}NO_4S$ (M + H⁺), 312.1264, found 312.1277. **15b**: ¹H NMR (500 HMz, CDCl₃) δ 1.05–1.28 (m, 4H), 1.58 (m, 1H), 1.65 (m, 1H), 1.73 (m, 1H), 2.01 (m, 1H), 2.43 (s, 3H, CH₃Ph), 2.76 (br s, 1H, OH), 2.86 (m, 1H, H-2), 3.31 (ddd, J = 4.3, 9.6, 9.6 Hz, 1H, H-1), 5.10 (d, J = 7.0 Hz, 1H, NH), 7.32 (d, J = 8.0 Hz, 2H, Ts), 7.80 (d, J = 8.4 Hz, 2H, Ts); ¹³C NMR (126 MHz, CDCl₃) δ 21.8 (CH₃Ph), 24.0, 24.8, 32.0, 33.5, 59.9 (C-2), 73.5 (C-1), 127.3, 130.0, 137.5, 143.8; HRMS (ESI) calcd for $C_{13}H_{20}NO_3S$ (M + H⁺), 270.1158, found 270.1129. **16b**: ¹H NMR (500 HMz, CDCl₃) δ 1.14 (m, 1H), 1.19–1.35 (m, 2H), 1.44 (m, 1H), 1.66 (m, 1H), 1.74 (m, 1H), 1.89 (s, 3H, CH₃CO), 1.92 (m, 1H), 2.01 (s, 3H, CH₃CO), 2.03 (m, 1H), 3.83 (dddd, J = 4.2, 4.4, 10.2, 10.2 Hz, 1H, H-2), 4.61 (ddd, J = 4.5, 10.7, 10.7 Hz, 1H, H-1), 5.81 (br s, 1H, NH); ¹³C NMR (126 MHz, CDCl₃) δ 21.3 (CH₃CO), 23.5 (CH₃CO), 24.2 and 24.3 (C-4, C-5), 31.2 (C-6), 32.2 (C-3), 52.9 (C-2), 74.8 (C-1), 169.9 (C=O), 172.1 (C=O); IR 3289, 1732, 1646, 1559, 1375, 1246, 1042 cm⁻¹; HRMS (ESI) calcd for $C_{10}H_{17}NNaO_3$ (M + Na⁺), 222.1101, found 222.1104.

Products of the tosylation of 2-acetamidoethanol (1c)

1-O-Acetyl-2-(tosylamido)ethanol (3c)

2-(Tosylamido)ethanol (15c)

2-Acetamido-1-*O*-acetyl-ethanol (16c)

The condition A was used for 2-acetamidoethanol (1.12 g, 10.9 mmol), and the crude product was purified by silica gel column chromatography (AcOEt/hexane, 1:3 to 1:0) to afford the product **3c** (1.01 g, 36%) as a colorless oil, the product **15c** (748 mg, 32%) as a white paste and the product **16c** (457 mg, 29%) as a colorless oil. **3c**: ¹H NMR (500 HMz, CDCl₃) δ 2.00 (s, 3H, *CH*₃CO), 2.43 (s, 3H, *CH*₃Ph), 3.22 (dt, *J* = 5.4, 5.7 Hz, 1H, H-2), 4.09 (t, *J* = 5.3 Hz, 1H, H-1), 5.06 (t, *J* = 5.6 Hz, 1H, NH), 7.32 (d, *J* = 7.8 Hz, 2H, Ts), 7.76 (d, *J* = 8.2 Hz, 2H, Ts); ¹³C NMR (126 MHz, CDCl₃) δ 20.9 (*C*H₃CO), 21.7 (*C*H₃Ph), 42.3 (C-2), 63.1 (C-1), 127.2, 130.0, 137.0, 143.9, 171.0 (*C*=O); IR 3237, 1736, 1719, 1596, 1441, 1331, 1252, 1160, 1091, 1051 cm⁻¹; HRMS (ESI) calcd for C₁₁H₁₅NNaO₄S (M + Na⁺), 280.0614, found 280.0603. **15c**: ¹H NMR (500 HMz, CDCl₃) δ 2.42 (s, 3H, *CH*₃Ph), 3.08 (m, 1H, H-2), 3.61 (m, 1H, *OH*), 3.68 (m, 1H, H-1), 5.42 (br m, 1H, NH), 7.31 (d, *J* = 7.8 Hz, 2H, Ts), 7.76 (d, *J* = 8.2 Hz, 2H, Ts); ¹³C NMR (126 MHz, CDCl₃) δ 21.7 (*C*H₃Ph), 45.4 (C-2), 61.4 (C-1), 127.3, 130.0, 136.7, 143.8; HRMS (ESI) calcd for C₉H₁₃NNaO₃S (M + Na⁺), 238.0508, found 238.0497. **16c**: ¹H NMR (500 HMz, CDCl₃) δ 1.98 (s, 3H, *CH*₃CO), 2.06 (s, 3H, *CH*₃CO), 3.49 (m, 1H, H-2), 4.13 (m, 1H, H-1), 5.07 (br s, 1H, NH); ¹³C NMR (126 MHz, CDCl₃) δ 21.0 (*C*H₃CO), 23.3 (*C*H₃CO), 38.9 (C-2), 63.4 (C-1), 171.3

² For compound **3b**: (a) Liu, Y-K.; Li, R.; Yue, L.; Li, B-J.; Chen, Y-C.; Wu, Y.; Ding, L-S. Org. Lett. **2006**, 8, 1521–1524. For compound **15b**: (b) Wang, Z.; Cui, Y-T.; Xu, Z-B.; Qu J. J. Org. Chem. **2008**, 73, 2270–2274. (c) Fan, R-H.; Hou, X-L. Org. Biomol. Chem. **2003**, 1, 1565–1567. For compound **16b**: (d) Miller, S. J.; Copeland, G. T.; Papaioannou, N.; Horstmann, T. E.; Ruel, E. M. J. Am. Chem. Soc. **1998**, 120, 1629–1630. (e) Tabanella, S.; Valancogne, I.; Jackson, R. F. W. Org. Biomol. Chem. **2003**, 1, 4254–4261.

(C=O); IR 3294, 1740, 1658, 1557, 1375, 1237, 1054 cm⁻¹; HRMS (ESI) calcd for C₆H₁₁NNaO₃ (M + Na⁺), 168.0631, found 168.0625.

1-(Acetamidomethyl)cyclohexanol (1d)

Sodium methoxide (489 mg, 9.1 mmol) was added to a solution of 1-aminomethyl-1-cyclohexanol hydrochloride (1.5 g, 9.1 mmol) in MeOH (30 mL), and the mixture was stirred for 15 min. After addition of acetic anhydride (0.94 mL, 10 mmol), the resulting mixture was stirred at room temperature for 25 h, and then the solvent and extra reagent were removed under reduced pressure. CH₂Cl₂ was added to the residue, and the resulting suspension was filtrated. The combined filtrate was concentrated to afford the product **1d** (1.55 g, quant.) as a white solid. **1d**: ¹H NMR (500 HMz, CDCl₃) δ 1.25–1.63 (m, 10H), 2.01 (s, 3H, CH₃CO), 2.80 (br s, 1H), 3.25 (d, *J* = 6.0 Hz, 2H, CH₂NHAc), 6.32 (br s, 1H); ¹³C NMR (126 MHz, CDCl₃) δ 22.2, 23.4, 25.9, 35.7, 49.3 (CH₂NHAc), 71.7 (C-1), 171.5 (*C*=O); IR 3391, 3290, 1652, 1552, 1421, 1292, 1274, 1166, 1127, 1044 cm⁻¹; HRMS (ESI) calcd for C₉H₁₇NNaO₂ (M + Na⁺), 194.1151, found 194.1147.

Product of the tosylation of the compound 1d 1-O-Acetyl-1-[(tosylamido)methyl]cyclohexanol (3d)

The condition Å was used for compound **1d** (150 mg, 0.88 mmol), and the crude product was purified by silica gel column chromatography (AcOEt) to afford the product **3d** (148 mg, 52%) as a white solid. The condition B gave the poduct **3d** in 96% from the same amount of starting material. **3d**: ¹H NMR (500 HMz, CDCl₃) δ 1.29 (m, 2H), 1.37–1.59 (m, 6H), 1.98 (s, 3H, CH₃CO), 2.08 (m, 2H), 2.42 (s, 3H, CH₃Ph), 3.25 (d, *J* = 6.6 Hz, 2H, CH₂NHTs), 5.53 (t, *J* = 6.6 Hz, 1H, NH), 7.29 (d, *J* = 7.8 Hz, 2H, Ts), 7.72 (d, *J* = 8.2 Hz, 2H, Ts); ¹³C NMR (126 MHz, CDCl₃) δ 21.7 (CH₃Ph), 21.7 (C-3, C-5), 22.3 (CH₃CO), 25.4 (C-4), 32.8 (C-2, C-6), 49.4 (CH₂NHTs), 83.0 (C-1), 127.1, 129.9, 137.3, 143.4, 171.4 (*C*=O); IR 3284, 1734, 1599, 1450, 1328, 1236, 1163, 1094, 1064 cm⁻¹; HRMS (ESI) calcd for C₁₆H₂₃NNaO₄S (M + Na⁺), 348.1240, found 348.1246.

2-Acetamido-1,5-anhydro-4,6-O-benzylidene-2-deoxy-D-glucitol (1e)

Sodium methoxide (110 mg, 2.1 mmol) was added to a solution of 2-acetamido-3-*O*-acetyl-1,5anhydro-4,6-*O*-benzylidene-2-deoxy-D-glucitol³ (700 mg, 2.1 mmol) in MeOH (15 mL), and the mixture was stirred for 3 h at room temperature. The reaction mixture was quenched by the addition of the Amberlite IR-120 (H⁺), and the resulting mixture was stirred for 5 min. The resin was filtered, and the filtrate was concentrated to dryness to afford the product **1e** (610 mg, quant.) as a white solid. **1e**: ¹H NMR (500 HMz, CDCl₃/CD₃OD = 95:5) δ 1.95 (s, 3H, CH₃CO), 3.15 (dd, *J* = 11.0, 11.0 Hz, 1H, H-1a), 3.32 (ddd, *J* = 5.2, 9.6, 9.6 Hz, 1H, H-5), 3.50 (dd, *J* = 9.2, 9.2 Hz, 1H, H-4), 3.62 (dd, *J* = 9.5, 9.5 Hz, 1H, H-3), 3.68 (dd, *J* = 10.4, 10.4 Hz, 1H, H-6a), 3.94 (ddd, *J* = 5.4, 10.4, 10.4 Hz, 1H, H-2), 4.05 (dd, *J* = 5.5, 11.1 Hz, 1H, H-1b), 4.26 (dd, *J* = 5.0, 10.6 Hz, 1H, H-6b), 5.52 (s, 1H, CHPh), 7.30–7.50 (m, 5H); ¹³C NMR (126 MHz, CDCl₃/CD₃OD = 95:5) δ 22.9 (CH₃CO), 52.2 (C-2), 68.7 (C-1), 68.8 (C-6), 71.5 (C-5), 72.4 (C-3), 82.1 (C-4), 102.0 (CHPh), 126.4, 128.4, 129.4, 137.2, 172.3 (C=O); IR 3426, 3277, 1655, 1544, 1377, 1127, 1100 cm⁻¹; HRMS (FAB) calcd for C₁₅H₂₀NO₅ (M + H⁺), 294.1336, found 294.1339.

Product of the tosylation of the compound **1e**

3-*O*-Acetyl-1,**5**-anhydro-4,**6**-*O*-benzylidene-2-deoxy-2-tosylamido-D-glucitol (3e)

The condition B was used for compound **1e** (50 mg, 0.17 mmol), and the crude product was purified by silica gel column chromatography (AcOEt/hexane, 1:3) to afford the product **3e** (93 mg, 94%) as a white solid. **3e**: ¹H NMR (500 HMz, CDCl₃) δ 1.70 (s, 3H, CH₃CO), 2.44 (s, 3H, CH₃Ph), 3.34 (dd, J = 11.3, 11.3 Hz, 1H, H-1a), 3.38 (ddd, J = 5.0, 9.6, 9.6 Hz, 1H, H-5), 3.49 (m, 1H, H-2), 3.56 (dd, J = 9.4,

⁵ Hesek, D.; Lee, M.; Yamaguchi, T.; Noll, B. C.; Mobashery, S. J. Org. Chem. 2008, 73, 7349-7352.

9.4 Hz, 1H, H-4), 3.67 (dd, J = 10.3, 10.3 Hz, 1H, H-6a), 4.13 (dd, J = 5.4, 11.6 Hz, 1H, H-1b), 4.30 (dd, J = 5.0, 10.6 Hz, 1H, H-6b), 4.96 (dd, J = 9.7, 9.7 Hz, 1H, H-3), 5.32 (d, J = 6.8 Hz, 1H, NH), 5.45 (s, 1H, CHPh), 7.33 (d, J = 7.8 Hz, 2H, Ts), 7.30–7.42 (m, 5H), 7.75 (d, J = 8.2 Hz, 2H, Ts); ¹³C NMR (126 MHz, CDCl₃) δ 20.8 (CH₃CO), 21.7 (CH₃Ph), 54.6 (C-2), 68.8 (C-6), 70.8 (C-1), 71.6 (C-5), 73.0 (C-3), 79.0 (C-4), 101.7 (CHPh), 126.3, 127.2, 128.4, 129.4, 130.0, 137.0, 137.7, 143.8, 172.3 (C=O); IR 3287, 1745, 1455, 1227, 1167, 1088 cm⁻¹; HRMS (FAB) calcd for C₂₂H₂₆NO₇S (M + H⁺), 448.1424, found 448.1421.

Product of the tosylation of 2-acetamidophenol (1f)

1-O-Tosyl-2-(acetamido)phenol (2f)

The condition A was used for 2-acetamidophenol (1.0 g, 6.6 mmol), and the crude product was purified by silica gel column chromatography (AcOEt/hexane, 1:1) to afford the product **2f** (1.98 g, 98%) as colorless crystals. The condition B gave the poduct **2f** in 98% from the same amount of starting material. **2f**: ¹H NMR (500 HMz, CDCl₃) δ 2.07 (s, 3H, CH₃CO), 2.47 (s, 3H, CH₃Ph), 6.91 (d, *J* = 8.0 Hz, 1H, C-3), 6.99 (dd, *J* = 7.8, 7.8 Hz, 1H, C-4), 7.24 (dd, *J* = 7.8, 7.8 Hz, 1H, C-5), 7.35 (d, *J* = 8.0 Hz, 2H, Ts), 7.56 (br s, 1H, NH), 7.73 (d, *J* = 8.4 Hz, 2H, Ts), 8.19 (d, *J* = 8.2 Hz, 1H, C-6); ¹³C NMR (126 MHz, CDCl₃) δ 22.0 (CH₃Ph), 24.8 (CH₃CO), 123.0, 124.5, 128.0, 128.7, 130.3, 131.3, 131.9, 139.1, 146.5, 168.3 (*C*=O); IR 3400, 3302, 1695, 1675, 1600, 1525, 1442, 1372, 1307, 1194, 1170, 1088 cm⁻¹; HRMS (ESI) calcd for C₁₅H₁₅NNaO₄S (M + Na⁺), 328.0614, found 328.0599.

Crystal was examined under Infineum V8512 oil. A suitable sample was glued to a glass capillary and then transferred to the 100 K N₂ stream for compound **2f** of a Bruker SMART Apex CCD diffractometer. Preliminary unit cell parameters and crystal system analysis were determined from the centroids of reflections with I > 20 σ (I) from three sets of 30 0.5° ω -scan frames ($\varphi = 0, 120, 240^{\circ}$); final unit cell parameters were determined from the centroids of reflections with I > 10 σ (I) from all data. An arbitrary, redundant, sphere of data was calculated using COSMO, included in the Apex2 suite of programs.⁴ Data are uncorrected for absorption and Lorentz polarization effects. Structure solution and refinement utilized the programs of the SHELXTL software package.⁵ Full detail of the X-ray structure determination is in the CIF file and ORTEP diagram is given in Figure S1.

trans-4-(Acetamido)cyclohexanol (1g)

Sodium methoxide (1.07 g, 19.8 mmol) was added to a solution of *trans*-4-aminocyclohexanol hydrochloride (3.0 g, 19.8 mmol) in MeOH (100 mL), and the mixture was stirred for 15 min. After addition of acetic anhydride (2.06 mL, 21.8 mmol), the mixture was stirred at room temperature for 26 h, and then the solvent and extra reagent were removed under reduced pressure. CHCl₃ was added to the residue, and the resulting suspension was filtrated. The combined filtrate was concentrated to dryness, and the resulting colorless paste was added hexane. The resulting suspension was filtrated to afford the product **1g** (3.11 g, quant.) as a white solid. **1g**: ¹H NMR (500 HMz, CD₃OD) δ 1.20–1.40 (m, 4H), 1.84–1.98 (m, 4H), 1.90 (s, 3H, CH₃CO), 3.51 (m, 1H), 3.59 (m, 1H), 7.92 (br d, *J* = 7.0 Hz, 1H, N*H*); ¹³C NMR (126 MHz, CD₃OD) δ 22.8 (CH₃CO), 31.6 and 34.9 (C-2, C-3, C-5 and C-6), 49.3 (C-4), 70.5 (C-1), 172.6 (*C*=O); IR 3277, 1636, 1562, 1456, 1371, 1328, 1148, 1112, 1065 cm⁻¹; HRMS (ESI) calcd for C₈H₁₆NO₂ (M + H⁺), 158.1176, found 158.1171.

Products of the tosylation of the compound 1g trans-4-Acetamido-1-O-tosyl-cyclohexanol (2g) trans-4-(Tosylamido)cyclohexanol (15g) trans-4-Acetamido-1-O-acetyl-cyclohexanol (16g)

The condition A was used for compound 1g (450 mg, 2.9 mmol), and the crude product was purified by silica gel column chromatography (AcOEt) to afford the product 2g (389 mg, 41%) as colorless

⁴ Apex2. Bruker-AXS: Madison, WI, 2008; Vol. 58.

⁵ Sheldrick, G. M., Acta Crystallogr. A. 2008, 64, 112–122.

crystals, the product 15g (69 mg, 9%) as a white solid and the product 16g (46 mg, 8%) as colorless crystals. **2g**: ¹H NMR (500 HMz, CDCl₃) δ 1.06–1.27 (m, 2H), 1.48–1.66 (m, 2H), 1.84–2.03 (m, 4H), 1.89 (s, 3H, CH_3CO), 2.43 (s, 3H, CH_3Ph), 3.68 (m, 1H, H-4), 4.40 (ddd, J = 3.9, 10.6, 10.6 Hz, 1H, H-1), 5.69 (d, J = 7.4 Hz, 1H, NH), 7.32 (d, J = 8.1 Hz, 2H, Ts), 7.75 (d, J = 8.1 Hz, 2H, Ts); ¹³C NMR (126 MHz, CDCl₃) δ 21.8 (CH₃Ph), 23.5 (CH₃CO), 30.2 and 30.9 (C-2, C-3, C-5, C-6), 46.8 (C-4), 80.4 (C-1), 127.6, 130.0, 134.4, 144.9, 169.7 (C=O); IR 3408, 3312, 1674, 1652, 1552, 1519, 1343, 1189, 1177, 1096 cm⁻¹; HRMS (ESI) calcd for $C_{15}H_{21}NNaO_4S$ (M + Na⁺), 334.1083, found 334.1072. **15g**: ¹H NMR (500 HMz, CDCl₃) δ 1.10–1.24 (m, 4H), 1.69–1.86 (m, 4H), 2.39 (s, 3H, CH₃Ph), 3.01 (m, 1H, H-4), 3.46 (m, 1H, H-1), 5.39 (d, J = 7.6 Hz, 1H, NH), 7.26 (d, J = 8.2 Hz, 2H, Ts), 7.72 (d, J = 8.2 Hz, 2H, Ts); ¹³C NMR (126 MHz, CDCl₃) δ 21.6 (CH₃Ph), 31.3 and 33.5 (C-2, C-3, C-5, C-6), 51.9 (C-4), 69.1 (C-1), 126.9, 129.8, 138.2, 143.4; HRMS (ESI) calcd for $C_{13}H_{19}NNaO_3S$ (M + Na⁺), 292.0978, found 292.0955. **16g**: ¹H NMR (500 HMz, CDCl₃) δ 1.13–1.31 (m, 2H), 1.35–1.53 (m, 2H), 1.86–2.03 (m, 4H), 1.91 (s, 3H, CH₃CO), 1.99 (s, 3H, CH₃CO), 3.72 (dtt, J = 4.0, 7.7, 11.3 Hz, 1H, H-4), 4.61 (tt, J = 4.0, 10.8 Hz, 1H, H-1), 5.81 (br d, J = 7.2 Hz, 1H, NH); ¹³C NMR (126 MHz, CDCl₃) δ 21.5 (CH₃CO), 23.5 (CH₃CO), 30.1 and 30.6 (C-2, C-3, C-4, C-5), 47.3 (C-4), 72.1 (C-1), 169.7 (C=O), 170.9 (C=O); IR 3440, 3251, 1726, 1638, 1562, 1365, 1250, 1035 cm⁻¹; HRMS (ESI) calcd for $C_{10}H_{17}NNaO_3 (M + Na^+)$, 222.1101, found 222.1087.

Product of the mesylation of the compound **1d**

1-O-Acetyl-1-[(mesylamido)methyl]cyclohexanol (19)

Methanesulfonyl chloride (90 µL, 1.2 mmol) was added to a solution of **1d** (100 mg, 0.58 mmol) in pyridine (1.0 mL) at 0 °C. The mixture was stirred at room temperature for 20 h, and then at 40 °C for 1 h. The solvent was removed under reduced pressure, and the crude product was purified by silica gel column chromatography (AcOEt/hexane, 2:3) to afford the product **19** (137 mg, 94%) as a white solid. **19**: ¹H NMR (500 HMz, CDCl₃) δ 1.33 (m, 2H), 1.43–1.63 (m, 6H), 2.07 (s, 3H, CH₃CO), 2.13 (m, 2H), 2.95 (s, 3H, CH₃SO₂), 3.48 (d, *J* = 6.6 Hz, 2H, CH₂NHMs), 5.15 (t, *J* = 6.2 Hz, 1H, NH); ¹³C NMR (126 MHz, CDCl₃) δ 21.7 (C-3, C-5), 22.3 (CH₃CO), 25.4 (C-4), 32.9 (C-2, C-6), 40.6 (Ms), 49.8 (CH₂NHMs), 83.1 (C-1), 171.5 (*C*=O); IR 3249, 1733, 1453, 1302, 1232, 1150 cm⁻¹; HRMS (ESI) calcd for C₁₀H₁₉NNaO₄S (M + Na⁺), 272.0927, found 272.0908.



Figure S1. The ORTEP diagram of compound 2f, shown at 30% probability level.

exp1 s2pul

	SAMPLE	DEC	2. & VT
date	Mar 22 2009	dfrq	499.866
solver	nt DMSO	dn	н1
file	exp	dpwr	30
ACC	UISITION	dof	0
sfrq	499.866	dm	nnn
tn	H1	dmm	c
at	5.016	dmf	200
np	65536	dseq	
sw	6533.3	dres	1.0
fb	4000	homo	n
bs	4		DEC2
tpwr	61	dfrg2	0
pw	13.5	dn2	
d1	0.100	dpwr2	1
tof	269.9	dof2	0
nt	16	dm 2	n
ct	16	dmm2	c
alock	n	dmf2	200
gain	not used	dsea2	
3	FLAGS	dres2	1.0
i1	n	homo2	n
in	n		DEC3
dp	v	dfro3	0
hs	nň	dn3	
1	DISPLAY	dpwr3	1
sp	-99.5	dof3	0
wp	5099.3	dm3	n
vs	29	dmm3	c
SC	0	dmf3	200
wc	250	dseg3	
hzmm	20.40	dres3	1.0
is	91.80	homo3	n
rf1	1752.7	PRO	CESSING
rfp	1249.7	wtfile	
th	7	proc	ft
ins	1.000	fn	65536
ai	ph	math	f
		werr	
		wexp	process p1H
		a share	





exp2 s2pu1 SAMPLE DEC. & VT date Jun 20 2009 dfrg 499.866 solvent DMS0 dn H1 file exp dpwr 40 ACQUISITION dof 0 TION dof 125.703 dm C13 dmm 1.215 dmf 65536 dseq 26963.3 dres sfrq УУУ tn at W 8787.35 np 1.0 sw 15000 homo n bs DEC2 4 4 52 dfrq2 10.2 dn2 1.800 dpwr2 144.5 dof2 1000 dm2 tpwr 0 pw d1 1 tof 0 nt ct alock n 48 dmm2 C not used dseq2 FLAGS . 10000 gain 1.0 11 n homo2 n DEC3 in n dp hs dfra3 y nn 0 DISPLAY -1480.6 26962.9 98 0 dn3 dpwr3 1 sp wp vs dof3 dm3 0 n dmm3 C 0 dmf3 250 dseq3 107.85 dres3 500.00 homo3 6447.1 PR SC 10000 WC 1.0 hzmm 15 п rf1 PROCESSING 1.00 rfp th ins 1 ai cdc ph ft 131072 math 4 werr wexp 4 wbs 39.67 wnt 30 39.841 .137.759 .137.733 .128.918 173 50 571 100.911 169,536 .233 82.119 891 96.963 22. 29 54. 180 160 140 120

80

60

40

20

0

ppm

100

Pulse Sequence: relayh Solvent: DMSO Ambient temperature INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 5533.3 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments OBSERVE H1, 499.8635420 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec

625



Pulse Sequence: relayh Solvent: DMS0 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec

Relax, delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments OBSERVE H1, 499.8635420 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec

16









TY2-318-B







TY2-318-B

Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments OBSERVE H1, 499.8611709 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec









TY2-318-A exp1 s2pul SAMPLE Di date Mar 21 2009 dfrq solvent CDCl3 dn file exp dour DEC. & VT 499.864 H1 exp dpwr dof ACQUISITION TION dof 499.864 dm H1 dmm 5.016 dmf 65536 dseq 6533.3 dres sfrq tn at np sw fb 6533.3 dres 4000 homo 4 61 dfrq2 13.5 dn2 0.100 dpwr2 269.9 dof2 16 dm2 16 dm2 0 dmf2 DEC2 bs tpwr pw d1 tof nt ct alock n dmf2 not used dseq2 FLAGS dres2 gain 11 n. homo2 in n DEC3 y dfrq3 nn dn3 dp hs DISPLAY dpwr3 sp wp vs sc wc hzmm -101.8 dof3 5099.3 dm3 37 dmm3 dmm3 0 dmf3 250 dseq3 20.40 dres3 120.00 homoo 4165 0 is rfl homo3 PROCESSING 4165.0 PR0 3634.0 wtfile 50 proc 3.000 fn math rfp th ins ai ph Werr wexp wnt



Ph-

'n

AcO-



TY2-318-A

TY2-318-A

Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 20 Width 6533.3 Hz 32 repetitions 256 increments 0BSERVE H1, 499.8611956 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec

15









CH3 carbons



- 3

exp1 s2pu1

SAMPLE			DE	C. & VT	
late	Jun	25 2009	dfra	499.	866
olve	nt	CD30D	dn	22.22	H1
ile	000	6XD	dowr		30
AC	OUISI	TON	dof		0
fra		499.866	dm		nnn
n		H1	dmm		C
it.		5.016	dmf		200
מר		65536	dsea		
w		6533.3	dres		1.0
fb		4000	homo		n
as		4	nome	DEC2	
towr		61	dfro2		0
ω		13.5	dn2		1
11		0.100	dowr2		1
tof		269 9	dof2		â
at		32	dm2		'n
		32	dmm2		
lock	10	56	dmf 2		200
ain		head used	deen2		200
Jam	FLAC		dree2		1 0
	FLAG	5	bomo2		1.0
14			10002	DECO	- 10
			dfrag	DEGS	0
ap		У	atrq3		0
15	DTODU	nn	data		
202	DISPLA	-07 0	upwr 3		
sp		-97.0	dot 3		U
мр		5099.3	am3		n
vs		15	dmm3		C
SC		0.5.0	amr 3		200
₩C.		250	aseq3		2.12
hZmm		20.40	dres3		1.0
15		148.04	homo3		n
rf1		509.1	PR	DCESSING	
rfp		0	wtfile		233
th		7	proc		ft
Ins	10.022	3.000	fn	6	5536
a 1	ph		math		f
			werr		
			wexp	process	p1H
			wbs	101001000000000	

AcHN 14





exp2 s2pu1

exp1 s2pu1

SAL	MPLE	DEC. & VT			
date A	рг 11 2009	dfrg	499.864		
solvent	CDC13	dn	H1		
file	exp	dpwr	30		
ACQUIS	SITION	dof	0		
sfrq	499.864	dm	nnn		
tn	H1	dmm	c		
at	5.016	dmf	200		
np	65536	dseq			
SW	6533.3	dres	1.0		
fb	4000	homo	n		
bs	4		DEC2		
tpwr	61	dfra2	0		
pw	13.5	dn2			
d1	0.100	dowr2	1		
tof	269.9	dof2	õ		
nt	16	dm2	n		
ct	16	dmm2	c		
alock	n	dmf2	200		
gain	not used	dseg2			
FLA	AGS	dres2	1.0		
11	n	homo2	n		
in	n	rione E	DEC3		
do	v	dfra3	0		
hs	nn	dn3			
DIS	PLAY	dowr3	1		
SD	-94.4	dof3	Ô		
WD	5090.8	dm3	ñ		
VS	52	dmm3	e e		
sc	ñ	dmf3	200		
wc	250	dsea3	200		
hzmm	20.36	dres3	1.0		
15	125.68	homo3			
rf1	4140.5	PR	DCESSING		
rfp	3634 0	wtfile	COLOOLING .		
th	50	DCOC	ft.		
ins	2 000	fo	65536		
ai ph	2.000	math	f		
		werr			
		wexp	process plH		
		wbs			
		wnt	wft		
		1.			





exp2 s2pul

SAMPLE date Apr 11 2009 solvent CDC13 file exp ACQUISITION sfrq 125.702 tn C13 at 1.215 np 65536 sw 26963.3 fb 15000 bs 4 tpwr 52 pW 10.2 d1 1.800 tof 144.5 nt 3000 ct 84 alock n gain not used fil n in n dp Y sc 0 vs 89 vs 89 vs 89 vs 89 vs 89 sc 0 wp 26962.9 vs 89 sc 0 vc 250 <th>DEC. & VT dfrq 499.864 dn H1 dpWr 40 dof 0 dm yyy dmm W dmf 8787.35 dseq dres 1.0 homo DEC2 dfrq2 0 dfrq2 0 dfrq2 1 dof2 0 dm2 n dmm2 c dmf2 10000 dseq2 1.0 homo Z dres2 1.0 homo DEC3 dfrq3 0 dres2 1.0 homo CEC3 dres3 1.0 homo DEC3 dres3 1.0 homo DEC3 dres3</th> <th></th> <th></th> <th></th> <th>- 24.755</th>	DEC. & VT dfrq 499.864 dn H1 dpWr 40 dof 0 dm yyy dmm W dmf 8787.35 dseq dres 1.0 homo DEC2 dfrq2 0 dfrq2 0 dfrq2 1 dof2 0 dm2 n dmm2 c dmf2 10000 dseq2 1.0 homo Z dres2 1.0 homo DEC3 dfrq3 0 dres2 1.0 homo CEC3 dres3 1.0 homo DEC3 dres3				- 24.755
	Mut		77.485	55,943	34.562 31.695 24.289

0 ppm

Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments OBSERVE H1, 499.8611711 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec



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TY2-360-C-crystal

exp1 s2pul SAMPLE D date Jun 3 2009 dfrq solvent CDC13 dn DEC. & VT a 499.864 H1 30 solvent file exp opw. ACQUISITION dof sfrq 499.864 dm tn H1 dmm at 5.016 dmf np 65536 dseq sw 6533.3 dres fb 4000 homo fb 4 0 nnn C 200 1.0 п DEC2 4 61 tpwr pw d1 tof dfrq2 0 61 13.5 0.100 269.9 32 32 dn2 dpwr2 dof2 dm2 1 0 nt n dmm2 С alock n dmf2 200 not used dseq2 GS dres2 n homo2 gain FLAGS 1.0 11 n DEC3 in n y dfrq3 dp hs 0 nn dn3 DISPLAY dpwr3 1 sp wp vs sc wc Y dpwF3 -103.0 dof3 5099.3 dm3 28 dmm3 0 dmf3 250 dseq3 Ô. п C 200 hzmm is rfl rfp th 20.40 dres3 1.0 20.40 dress 113.77 homo3 4140.5 PR0 3634.0 wtfile 50 proc 3.000 fn n PROCESSING ft 65536 ins at ph math f werr wexp wbs process plH wft wnt




TY2-360-C-crystal



TY2-360-C-crystal

Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "mmr2a.chem.nd.edu"

Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments OBSERVE H1, 499.86511711 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec









TY2-337-A

exp1 s2pul DEC. & VT 499.864 H1 SAMPLE Dr date Apr 13 2009 dfrq colvent CDC13 dn Avn dpwr APLE Apr 13 2005 (vent CDC13 ACQUISITION dos 3 499.864 dm 5.016 dmf 65536 dseq 6533.3 dres 4000 homo 4 61 dfr 5 r 30 0 sfrq tn at nnn C 200 np sw fb 1.0 n bs tpwr pw d1 4 61 dfrq2 13.5 dn2 0.100 dpwr2 269.9 dof2 16 dm2 16 dm2 n dmf2 FLAGS dres2 fLAGS p bomo2 DEC2 0 1 tof 0 nt ct alock n C 200 gain 1.0 11 n homo2 n n y dfrq3 nn dn3 dpwr3 in DEC3 dp hs 0 DISPLAY 1 Y dpwr3 -102.8 dof3 5099.3 dm3 28 dmm3 0 dmf3 250 dseq3 20.40 dres3 sp wp vs sc wc hzmm is rf1 Ū n C 200 1.0 87.50 homo3 n PROCESSING 4140.3 rfp th ins 3634.0 wtfile 7 proc 7 proc 3.000 fn ft 65536 ph ai math f Werr process p1H wexp wbs wnt wft





TY2-337-A

exp2 s2pul

•••••••••••••••••••••••••••••			···/····						<u></u>
		143.240				57.138	33.685		
	werr wexp wbs wnt				77.485 77.230 76.975 -74.261			1.382 3.858 21.672 -21.118	
wp 26962.9 vs 168 sc 0 wc 250 hzmm 107.85 is 500.00 rfl 11112.2 rfp 9707.1 th 7 ins 100.000 ai cdc	dm3 n dmm3 c dmf3 1000 dseq3 1.0 homo3 n PROCESSING b 1.00 wtfile proc ft fn 131072 math f	024	127.083						
FLAGS i1 n in n dp y hs nn DISPLAY	dres2 1.0 homo2 n dfrq3 0 dn3 dpwr3 1 dof3 0								
d1 1.800 tof 144.5 nt 3000 ct 99 alock n gain not used	dpwr2 1 dof2 0 dm2 n dmm2 c dmf2 10000 dseq2 1			*					
sw 26963.3 fb 15000 bs 4 tpwr 52 pw 10.2	dres 1.0 homo n DEC2 dfrq2 0 dn2								
ACCONSISTION sfrq 125.702 tn C13 at 1.215 np 65536	don yyy dm yyy dm w dmf 8787.35 dseq								
date Apr 13 2009 solvent CDC13 file exp	dfrq 499.864 dn H1 dpwr 40								

TY2-337-A

Pulse Sequence: relayh Solvent: CDC13 Amblent temperature INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 20 Width 6533.3 Hz 16 repetitions 256 increments DBSERVE H1, 499.8611709 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 1 hr, 41 min, 40 sec







CH3 carbons



TY2-337-B

exp1 s2pul

	SAMPLE		DE	C. & VT	
date	Apr 1	13 2009	dfrq	499	.864
solve	int	CDC13	dn		H1
file		exp	dpwr		30
AC	QUISITI	ON	dof		0
sfrq	4	199.864	dm		nnn
tn		H1	dmm		c
at		5.016	dmf		200
np		65536	dseq		
SW		6533.3	dres		1.0
fb		4000	homo		n
bs		4		DEC2	
tpwr		61	dfrg2		0
pW		13.5	dn2		- 6
d1		0.100	dpwr2		1
tof		269.9	dof2		0
nt		16	dm2		n
ct		16	dmm2		c
alock	8	n	dmf2		200
gain	nc	ot used	dseq2		
1000	FLAGS	0.000	dres2		1.0
11	0.511600.000	n	homo2		n
in		n		DEC3	
dp		v	dfra3		0
hs		nn	dn3		- 23
	DISPLAY	1	dowr3		1
SD		-102.8	dof3		õ
WD		5099.3	dm3		n
VS		43	dmm3		c
sc		0	dmf3		200
WC		250	dsea3		~ * * *
hzmm		20.40	dres3		1.0
15		140.00	homo3		
rf1		4140.3	PR	OCESSING	. . .
rfp		3634.0	wtfile		
th		7	DEDC		ft
ins		3.000	fn	6	5536
aí	ph		math		f
			werr		
			wexp	process	p1H
			wbs		1000
			wnt		wft

9

8

ዋ 1.78 7

누 1.82 6



1 1

ppm

S48

5

0.98

4

3

ب 0.96 0.95 3.00 0.91 2

3.41 1.10 4.83

1

TY2-337-B

exp2 s2pul

				143.8			2		89 8,9 5,			
		wbs Wnt		13				77.482 77.230 76.975 73.508		33.537 846	24.830	
vs wc hzmmn is rfl rfp th ins ai co	250 250 107.85 500.00 11111.4 9707.1 5 100.000 ic ph	dmf3 dseq3 dres3 homo3 PROCES 1b wtfile proc fn math werr werr	10000 1.0 n ssing not used ft 131072 f		129.996 127.335							
il in dp hs Sp WP	FLAGS n y DISPLAY -1403.9 26962.9	dres2 homo2 DE(dfrq3 dn3 dpwr3 dof3 dm3 dm3	1.0 n 0 1 0									
pw d1 tof nt ct alock gain	10.2 1.800 144.5 3000 78 n not used	dn2 dpwr2 dof2 dm2 dm2 dmf2 dseq2	1 0 n c 10000									
tn at np sw fb bs tpwr	C13 1.215 65536 26963.3 15000 4 52	dmm dmf dseq dres homo DE(dfra2	8787.35 1.0 12									
date solven file ACQ sfrq	Apr 13 2009 at CDC13 exp UISITION 125.702	dfrq dn dpwr dof dm	499.864 H1 40 9 9 9 9 9									

TY2-337-B Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments OBSERVE H1, 499.8611905 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec







CH3 carbons



exp2 s2pu1

SAMPLE	DEC. & VT					
te Jun 29 2009	dfrg 499.864					
lvent CDC13	dn H1					
ACOUISITION	dof 0					
q 125.702	dm yyy					
C13	dmm W					
65536	dmr 0/0/.33 dsea					
26963.3	dres 1.0					
15000	homo n					
4 97 52	dfra2 0					
10.2	dn2					
1.800	dpwr2 1					
3000	d012 U dm2 n					
139	dmm2 C					
ock n	dmf2 10000					
FLAGS	dseq2 dres2 1.0					
n	homo2 n					
п	DEC3					
y no	dtrq3 D dp3					
DISPLAY	dpwr3 1					
-1408.5	dof3 0					
26962.9	dm3 n dmp3 c					
124	dmf3 10000					
250	dseq3					
m 107.85	dres3 1.0				0	
1 11115.9	PROCESSING		10 P		32	
p 9707.1	1b 1.00		97		24.	
e 100.000	wtfile proc ft				4 1 H	
cdc ph	fn 131072		77		11.	
	math f		الم لم		3.5	
	werr				m l	
	wexp				22	
	wbs			5	5 m	
	wnt			6	213	
				2		
			0 5			
	00		23			
	50 00		22		14 B	
	6		77			
	11.					
	1					
	[]			1		
					200	
			5 C			
	245					
			11	1		
	1					
	1					
sectors down and the former sectors in the sector of		and the second		Contraction of the local division of the loc	A Designation of the second seco	

Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec COSY 90-90

Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments OBSERVE H1, 499.86611705 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec





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CH carbons									
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CH2 carbons									
		and a subsection of subsection			 			20130-09 20130-09 20130-09	
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exp1 s2pul

	SAMPLE	DEC. & VT			
date	Apr 23 2009	dfrq	499.864		
solven	t CDC13	dn	H1		
file	exp	dpwr	30		
ACQ	UISITION	dof	0		
sfrq	499.864	dm	nnn		
tn	H1	dmm	c		
at	5.016	dmf	200		
np	65536	dseq			
SW	6533.3	dres	1.0		
fb	4000	homo	n		
bs	4		DEC2		
tpwr	61	dfrq2	0		
pw	13.5	dn2			
d1	0.100	dpwr2	1		
tof	269.9	dof2	0		
nt	4	dm2	n		
ct	4	dmm2	с		
alock	n	dmf2	200		
gain	not used	dsea2			
- 10 A	FLAGS	dres2	1.0		
11	n	homo2	n		
in	n		DEC3		
dp	Y	dfra3	0		
hs	nn	dn3			
D	ISPLAY	dpwr3	1		
Sp	-102.2	dof3	0		
WD	5099.3	dm3	n		
VS	15	dmm3	C		
SC	0	dmf3	200		
WC	250	dseg3			
hzmm	20.40	dres3	1.0		
is	117.48	homo3	n		
rf]	4139.7	PR	OCESSING		
rfp	3634.0	wtfile			
th	7	DEOC	ft		
105	1.000	fn	65536		
ai	ph	math	Ť		
		werr			
		wexp	process p1H		
		wbs	an a		







Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec CDSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments OBSERVE H1, 499.8611703 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec









TY2-362-B-c15-22

exp1 s2pul SAMPLE date Jun 6 2009 dfrq solvent CDC13 dn DEC. & VT q 499.864 H1 30 solvent file exp apw, ACQUISITION dof sfrq 499.864 dm tn H1 dmm at 5.016 dmf np 65536 dseq sw 6533.3 dres fb 4000 homo fb 4000 homo 61 dfrq2 0 nnn C 200 1.0 n 4 61 dfrq2 13.5 dn2 0.100 dpwr2 269.9 dof2 16 dm2 n dm72 not used dseq2 dres2 boxo2 DEC2 tpwr pw d1 0 1 tof nt ct õ n c alock 200 gain il in dp hs FLAGS 1.0 n y nn homo2 n DECS dfrq3 0 dn3 nn dn3 Y dpwr3 -103.0 dof3 5099.3 dm3 29 dmm3 0 dmf3 250 dseq3 20.40 dres3 136.39 bores2 DISPLAY 1 sp wp vs sc wc hzmm is rfl rfp th õ n C 200 250 dseq3 20.40 dres3 136.39 homo3 4140.5 PR0 3634.0 wtfile 50 proc 3.000 fn 1.0 n PROCESSING ft 65536 ins ai ph math f werr wexp process p1H

wnt

wft





TY2-362-B-c15-22

exp2 s2pul SAMPLE DEC. & VT
 SAMPLE
 Diamon Sample

 date
 Jun 6 2009
 dfrq

 solvent
 CDC13
 dn

 file
 exp
 dpwr

 ACQUISITION
 dof

 sfrq
 125.702
 dm

 tn
 C13
 dmm

 at
 1.215
 dmf

 np
 65536
 dseq

 SW
 26963.3
 dres

 fb
 15000
 homo

 bs
 4
 45000
499.864 H1 40 0 ууу W 8787.35 1.0 n DEC2 bs 4 52 dfrq2 10.2 dn2 1.800 dpwr2 tpwr 0 pw d1 1 144.5 dof2 2000 dm2 84 dmm2 tof nt ct ō. n dmm2 dmf2 c 10000 alock n not used dseq2 FLAGS dres2 gain 1.0 i1 in n homo2 n DISPLAY -1403.5 dot_ 26962.9 dm3 207 dmm3 0 dmf3 50 dseq3 dres? ~mr n y nn DEC3 dp hs 0 1 sp wp vs sc õ п C 10000 WC 1.0 hzmm 107.85 dres3 500.00 homo3 11111.0 PRC 9707.1 lb 4 wtfile 100.000 proc 15 n rf1 rfp th PROCESSING 1.00 129.986 ft ins 1 ai cdc ph 131072 fn math f 127.302 Werr wexp wbs .485 975 wnt NOW 53 449 61.446 5 136.749 143.784 1.1.1.1 1 1 1 80 60 200 180 160 140 120 100

21.734

11111

40

20

1111111

0

ppm

TY2-362-8-c15-22 Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 2D Width 6533.3 Hz 32 repetitions 32 repetitions 256 increments OBSERVE H1, 499.8611711 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec F 2 . (ppm) 2.5-3.0 < 8 ۰ 3.5-3 28 . 4.0-4.5-5.0-5.5-6.0-6.5-7.0--0 . 7.5-0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 F1 (ppm)

TY2-362-C-c24-28

e×p1	s2pu1			
	SAMPL	E	DE	C. & VT
date	Jun	6 2009	dfra	499.864
solve	nt	CDC13	dn	H1
file	88	exp	down	30
AC	OUTSIT	TON	dof	0
sfra	******	499.864	dm	nnn
tn		H1	dmm	c
at		5.016	dmf	200
np		65536	dseq	(71.70.70)
sw		6533.3	dres	1.0
fh		4000	homo	
bs		4		DEC2
towr		61	dfra2	0
nw		13 5	dn2	•
di		0 100	dowr2	1
tof		269 9	dof2	0
nt		200.0	dm2	
c+			dmm2	
alock			dmf2	200
anto	S	nt used	de ea 2	200
gain	FLACE	or used	deec2	1.0
41	TLAGS		u1652	1.0
1.0			1101802	0503
110			dfrag	0000
ap		y	dag	U
ns	-	u nn	dat in 2	
	DISLIN	100 0	upwr 3	1
sp		-102.2 E000 2	dm2	0
wp		5099.3	din 3	<u>n</u>
VS		10	amm3	C 0.00
SC		0.50	dm13	200
wc		250	aseq3	
nzmm		20.40	ares3	1.0
15		31.00	homo3	n
LT I		4139.7	PR	UCESSING
гтр		3634.0	wttile	52.2
th		50	proc	ft
105		100.000	fn	65536
al	ph		math	f
			werr	
			wexp	process plH
			wbs	
			wnt	wft



3.6

18

AcO 16c

TY2-362-C-c24-28

exp2 s2pu1

	190	er et an de Alan	160	140	120

		3			
		-171.			
		280			
		wnt			
		werr wexp			
	198 - 1 99	math	ŕ		
ins ai c	100.000 dc ph	proc	ft 131072		
rfp th	9707.1	1b wtfile	1.00		
is rfl	500.00 11114.7	homo3 PF	n ROCESSING		
wc hzmm	250 107.85	dseq3 dres3	1.0		
sc	123	dmf3	10000		
Wp	26962.9	dm3 dmm3	ņ		
sp	DISPLAY -1407.2	dpwr3 dof3	1		
hs	y nn	dn3	0		
in	n	dfra2	DEC3		
11	FLAGS n	dres2 homo2	1.0		
gain	not used	dseq2	10000		
ct alock	257	dmm2	C 10000		
nt	300	dm2	0 N		
d1 tof	1.800	dpwr2	1		
pw	10.2	dn2	U		
bs	4	dfrag	DEC2		
fb	26963.3	homo	1.0 n		
np	65536	dseq	1000 C C C C C C C C C C C C C C C C C C		
at	1.215	dmin	8787.35		
sfrq	125.702	dm	ууу		
AC	QUISITION	dof	40		
solve	nt CDC13	dn	H1		
14 4 A A A	Jun 6 2009	dfrq	433.004		

		1	I			
	1			1		
100	80	60	40	20	0 1	ppm

77.230

63.430

38,889

23.338







CH3 carbons


exp2 s2pul

	SAMPLE	DEC. & VT			
date	Jul 2 2009	dfrq	499.864		
solven	t CDC13	dn	H1		
file	exp	dpwr	30		
ACO	UISITION	dof	0		
sfra	499.864	dm	nnn		
tn	H1	dmm	C		
at	5.016	dmf	200		
np	65536	dseq			
SW	6533.3	dres	1.0		
fb	4000	homo	n		
bs	4		DEC2		
tpwr	61	dfrq2	0		
pw	13.5	dn2			
d1	0.100	dpwr2	1		
tof	269.9	dof2	0		
nt	32	dm2	n		
ct	32	dmm2	C		
alock	n	dmf2	200		
gain	not used	dseq2			
	FLAGS	dres2	1.0		
11	n	homo2	n		
in	n		DEC3		
dp	У	dfrq3	0		
hs	nn	dn3			
0	ISPLAY	dpwr3	1		
Sp	-94.0	dof3	0		
WD	5090.8	dm3	n		
VS	21	dmm3	C		
SC	0	dmf3	200		
WC	250	dseq3			
hzmm	20.36	dres3	1.0		
15	46.51	homo3	n		
rf1	4140.1	PR	OCESSING		
rfp	3634.0	wtfile			
th	7	ргос	ft		
ins	2.000	fn	65536		
ai	ph	math	f		
		werr			
		wexp	process plH		
		WDS			





exp3 s2pul

	171.476	Rea II , dan ak as usud	7 230 7 7.728	49,312		
SAMPLE date Jul 2 2009 solvent CDC13 file exp ACQUISITION sfrq it 125.702 tn C13 at 1.215 np 65536 sw 26963.3 fb 15000 bs 4 tpwr 502 pw 10.2 d1 1.800 tof 144.5 nt 1000 ct 52 gain not used il n in n gain not used il n off 1412.6 in n owc 250 hzmm 107.85 is 500.000 off 14112.6 rfp 9707.1 th 4 ins 100.000 ai cdc	DEC. & VT dfrq 499.864 dn H1 dpwr 40 dof 0 dm Yyy dmm 8787.35 dseq dres 1.0 homo DEC2 dfrq2 0 dfrq2 0 dfrq2 1 dof2 0 dm2 n dmm2 c dmf2 10000 dseq2 1.00 homo2 n DEC3 dfrq3 0 dfrq3 0 dfrq3 0 dfrq3 1 dof3 0 dfrq3 10000 dseq3 1.00 homo3 n PROCESSING lb 1.00 wtfile proc ft fn 131072 math f		-77.485 130 - 76.975 8		35.681 25.855 23.367	



TY2-398-c2

exp2 s2pul date Jul 7 2009 dfrq -luent CDCl3 dn -luent dpwr DEC. & VT date Ju. solvent CDC:5 file exp dpwr ACQUISITION dof --- 499.864 dm H1 dmw 499.864 H1 30 0 nnn 499.864 dm H1 dmm 5.016 dmf 65536 dseq 6533.3 dres 4000 homo tn at sw fb bs tpwr d1 tof nt ct C 200 1.0 n DEC2 4 4 61 dfrq2 13.5 dn2 0.100 dpwr2 269.9 dof2 32 dm2 32 dm2 32 dm2 0 1 õ n C not used dseq2 FLAGS alock gain 200 1.0 il in dp hs n homo2 n DEC3 n y dfrq3 0 nn dn3 DISPLAY dpwr3 1 sp vs sc wc hzmm is rf1 rfp th ins -102.4 dof3 5099.3 dm3 ō n 20 dmm3 С dmf3 200 250 dseq3 20.40 60.56 dres3 1.0 homo3 n 4139.9 PROCESSING 3634.0 wtfile 7 proc 7 proc 2.000 fn ft 65536 ai ph math f werr wexp process p1H wbs wft wnt





TY2-398-c2

exp3 s2pul





S79



exp2 s2pul

	SAMPL	E	DE	C. & VT	
date	Jan	4 2010	dfrg	499.	.864
solver	nt	CDC13	dn		H1
file .		exp	dpwr		30
ACC	DUISIT	ION	dof		0
sfrg		499.864	dm		nnn
tn		H1	dmm		C
at		5.016	dmf		200
np		65536	dseq		
SW		6533.3	dres		1.0
fb		4000	homo		n
bs		4		DEC2	
tpwr		61	dfrg2		0
pw		13.5	dn2		
d1		0.100	dpwr2		1
tof		269.9	dof2		0
nt		16	dm2		n
ct		16	dmm2		c
alock		n	dmf2		200
qain	n	ot used	dseq2		
5	FLAGS		dres2		1.0
11		n	homo2		n
in		n		DEC3	
dp		Y	dfrq3		0
hs		nn	dn3		
x	DISPLA	Y	dpwr3		1
SD		-103.2	dof3		0
WD		5099.3	dm3		n
vs		36	dmm3		c
SC		0	dmf3		200
WC		250	dseg3		
hzmm		20.40	dres3		1.0
is		177.14	homo3		n
rf1		4140.7	PR	OCESSING	
rfp		3634.0	wtfile		
th		50	proc		ft
ins		3.000	fn	6	5536
ai	ph		math		f
			werr		
			wexp	process	p1H
			wbs		See.
			wnt		wft





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exp3 s2pu1



Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 20 Width 6533.3 Hz 32 repetitions 256 increments OBSERVE H1, 499.8611713 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec









CH3 carbons



%Transmittance

exp2 s2pul

	CAND	I F	DEC & VT			
date Dec 23 2009			dfra 499 864			
SOLV	ent	CDC13	dn		H1	
file	900	exp	dowr		30	
A	COUISI	TION	dof		0	
sfrg		499.864	dm		nnn	
tn		H1	dmm		C	
at		5.016	dmf		200	
np		65536	dseq			
SW		6533.3	dres		1.0	
fb		4000	homo		n	
bs		4		DEC2		
tpwr		61	dfrq2		0	
pw		13.5	dn2			
d1		0.100	dpwr2		1	
tof		269.9	dof2		0	
nt		16	dm2		п	
ct		16	dmm2		C	
aloc	ĸ	n	dmf2		200	
gain		not used	dseq2			
	FLAG	S	dres2		1.0	
11		n	homo2	100000	n	
in		n		DEC3		
dp		У	dfrq3		0	
hs	2828333	nn	dn3		16 C	
	DISPL	AY	dpwr3		1	
sp		-102.8	dof3		0	
wp		5099.3	dm3		n	
vs		22	dmm3		C	
sc		0	dmf3		200	
wc		250	dseq3		806201	
hzmm		20.40	dres3		1.0	
15		127.97	homo3		n	
rf)		4140.3	PR	OCESSING		
rfp		3634.0	wtfile			
th			proc		TL	
ins	2003 a.M.2	1.000	tn	65	536	
aı	ph		math		1	
			werr			
			wexp	process	plH	
			wbs		Sec.	
			wint		WEt	





exp3 s2pu1



TY2-501 Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 32 repetitions 256 increments OBSERVE H1, 499.8611709 MHz DATA PROCESSING Sine bell 0.039 sec

Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec



TY2-501 Pulse Sequence: hetcor Solvent: CDC13 Ambient temperature User: 1-14-87 INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.500 sec Acq. time 0.111 sec Width 18403.5 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments OBSERVE C13, 125.6901681 MHz DECOUPLE H1, 499.8639312 MHz Power 40 dB on during acquisition off during delay WALTZ-16 modulated DATA PROCESSING Line broadening 1.0 Hz F1 DATA PROCESSING Line broadening 0.3 Hz FT size 4096 x 1024 Total time 3 hr, 46 min, 5 sec F1 (ppm) v 2.0-2.5-3.0 1 3.5 ۱ 1 4.0-1 4.5







S92

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exp1 s2pu1

SAMP	LE	DE	C. & VT
date Jun	9 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	30
ACQUISI	TION	dof	0
sfrq	499.864	dm	nnn
tn	H1	dmm	c
at	5.016	dmf	200
np	65536	dseq	
SW	6533.3	dres	1.0
fb	4000	homo	п
bs	4		DEC2
tpwr	61	dfrq2	0
pw	13.5	dn2	
d1	0.100	dpwr2	1
tof	269.9	dof2	0
nt	32	dm2	n
ct	32	dmm2	С
alock	n	dmf2	200
gain	not used	dseq2	
FLAG	S	dres2	1.0
11	n	homo2	n
in	n		DEC3
dp	У	dfrq3	0
hs	nn	dn3	
DISPL	AY	dpwr3	1
sp	-102.6	dof3	0
Wp	5099.3	dm3	n
VS	34	dmm3	с
SC	0	dmf3	200
WC	250	dseq3	
hzmm	20.40	dres3	1.0
15	123.23	homo3	n
rf1	4140.1	PR	OCESSING
rfp	3634.0	wtfile	
th	50	proc	ft
ins	1.000	fn	65536
ai ph		math	f
		werr	
		wexp wbs	process plH





exp2 s2pul



Pulse Sequence: relayh Solvent: CDC13 Ambient temperature IN0VA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec CDSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 20 Width 6533.3 Hz 32 repetitions 258 increments OBSERVE H1, 499.8611707 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 24 min, 46 sec





S96

Pulse Sequence: hetcor Solvent: CDC13 Ambient temperature User: 1-14-87 INOVA-500 "nmr2@.chem.nd.edu" Relax. delay 1.500 sec Acq. time 0.111 sec Width 18403.5 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments DBSERVE C13, 125.6901591 MHz DECOUPLE H1, 499.8639312 MHz Power 40 dB on during acquisition off during delay WALTZ-16 modulated DATA PROCESSING Line broadening 1.0 Hz F1 DATA PROCESSING Line broadening 0.3 Hz FT size 4096 x 1024 Total time 3 hr, 46 min, 5 sec F1 (ppm] 7.0-7.2-7.4 7.6-7.8 8.0-8.2-131 130 129 127 125 124 122 128 126 123 F2 (ppm)







1

9

exp1 s2pul

S	AMPLE	DEC. & VT		
date	date Jun 21 2009		499.866	
solvent	CD30D	dn	H1	
file	éxp	dpwr	30	
ACQU	ISITION	dof	0	
sfra	499.866	dm	nnn	
tn	H1	dmm	c	
at	5.016	dmf	200	
np	65536	dseq		
SW	6533.3	dres	1.0	
fb	4000	homo	n	
bs	4		DEC2	
tpwr	61	dfrq2	0	
pw	13.5	dn2		
d1	0.100	dpwr2	1	
tof	269.9	dof2	0	
nt	16	dm2	n	
ct	16	dmm2	c	
alock	n	dmf2	200	
gain	not used	dseg2		
F	LAGS	dres2	1.0	
11	n	homo2	n	
in	n		DEC3	
dp	Y	dfrq3	0	
hs	nn	dn3		
DI	SPLAY	dpwr3	1	
Sp	-96.8	dof3	0	
WD	5099.3	dm3	n	
VS	36	dmm3	c	
SC	0	dmf3	200	
WC	250	dseg3		
hzmm	20.40	dres3	1.0	
is	44.25	homo3	n	
rf1	2163.5	PR	OCESSING	
rfp	1654.5	wtfile		
th	13	proc	ft	
ins	7.000	fn	65536	
ai p	h	math	f	
		werr		
		wexp wbs	process p1H	
		wnt	wft	

8

0.38

7

6



1

ppm

5

TA

1.80

4

3

2

7.00

1

4.14

TY2-376 exp2 s2pul SAMPLE DEC. & VT date Jun 21 2009 dfrq 499.866 solvent file exp apw, ACQUISITION dof sfrq 125.703 dm tn C13 dmm at 1.215 dmf np 65536 dseq sw 26963.3 dres fb 15000 homo 4 dfrg7 cd3od dn exp dpwr 0N dof solvent 8787.35 DEC2 tpwr pw d1 tof nt ct alock not used dseq2 gain FLAGS dres2 11 n homo2 n y dfrq3 nn dn3 dpwr3 in dp DEC3 hs DISPLAY AY dpwr3 -1234.2 dof3 26962.9 dm3 61 dmm3 0 dmf3 sp wp vs sc wc hzmm is rf1 rfp th ins 1 ai cdc ph werr wexp wbs wnt

H1 40 0 УУУ w

1.0 n

0





TY2-381-c5-24-recrystalization

exp2 s2pul

	SAMPLE	DEC. & VT		
date	Jul 7 2009	dfrq	499.864	
solver	nt CDC13	dn	H1	
file	exp	dpwr	30	
ACC	UISITION	dof	0	
sfrq	499.864	dm	nnn	
tn	H1	dmm	c	
at	5.016	dm f	200	
np	65536	dseq		
sw	6533.3	dres	1.0	
fb	4000	homo	n	
bs	4		DEC2	
tpwr	61	dfrq2	0	
pw	13.5	dn2		
d1	0.100	dpwr2	1	
tof	269.9	dof2	0	
nt	32	dm 2	n	
ct	32	dmm2	с	
alock	n	dmf2	200	
gain	not used	dseq2		
	FLAGS	dres2	1.0	
i1	n	homo2	n	
in	n		DEC3	
dp	v	dfra3	0	
ĥs	กก้	dn3		
	DISPLAY	dpwr3	1	
SD	-102.0	dof3	ō	
wb	5099.3	dm3	'n	
vs	33	dmm3	c	
sc	0	dmf3	200	
wc	250	dsea3	200	
hzmm	20.40	dres3	1.0	
is	122.51	homo3		
rf1	4139.5	PRO	DCESSING	
rfp	3634.0	wtfile		
th	7	ргос	ft	
ins	3.000	fn	65536	
ai	ph	math	f	
		werr		
		wexp	process p1H	
		wbs		
		wnt	wft	





TY2-381-c5-24-recrystalization

exp3 s2pul SAMPLE DEC. & VT Jul 7 2009 dfrq date 499.864 solvent file CDC13 dn HI 40 exp dpwr ACQUISITION dof rq 125.702 dm C13 dmm 1.215 dmf 0 sfrq ууу tn at np sw fb ŵ 8787.35 65536 dseq 26963.3 dres 15000 homo 1.0 n bs DEC2 4 4 52 dfrq2 10.2 dn2 1.800 dpwr2 144.5 dof2 1000 dm2 67 dmm2 tpwr pw d1 0 1 0 tof nt n ct C alock gain п dmf2 10000 not used dseq2 dres2 FLAGS 1.0 11 n homo2 n in DEC3 п dfrq3 y dfrq nn dn3 dp 0 DISPLAY -1411.3 / 26962.9 98 hs dpwr3 dof3 1 sp wp vs sc 0 dm3 dmm3 п C dmf3 10000 0 127.633 250 wc dseq3 107.85 dre 500.00 hom 11118.8 9707.1 lb hzmm dres3 homo3 1.0 is rfl n PROCESSING 1.00 rfp wtfile th 6 100.000 proc ins ft 131072 f ai cdc ph fn math Werr wexp wbs 122 23.485 wnt .785 46. 9 144.870 169.682 134.445 ****************** -----200 180 160 140 120 100 40 0 ppm 80 60 20

TY2-381-c5-24-recrystalization

Puise Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments OBSERVE H1, 499.8611619 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec



S105



S106



exp2 s2pul

	SAMPLE	DEC. & VT		
date	Jul 20 2009	dfrq	499.864	
solver	nt CDC13	dn	H1	
file	exp	dpwr	30	
AC	QUISITION	dof	0	
sfrq	499.864	dm	nnn	
tn	H1	dmm	c	
at	5.016	dmf	200	
np	65536	dseq		
sw	6533.3	dres	1.0	
fb	4000	homo	n	
bs	4		DEC2	
tpwr	61	dfrq2	0	
pw	13.5	dn2		
d1	0.100	dpwr2	1	
tof	269.9	dof2	0	
nt	16	dm2	n	
ct	16	dmm2	c	
alock	n	dm+2	200	
gaın	not used	dseq2		
	FLAGS	dres2	1.0	
11	n	homo2	n	
10	n		DEC3	
dp	У	dfrq3	U	
hs	nn	dn3		
-	DISPLAY	dpwr3	1	
sp	-111.2	dof 3	U	
wp	5107.9	dma 2		
VS	28	amm 3	200	
sc	050	dm t 3	200	
wc	250	dseq3	1.0	
nzmm	20.43	ures3	1.0	
18	97.17	10000	OCESSING	
ET I	500.1	utfile.	JCESSING	
*5	2	WLTTIG	£+	
ine	1 000	fn	65536	
1115	nh 1.000	math	655550 F	
ai	pn -	1000 611		
		werr		
		wexp	process plH	
		wbs		
		wnt	wft	




exp3 s2pu1



TY2-411 Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 2D Width 6533.3 Hz 16 repetitions 256 increments DBSERVE H1, 499.8611707 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 1 hr, 41 min, 40 sec F2 0 23 Ø (ppm) -1.5-:: 5 : 2.0-. 1.0 2.5-0 3.0-8 D ۵ ... 0 ~ 3.5-4.0-4.5-5.0 00 -5.5 1 1.5 5.5 3.5 3.0 2.5 2.0 5.0 4.5 4.0 F1 (ppm)



CH3 carbons

CH2 carbons



exp2 s2pul

	SAMPLE	DE	C. & VT		
date	Jul 21 2009	dfra	499	864	
solve	nt CDC13	dn		H1	
file	exp	dpwr		30	
AC	OUISITION	dof		0	
sfra	499.864	dm		nnn	
tn	H1	dmm		C	
at	5.016	dmf		200	
nn	65536	dsea		124.65	
SW	6533.3	dres		1.0	
fb	4000	homo			
bs	4		DEC2		
towr	61	dfra2		0	
DW	13.5	dn2			
di	0.100	dowr2		1	
tof	269.9	dof2		õ	
nt	16	dm2		n	
ct	16	dmm2		c	
alock	'n	dmf2		200	
gain	not used	dseq2		100.0	
3	FLAGS	dres2		1.0	
11	n	homo2			
in	n		DEC3	53	
do	v	dfra3		0	
hs	, j nn	dn3			
10 A	DISPLAY	dowr3		1	
SD	-110.8	dof3		ô	
wn	5107 9	dm3		ñ	
VS	29	dmm3		č	
sc	-0	dmf3		200	
we	250	dsen3			
h Zmm	20.43	dres3		1.0	
is	153.36	homo3			
rf1	4139.7	PR	OCESSING		
rfo	3634.0	wtfile			
th	7	DLOC		ft	
ins	1.000	fn	6	5536	
ai	nh	math		f	
	33 T				
		WELL			
		Wexp	nrocess	n1H	
		wbs	p. 00000	Plat	
		wnt		wft	





exp3 s2pu1

	100	150	1.40	120	100	on				• • • • •
	hengen skon stor stat Managara des trais						ana ana amin'ny sorana amin'ny sorana amin'ny sorana amin'ny sorana amin'ny sorana amin'ny sorana amin'ny sora			
						Ť				
		170. 169.				77.48 230 76.97				
		731				5 2.068		47.338	23.4	
		wbs wnt							62 62	
		math werr wexp	f						0.127 0.127	
rfp th ins ai cdo	9707.1 6 100.000 c ph	lb 1.0 wtfile proc f fn 1310	00 ft 72							
sc wc hzmm is rfl	250 107.85 500.00 11119.2	dm T3 1000 dseq3 dres3 1 homo3 PROCESSING	.0 n							
Sp Wp VS	ISPLAY -1411.8 26962.9 47	dpwr3 dof3 dm3 dm3 dm3	1 0 n c							
il in dp hs	n n Y nn	homo2 DEC3 dfrq3 dn3	n 0							
ct alock gain f	69 n not used FLAGS	dmm2 dmf2 1000 dseq2 dres2 1.	с)0 .0							
pw d1 tof nt	10.2 1.800 144.5 512	dn2 dpwr2 dof2 dm2	1 0							
fb bs tpwr	15000 4 52	homo DEC2 dfrq2	0							
tn at np	C13 1.215 65536 26963 3	dmm dmf 8787.3 dseq dres 1	35 0							
solvent file ACQU sfra	t CDC13 exp UISITION 125.702	dn h dpwr 4 dof dm VV	11 10 0							
date	SAMPLE Jul 21 2009	DEC. & VT dfrq 499.86	4							

Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 20 Width 6533.3 Hz 20 Width 6533.3 Hz 20 Width 6533.3 Hz 256 increments 0BSERVE H1, 499.8611703 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 1 hr, 41 min, 40 sec









exp2 s2pul exp2 SAMPLE DE date Jul 18 2009 dfrq solvent CDC13 dn file exp dpwr ACQUISITION dof sfrq 499.864 dm tn H1 dmm at 5.016 dmf 55536 dse DEC. & VT 499.864 H1 30 dpwr dof dm dmm dmf dseq dres homo 0 nnn tn at np sw fb bs 200 5.016 dmf 6553.6 dseq 6533.3 dres 4000 homo 4 61 dfrq2 13.5 dn2 0.100 dpwr2 269.9 dof2 16 dmm2 16 dmm2 n dmf2 t used dseq2 dres2 n homo2 1.0 n DEC2 tpwr pw d1 tof nt ct alock gain 0 1 ō п С not used 200 FLAGS 1.0 il in dp hs homo2 n n n y nn DEC3 y dfrq3 nn dn3 dpwr3 -102.2 dof3 5099.3 dm3 0 dmf3 21 dmm3 0 dmf3 250 dseq3 20.40 dres3 69.57 homo3 4139.7 PROCESSING 3634.0 wtfile 7 proc 2.000 fn math 0 Sp Wp SC WC hzmm is rfl rfp th ins aj 0 n C 200 1.0 n ft 65536 f werr wexp wbs process p1H wnt wft





exp3 s2pul

SAMPLE date Jul 18 2009 solvent CDC13 file exp ACQUISITION sfrq 125.702 tn C13 at 1.215 np 65536 sw 26963.3 fb 15000 bs 4 tpwr 52 pw 10.2 d1 1.800 tof 144.5 nt 1000 ct 122 alock n gain not used FLAGS i1 n dp y hs nn DISPLAY sp -1401.5 wp 26962.9 vs 95 sc 0 wc 250 hzmm 107.85 is 50.00 rfl 11108.9 rfp 9707.1 th 3 ins 100.000 ai cdc ph	DEC. & VT dfrq 499.864 dn H1 dpwr 40 dof 0 dm yyy dmm w dm dmf 8787.35 dseq 1.0 homo DEC2 dfrq2 0 dn2 1 dof2 0 dm2 1 dof2 0 dm2 n dmf2 10000 dseq2 1.0 homo2 n DEC3 dfrq3 0 dm3 0 dm3 10000 dseq3 1.0 homo3 1 dof3 0 dm3 10000 dseq3 1.0 homo4 frq3 10000 dseq3 1.0 homo5 n PROCESSING 1 b not used wtf11 fr 131072 math f werr wexp wbs wnt			83.093 77.230 76.978	49.763	40.595	25.400	
200 180	160	140 120	100	80	60	40	20	0 ppm

TY2-416 Pulse Sequence: relayh Solvent: CDC13 Ambient temperature INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.300 sec COSY 90-90 Acq. time 0.157 sec Width 6533.3 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments OBSERVE H1, 499.8611703 MHz DATA PROCESSING Sine bell 0.078 sec F1 DATA PROCESSING Sine bell 0.039 sec FT size 2048 x 2048 Total time 3 hr, 23 min, 10 sec F2 (ppm) 1.5-8 2.0-• :000 1 2.5-35 35 0 3.0 6 G . -3.5-• 4.0-4.5 5.0 0 B 111 1 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 F1 (ppm)

Pulse Sequence: hetcor Solvent: CDC13 Ambient temperature User: 1-14-87 File: TY2-416-CH INOVA-500 "nmr2a.chem.nd.edu" Relax. delay 1.500 sec Acq. time 0.111 sec Width 18403.5 Hz 2D Width 6533.3 Hz 32 repetitions 256 increments OBSERVE C13, 125.6901591 MHz DECOUPLE H1, 499.8639312 MHz POwer 40 dB on during acquisition off during delay WALTZ-16 modulated DATA PROCESSING Line broadening 1.0 Hz F1 DATA PROCESSING Line broadening 0.3 Hz FT size 4096 x 1024 Total time 3 hr, 46 min, 5 sec





CH3 carbons

