

# Supporting Information

## Sulfonylation-Induced *N*- to *O*-Acetyl Migration in 2-Acetamidoethanol Derivatives

Takao Yamaguchi, Dusan Heseck, Mijoon Lee, Allen G. Oliver, and Shahriar Mobashery\*

*Department of Chemistry and Biochemistry, University of Notre Dame, Notre Dame, Indiana 46556*

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## Experimental Procedures

### Benzyl 2-acetamido-4,6-*O*-benzylidene-2-deoxy- $\alpha$ -D-glucopyranoside (**1a**)

The compound **1a** was prepared from D-(+)-glucosamine by a variation of the known procedures.<sup>1</sup> Sodium methoxide (32.6 g, 0.60 mol) was added to a suspension of D-(+)-glucosamine hydrochloride (100 g, 0.46 mol) in MeOH (1 L), and the mixture was stirred for 30 min at 40 °C. After addition of acetic anhydride (79 mL, 0.83 mol), the resulting suspension was stirred vigorously for 22 h at 40 °C, and then cooled to 0 °C. After filtration of the reaction mixture, the filtered white solid was washed with cold MeOH and dried to afford *N*-acetyl D-glucosamine (100 g, 97%) as a white powder.

Acetyl chloride (27.4 mL, 0.39 mol) was slowly added to a suspension of *N*-acetyl D-glucosamine (85 g, 0.39 mol) in anhydrous benzyl alcohol (300 mL) under nitrogen atmosphere. The suspension was stirred at room temperature for 2 h, heated to 50 °C for 4 h, and then cooled to room temperature. The resulting yellow solution was slowly poured onto Et<sub>2</sub>O (3 L) in ice-water bath, and the mixture was vigorously stirred for 2 h at 0 °C. The precipitate was recovered by filtration and dried under vacuum to afford benzyl 2-acetamido-2-deoxy- $\alpha$ -D-glucopyranoside (113 g, 94%) as a white solid.

Benzaldehyde dimethylacetal (58 mL, 0.39 mol) and *p*-toluenesulfonic acid monohydrate (1.8 g, 9.6 mmol) were added to a solution of benzyl 2-acetamido-2-deoxy- $\alpha$ -D-glucopyranoside (60 g, 0.19 mol) in anhydrous DMF (500 mL), and the mixture was stirred at 70 °C for 24 h. The resulting mixture was cooled to 0 °C and then triethylamine (8.1 mL, 56 mmol) was added. After stirring for 30 min at room temperature, the solvent and extra reagent were removed under reduced pressure to give a white solid. MeOH was added to the solid and the resulting suspension was stirred vigorously for 5 min. After filtration, filtered white solid was washed well with MeOH and dried under vacuum to afford **1a** (67.7 g, 88%) as a white powder. **1a**: <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  1.85 (s, 3H, CH<sub>3</sub>CO), 3.51 (dd, *J* = 8.5, 8.5 Hz, 1H, H-4), 3.63–3.80 (m, 3H, H-3, H-5, H-6a), 3.85 (ddd, *J* = 3.6, 8.2, 8.4 Hz, 1H, H-2), 4.14 (dd, *J* = 2.8, 8.6 Hz, 1H, H-6b), 4.48 and 4.70 (AB, *J* = 12.7 Hz, 2H, OCH<sub>2</sub>Ph), 4.82 (d, *J* = 2.4 Hz, 1H, H-1), 5.19 (d, *J* = 5.8 Hz, 1H, OH), 5.61 (s, 1H, CHPh), 7.26–7.49 (m, 10H), 8.00 (d, *J* = 8.0 Hz, 1H, NH); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  22.5, 54.2, 62.8, 67.2, 68.5, 82.1, 96.9, 100.9, 126.4, 127.6, 127.6, 128.0, 128.3, 137.7, 137.7, 169.4; IR 3399, 3302, 1651, 1552, 1452, 1374, 1129, 1087, 1059 cm<sup>-1</sup>; HRMS (FAB) calcd for C<sub>22</sub>H<sub>25</sub>NO<sub>6</sub> (M + H<sup>+</sup>), 400.1760, found 400.1763.

### General procedures for the tosylation of 2-acetamidoethanol derivatives (Condition A or B)

**Condition A:** *p*-Toluenesulfonyl chloride (1.05 equiv) and pyridine (3.0 equiv) were added to a solution of substrate in CH<sub>2</sub>Cl<sub>2</sub> at 0 °C, and the mixture was stirred at room temperature for 24 h. The solvent was removed under reduced pressure, and the crude product was purified by silica gel column chromatography. **Condition B:** *p*-Toluenesulfonyl chloride (2.0 equiv) was added to a solution of substrate in pyridine, and the mixture was refluxed for 30 h. The mixture was cooled to room temperature, and the solvent was removed under reduced pressure. The crude product was purified by silica gel column chromatography.

Products of the tosylation of the compound **1a**

### Benzyl 2-acetamido-4,6-*O*-benzylidene-2-deoxy-3-*O*-*p*-toluenesulfonyl- $\alpha$ -D-glucopyranoside (**2a**)

### Benzyl 3-*O*-acetyl-4,6-*O*-benzylidene-2-deoxy-2-*p*-toluenesulfoneamido- $\alpha$ -D-glucopyranoside (**3a**)

The condition B was used for compound **1a** (100 mg, 0.25 mmol), and the crude product was purified by silica gel column chromatography (AcOEt/hexane, 1:3) to afford the product **2a** (46 mg, 33%) as a white solid and the product **3a** (74 mg, 53%) as a white solid. **2a**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  2.02 (s, 3H, CH<sub>3</sub>CO), 2.28 (s, 3H, CH<sub>3</sub>Ph), 3.68 (dd, *J* = 9.6, 9.6 Hz, 1H, H-4), 3.72 (dd, *J* = 10.7, 10.7 Hz, 1H, H-6a), 3.88 (ddd, *J* = 4.8, 9.8, 9.8 Hz, 1H, H-5), 4.21 (dd, *J* = 5.0, 10.4 Hz, 1H, H-6b), 4.46 (ddd, *J* = 3.6, 9.2, 10.4 Hz, 1H, H-2), 4.53 and 4.73 (AB, *J* = 11.9 Hz, 2H, OCH<sub>2</sub>Ph), 4.92 (dd,

<sup>1</sup> (a) Berger, I.; Nazarov, A. A.; Hartinger, C. G.; Groessl, M.; Valiahdi, S-M.; Jakupec, M. A.; Keppler, B. K. *ChemMedChem* **2007**, *2*, 505–514. (b) Babic, A.; Pecar, S. *Tetrahedron: Asymmetry* **2008**, *19*, 2265–2271.

$J = 10.0, 10.0$  Hz, 1H, H-3), 5.02 (d,  $J = 3.8$  Hz, 1H, H-1), 5.37 (s, 1H, *CHPh*), 6.00 (d,  $J = 9.0$  Hz, 1H, *NH*), 6.97 (d,  $J = 8.0$  Hz, 2H, Ts), 7.19–7.44 (m, 10H), 7.67 (d,  $J = 8.4$  Hz, 2H, Ts);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  21.9 (*CH}\_3\text{Ph}*), 23.4 (*CH}\_3\text{CO}*), 52.5 (C-2), 63.6 (C-5), 68.9 (C-6), 70.6 (*OCH}\_2\text{Ph}*), 78.4 (C-3), 79.2 (C-4), 98.0 (C-1), 101.8 (*CHPh*), 126.4, 128.1, 128.3, 128.4, 128.6, 128.9, 129.2, 129.5, 133.9, 136.7, 136.8, 144.6, 170.6 (C=O); IR 3324, 1656, 1547, 1362, 1182, 1123, 1097  $\text{cm}^{-1}$ ; HRMS (FAB) calcd for  $\text{C}_{29}\text{H}_{32}\text{NO}_8\text{S}$  ( $\text{M} + \text{H}^+$ ), 554.1849, found 554.1848. **3a**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.72 (s, 3H, *CH}\_3\text{CO}*), 2.37 (s, 3H, *CH}\_3\text{Ph}*), 3.53 (ddd,  $J = 3.8, 10.2, 10.2$  Hz, 1H, H-2), 3.56 (dd,  $J = 9.7, 9.7$  Hz, 1H, H-4), 3.67 (dd,  $J = 10.3, 10.3$  Hz, 1H, H-6a), 3.83 (ddd,  $J = 4.8, 9.8, 9.8$  Hz, 1H, H-5), 4.15 (dd,  $J = 4.8, 10.4$  Hz, 1H, H-6b), 4.37 and 4.62 (AB,  $J = 11.6$  Hz, 2H, *OCH}\_2\text{Ph}*), 4.71 (d,  $J = 3.8$  Hz, 1H, H-1), 5.00 (d,  $J = 10.2$  Hz, 1H, *NH*), 5.24 (dd,  $J = 10.1, 10.1$  Hz, 1H, H-3), 5.42 (s, 1H, *CHPh*), 7.21 (d,  $J = 8.4$  Hz, 2H, Ts), 7.26–7.39 (m, 10H), 7.62 (d,  $J = 8.4$  Hz, 2H, Ts);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  20.8 (*CH}\_3\text{CO}*), 21.7 (*CH}\_3\text{Ph}*), 56.8 (C-2), 63.1 (C-5), 68.9 (C-6), 69.4 (C-3), 70.5 (*OCH}\_2\text{Ph}*), 79.4 (C-4), 98.0 (C-1), 101.7 (*CHPh*), 126.3, 127.1, 128.4, 128.6, 128.6, 128.9, 129.3, 129.9, 136.4, 137.0, 138.4, 143.7, 170.8 (C=O); IR 3356, 1734, 1498, 1337, 1234, 1163, 1090  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{29}\text{H}_{31}\text{NNaO}_8\text{S}$  ( $\text{M} + \text{Na}^+$ ), 576.1659, found 576.1662.

#### ***N*-(4-Acetamidobenzyl)acetamide (14)**

Acetic anhydride (3.4 mL, 36 mmol) and pyridine (6.6 mL, 82 mmol) were added to a solution of 4-aminobenzylamine (2.0 g, 16 mmol) in  $\text{CH}_2\text{Cl}_2$  (100 mL), and the mixture was stirred for 40 h. After removal of the solvent and extra reagent under reduced pressure, the resulting yellow solid was washed with  $\text{Et}_2\text{O}$  to afford the product **14** (3.3 g, 99%) as a light yellow solid. **14**:  $^1\text{H}$  NMR (500 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  1.97 (s, 3H, *CH}\_3\text{CO}*), 2.11 (s, 3H, *CH}\_3\text{CO}*), 4.30 (s, 2H, *CH}\_2*), 7.22 (d,  $J = 8.6$  Hz, 2H), 7.49 (d,  $J = 8.6$  Hz, 2H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  22.7 (*CH}\_3\text{CO}*), 23.9 (*CH}\_3\text{CO}*), 43.9 (*CH}\_2*), 121.4, 129.3, 135.8, 139.2, 171.8 (C=O), 173.2 (C=O); HRMS (ESI) calcd for  $\text{C}_{11}\text{H}_{14}\text{N}_2\text{NaO}_2$  ( $\text{M} + \text{Na}^+$ ), 229.0947, found 229.0966.

#### ***trans*-2-(Acetamido)cyclohexanol (1b)**

Sodium methoxide (356 mg, 6.6 mmol) was added to a solution of *trans*-2-aminocyclohexanol hydrochloride (1.0 g, 6.6 mmol) in MeOH (30 mL), and the mixture was stirred for 15 min. After addition of acetic anhydride (0.69 mL, 7.3 mmol), the resulting mixture was stirred at room temperature for 40 h, and then the solvent and extra reagent were removed under reduced pressure.  $\text{CHCl}_3$  was added to the residue and filtrated. The combined filtrate was concentrated to dryness, and the resulting colorless paste was added hexane. The resulting suspension was filtrated to afford the product **1b** (1.03 g, quant.) as a white solid. **1b**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.11–1.38 (m, 4H), 1.64–1.77 (m, 2H), 1.94 (m, 1H), 2.01 (s, 3H, *CH}\_3\text{CO}*), 2.05 (m, 1H), 3.31 (ddd,  $J = 4.4, 10.0, 10.0$ , 1H, H-2), 3.60 (m, 1H, H-1), 5.64 (br s, 1H, *OH*), 6.16 (br s, 1H, *NH*);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  23.5 (*CH}\_3\text{CO}*), 24.3 and 24.8 (C-4, C-5), 31.7 (C-3), 34.6 (C-6), 55.9 (C-2), 75.0 (C-1), 172.3 (C=O); IR 3275, 1646, 1562, 1450, 1376, 1140, 1069  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_8\text{H}_{16}\text{NO}_2$  ( $\text{M} + \text{H}^+$ ), 158.1176, found 158.1197.

Products of the tosylation of the compound **1b**

#### ***trans*-2-Acetamido-1-*O*-tosyl-cyclohexanol (2b)**

#### ***trans*-1-*O*-Acetyl-2-(tosylamido)cyclohexanol (3b)**

#### ***trans*-2-(Tosylamido)cyclohexanol (15b)**

#### ***trans*-2-Acetamido-1-*O*-acetyl-cyclohexanol (16b)**

The condition A was used for compound **1b** (150 mg, 0.93 mmol), and the crude product was purified by silica gel column chromatography (AcOEt/hexane, 1:3 to 1:0) to afford the product **2b** (27 mg, 9%) as a white solid, the product **3b** (107 mg, 36%) as a white solid, the product **15b** (64 mg, 25%) as a white solid and the product **16b** (36 mg, 19%) as a white solid. The structures of the products **3b**,

**15b** and **16b** were confirmed by comparison to the reported data.<sup>2</sup> **2b**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.10–1.37 (m, 3H), 1.54 (m, 1H), 1.64 (m, 1H), 1.73 (m, 1H), 1.88 (m, 1H), 1.94 (s, 3H, CH<sub>3</sub>CO), 2.19 (m, 1H), 2.46 (s, 3H, CH<sub>3</sub>Ph), 3.85 (m, 1H, H-2), 4.34 (ddd, *J* = 4.6, 10.8, 10.8 Hz, 1H, H-1), 5.78 (d, *J* = 7.2 Hz, 1H, NH), 7.35 (d, *J* = 8.6 Hz, 2H, Ts), 7.78 (d, *J* = 8.0 Hz, 2H, Ts); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 21.9 (CH<sub>3</sub>Ph), 23.7 (CH<sub>3</sub>CO), 24.0 (C-4), 24.4 (C-5), 32.1 (C-6), 32.2 (C-3), 52.5 (C-2), 82.5 (C-1), 127.8, 130.1, 134.2, 145.1, 170.2 (C=O); IR 3436, 1638, 1561, 1370, 1188, 1175 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>15</sub>H<sub>21</sub>NNaO<sub>4</sub>S (M + Na<sup>+</sup>), 334.1083, found 334.1075. **3b**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.14–1.38 (m, 4H), 1.64 (m, 1H), 1.69 (m, 1H), 1.76 (s, 3H, CH<sub>3</sub>CO), 1.92 (m, 1H), 2.02 (m, 1H), 2.42 (s, 3H, CH<sub>3</sub>Ph), 3.19 (m, 1H, H-2), 4.55 (ddd, *J* = 4.5, 10.2, 10.2 Hz, 1H, H-1), 4.95 (d, *J* = 7.6 Hz, 1H, NH), 7.29 (d, *J* = 8.0 Hz, 2H, Ts), 7.74 (d, *J* = 8.2 Hz, 2H, Ts); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 21.1 (CH<sub>3</sub>CO), 21.7 (CH<sub>3</sub>Ph), 23.9 and 24.4 (C-4, C-5), 31.3 (C-6), 33.7 (C-3), 57.1 (C-2), 74.3 (C-1), 127.1, 129.8, 138.8, 143.2, 171.6 (C=O); IR 3261, 1715, 1599, 1453, 1335, 1257, 1163, 1094, 1046 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>15</sub>H<sub>22</sub>NO<sub>4</sub>S (M + H<sup>+</sup>), 312.1264, found 312.1277. **15b**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.05–1.28 (m, 4H), 1.58 (m, 1H), 1.65 (m, 1H), 1.73 (m, 1H), 2.01 (m, 1H), 2.43 (s, 3H, CH<sub>3</sub>Ph), 2.76 (br s, 1H, OH), 2.86 (m, 1H, H-2), 3.31 (ddd, *J* = 4.3, 9.6, 9.6 Hz, 1H, H-1), 5.10 (d, *J* = 7.0 Hz, 1H, NH), 7.32 (d, *J* = 8.0 Hz, 2H, Ts), 7.80 (d, *J* = 8.4 Hz, 2H, Ts); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 21.8 (CH<sub>3</sub>Ph), 24.0, 24.8, 32.0, 33.5, 59.9 (C-2), 73.5 (C-1), 127.3, 130.0, 137.5, 143.8; HRMS (ESI) calcd for C<sub>13</sub>H<sub>20</sub>NO<sub>3</sub>S (M + H<sup>+</sup>), 270.1158, found 270.1129. **16b**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.14 (m, 1H), 1.19–1.35 (m, 2H), 1.44 (m, 1H), 1.66 (m, 1H), 1.74 (m, 1H), 1.89 (s, 3H, CH<sub>3</sub>CO), 1.92 (m, 1H), 2.01 (s, 3H, CH<sub>3</sub>CO), 2.03 (m, 1H), 3.83 (dddd, *J* = 4.2, 4.4, 10.2, 10.2 Hz, 1H, H-2), 4.61 (ddd, *J* = 4.5, 10.7, 10.7 Hz, 1H, H-1), 5.81 (br s, 1H, NH); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 21.3 (CH<sub>3</sub>CO), 23.5 (CH<sub>3</sub>CO), 24.2 and 24.3 (C-4, C-5), 31.2 (C-6), 32.2 (C-3), 52.9 (C-2), 74.8 (C-1), 169.9 (C=O), 172.1 (C=O); IR 3289, 1732, 1646, 1559, 1375, 1246, 1042 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>10</sub>H<sub>17</sub>NNaO<sub>3</sub> (M + Na<sup>+</sup>), 222.1101, found 222.1104.

Products of the tosylation of 2-acetamidoethanol (**1c**)

**1-O-Acetyl-2-(tosylamido)ethanol (3c)**

**2-(Tosylamido)ethanol (15c)**

**2-Acetamido-1-O-acetyl-ethanol (16c)**

The condition A was used for 2-acetamidoethanol (1.12 g, 10.9 mmol), and the crude product was purified by silica gel column chromatography (AcOEt/hexane, 1:3 to 1:0) to afford the product **3c** (1.01 g, 36%) as a colorless oil, the product **15c** (748 mg, 32%) as a white paste and the product **16c** (457 mg, 29%) as a colorless oil. **3c**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 2.00 (s, 3H, CH<sub>3</sub>CO), 2.43 (s, 3H, CH<sub>3</sub>Ph), 3.22 (dt, *J* = 5.4, 5.7 Hz, 1H, H-2), 4.09 (t, *J* = 5.3 Hz, 1H, H-1), 5.06 (t, *J* = 5.6 Hz, 1H, NH), 7.32 (d, *J* = 7.8 Hz, 2H, Ts), 7.76 (d, *J* = 8.2 Hz, 2H, Ts); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 20.9 (CH<sub>3</sub>CO), 21.7 (CH<sub>3</sub>Ph), 42.3 (C-2), 63.1 (C-1), 127.2, 130.0, 137.0, 143.9, 171.0 (C=O); IR 3237, 1736, 1719, 1596, 1441, 1331, 1252, 1160, 1091, 1051 cm<sup>-1</sup>; HRMS (ESI) calcd for C<sub>11</sub>H<sub>15</sub>NNaO<sub>4</sub>S (M + Na<sup>+</sup>), 280.0614, found 280.0603. **15c**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 2.42 (s, 3H, CH<sub>3</sub>Ph), 3.08 (m, 1H, H-2), 3.61 (m, 1H, OH), 3.68 (m, 1H, H-1), 5.42 (br m, 1H, NH), 7.31 (d, *J* = 7.8 Hz, 2H, Ts), 7.76 (d, *J* = 8.2 Hz, 2H, Ts); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 21.7 (CH<sub>3</sub>Ph), 45.4 (C-2), 61.4 (C-1), 127.3, 130.0, 136.7, 143.8; HRMS (ESI) calcd for C<sub>9</sub>H<sub>13</sub>NNaO<sub>3</sub>S (M + Na<sup>+</sup>), 238.0508, found 238.0497. **16c**: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 1.98 (s, 3H, CH<sub>3</sub>CO), 2.06 (s, 3H, CH<sub>3</sub>CO), 3.49 (m, 1H, H-2), 4.13 (m, 1H, H-1), 5.07 (br s, 1H, NH); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 21.0 (CH<sub>3</sub>CO), 23.3 (CH<sub>3</sub>CO), 38.9 (C-2), 63.4 (C-1), 171.3

<sup>2</sup> For compound **3b**: (a) Liu, Y-K.; Li, R.; Yue, L.; Li, B-J.; Chen, Y-C.; Wu, Y.; Ding, L-S. *Org. Lett.* **2006**, *8*, 1521–1524. For compound **15b**: (b) Wang, Z.; Cui, Y-T.; Xu, Z-B.; Qu J. *J. Org. Chem.* **2008**, *73*, 2270–2274. (c) Fan, R-H.; Hou, X-L. *Org. Biomol. Chem.* **2003**, *1*, 1565–1567. For compound **16b**: (d) Miller, S. J.; Copeland, G. T.; Papaioannou, N.; Horstmann, T. E.; Ruel, E. M. *J. Am. Chem. Soc.* **1998**, *120*, 1629–1630. (e) Tabanella, S.; Valancogne, I.; Jackson, R. F. W. *Org. Biomol. Chem.* **2003**, *1*, 4254–4261.

(C=O); IR 3294, 1740, 1658, 1557, 1375, 1237, 1054  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_6\text{H}_{11}\text{NNaO}_3$  ( $\text{M} + \text{Na}^+$ ), 168.0631, found 168.0625.

### 1-(Acetamidomethyl)cyclohexanol (**1d**)

Sodium methoxide (489 mg, 9.1 mmol) was added to a solution of 1-aminomethyl-1-cyclohexanol hydrochloride (1.5 g, 9.1 mmol) in MeOH (30 mL), and the mixture was stirred for 15 min. After addition of acetic anhydride (0.94 mL, 10 mmol), the resulting mixture was stirred at room temperature for 25 h, and then the solvent and extra reagent were removed under reduced pressure.  $\text{CH}_2\text{Cl}_2$  was added to the residue, and the resulting suspension was filtrated. The combined filtrate was concentrated to afford the product **1d** (1.55 g, quant.) as a white solid. **1d**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.25–1.63 (m, 10H), 2.01 (s, 3H,  $\text{CH}_3\text{CO}$ ), 2.80 (br s, 1H), 3.25 (d,  $J = 6.0$  Hz, 2H,  $\text{CH}_2\text{NHAc}$ ), 6.32 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  22.2, 23.4, 25.9, 35.7, 49.3 ( $\text{CH}_2\text{NHAc}$ ), 71.7 (C-1), 171.5 (C=O); IR 3391, 3290, 1652, 1552, 1421, 1292, 1274, 1166, 1127, 1044  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_9\text{H}_{17}\text{NNaO}_2$  ( $\text{M} + \text{Na}^+$ ), 194.1151, found 194.1147.

Product of the tosylation of the compound **1d**

### 1-O-Acetyl-1-[(tosylamido)methyl]cyclohexanol (**3d**)

The condition A was used for compound **1d** (150 mg, 0.88 mmol), and the crude product was purified by silica gel column chromatography (AcOEt) to afford the product **3d** (148 mg, 52%) as a white solid. The condition B gave the product **3d** in 96% from the same amount of starting material. **3d**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.29 (m, 2H), 1.37–1.59 (m, 6H), 1.98 (s, 3H,  $\text{CH}_3\text{CO}$ ), 2.08 (m, 2H), 2.42 (s, 3H,  $\text{CH}_3\text{Ph}$ ), 3.25 (d,  $J = 6.6$  Hz, 2H,  $\text{CH}_2\text{NHTs}$ ), 5.53 (t,  $J = 6.6$  Hz, 1H,  $\text{NH}$ ), 7.29 (d,  $J = 7.8$  Hz, 2H, Ts), 7.72 (d,  $J = 8.2$  Hz, 2H, Ts);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  21.7 ( $\text{CH}_3\text{Ph}$ ), 21.7 (C-3, C-5), 22.3 ( $\text{CH}_3\text{CO}$ ), 25.4 (C-4), 32.8 (C-2, C-6), 49.4 ( $\text{CH}_2\text{NHTs}$ ), 83.0 (C-1), 127.1, 129.9, 137.3, 143.4, 171.4 (C=O); IR 3284, 1734, 1599, 1450, 1328, 1236, 1163, 1094, 1064  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{16}\text{H}_{23}\text{NNaO}_4\text{S}$  ( $\text{M} + \text{Na}^+$ ), 348.1240, found 348.1246.

### 2-Acetamido-1,5-anhydro-4,6-O-benzylidene-2-deoxy-D-glucitol (**1e**)

Sodium methoxide (110 mg, 2.1 mmol) was added to a solution of 2-acetamido-3-O-acetyl-1,5-anhydro-4,6-O-benzylidene-2-deoxy-D-glucitol<sup>3</sup> (700 mg, 2.1 mmol) in MeOH (15 mL), and the mixture was stirred for 3 h at room temperature. The reaction mixture was quenched by the addition of the Amberlite IR-120 ( $\text{H}^+$ ), and the resulting mixture was stirred for 5 min. The resin was filtered, and the filtrate was concentrated to dryness to afford the product **1e** (610 mg, quant.) as a white solid. **1e**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3/\text{CD}_3\text{OD} = 95:5$ )  $\delta$  1.95 (s, 3H,  $\text{CH}_3\text{CO}$ ), 3.15 (dd,  $J = 11.0, 11.0$  Hz, 1H, H-1a), 3.32 (ddd,  $J = 5.2, 9.6, 9.6$  Hz, 1H, H-5), 3.50 (dd,  $J = 9.2, 9.2$  Hz, 1H, H-4), 3.62 (dd,  $J = 9.5, 9.5$  Hz, 1H, H-3), 3.68 (dd,  $J = 10.4, 10.4$  Hz, 1H, H-6a), 3.94 (ddd,  $J = 5.4, 10.4, 10.4$  Hz, 1H, H-2), 4.05 (dd,  $J = 5.5, 11.1$  Hz, 1H, H-1b), 4.26 (dd,  $J = 5.0, 10.6$  Hz, 1H, H-6b), 5.52 (s, 1H,  $\text{CHPh}$ ), 7.30–7.50 (m, 5H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3/\text{CD}_3\text{OD} = 95:5$ )  $\delta$  22.9 ( $\text{CH}_3\text{CO}$ ), 52.2 (C-2), 68.7 (C-1), 68.8 (C-6), 71.5 (C-5), 72.4 (C-3), 82.1 (C-4), 102.0 ( $\text{CHPh}$ ), 126.4, 128.4, 129.4, 137.2, 172.3 (C=O); IR 3426, 3277, 1655, 1544, 1377, 1127, 1100  $\text{cm}^{-1}$ ; HRMS (FAB) calcd for  $\text{C}_{15}\text{H}_{20}\text{NO}_5$  ( $\text{M} + \text{H}^+$ ), 294.1336, found 294.1339.

Product of the tosylation of the compound **1e**

### 3-O-Acetyl-1,5-anhydro-4,6-O-benzylidene-2-deoxy-2-tosylamido-D-glucitol (**3e**)

The condition B was used for compound **1e** (50 mg, 0.17 mmol), and the crude product was purified by silica gel column chromatography (AcOEt/hexane, 1:3) to afford the product **3e** (93 mg, 94%) as a white solid. **3e**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.70 (s, 3H,  $\text{CH}_3\text{CO}$ ), 2.44 (s, 3H,  $\text{CH}_3\text{Ph}$ ), 3.34 (dd,  $J = 11.3, 11.3$  Hz, 1H, H-1a), 3.38 (ddd,  $J = 5.0, 9.6, 9.6$  Hz, 1H, H-5), 3.49 (m, 1H, H-2), 3.56 (dd,  $J = 9.4,$

<sup>3</sup> Heseck, D.; Lee, M.; Yamaguchi, T.; Noll, B. C.; Mobashery, S. *J. Org. Chem.* **2008**, *73*, 7349–7352.

9.4 Hz, 1H, H-4), 3.67 (dd,  $J = 10.3, 10.3$  Hz, 1H, H-6a), 4.13 (dd,  $J = 5.4, 11.6$  Hz, 1H, H-1b), 4.30 (dd,  $J = 5.0, 10.6$  Hz, 1H, H-6b), 4.96 (dd,  $J = 9.7, 9.7$  Hz, 1H, H-3), 5.32 (d,  $J = 6.8$  Hz, 1H, NH), 5.45 (s, 1H, CHPh), 7.33 (d,  $J = 7.8$  Hz, 2H, Ts), 7.30–7.42 (m, 5H), 7.75 (d,  $J = 8.2$  Hz, 2H, Ts);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  20.8 ( $\text{CH}_3\text{CO}$ ), 21.7 ( $\text{CH}_3\text{Ph}$ ), 54.6 (C-2), 68.8 (C-6), 70.8 (C-1), 71.6 (C-5), 73.0 (C-3), 79.0 (C-4), 101.7 (CHPh), 126.3, 127.2, 128.4, 129.4, 130.0, 137.0, 137.7, 143.8, 172.3 (C=O); IR 3287, 1745, 1455, 1227, 1167, 1088  $\text{cm}^{-1}$ ; HRMS (FAB) calcd for  $\text{C}_{22}\text{H}_{26}\text{NO}_7\text{S}$  ( $\text{M} + \text{H}^+$ ), 448.1424, found 448.1421.

Product of the tosylation of 2-acetamidophenol (**1f**)

#### **1-O-Tosyl-2-(acetamido)phenol (2f)**

The condition A was used for 2-acetamidophenol (1.0 g, 6.6 mmol), and the crude product was purified by silica gel column chromatography (AcOEt/hexane, 1:1) to afford the product **2f** (1.98 g, 98%) as colorless crystals. The condition B gave the product **2f** in 98% from the same amount of starting material. **2f**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.07 (s, 3H,  $\text{CH}_3\text{CO}$ ), 2.47 (s, 3H,  $\text{CH}_3\text{Ph}$ ), 6.91 (d,  $J = 8.0$  Hz, 1H, C-3), 6.99 (dd,  $J = 7.8, 7.8$  Hz, 1H, C-4), 7.24 (dd,  $J = 7.8, 7.8$  Hz, 1H, C-5), 7.35 (d,  $J = 8.0$  Hz, 2H, Ts), 7.56 (br s, 1H, NH), 7.73 (d,  $J = 8.4$  Hz, 2H, Ts), 8.19 (d,  $J = 8.2$  Hz, 1H, C-6);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  22.0 ( $\text{CH}_3\text{Ph}$ ), 24.8 ( $\text{CH}_3\text{CO}$ ), 123.0, 123.0, 124.5, 128.0, 128.7, 130.3, 131.3, 131.9, 139.1, 146.5, 168.3 (C=O); IR 3400, 3302, 1695, 1675, 1600, 1525, 1442, 1372, 1307, 1194, 1170, 1088  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{15}\text{H}_{15}\text{NNaO}_4\text{S}$  ( $\text{M} + \text{Na}^+$ ), 328.0614, found 328.0599.

Crystal was examined under Infineum V8512 oil. A suitable sample was glued to a glass capillary and then transferred to the 100 K  $\text{N}_2$  stream for compound **2f** of a Bruker SMART Apex CCD diffractometer. Preliminary unit cell parameters and crystal system analysis were determined from the centroids of reflections with  $I > 20\sigma(I)$  from three sets of 30  $0.5^\circ$   $\omega$ -scan frames ( $\varphi = 0, 120, 240^\circ$ ); final unit cell parameters were determined from the centroids of reflections with  $I > 10\sigma(I)$  from all data. An arbitrary, redundant, sphere of data was calculated using COSMO, included in the Apex2 suite of programs.<sup>4</sup> Data are uncorrected for absorption and Lorentz polarization effects. Structure solution and refinement utilized the programs of the SHELXTL software package.<sup>5</sup> Full detail of the X-ray structure determination is in the CIF file and ORTEP diagram is given in Figure S1.

#### **trans-4-(Acetamido)cyclohexanol (1g)**

Sodium methoxide (1.07 g, 19.8 mmol) was added to a solution of *trans*-4-aminocyclohexanol hydrochloride (3.0 g, 19.8 mmol) in MeOH (100 mL), and the mixture was stirred for 15 min. After addition of acetic anhydride (2.06 mL, 21.8 mmol), the mixture was stirred at room temperature for 26 h, and then the solvent and extra reagent were removed under reduced pressure.  $\text{CHCl}_3$  was added to the residue, and the resulting suspension was filtrated. The combined filtrate was concentrated to dryness, and the resulting colorless paste was added hexane. The resulting suspension was filtrated to afford the product **1g** (3.11 g, quant.) as a white solid. **1g**:  $^1\text{H}$  NMR (500 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  1.20–1.40 (m, 4H), 1.84–1.98 (m, 4H), 1.90 (s, 3H,  $\text{CH}_3\text{CO}$ ), 3.51 (m, 1H), 3.59 (m, 1H), 7.92 (br d,  $J = 7.0$  Hz, 1H, NH);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  22.8 ( $\text{CH}_3\text{CO}$ ), 31.6 and 34.9 (C-2, C-3, C-5 and C-6), 49.3 (C-4), 70.5 (C-1), 172.6 (C=O); IR 3277, 1636, 1562, 1456, 1371, 1328, 1148, 1112, 1065  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_8\text{H}_{16}\text{NO}_2$  ( $\text{M} + \text{H}^+$ ), 158.1176, found 158.1171.

Products of the tosylation of the compound **1g**

#### **trans-4-Acetamido-1-O-tosyl-cyclohexanol (2g)**

#### **trans-4-(Tosylamido)cyclohexanol (15g)**

#### **trans-4-Acetamido-1-O-acetyl-cyclohexanol (16g)**

The condition A was used for compound **1g** (450 mg, 2.9 mmol), and the crude product was purified by silica gel column chromatography (AcOEt) to afford the product **2g** (389 mg, 41%) as colorless

<sup>4</sup> Apex2. Bruker-AXS: Madison, WI, 2008; Vol. 58.

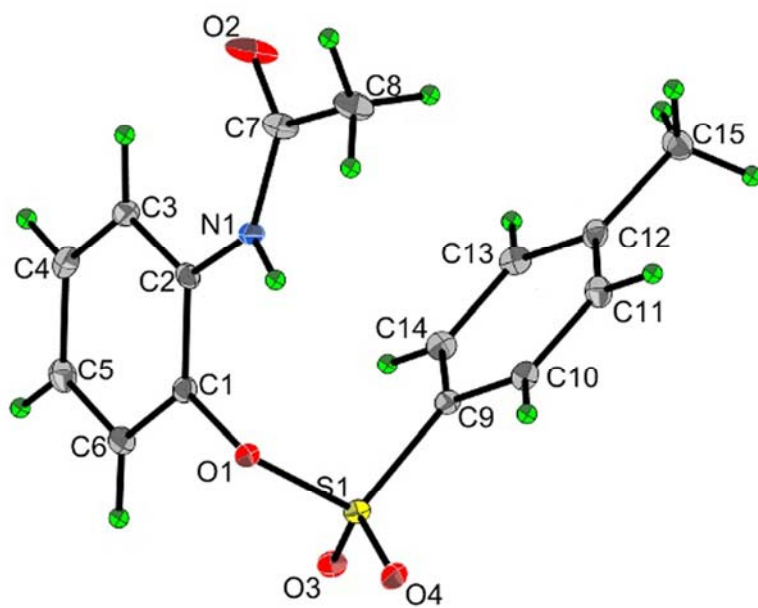
<sup>5</sup> Sheldrick, G. M., *Acta Crystallogr. A*. **2008**, *64*, 112–122.

crystals, the product **15g** (69 mg, 9%) as a white solid and the product **16g** (46 mg, 8%) as colorless crystals. **2g**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.06–1.27 (m, 2H), 1.48–1.66 (m, 2H), 1.84–2.03 (m, 4H), 1.89 (s, 3H,  $\text{CH}_3\text{CO}$ ), 2.43 (s, 3H,  $\text{CH}_3\text{Ph}$ ), 3.68 (m, 1H, H-4), 4.40 (ddd,  $J = 3.9, 10.6, 10.6$  Hz, 1H, H-1), 5.69 (d,  $J = 7.4$  Hz, 1H,  $\text{NH}$ ), 7.32 (d,  $J = 8.1$  Hz, 2H, Ts), 7.75 (d,  $J = 8.1$  Hz, 2H, Ts);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  21.8 ( $\text{CH}_3\text{Ph}$ ), 23.5 ( $\text{CH}_3\text{CO}$ ), 30.2 and 30.9 (C-2, C-3, C-5, C-6), 46.8 (C-4), 80.4 (C-1), 127.6, 130.0, 134.4, 144.9, 169.7 (C=O); IR 3408, 3312, 1674, 1652, 1552, 1519, 1343, 1189, 1177, 1096  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{15}\text{H}_{21}\text{NNaO}_4\text{S}$  ( $\text{M} + \text{Na}^+$ ), 334.1083, found 334.1072. **15g**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.10–1.24 (m, 4H), 1.69–1.86 (m, 4H), 2.39 (s, 3H,  $\text{CH}_3\text{Ph}$ ), 3.01 (m, 1H, H-4), 3.46 (m, 1H, H-1), 5.39 (d,  $J = 7.6$  Hz, 1H,  $\text{NH}$ ), 7.26 (d,  $J = 8.2$  Hz, 2H, Ts), 7.72 (d,  $J = 8.2$  Hz, 2H, Ts);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  21.6 ( $\text{CH}_3\text{Ph}$ ), 31.3 and 33.5 (C-2, C-3, C-5, C-6), 51.9 (C-4), 69.1 (C-1), 126.9, 129.8, 138.2, 143.4; HRMS (ESI) calcd for  $\text{C}_{13}\text{H}_{19}\text{NNaO}_3\text{S}$  ( $\text{M} + \text{Na}^+$ ), 292.0978, found 292.0955. **16g**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.13–1.31 (m, 2H), 1.35–1.53 (m, 2H), 1.86–2.03 (m, 4H), 1.91 (s, 3H,  $\text{CH}_3\text{CO}$ ), 1.99 (s, 3H,  $\text{CH}_3\text{CO}$ ), 3.72 (dt,  $J = 4.0, 7.7, 11.3$  Hz, 1H, H-4), 4.61 (tt,  $J = 4.0, 10.8$  Hz, 1H, H-1), 5.81 (br d,  $J = 7.2$  Hz, 1H,  $\text{NH}$ );  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  21.5 ( $\text{CH}_3\text{CO}$ ), 23.5 ( $\text{CH}_3\text{CO}$ ), 30.1 and 30.6 (C-2, C-3, C-4, C-5), 47.3 (C-4), 72.1 (C-1), 169.7 (C=O), 170.9 (C=O); IR 3440, 3251, 1726, 1638, 1562, 1365, 1250, 1035  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{10}\text{H}_{17}\text{NNaO}_3$  ( $\text{M} + \text{Na}^+$ ), 222.1101, found 222.1087.

Product of the mesylation of the compound **1d**

#### **1-O-Acetyl-1-[(mesylamido)methyl]cyclohexanol (19)**

Methanesulfonyl chloride (90  $\mu\text{L}$ , 1.2 mmol) was added to a solution of **1d** (100 mg, 0.58 mmol) in pyridine (1.0 mL) at 0  $^\circ\text{C}$ . The mixture was stirred at room temperature for 20 h, and then at 40  $^\circ\text{C}$  for 1 h. The solvent was removed under reduced pressure, and the crude product was purified by silica gel column chromatography (AcOEt/hexane, 2:3) to afford the product **19** (137 mg, 94%) as a white solid. **19**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.33 (m, 2H), 1.43–1.63 (m, 6H), 2.07 (s, 3H,  $\text{CH}_3\text{CO}$ ), 2.13 (m, 2H), 2.95 (s, 3H,  $\text{CH}_3\text{SO}_2$ ), 3.48 (d,  $J = 6.6$  Hz, 2H,  $\text{CH}_2\text{NHMs}$ ), 5.15 (t,  $J = 6.2$  Hz, 1H,  $\text{NH}$ );  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  21.7 (C-3, C-5), 22.3 ( $\text{CH}_3\text{CO}$ ), 25.4 (C-4), 32.9 (C-2, C-6), 40.6 (Ms), 49.8 ( $\text{CH}_2\text{NHMs}$ ), 83.1 (C-1), 171.5 (C=O); IR 3249, 1733, 1453, 1302, 1232, 1150  $\text{cm}^{-1}$ ; HRMS (ESI) calcd for  $\text{C}_{10}\text{H}_{19}\text{NNaO}_4\text{S}$  ( $\text{M} + \text{Na}^+$ ), 272.0927, found 272.0908.



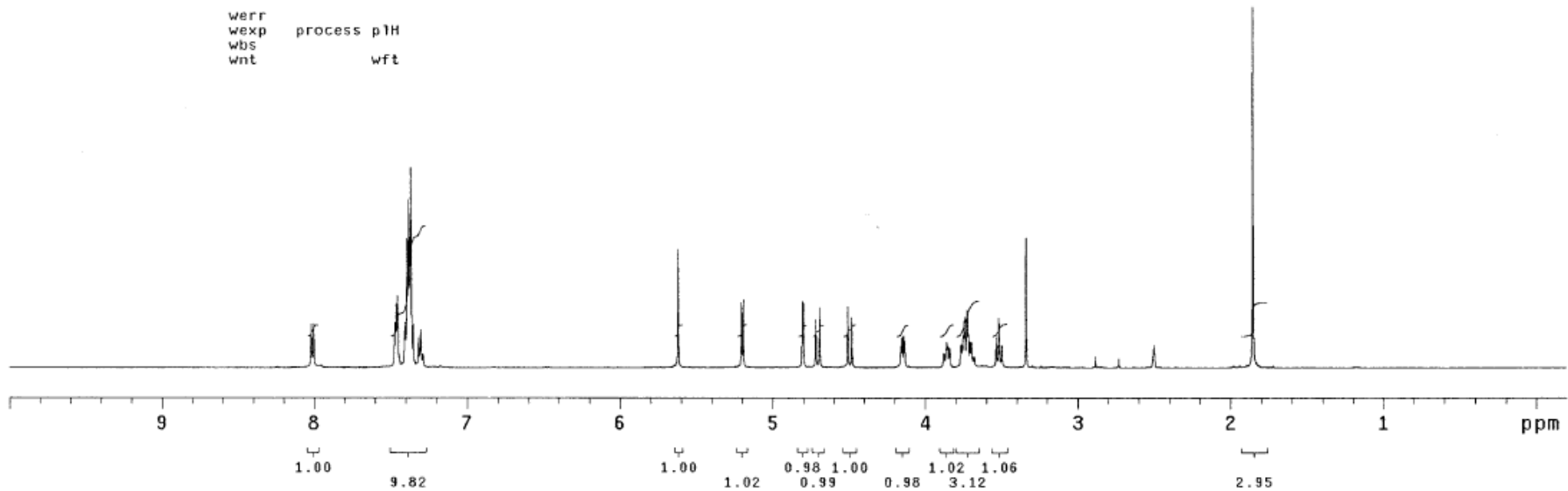
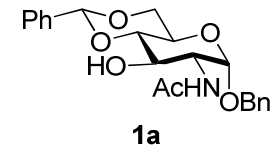
**Figure S1.** The ORTEP diagram of compound **2f**, shown at 30% probability level.



TY2-297

exp1 s2pu1

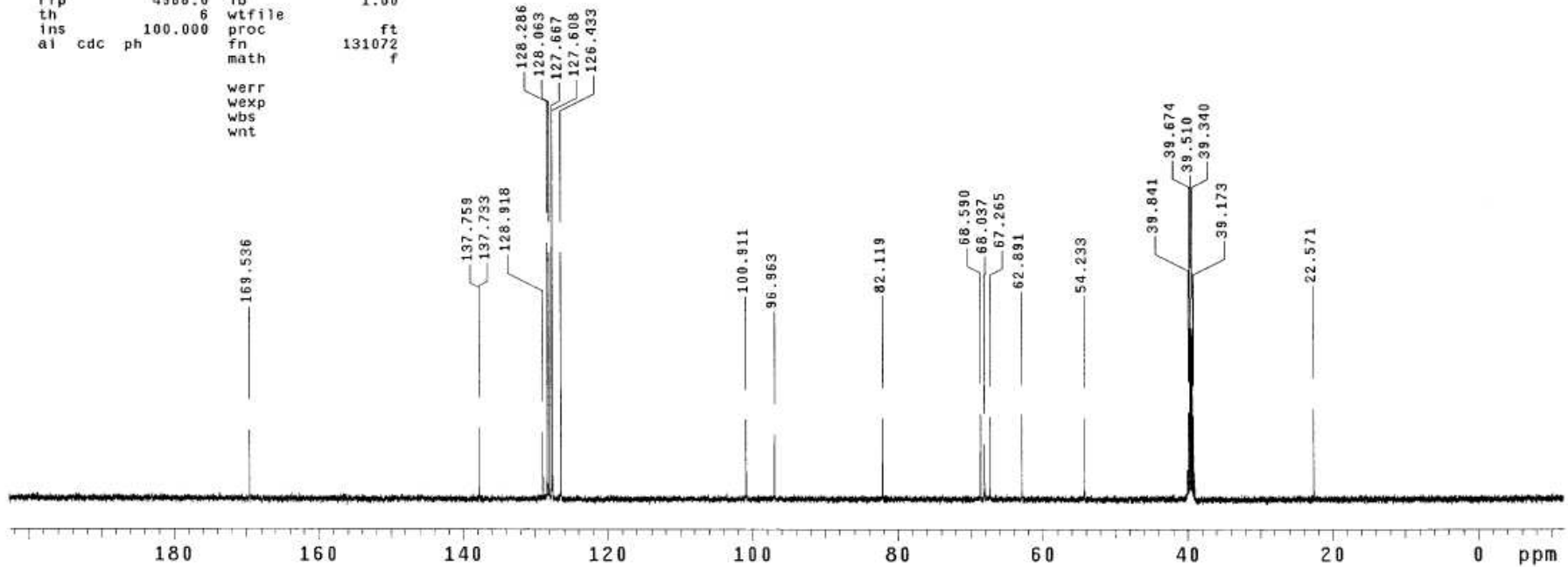
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TY2-297

exp2 s2pu1

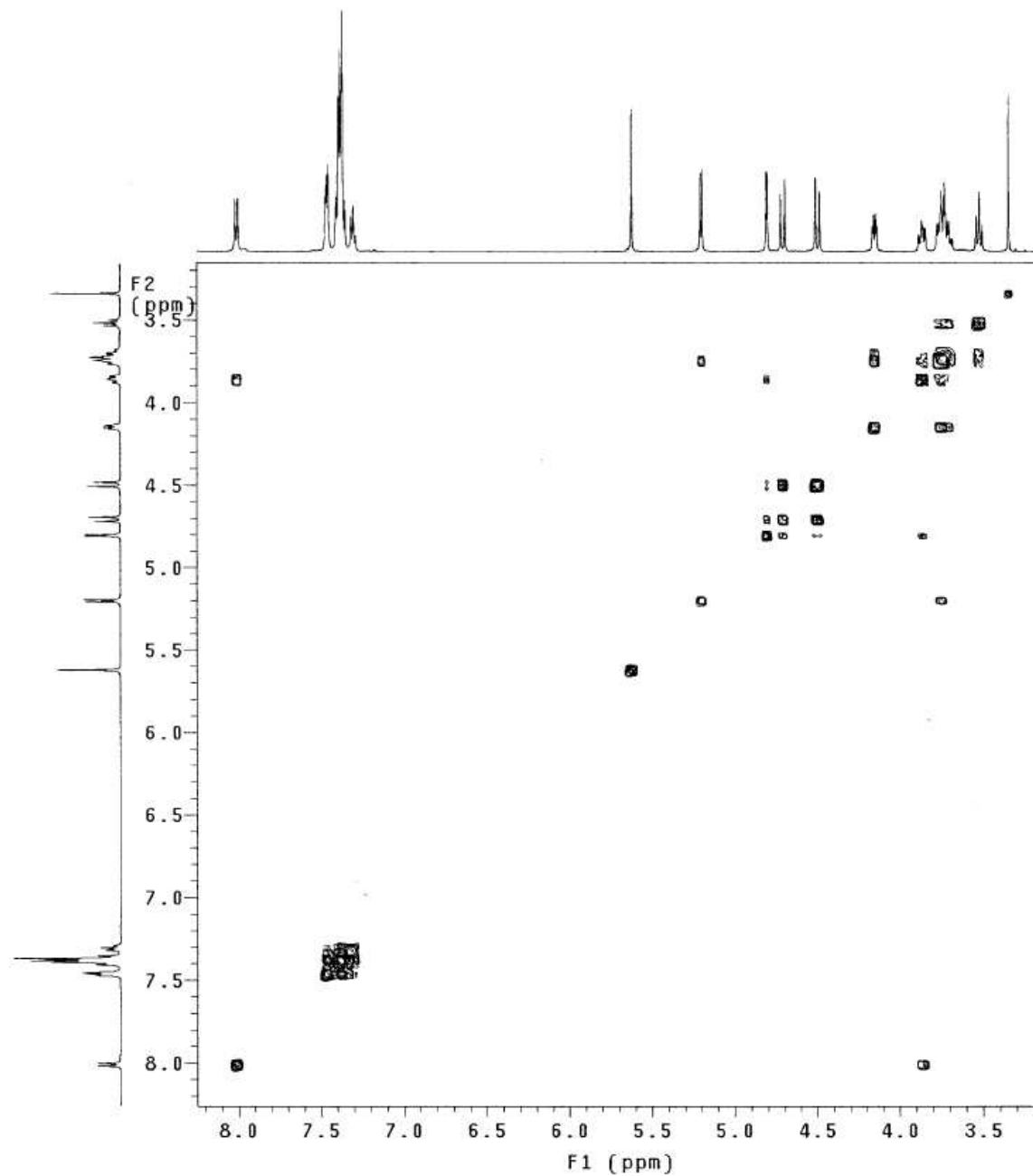
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TY2-297

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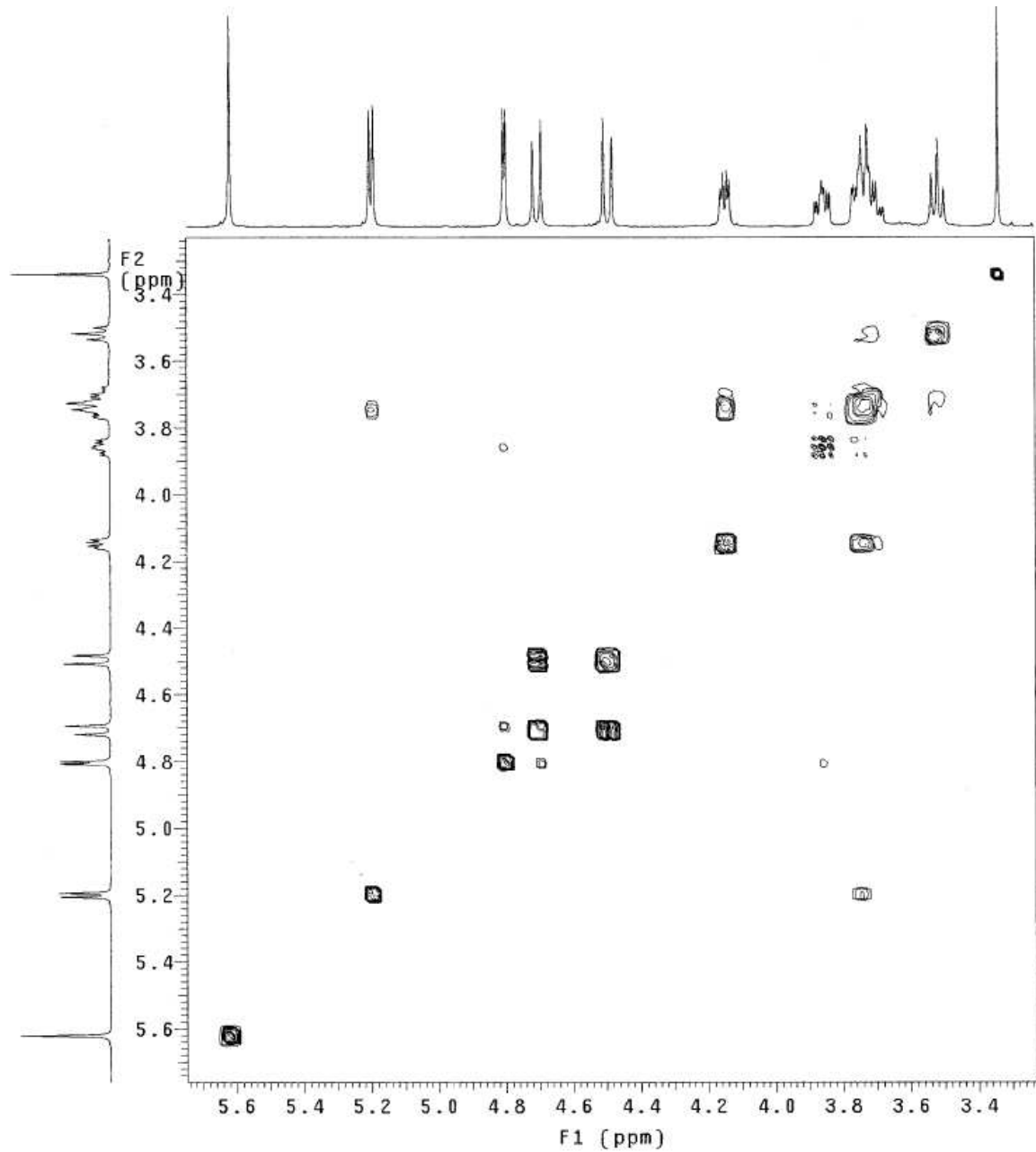
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TY2-297

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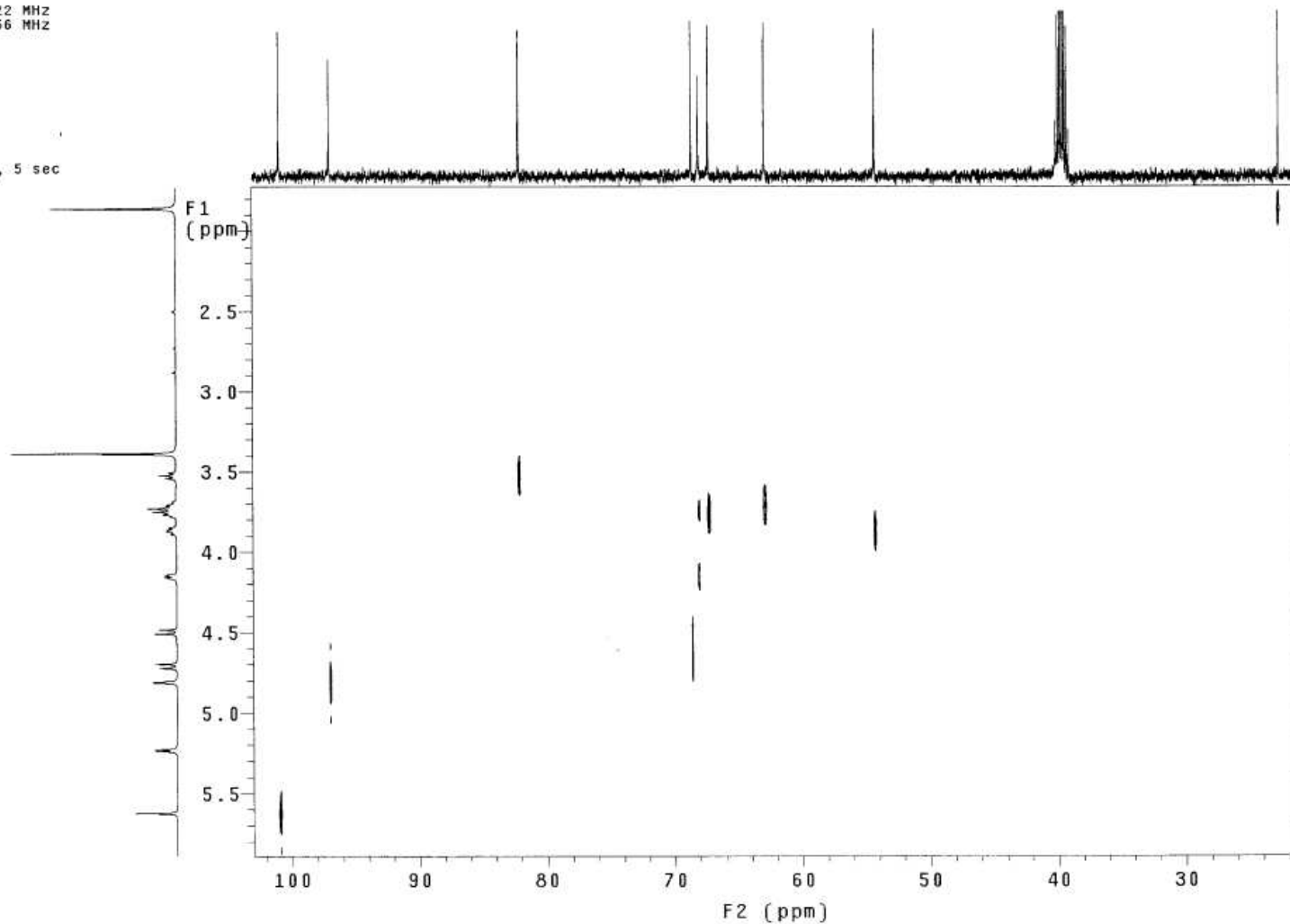
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TY2-297

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CH3 carbons



CH2 carbons



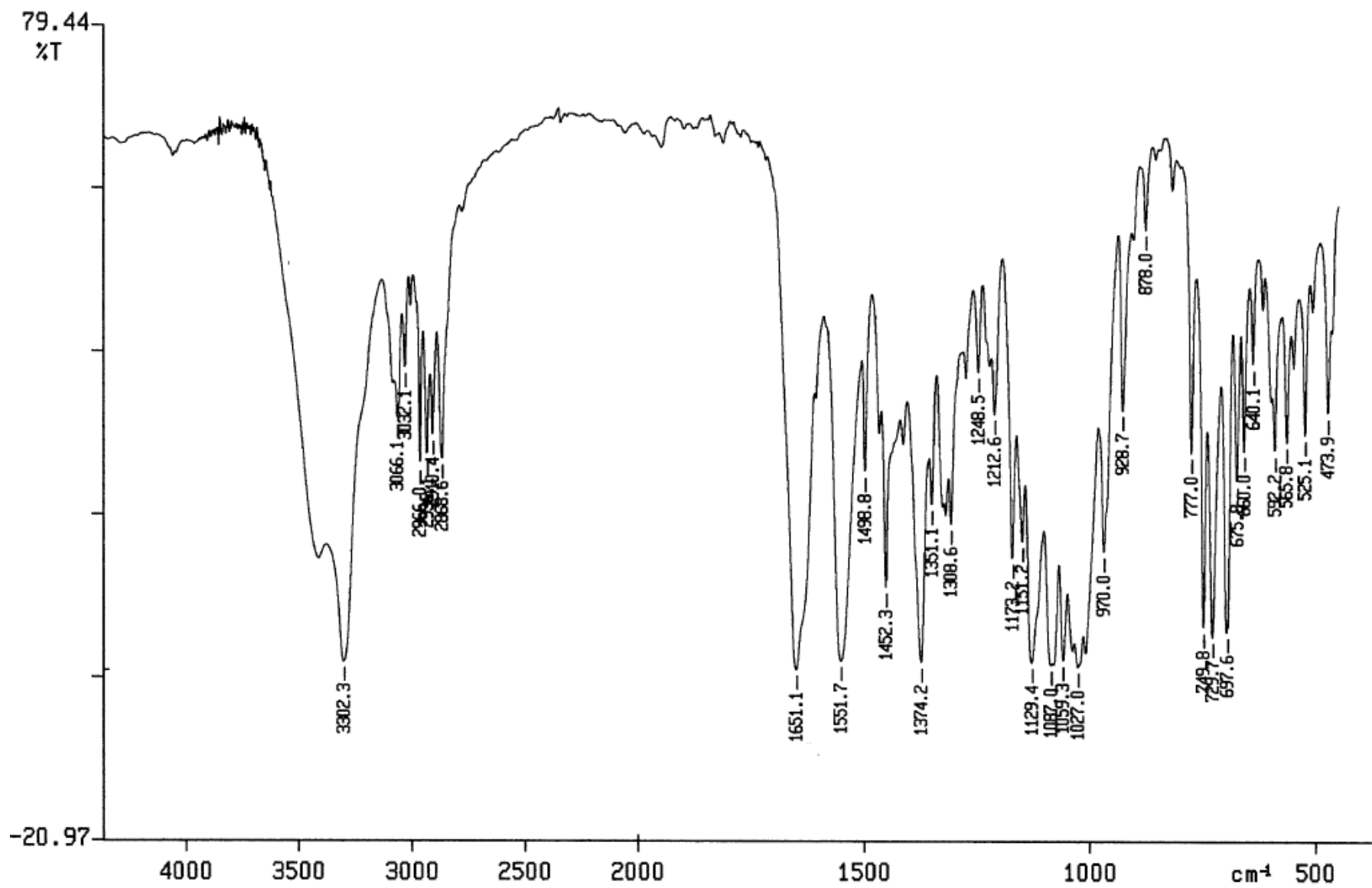
CH carbons



all protonated carbons



200 180 160 140 120 100 80 60 40 20 0 ppm

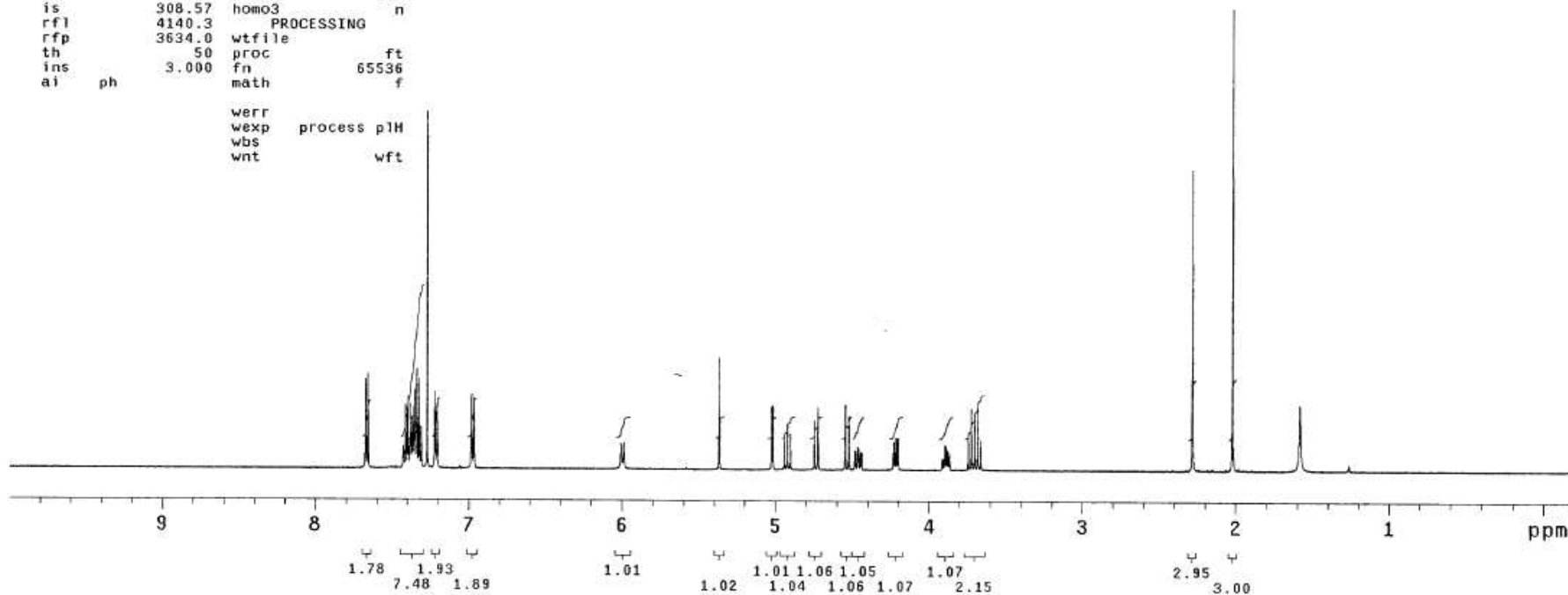
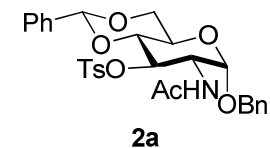


TY2-318-B

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gain	not used	dseq2	
FLAGS			
i1	n	homo2	1.0
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY			
sp	-102.8	dpwr3	1
wp	5099.3	dof3	0
vs	37	dm3	n
sc	0	dmm3	c
wc	250	dmf3	200
hzmm	20.40	dseq3	
is	308.57	dres3	1.0
rfl	4140.3	homo3	n
PROCESSING			
rff	3634.0	wfile	
th	50	proc	ft
ins	3.000	fn	65536
ai	ph	math	f

werr  
wexp process pH  
wbs  
wnt wft

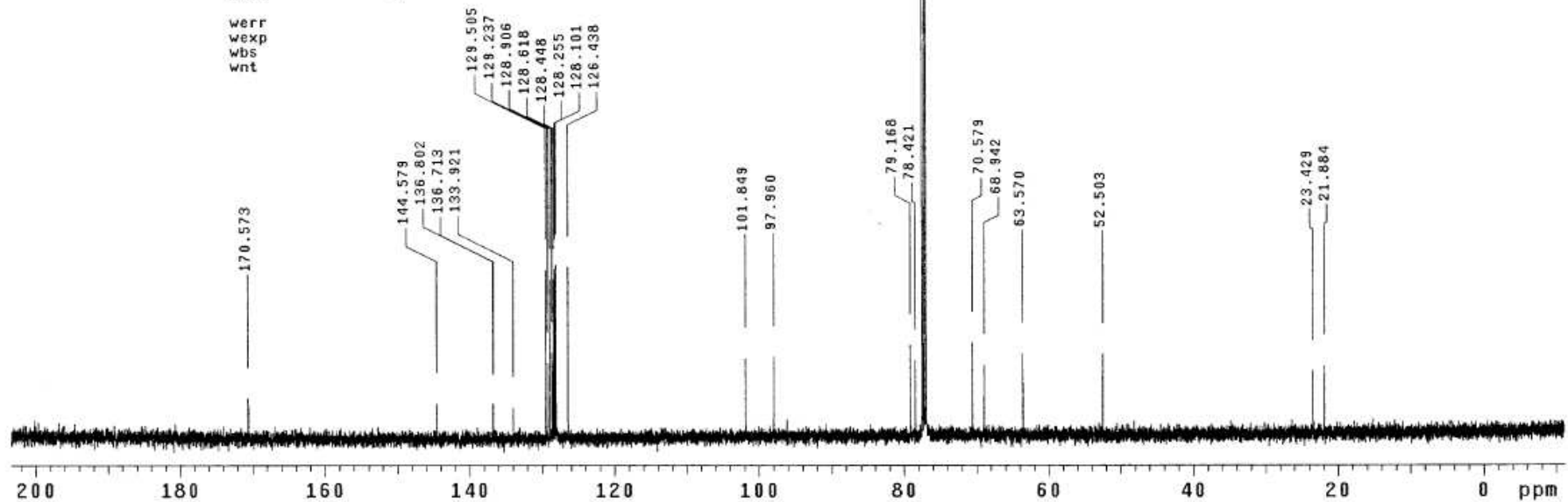




TY2-318-B

exp2 s2pu1

SAMPLE		DEC. & VT	
date	Mar 21 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION		dof	0
sfrq	125.702	dm	yyy
tn	C13	dmm	w
at	1.215	dmf	8787.35
np	65536	dseq	
sw	26963.3	dres	1.0
fb	15000	homo	n
bs	4	DEC2	
tpwr	52	dfrq2	0
pw	10.2	dn2	
d1	1.800	dpwr2	1
tof	144.5	dof2	0
nt	3000	dm2	n
ct	730	dmm2	c
alock	n	dmf2	10000
gain	not used	dseq2	
FLAGS		dres2	1.0
il	n	homo2	n
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY		dpwr3	1
sp	-1400.2	dof3	0
wp	26962.9	dm3	n
vs	771	dmm3	c
sc	0	dmf3	10000
wc	250	dseq3	
hzmm	107.85	dres3	1.0
is	500.00	homo3	n
PROCESSING		lb	1.00
rfl	11107.7	wtfile	
rfp	9707.1	proc	ft
th	4	fn	131072
ins	100.000	math	f
ai	cdc ph		

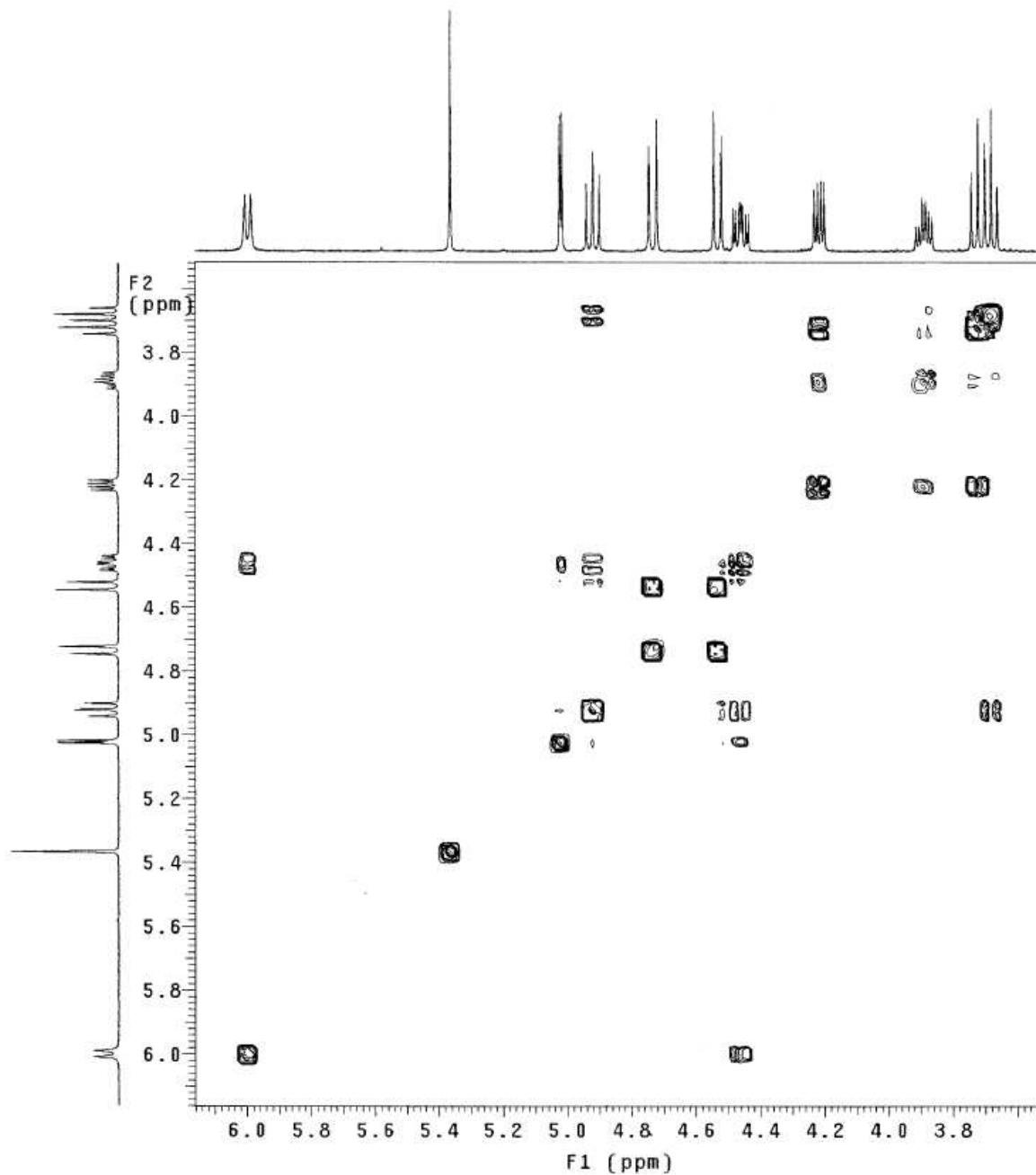


TY2-318-B

Pulse Sequence: relayh

Solvent: CDC13  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE H1, 499.8611709 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 3 hr, 23 min, 10 sec

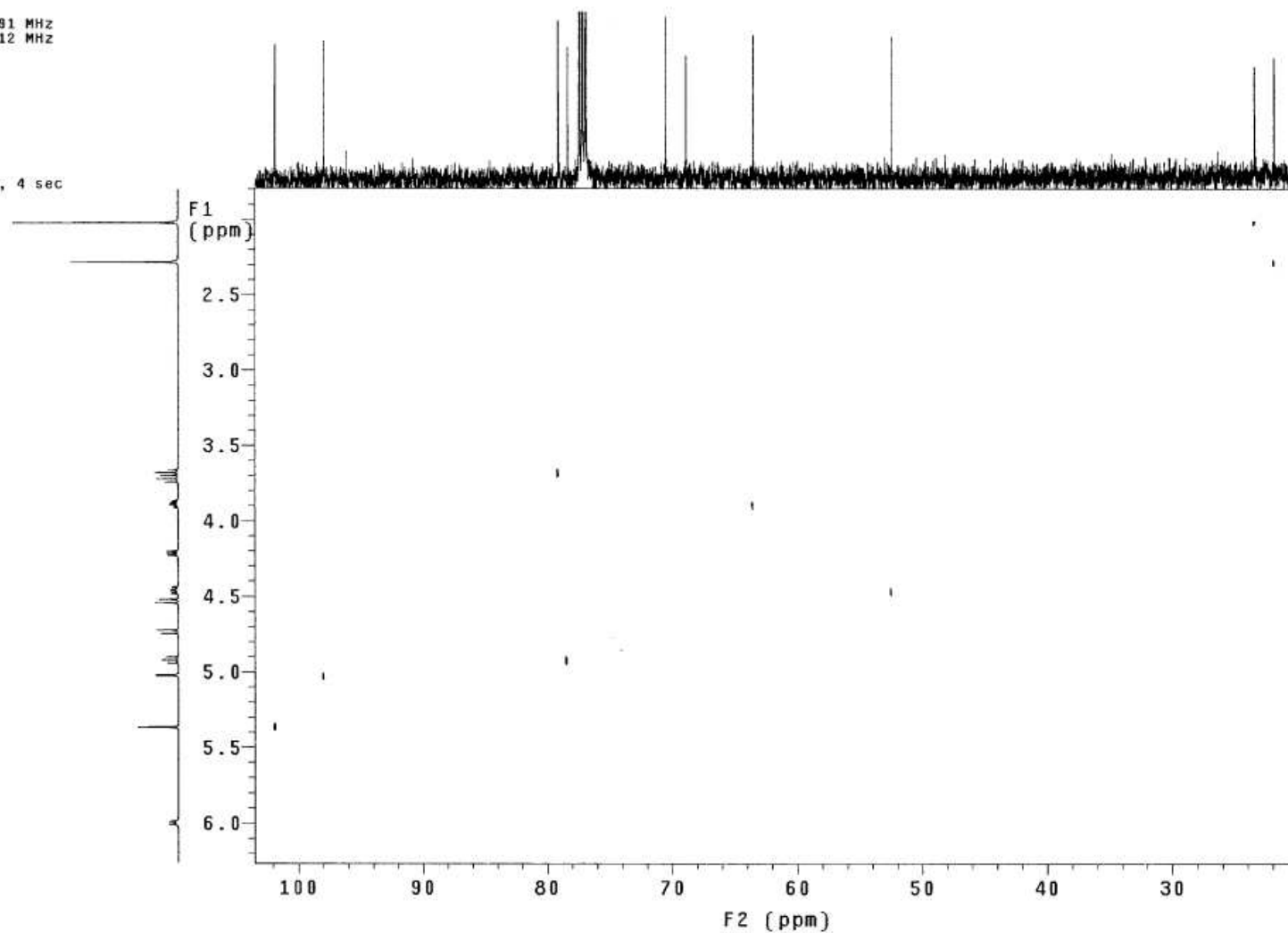


TY2-318-B

Pulse Sequence: hetcor

Solvent: CDCl<sub>3</sub>  
Ambient temperature  
User: 1-14-87  
File: TY2-318-B-CH  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
64 repetitions  
256 increments  
OBSERVE C13, 125.6901591 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 7 hr, 32 min, 4 sec



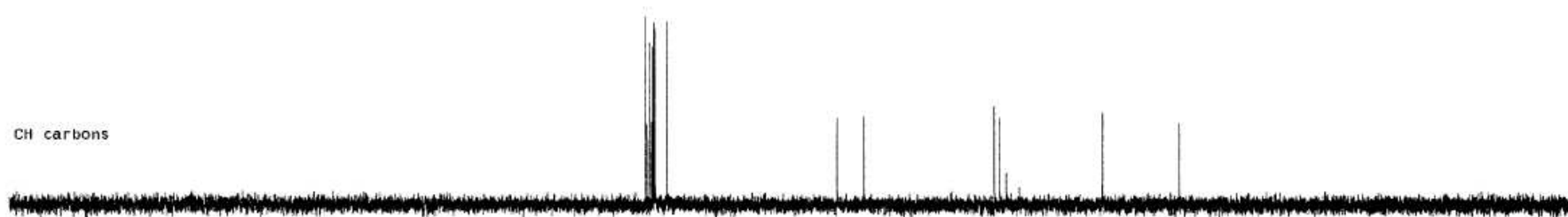
CH3 carbons



CH2 carbons



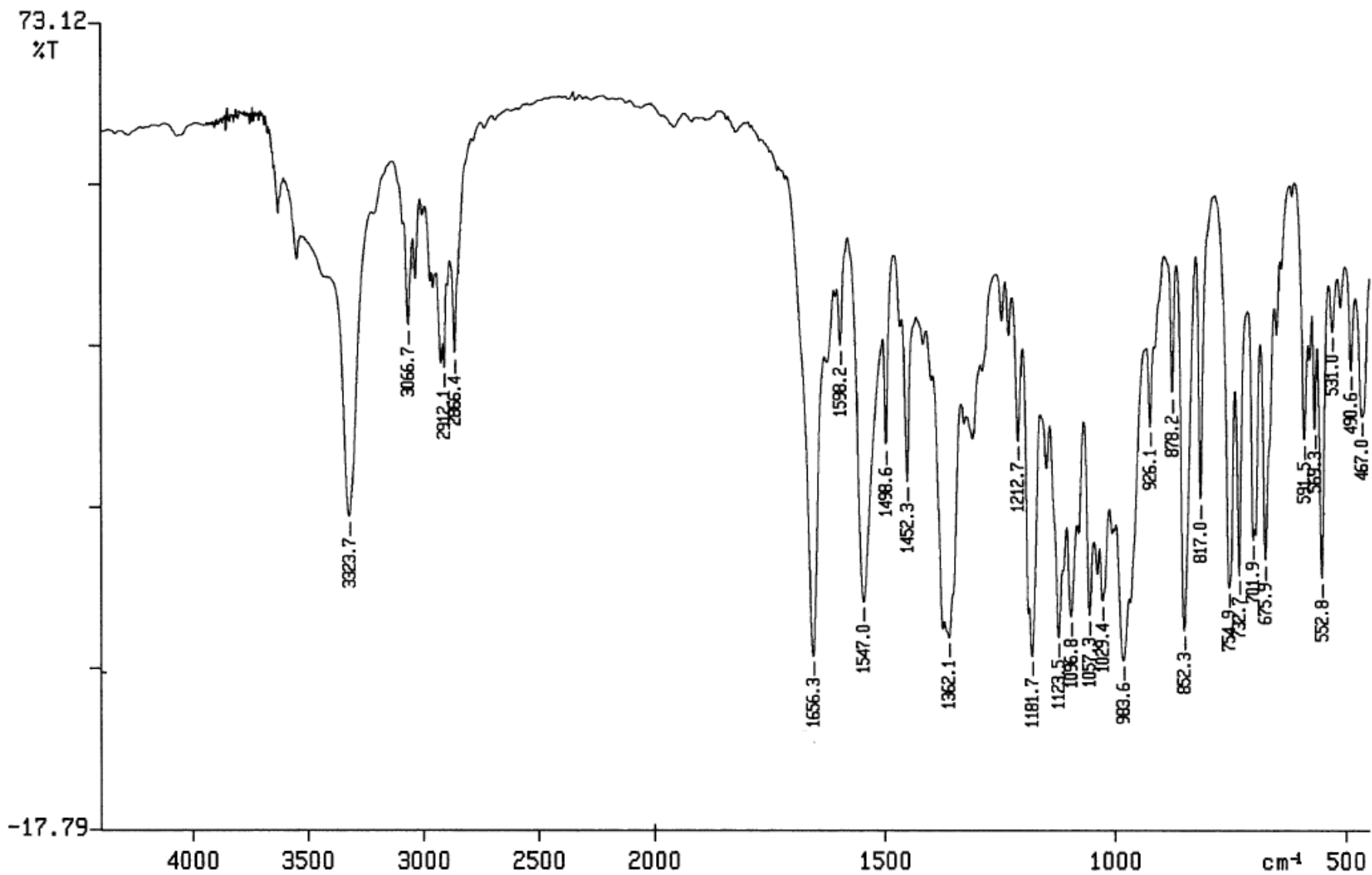
CH carbons



all protonated carbons



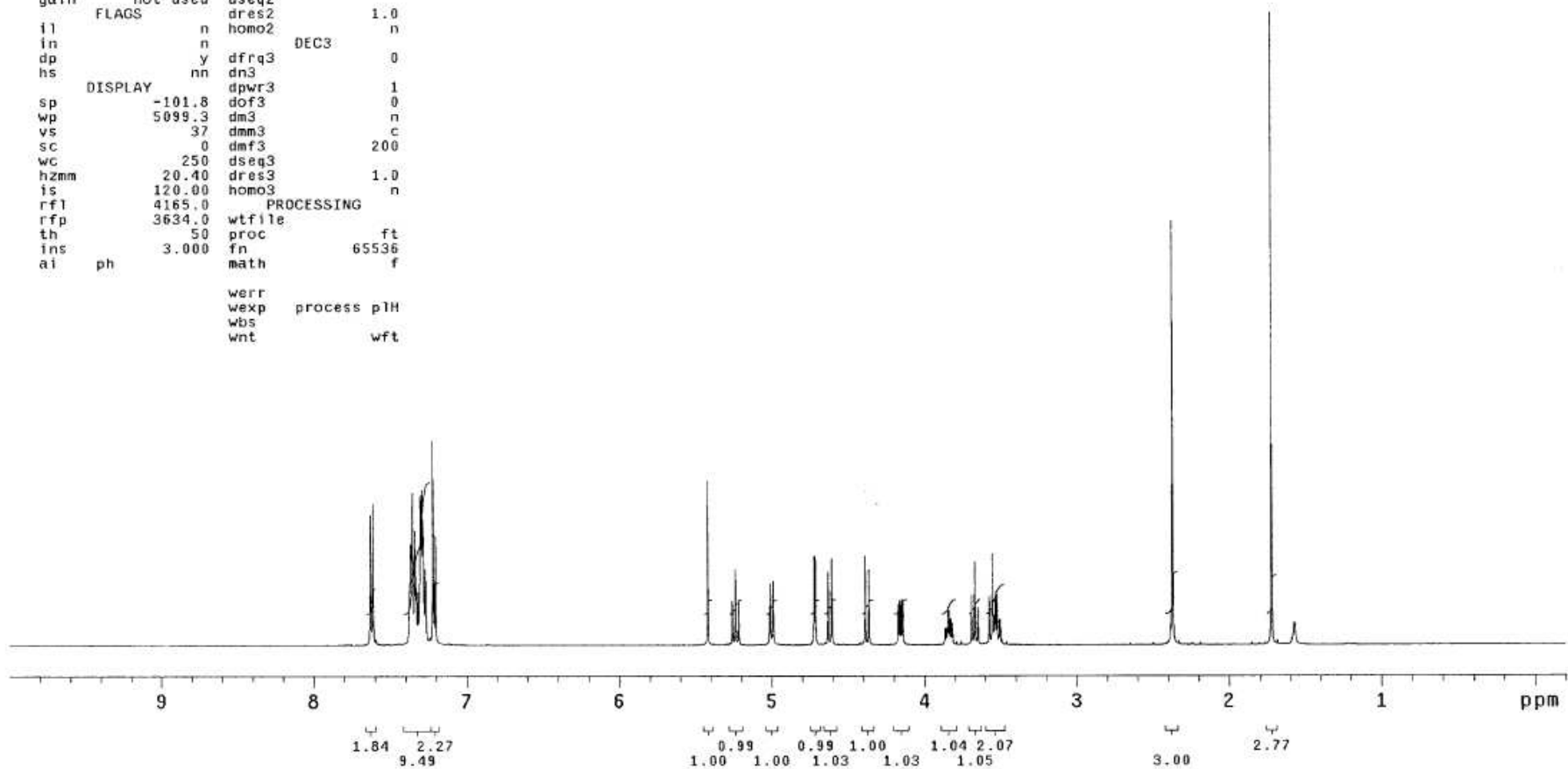
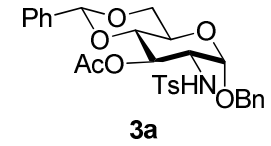
200 180 160 140 120 100 80 60 40 20 ppm



TY2-318-A

exp1 s2pu1

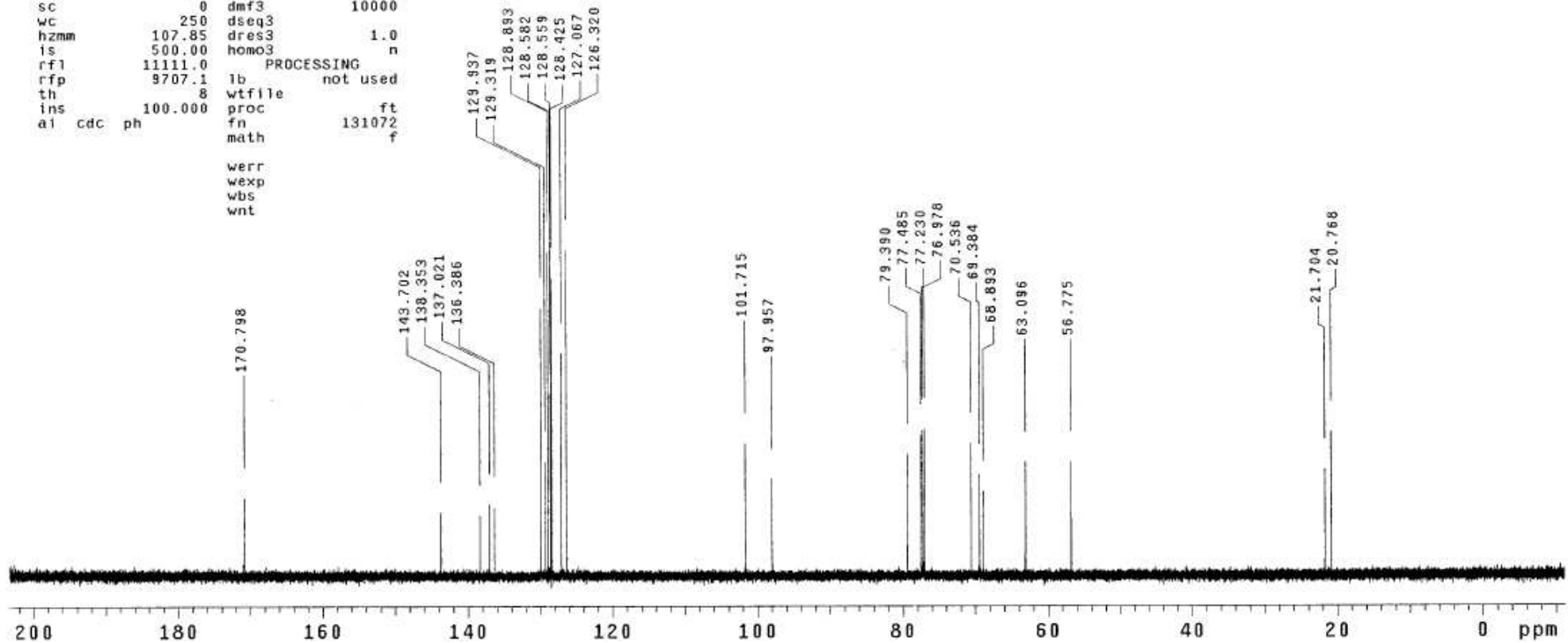
SAMPLE		DEC. & VT	
date	Mar 21 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	30
ACQUISITION			
sfrq	499.864	dm	nnn
tn	H1	dmm	c
at	5.016	dmf	200
np	65536	dseq	
sw	6533.3	dres	1.0
fb	4000	homo	n
bs	4	DEC2	
tpwr	61	dfrq2	0
pw	13.5	dn2	
d1	0.100	dpwr2	1
tof	269.9	dof2	0
nt	16	dm2	n
ct	16	dmm2	c
alock	n	dmf2	200
gain	not used	dseq2	
FLAGS			
il	n	homo2	1.0
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY			
sp	-101.8	dof3	0
wp	5099.3	dm3	n
vs	37	dmm3	c
sc	0	dmf3	200
wc	250	dseq3	
h2mm	20.40	dres3	1.0
is	120.00	homo3	n
PROCESSING			
rfl	4165.0	wfille	ft
rfp	3634.0	proc	
th	50	fn	65536
ins	3.000	math	f
ai	ph		
		werr	
		wexp	process pH
		wbs	
		wnt	wft



TY2-318-A

exp2 s2pu1

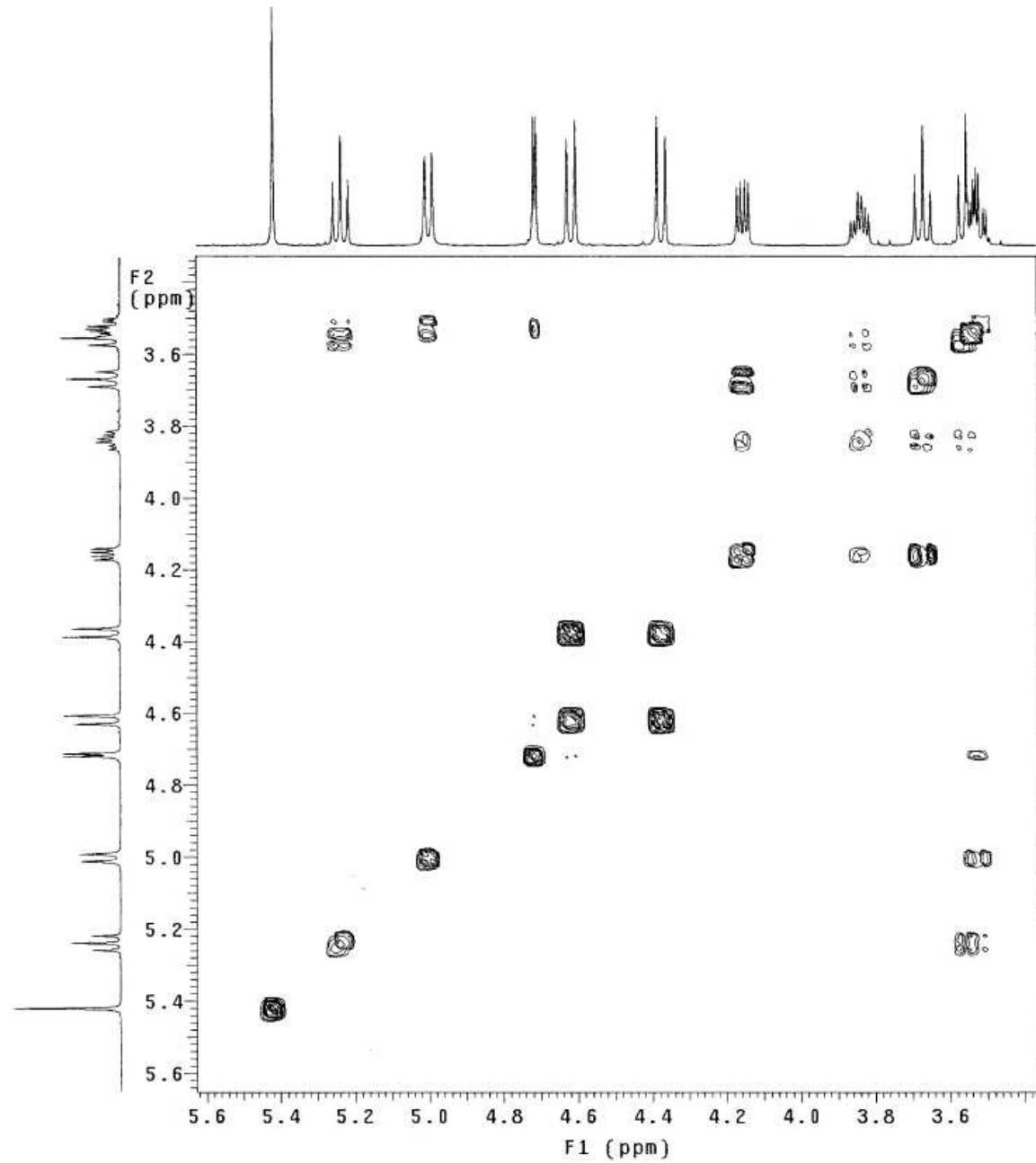
```
SAMPLE          DEC. & VT
date Mar 21 2009 dfrq      499.864
solvent CDC13      dn       H1
file      exp      dpwr     40
ACQUISITION     dof       0
sfrq      125.702  dm       yyY
tn         C13      dmm      W
at         1.215    dmf      8787.35
np         65536    dseq
sw         26963.3 dres     1.0
fb         15000    homo     n
bs         4        DEC2
tpwr       52      dfrq2    0
pw         10.2    dn2
d1         1.800   dpwr2    1
tof        144.5   dof2     0
nt         3000    dm2      n
ct         196     dmm2     c
alock      not used dmf2     10000
gain       FLAGS  dseq2    1.0
           n      dres2    1.0
           n      homo2   n
           n      DEC3
           y      dfrq3    0
           nn     dn3
           nn     dpwr3    1
DISPLAY   -1403.5  dof3     0
wp         26962.9 dm3      n
vs         105     dmm3     c
sc         0       dmf3     10000
wc         250     dseq3
hzmm       107.85  dres3    1.0
is         500.00  homo3    n
rf1        11111.0 PROCESSING
rfp         9707.1 lb       not used
th          8      wfile
ins         100.000 proc     ft
ai cdc ph      131072 fn
           math     f
```



TY2-318-A

Pulse Sequence: relayh  
Solvent: CDC13  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE H1, 499.8611956 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 3 hr, 23 min, 10 sec



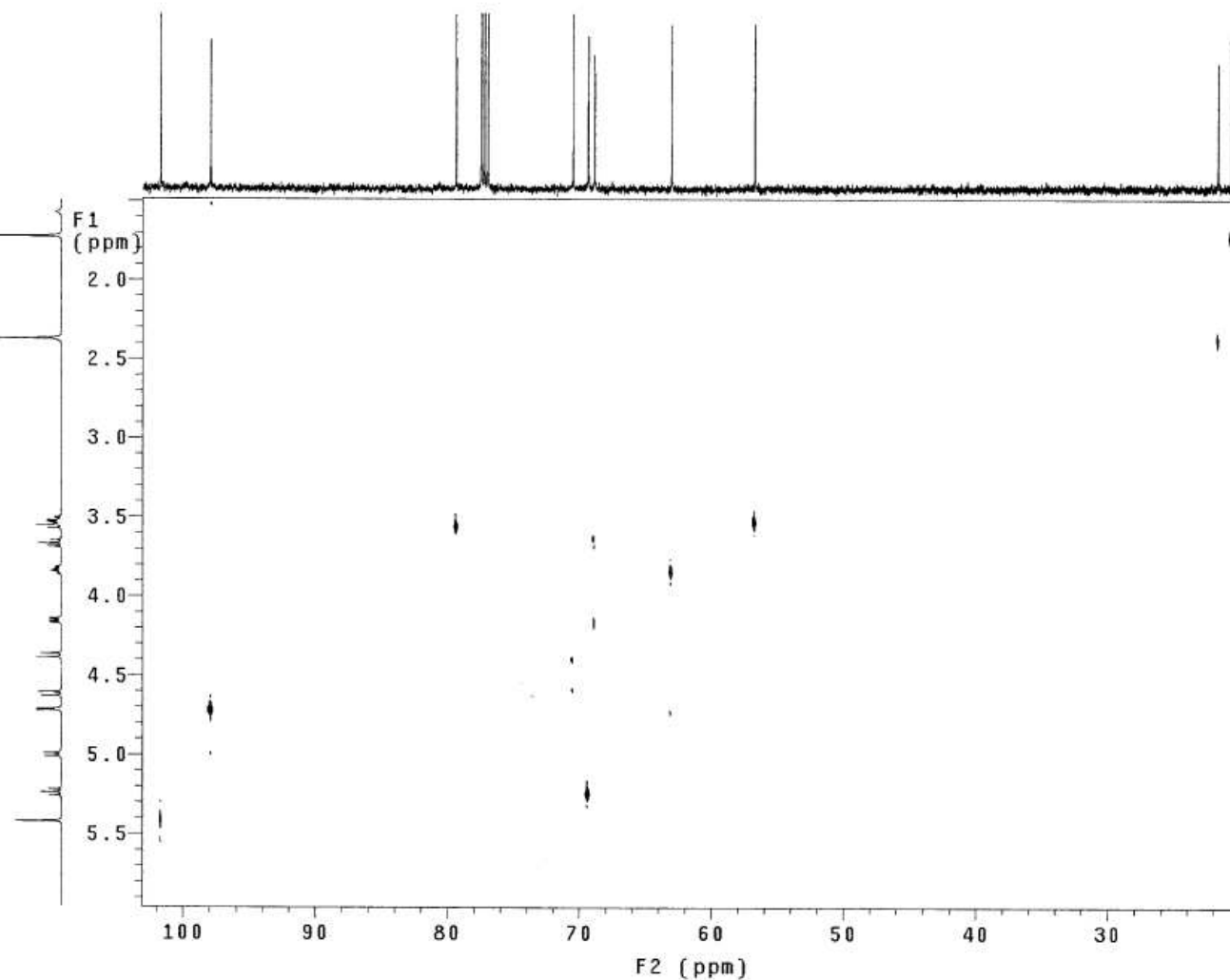


TY2-318-A

Pulse Sequence: hetcor

Solvent: CDC13  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
64 repetitions  
256 increments  
OBSERVE C13, 125.6901681 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 7 hr, 32 min, 4 sec



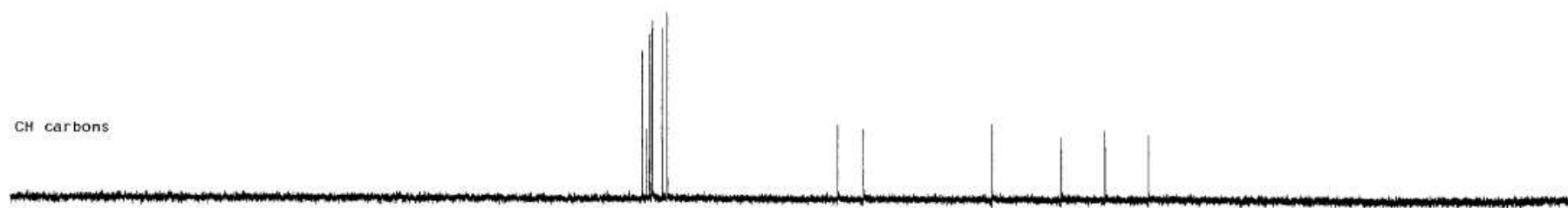
CH3 carbons



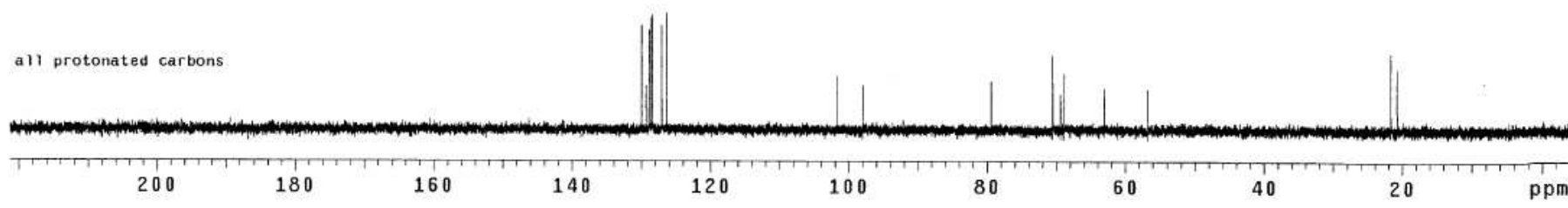
CH2 carbons



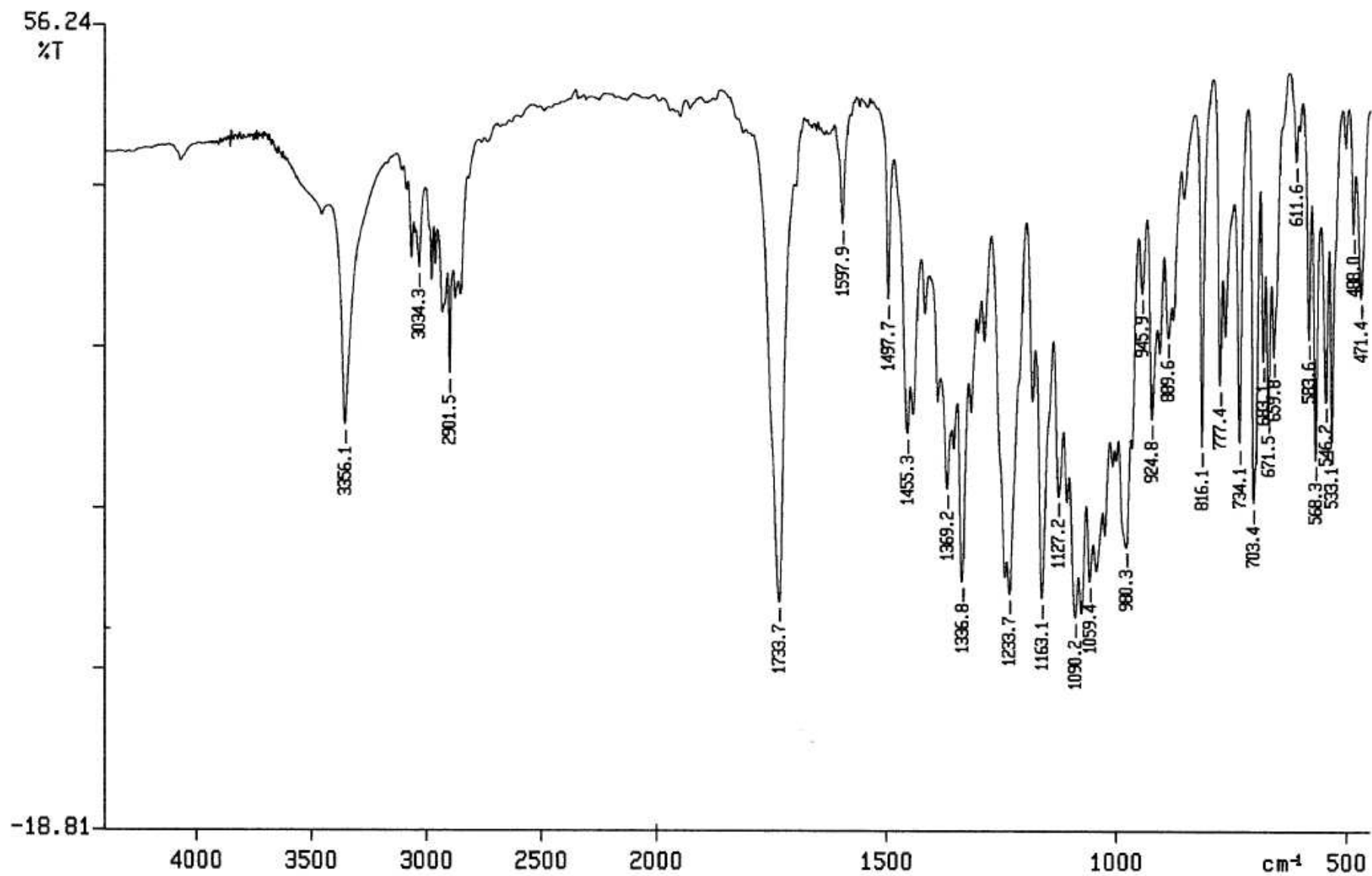
CH carbons



all protonated carbons



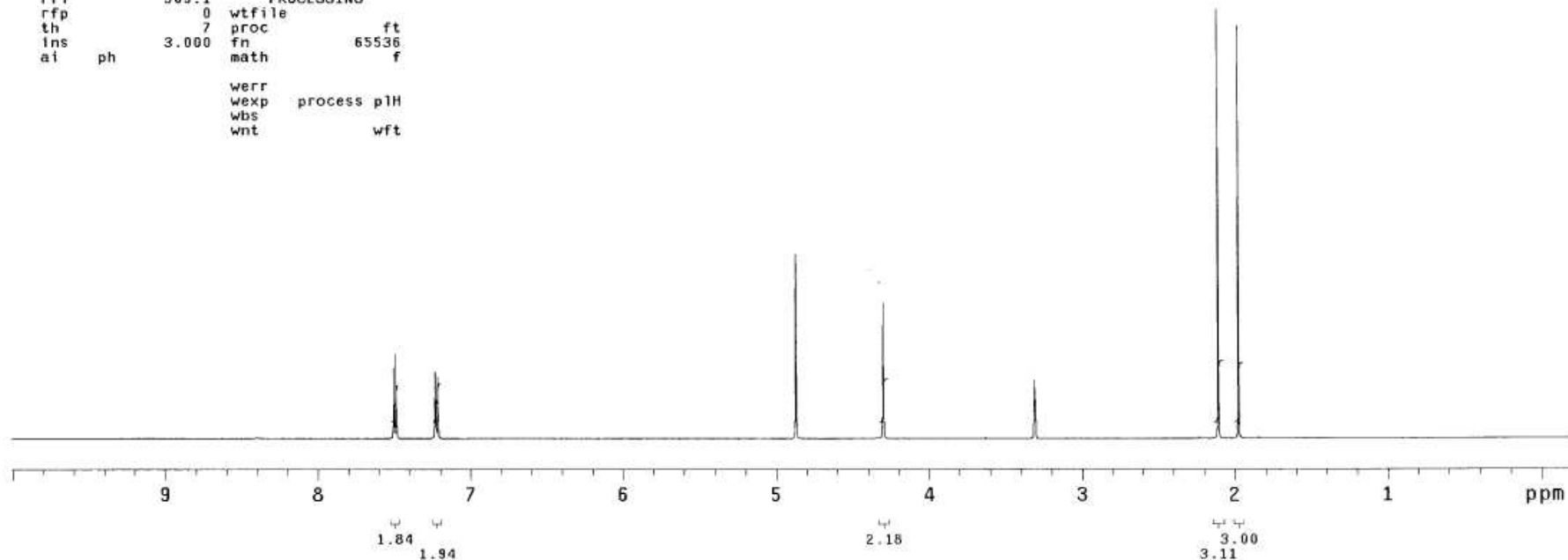
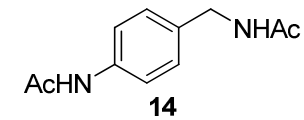
200 180 160 140 120 100 80 60 40 20 ppm



TY2-380

expl s2pu1

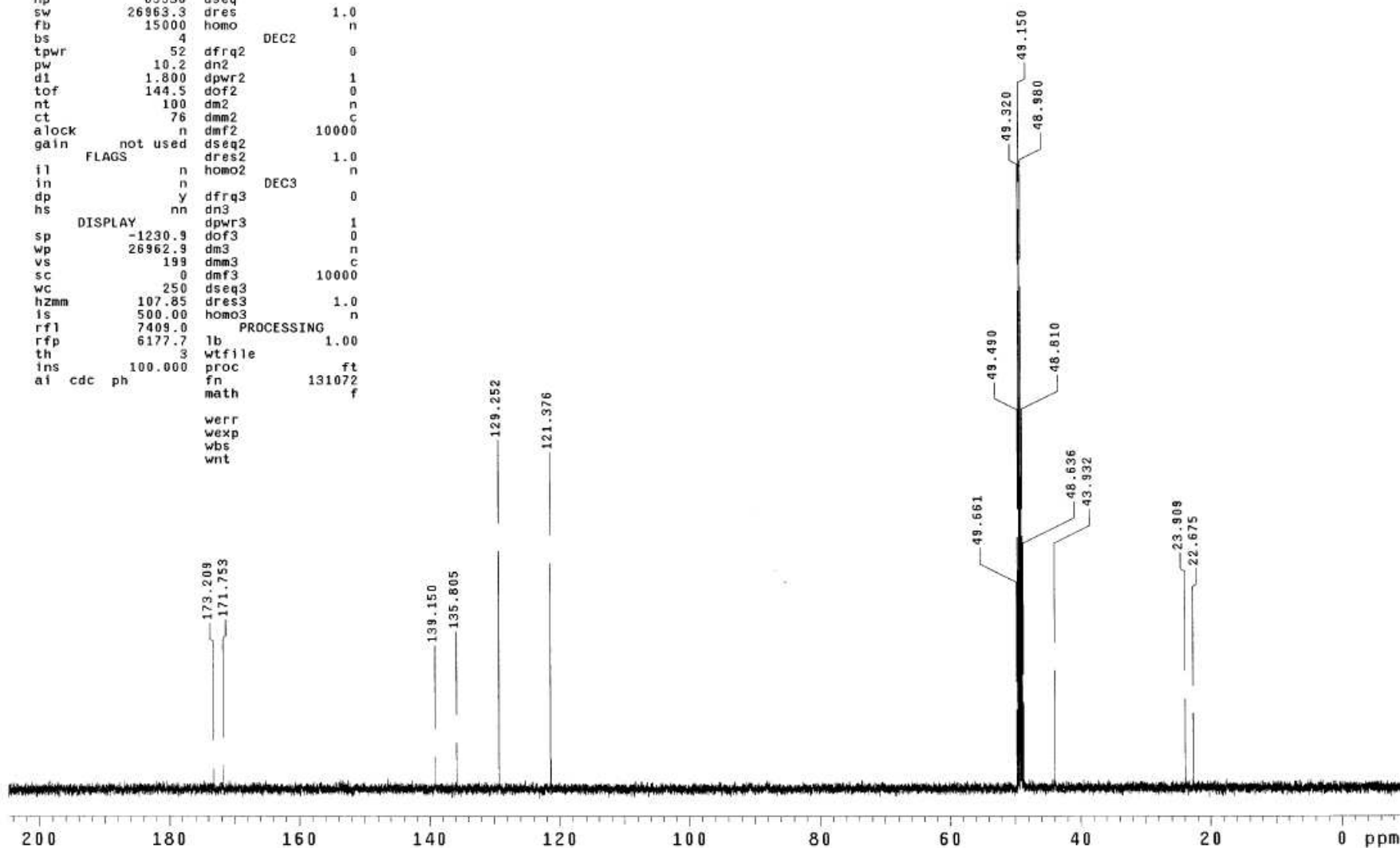
```
SAMPLE          DEC. & VT
date Jun 25 2009 dfrq          499.866
solvent CD300      dn           H1
file      exp      dpwr          30
ACQUISITION      dof           0
sfrq      499.866 dm            nnn
tn         H1      dmm           c
at         5.016  dmf           200
np         65536  dseq
sw         6533.3 dres          1.0
fb         4000  homo
bs         4      DEC2
tpwr       61     dfrq2         0
pw        13.5   dn2
dl         0.100 dpwr2         1
tof        269.9 dof2          0
nt         32    dm2            n
ct         32    dmm2           c
alock      n     dmf2           200
gain      not used dseq2
FLAGS      n     dres2          1.0
           n     homo2         n
           n     DEC3
in         n     dfrq3         0
dp         y     dn3
hs         nn    dpwr3         1
DISPLAY    dof3          0
sp        -97.0  dm3            n
wp        5099.3 dmm3           c
vs         15    dmf3           200
sc         0     dseq3
wc         250   dres3          1.0
hzmm       20.40 homo3         n
is         148.04
rf1        509.1 PROCESSING
rfp         0    wtfile
th          7    proc
ins        3.000 fn           65536
ai         ph    math          f
werr
wexp      process pH
wbs
wnt       wft
```



TY2-380

exp2 s2pu1

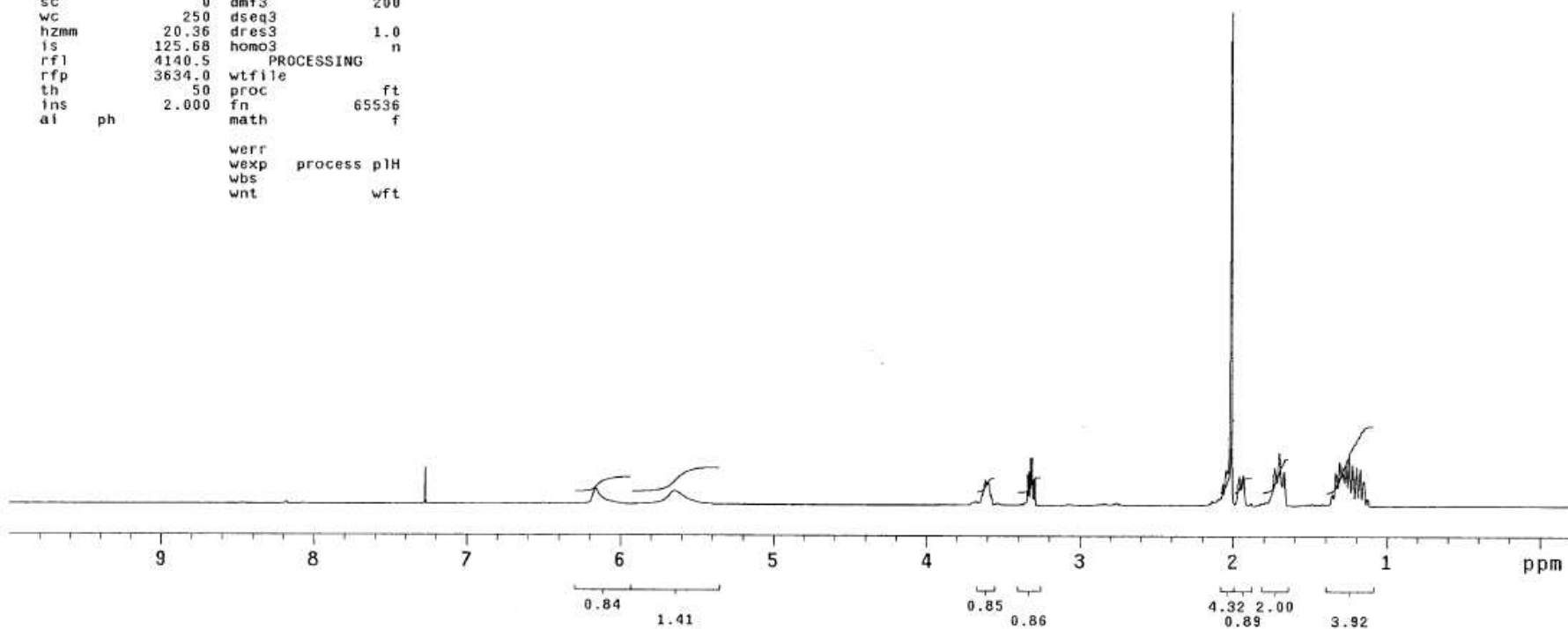
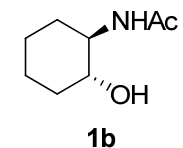
```
SAMPLE          DEC. & VT
date Jun 25 2009 dfrq 499.866
solvent cd3od   dn  H1
file      exp   dpwr 40
ACQUISITION dof  0
sfrq 125.703   dm   yy
tn C13         dmm   w
at 1.215      dmf   8787.35
np 65536      dseq
sw 26963.3    dres 1.0
fb 15000      homo n
bs 4          DEC2
tpwr 52       dfrq2 0
pw 10.2      dn2
d1 1.800     dpwr2 1
tof 144.5    dof2  0
nt 100      dm2   n
ct 76       dmm2  c
alock not used dmf2 10000
gain      dseq2 1.0
FLAGS     dres2 1.0
          homo2 n
il n       DEC3
in n
dp y       dfrq3 0
hs nn      dn3
DISPLAY   dpwr3 1
sp -1230.9 dof3  0
wp 26962.9 dm   n
vs 199     dmm3  c
sc 0       dmf3 10000
wc 250     dseq3
hzmm 107.85 dres3 1.0
is 500.00  homo3 n
rfl 7409.0 PROCESSING
rfp 6177.7 lb 1.00
th 3       wtfile
ins 100.000 proc ft
ai cdc ph  fn 131072
          math f
```



TY2-334

exp1 s2pu1

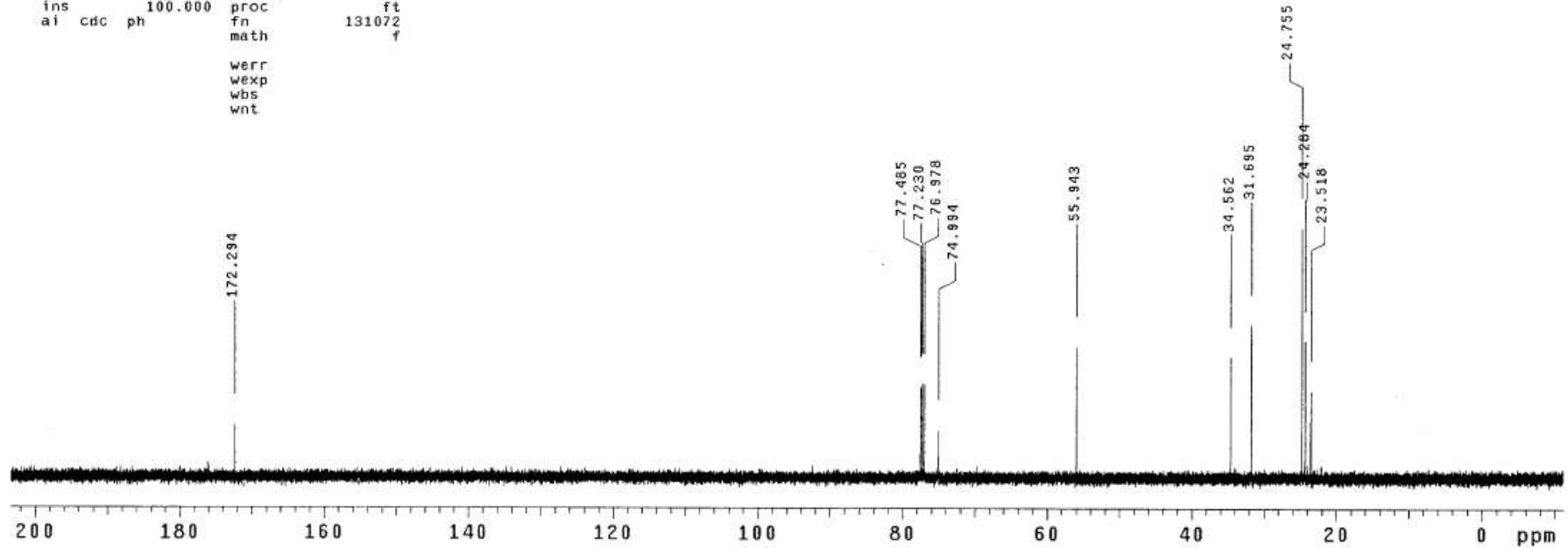
```
SAMPLE          DEC. & VT
date Apr 11 2009 dfrq      499.864
solvent CDC13     dn       H1
file exp        dpwr     30
ACQUISITION    dof      0
sfrq 499.864    dm       nnn
tn    H1        dmm      c
at    5.016     dmf     200
np    65536     dseq    1.0
sw    6533.3    dres    n
fb    4000     homo
bs    4         DEC2
tpwr  61       dfrq2   0
pw    13.5     dn2     1
d1    0.100    dpwr2   0
tof   269.9    dof2    n
nt    16      dm2     c
ct    16      dmm2    200
alock n         dmf2
gain  not used dseq2   1.0
      FLAGS    n     homo2  n
      in       n     DEC3
      dp       y     dfrq3   0
      hs       nn    dn3
      DISPLAY  dpwr3  1
      sp      -94.4  dof3    0
      wp      5090.8 dm3     n
      vs      52    dmm3    c
      sc      0     dmf3    200
      wc      250   dseq3   1.0
      hzmm    20.36 dres3   n
      is      125.68 homo3
      rfl     4140.5 PROCESSING
      rfp     3634.0 wtfile
      th      50    proc
      ins     2.000 fn      65536
      ai      ph   math
                        werr
                        wexp  process pH
                        wbs
                        wnt   wft
```



TY2-334

exp2 s2pu1

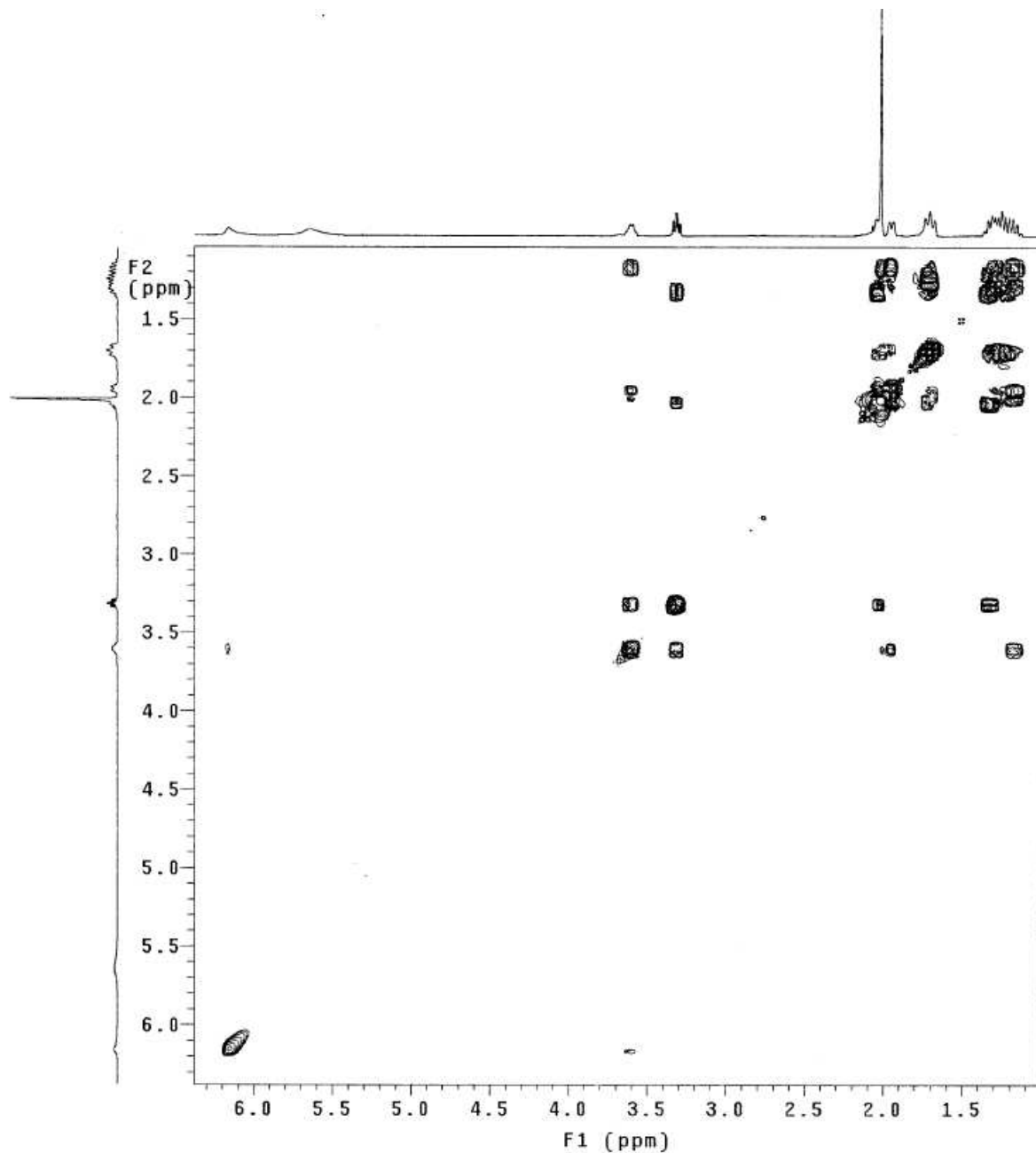
SAMPLE		DEC. & VT	
date	Apr 11 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION		dof	0
sfrq	125.702	dm	yyy
tn	C13	dmm	w
at	1.215	dmf	8787.35
np	65536	dseq	
sw	26963.3	dres	1.0
fb	15000	homo	n
bs	4	DEC2	
tpwr	52	dfrq2	0
pw	10.2	dn2	
d1	1.800	dpwr2	1
tof	144.5	dof2	0
nt	3000	dm2	n
ct	84	dmm2	c
alock	n	dmf2	10000
gain	not used	dseq2	
FLAGS		dres2	1.0
il	n	homo2	n
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY		dpwr3	1
sp	-1404.4	dof3	0
wp	26962.9	dm3	n
vs	89	dmm3	c
sc	0	dmf3	10000
wc	250	dseq3	
hzmm	107.85	dres3	1.0
is	500.00	homo3	n
rfl	11111.8	PROCESSING	
rfp	9707.1	lb	not used
th	6	wfile	
ins	100.000	proc	ft
ai	cdc ph	fn	131072
		math	f



TY2-334

Pulse Sequence: relayh  
Solvent: CDC13  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE H1, 499.8611711 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 3 hr, 23 min, 10 sec





TY2-334

Pulse Sequence: hetcor

Solvent: CDCl<sub>3</sub>

Ambient temperature

User: 1-14-87

INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec

Acq. time 0.111 sec

Width 18403.5 Hz

2D Width 6533.3 Hz

32 repetitions

256 increments

OBSERVE C13, 125.6901689 MHz

DECOUPLE H1, 499.8639312 MHz

Power 40 dB

on during acquisition

off during delay

WALTZ-16 modulated

DATA PROCESSING

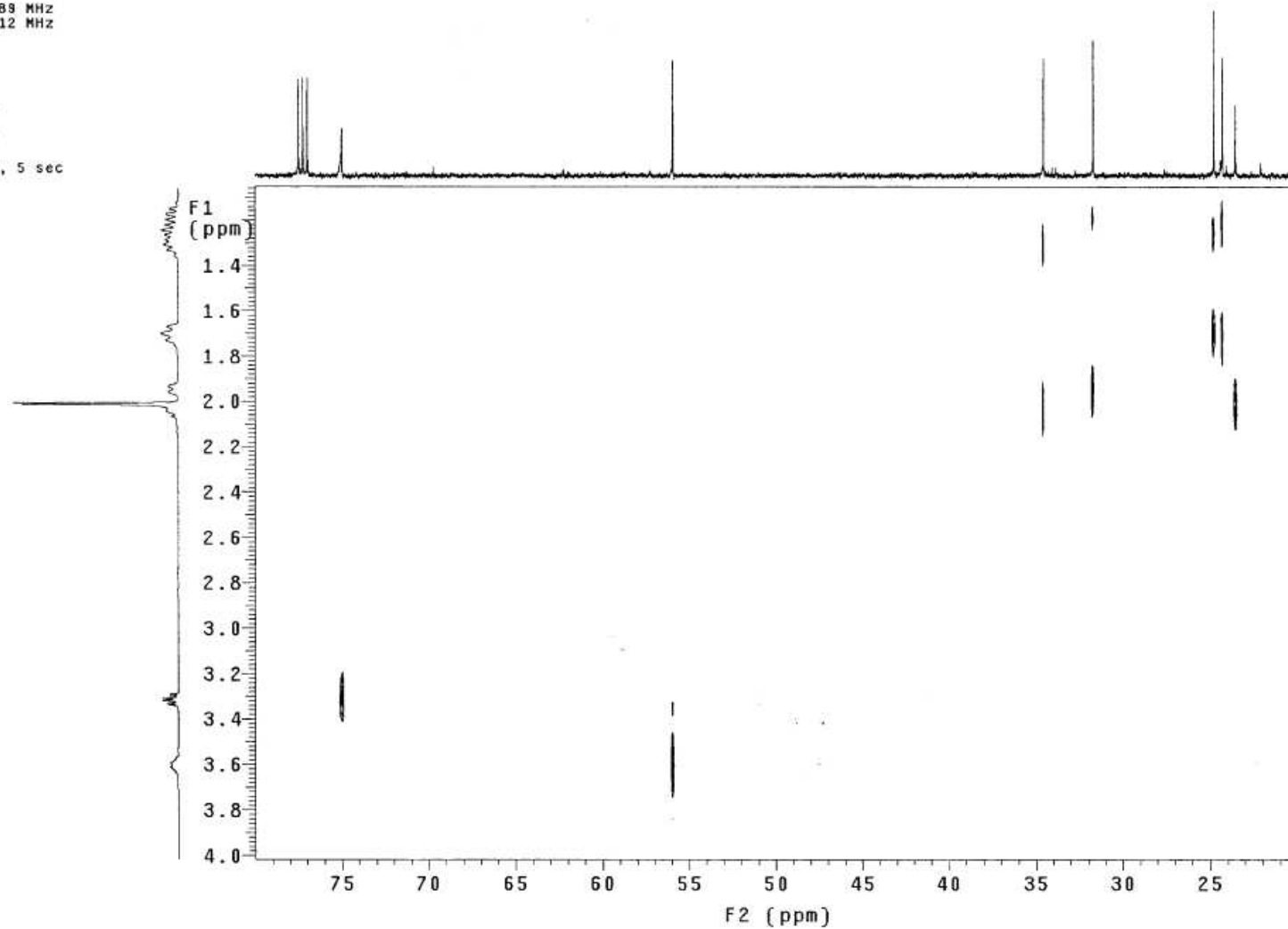
Line broadening 1.0 Hz

F1 DATA PROCESSING

Line broadening 0.3 Hz

FT size 4096 x 1024

Total time 3 hr, 46 min, 5 sec



CH3 carbons



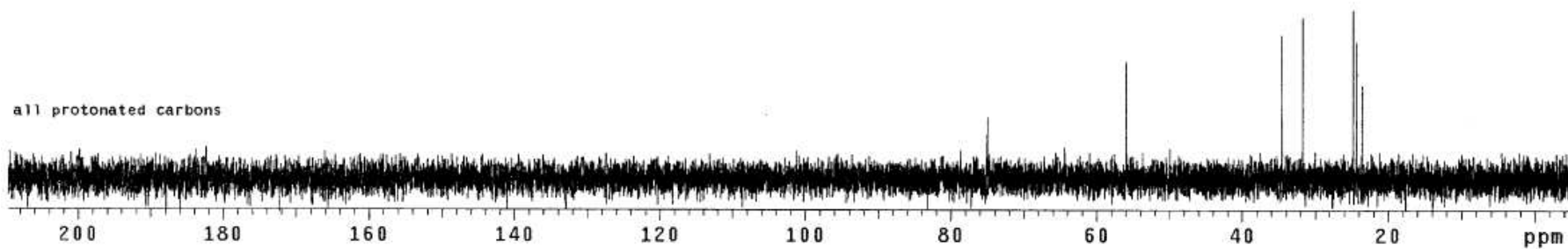
CH2 carbons

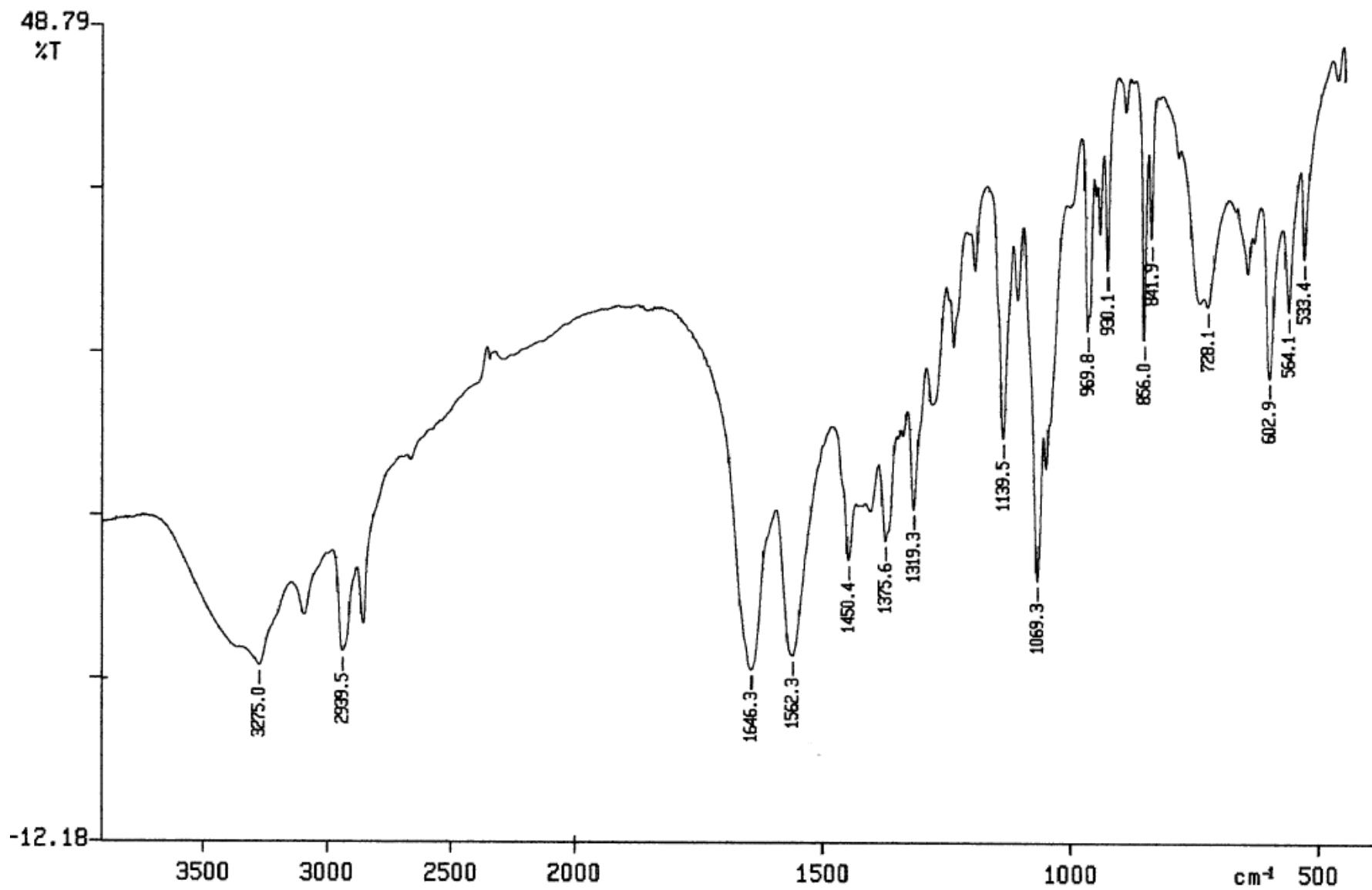


CH carbons



all protonated carbons





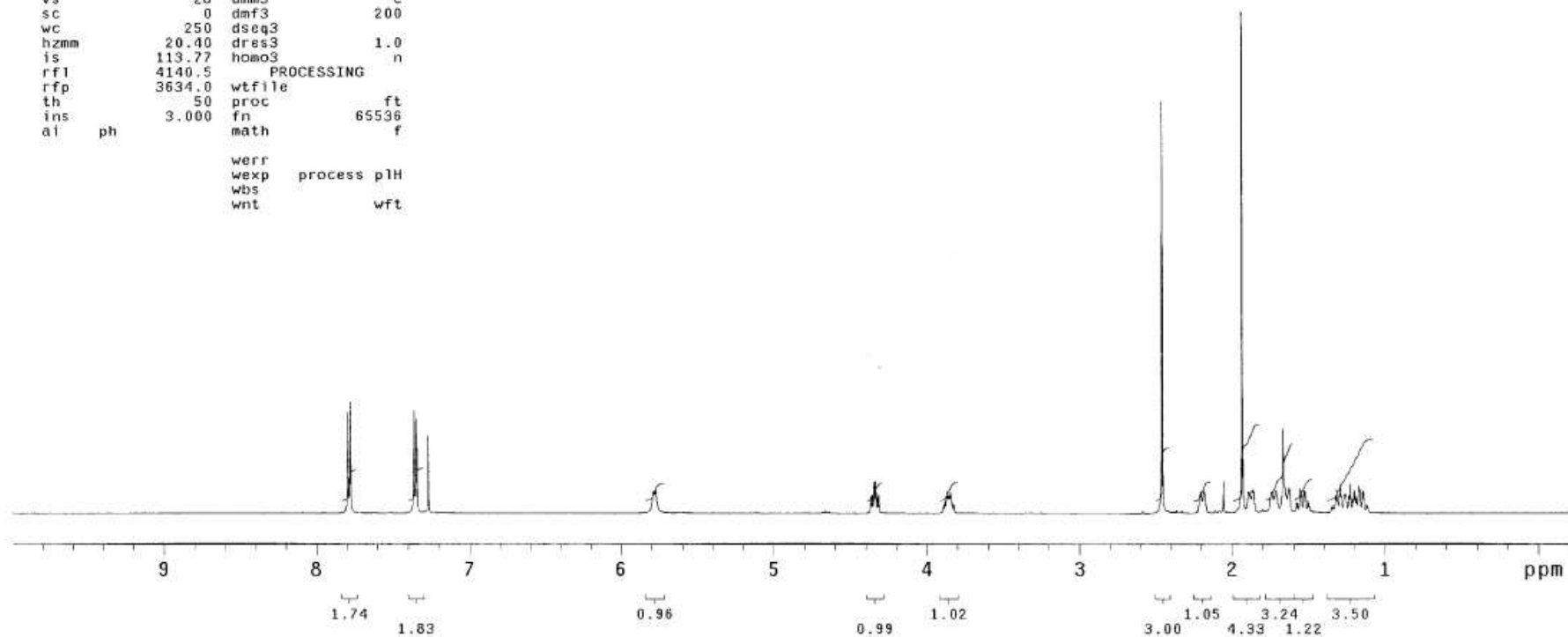
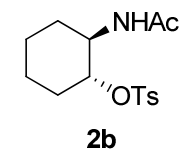
TY2-360-C-crystal

exp1 s2pu1

```

SAMPLE          DEC. & VT
date Jun 3 2009 dfrq          499.864
solvent CDC13      dn          H1
file exp         dpwr         30
ACQUISITION    dof          0
sfrq 499.864    dm           nnn
tn H1          dmm           c
at 5.016       dmf          200
np 65536      dseq           1.0
sw 6533.3     dres           n
fb 4000       homo
bs 4          DEC2
tpwr 61       dfrq2          0
pw 13.5      dn2
d1 0.100     dpwr2          1
tof 269.9    dof2           0
nt 32       dm2           n
ct 32       dmm2           c
alock n       dmf2          200
gain not used dseq2           1.0
FLAGS        dres2           n
il n         homo2
in n         DEC3
dp y         dfrq3          0
hs nn        dn3
DISPLAY      dpwr3          1
sp -103.0    dof3           0
wp 5099.3   dm3           n
vs 28      dmm3           c
sc 0       dmf3          200
wc 250     dseq3           1.0
hzmm 20.40 dres3           n
is 113.77 homo3
rfl 4140.5 PROCESSING
rfp 3634.0 wtf file
th 50      proc
ins 3.000  fn          65536
al ph      math          f
werr
wexp process pH
wbs
wnt wft

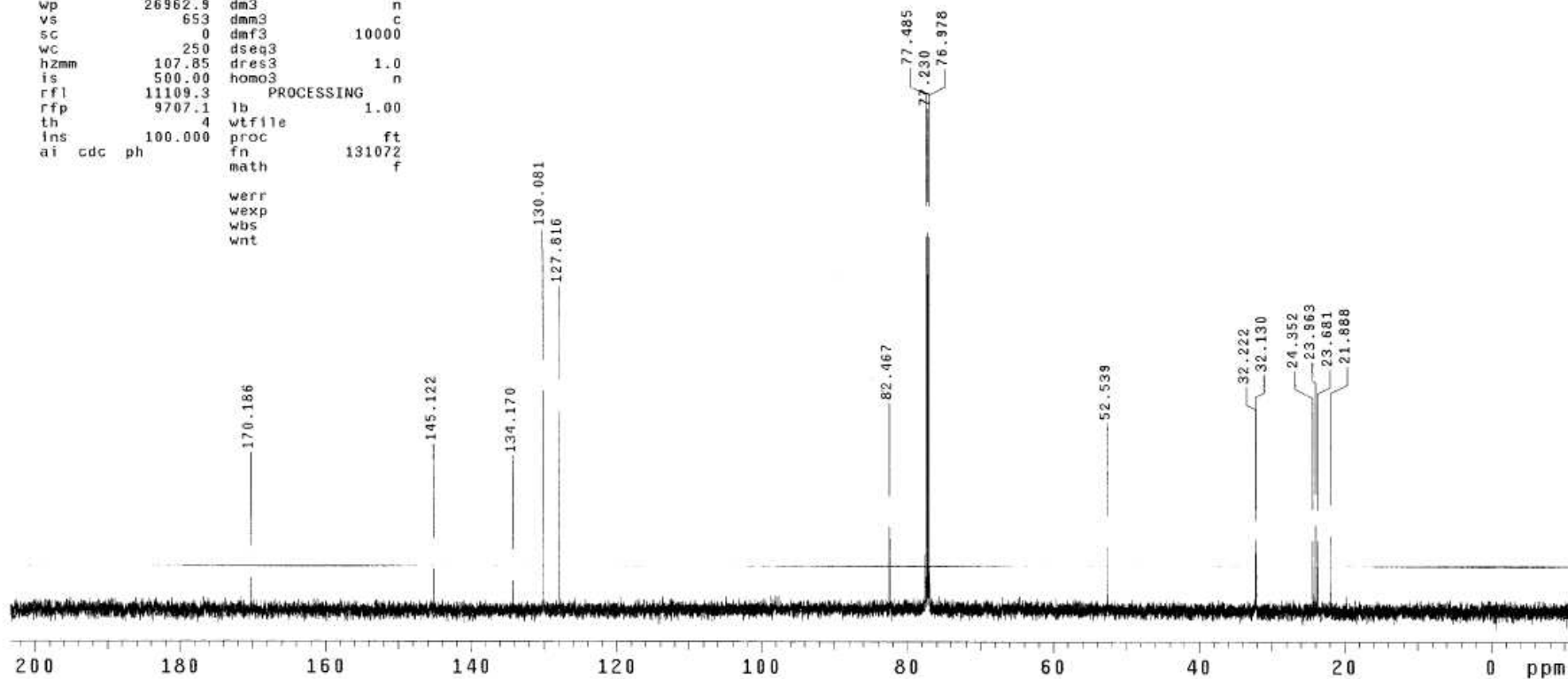
```



TY2-360-C-crystal

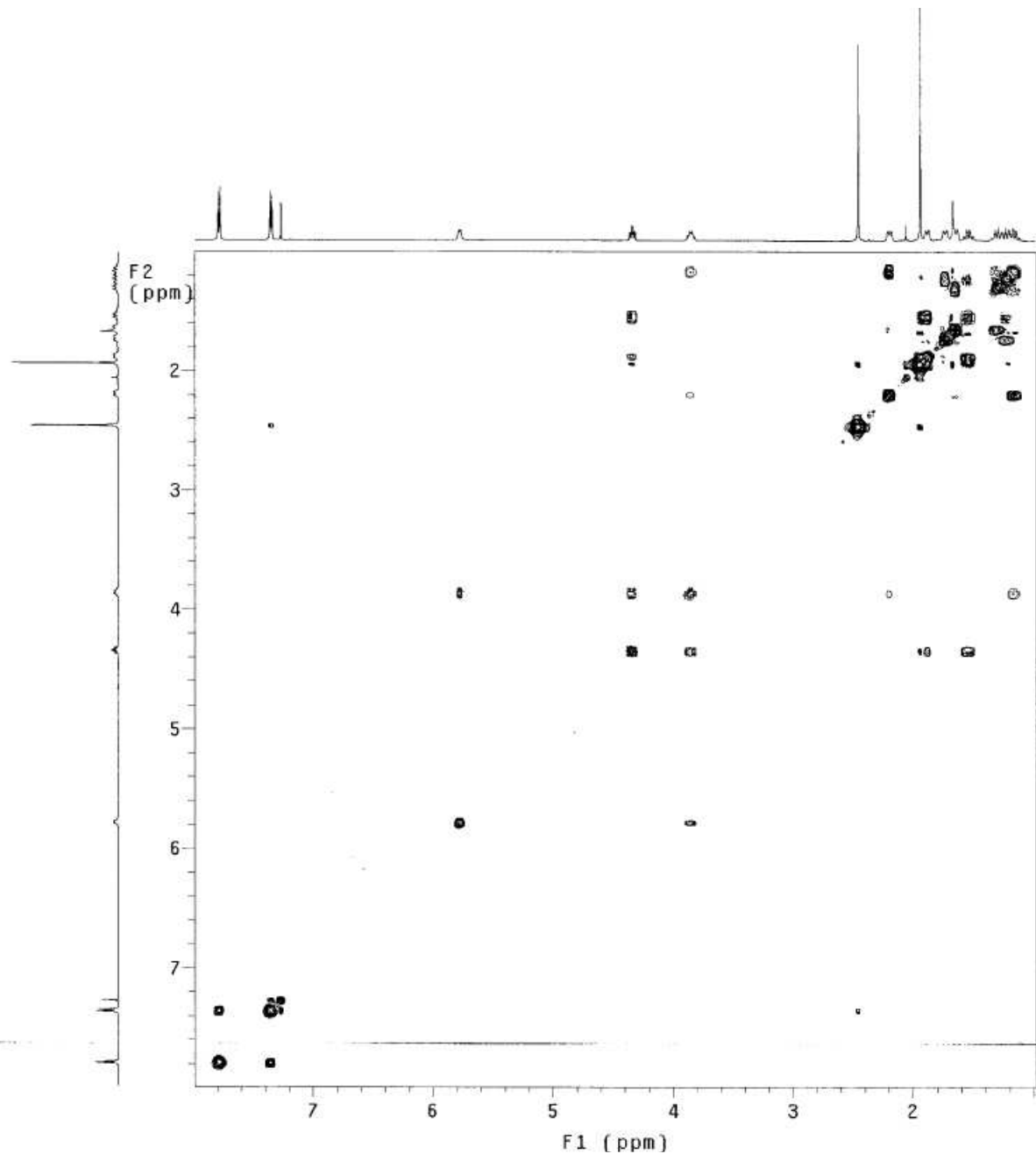
exp2 s2pu1

SAMPLE		DEC. & VT	
date	Jun 3 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION		dof	0
sfrq	125.702	dm	yy
tn	C13	dmm	w
at	1.215	dmf	8787.35
np	65536	dseq	
sw	26963.3	dres	1.0
fb	15000	homo	n
bs	4	DEC2	
tpwr	52	dfrq2	0
pw	10.2	dn2	
d1	1.800	dpwr2	1
tof	144.5	dof2	0
nt	3000	dm2	n
ct	278	dmm2	c
alock	not used	dmf2	10000
gain		dseq2	
FLAGS		dres2	1.0
il	n	homo2	n
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY		dpwr3	1
sp	-1401.9	dof3	0
wp	26962.9	dm3	n
vs	653	dmm3	c
sc	0	dmf3	10000
wc	250	dseq3	
hzmm	107.85	dres3	1.0
is	500.00	homo3	n
rfl	11109.3	PROCESSING	
rff	9707.1	lb	1.00
th	4	wtfile	
ins	100.000	proc	ft
ai	cdc ph	fn	131072
		math	f



TY2-360-C-crystal1  
Pulse Sequence: relayh  
Solvent: CDC13  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE H1, 499.8611711 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 3 hr, 23 min, 10 sec

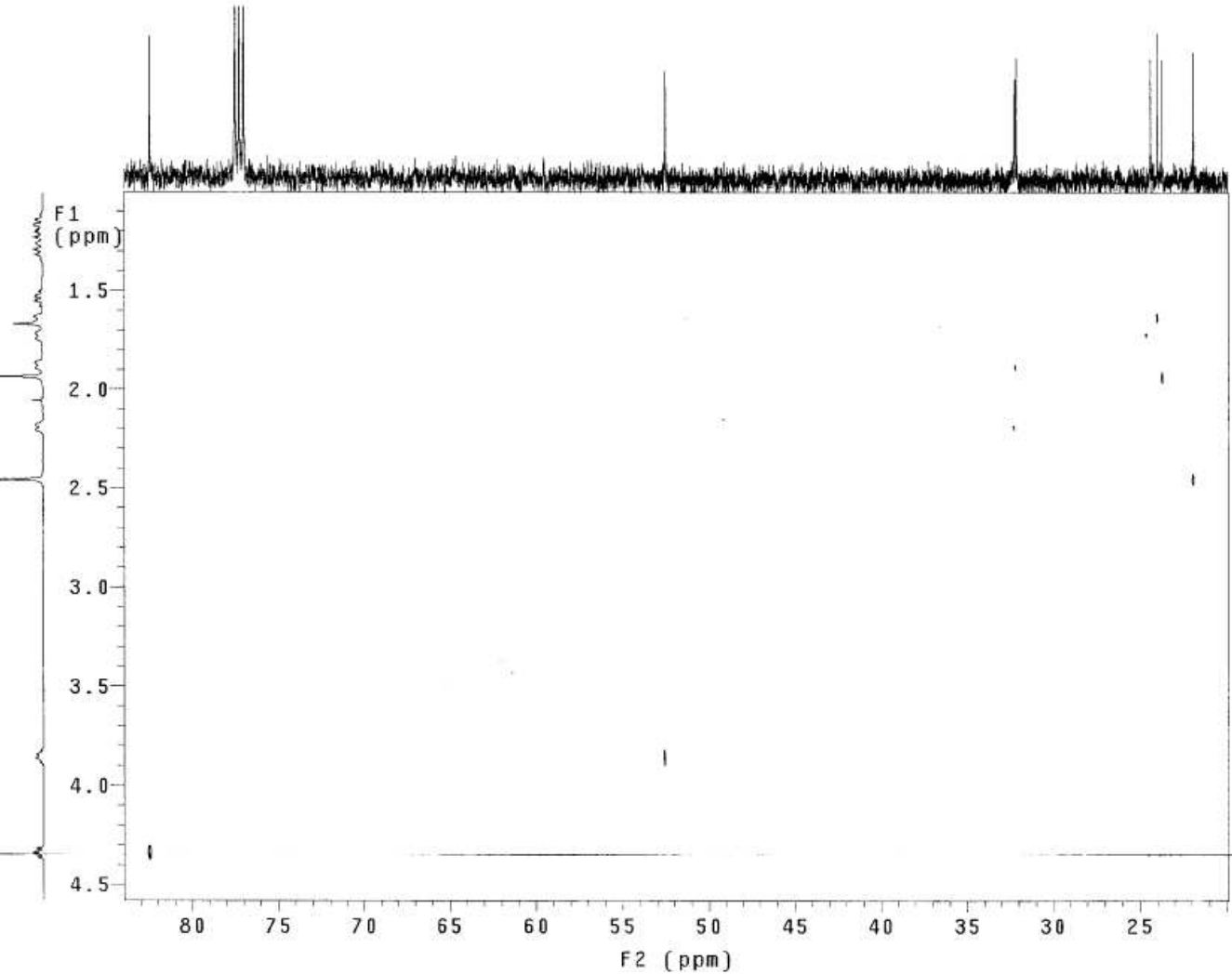


TY2-360-C-crystal

Pulse Sequence: hetcor

Solvent: CDC13  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE C13, 125.6901665 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 3 hr, 46 min, 5 sec



CH3 carbons



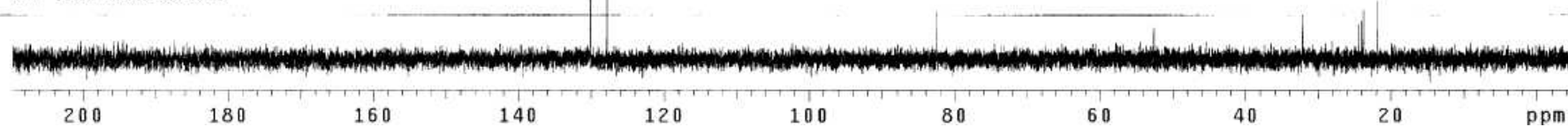
CH2 carbons



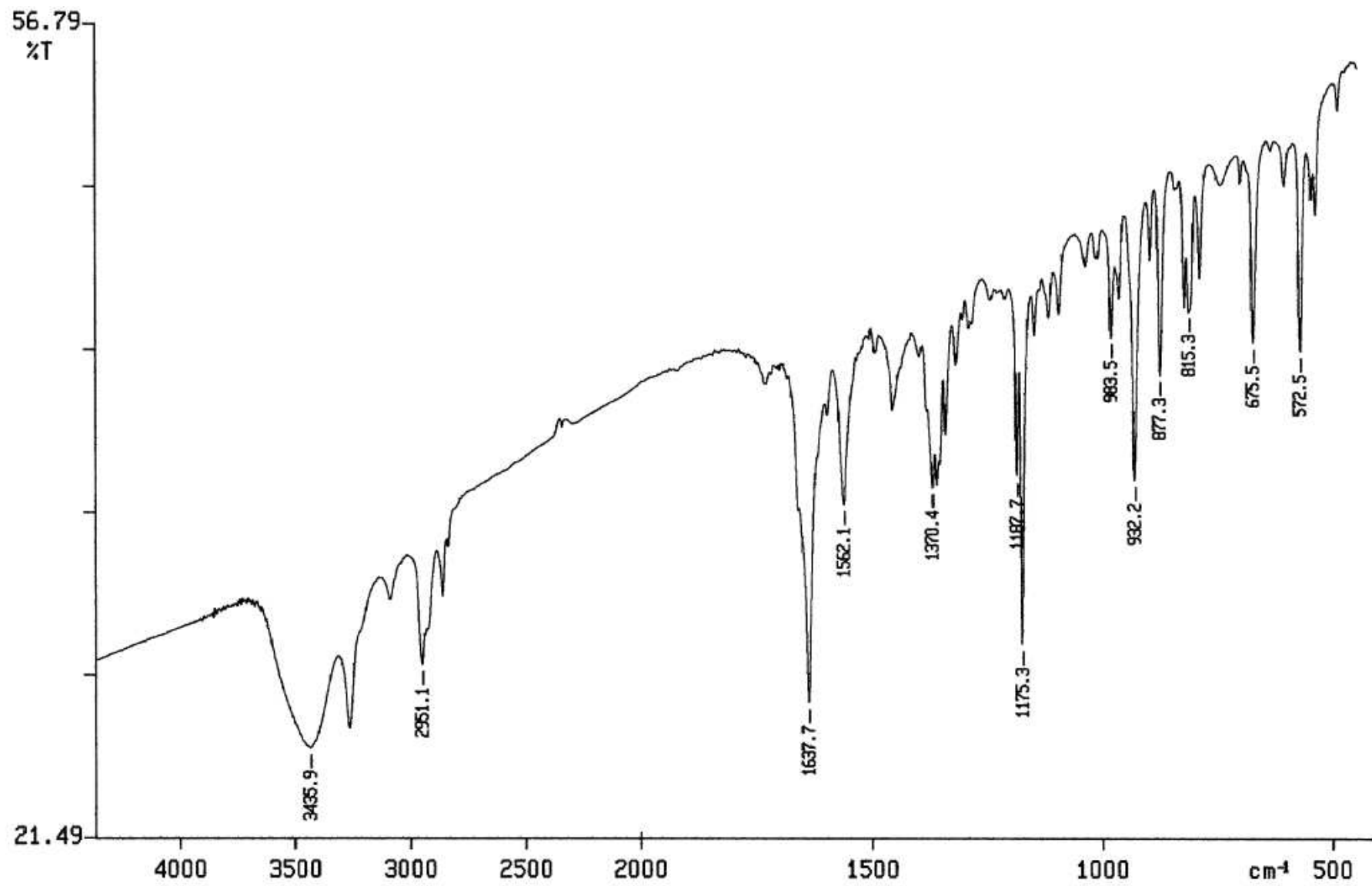
CH carbons



all protonated carbons



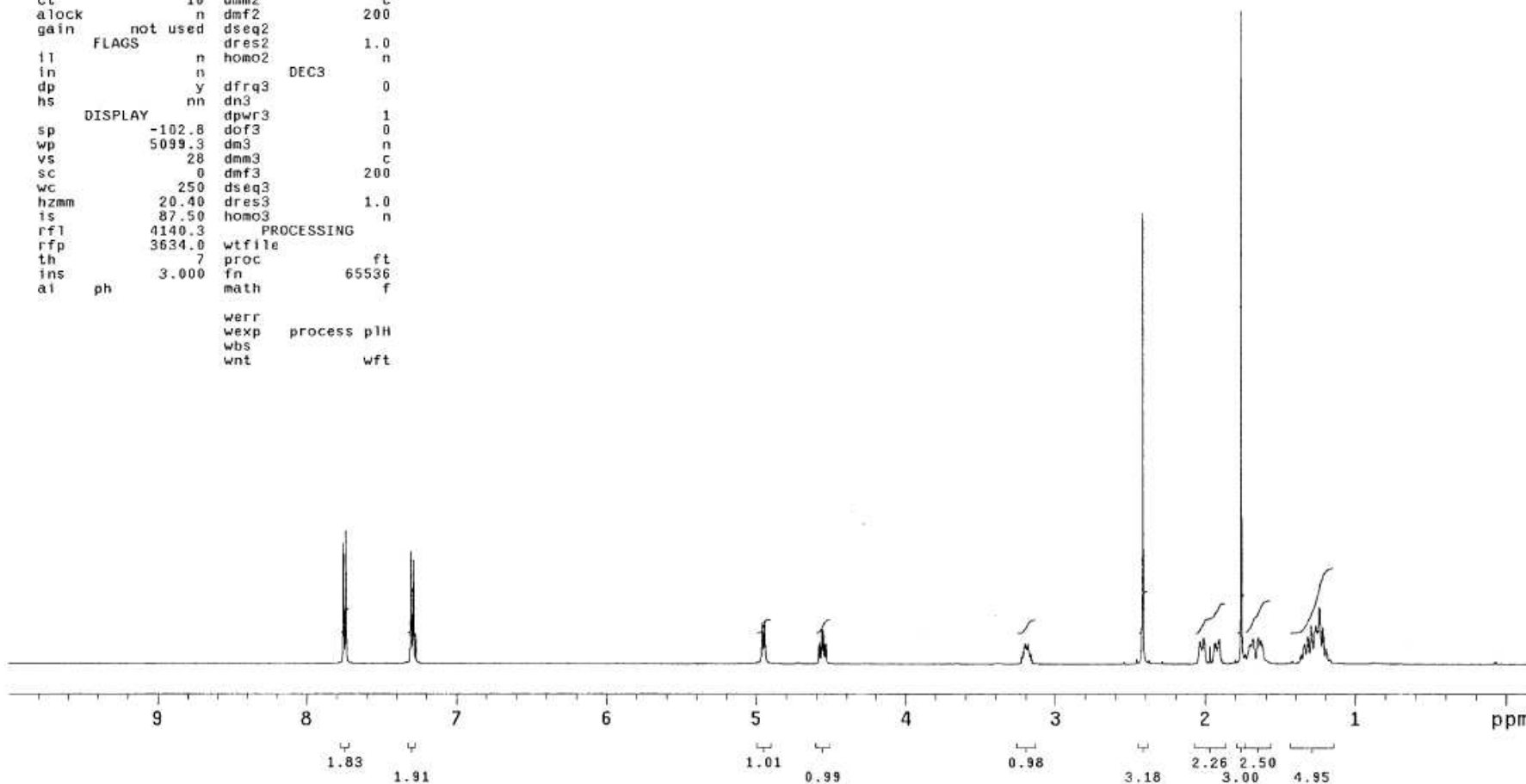
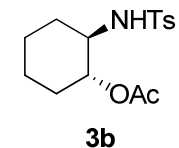




TY2-337-A

exp1 s2pu1

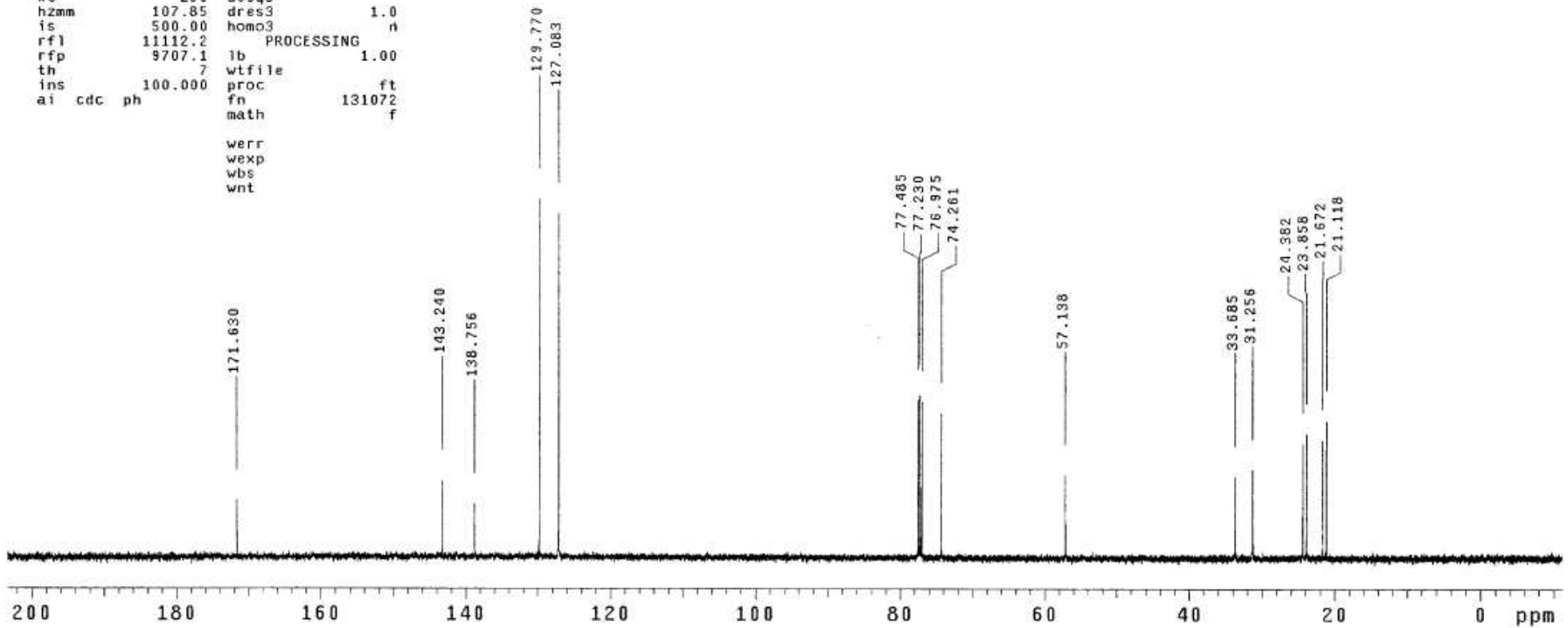
```
SAMPLE          DEC. & VT
date Apr 13 2009 dfrq      499.864
solvent CDC13      dn       H1
file      exp      dpwr     30
ACQUISITION     dof       0
sfrq      499.864  dm       nnn
tn         H1      dmm       c
at         5.016   dmf       200
np         65536   dseq
sw         6533.3 dres     1.0
fb         4000   homo
bs         4       DEC2
tpwr       61     dfrq2    0
pw         13.5   dn2
d1         0.100 dpwr2    1
tof        269.9 dof2     0
nt         16     dm2       n
ct         16     dmm2     c
alock      n     dmf2     200
gain       not used dseq2
FLAGS      n     homo2    1.0
           n     DEC3
           y     dfrq3    0
           nn    dn3
DISPLAY    dpwr3    1
sp         -102.8 dof3     0
wp         5099.3 dm3       n
vs         28     dmm3     c
sc         0     dmf3     200
wc         250   dseq3
hzmm       20.40 dres3    1.0
is         87.50 homo3    n
rf1        4140.3 PROCESSING
rfp        3634.0 wtfile
th         7     proc
ins        3.000 fn       65536
al         ph    math     f
                    werr
                    wexp   process plH
                    wbs
                    wnt     wft
```



TY2-337-A

exp2 s2pu1

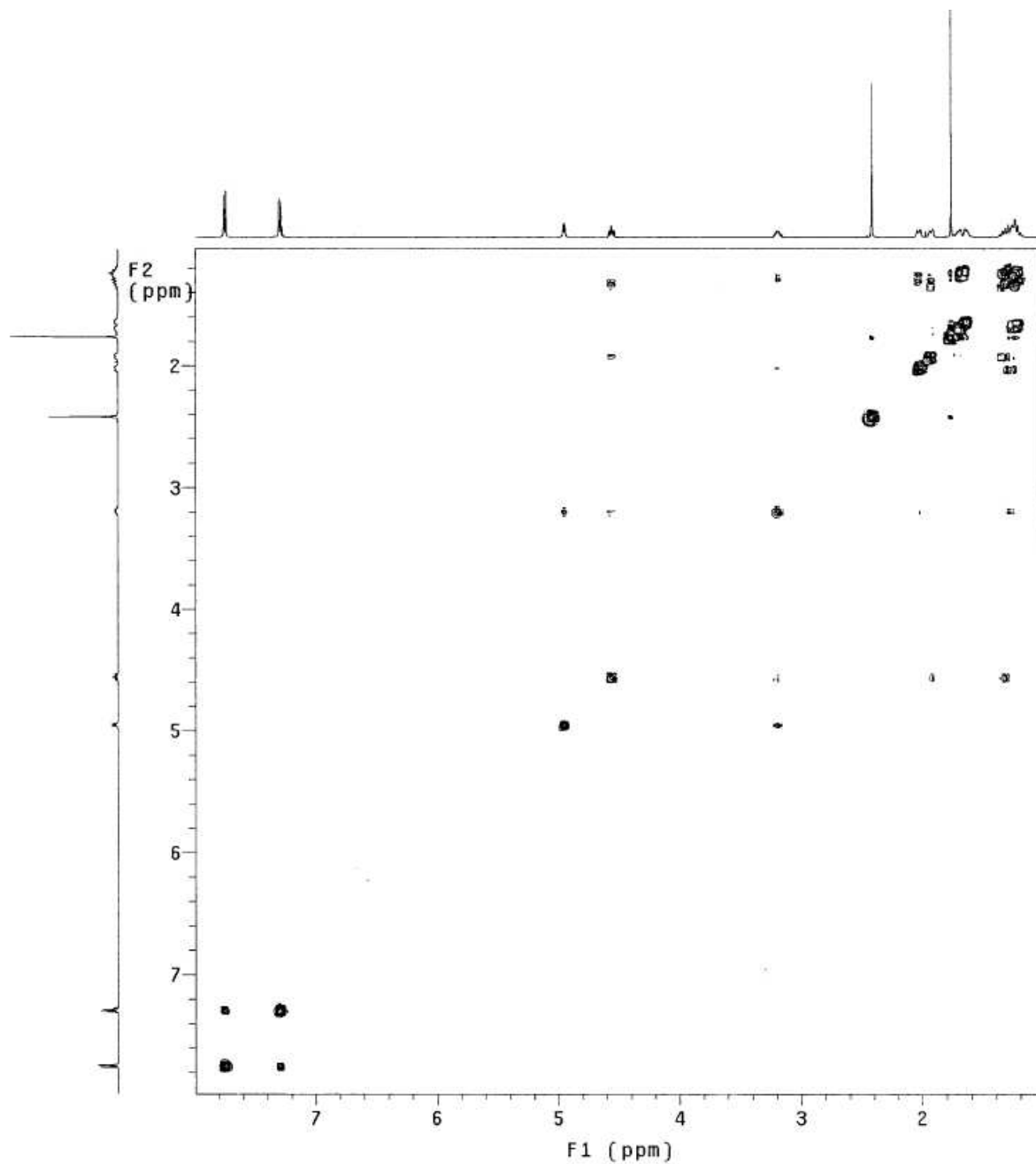
```
SAMPLE          DEC. & VT
date Apr 13 2009 dfrq 499.864
solvent CDC13    dn      H1
file  exp      dpwr    40
ACQUISITION     dof      0
sfrq 125.702   dm       yyy
tn    C13      dmm      w
at    1.215    dmf      8787.35
np    65536    dseq
sw    26963.3 dres      1.0
fb    15000    homo
bs    4        DEC2
tpwr  52      dfrq2   0
pw    10.2    dn2
d1    1.800   dpwr2   1
tof   144.5   dof2    0
nt    3000    dm2     n
ct    99     dmm2    c
alock n        dmf2    10000
gain  not used dseq2
FLAGS          dres2   1.0
il            n        homo2  n
ln            n        DEC3
dp            y        dfrq3  0
hs            nn       dn3
DISPLAY       dpwr3   1
sp            -1404.8  dof3   0
wp            26962.9 dm3     n
vs            168    dmm3    c
sc            0      dmf3    10000
wc            250    dseq3
hzmm         107.85 dres3   1.0
is           500.00 homo3   n
rf1          11112.2 PROCESSING
rfp          9707.1  lb      1.00
th           7      wtfile
ins          100.000 proc    ft
ai cdc ph     math    fn      131072
                               f
                               werr
                               wexp
                               wbs
                               wnt
```



TY2-337-A

Pulse Sequence: relayh  
Solvent: CDC13  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

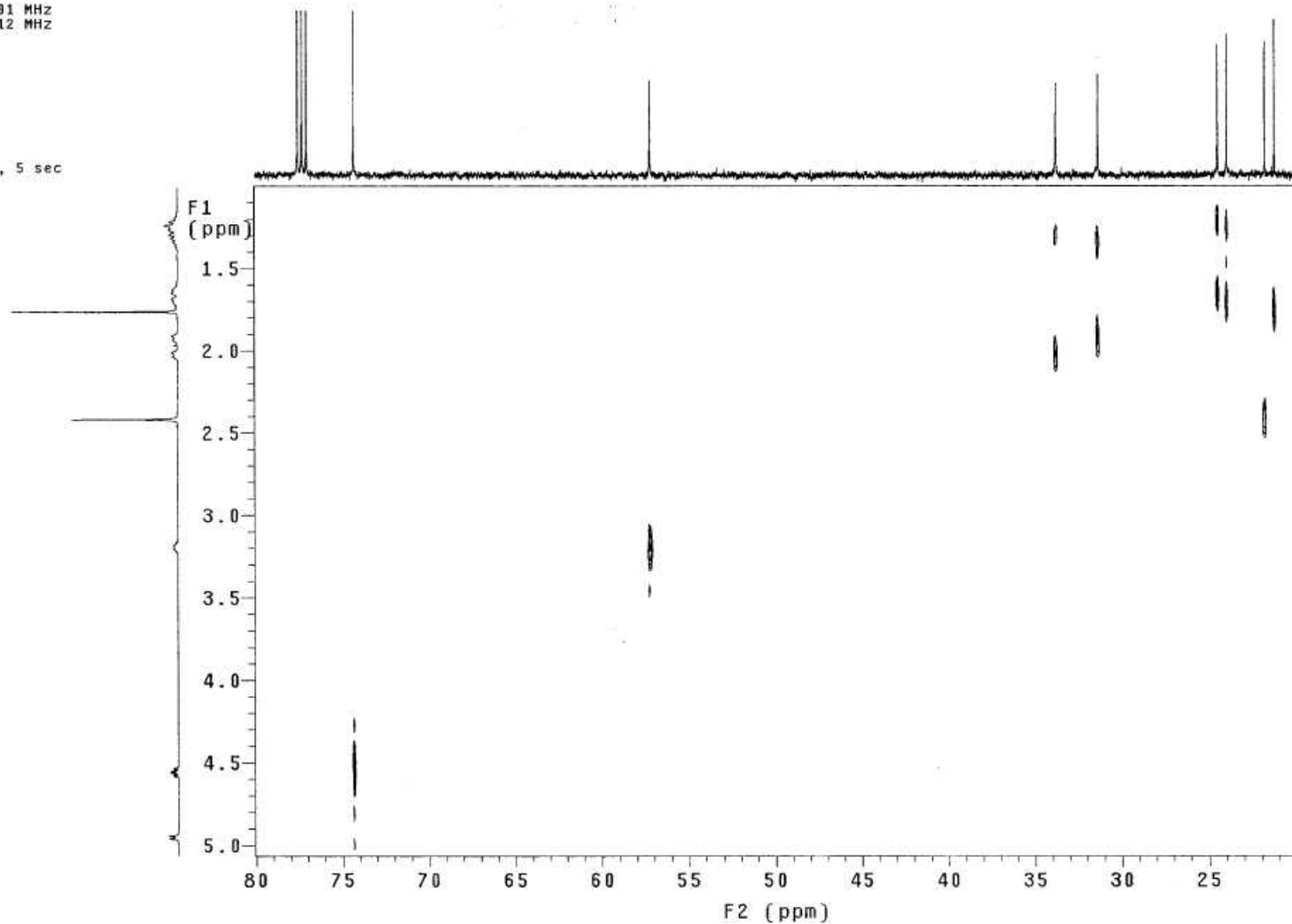
Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
16 repetitions  
256 increments  
OBSERVE H1, 499.8611709 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 1 hr, 41 min, 40 sec



TY2-337-A

Pulse Sequence: hetcor  
Solvent: CDC13  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE C13, 125.6901591 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 3 hr, 46 min, 5 sec



CH3 carbons



CH2 carbons



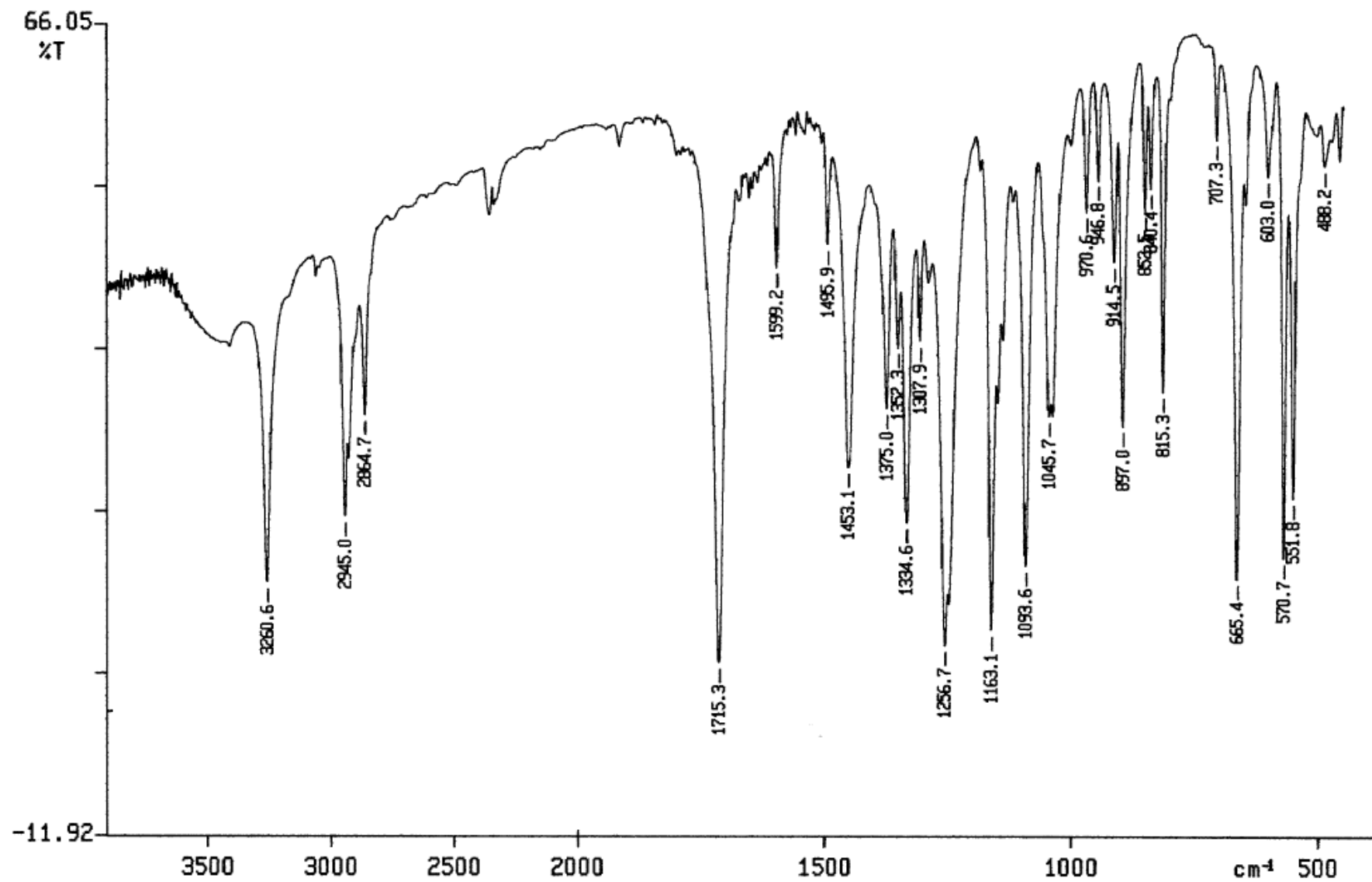
CH carbons



all protonated carbons



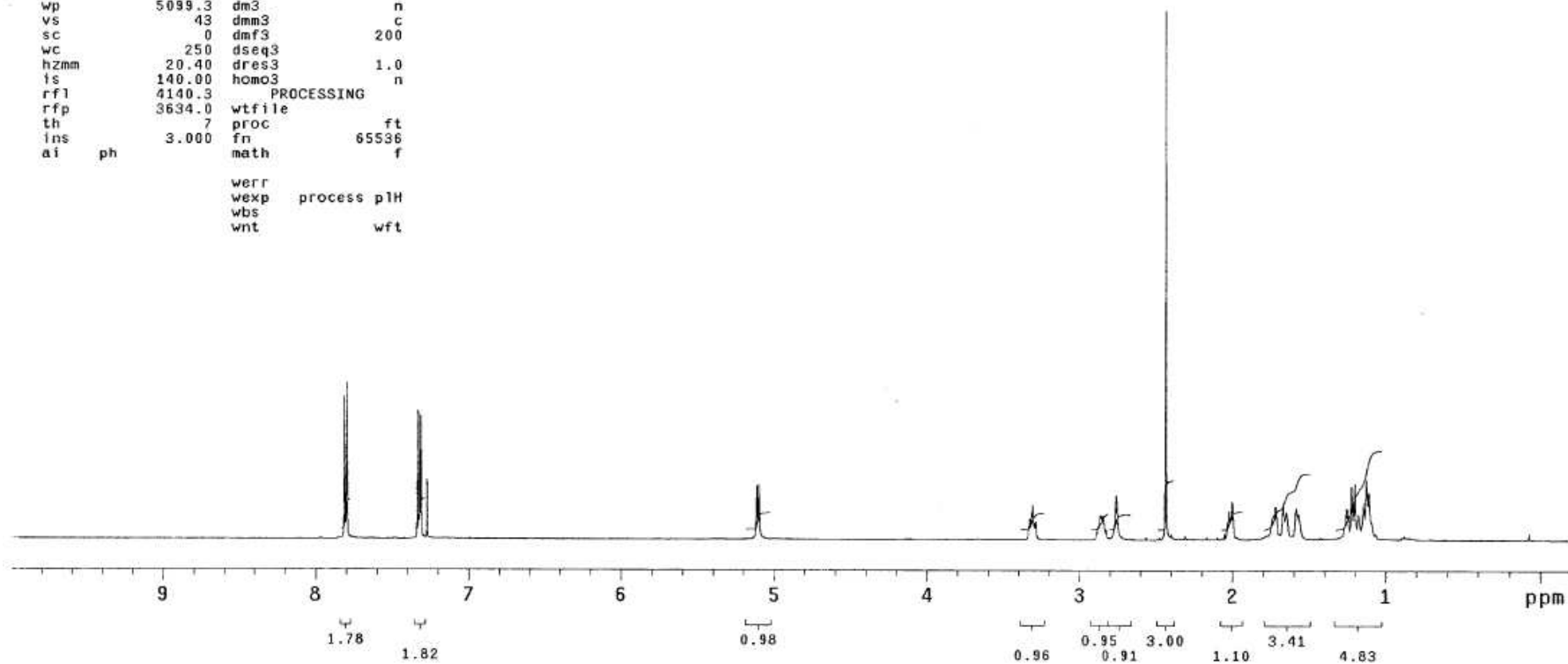
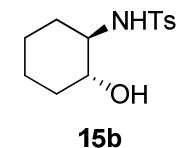
200 180 160 140 120 100 80 60 40 20 ppm



TY2-337-B

exp1 s2pu1

```
SAMPLE          DEC. & VT
date  Apr 13 2009  dfrq      499.864
solvent  CDC13      dn        H1
file     exp       dpwr      30
ACQUISITION  dof        0
sfrq     499.864  dm         nnn
tn       H1       dmm        c
at       5.015    dmf        200
np       65536    dseq
sw       6533.3   dres      1.0
fb       4000     homo
bs       4        DEC2
tpwr     61       dfrq2     0
pw       13.5     dn2
d1       0.100    dpwr2     1
tof      269.9    dof2      0
nt       16      dm2        n
ct       16      dmm2       c
alock    n        dmf2       200
gain     not used dseq2
FLAGS    n        dres2     1.0
         n        homo2     n
         n        DEC3
dp       y        dfrq3     0
hs       nn       dn3
DISPLAY  dpwr3     1
sp       -102.8   dof3      0
wp       5099.3   dm3        n
vs       43      dmm3       c
sc       0        dmf3       200
wc       250     dseq3
hzmm     20.40   dres3     1.0
is       140.00  homo3     n
rfl      4140.3  PROCESSING
rfp      3634.0  wfile
th       7       proc       ft
ins     3.000   fn        65536
ai      ph     math       f
                werr
                wexp  process pH
                wbs
                wnt   wft
```

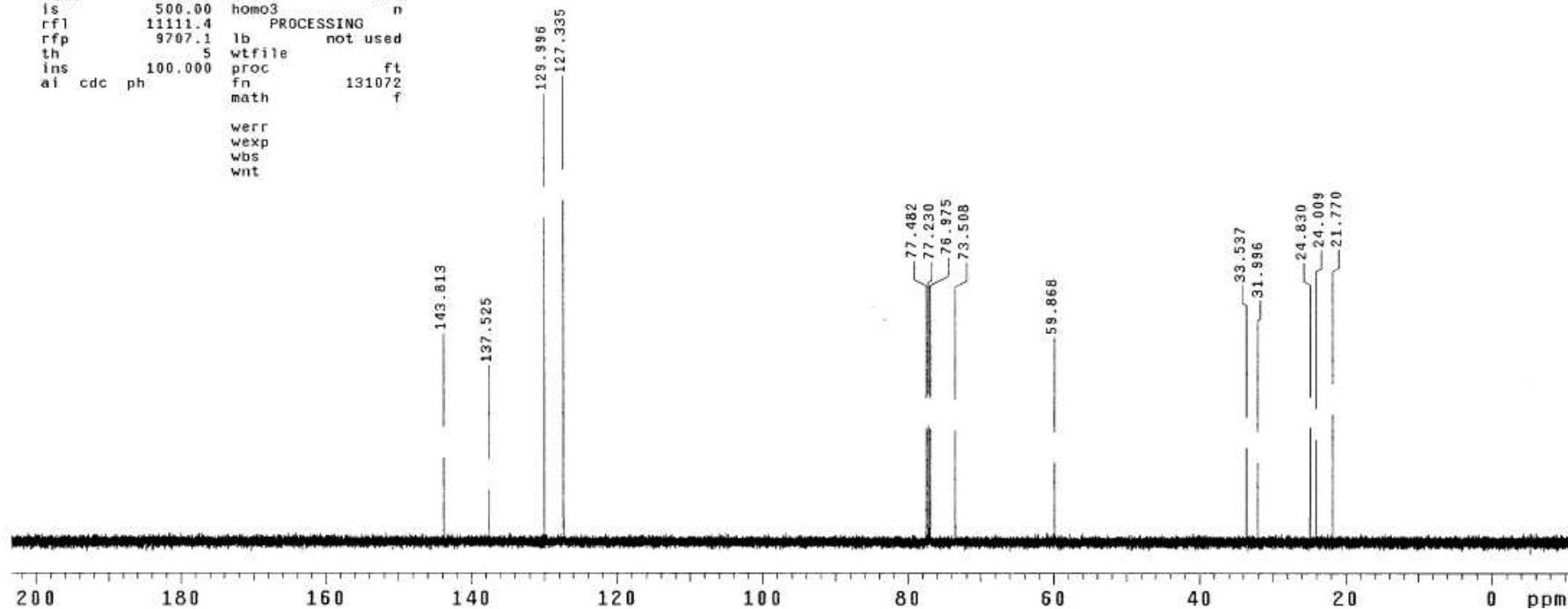




TY2-337-B

exp2 s2pu1

SAMPLE		DEC. & VT	
date	Apr 13 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION			
sfrq	125.702	dof	0
tn	C13	dm	yyy
at	1.215	dmm	w
np	65536	dmf	8787.35
sw	26963.3	dseq	
fb	15000	dres	1.0
bs	4	homo	n
tpwr	52	DEC2	
pw	10.2	dfrq2	0
d1	1.800	dn2	
tof	144.5	dpwr2	1
nt	3000	dof2	0
ct	78	dm2	n
alock	78	dmm2	c
gain	not used	dmf2	10000
FLAGS			
il	n	dseq2	1.0
in	n	dres2	
dp	y	homo2	n
hs	nn	DEC3	
DISPLAY			
sp	-1403.9	dfrq3	0
wp	26962.9	dn3	
vs	78	dpwr3	1
sc	0	dof3	0
wc	250	dm3	n
hzmm	107.85	dmm3	c
ls	500.00	dmf3	10000
rfl	11111.4	dseq3	
rfp	9707.1	dres3	1.0
th	5	homo3	n
ins	100.000	PROCESSING	
ai	cdc ph	lb	not used
		wfile	
		proc	ft
		fn	131072
		math	f
		werr	
		wexp	
		wbs	
		wnt	



TY2-337-B

Pulse Sequence: relayh

Solvent: CDCl<sub>3</sub>

Ambient temperature

INDVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec

COSY 90-90

Acq. time 0.157 sec

Width 6533.3 Hz

2D Width 6533.3 Hz

32 repetitions

256 increments

OBSERVE H1, 499.8611905 MHz

DATA PROCESSING

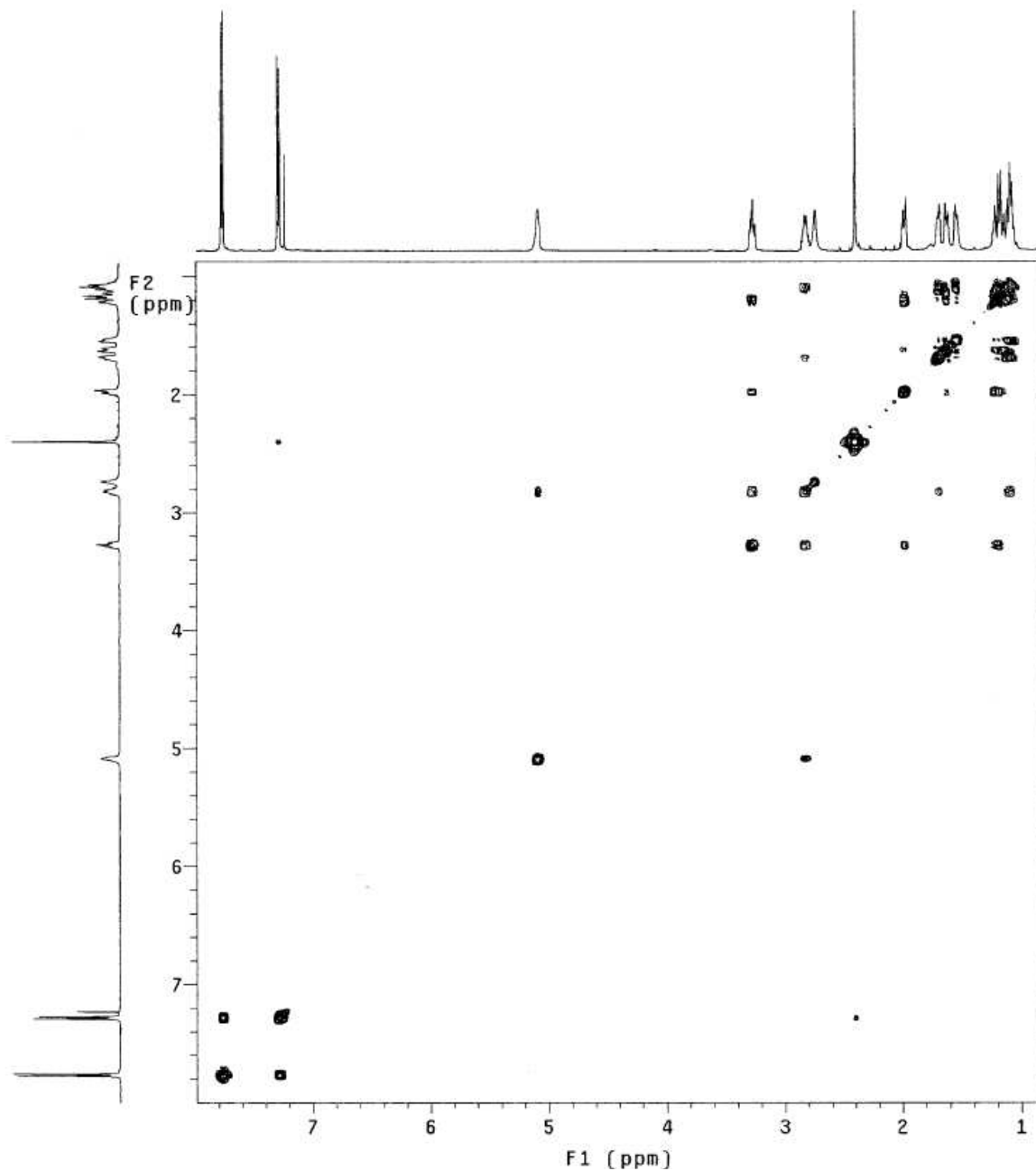
Sine bell 0.078 sec

F1 DATA PROCESSING

Sine bell 0.039 sec

FT size 2048 x 2048

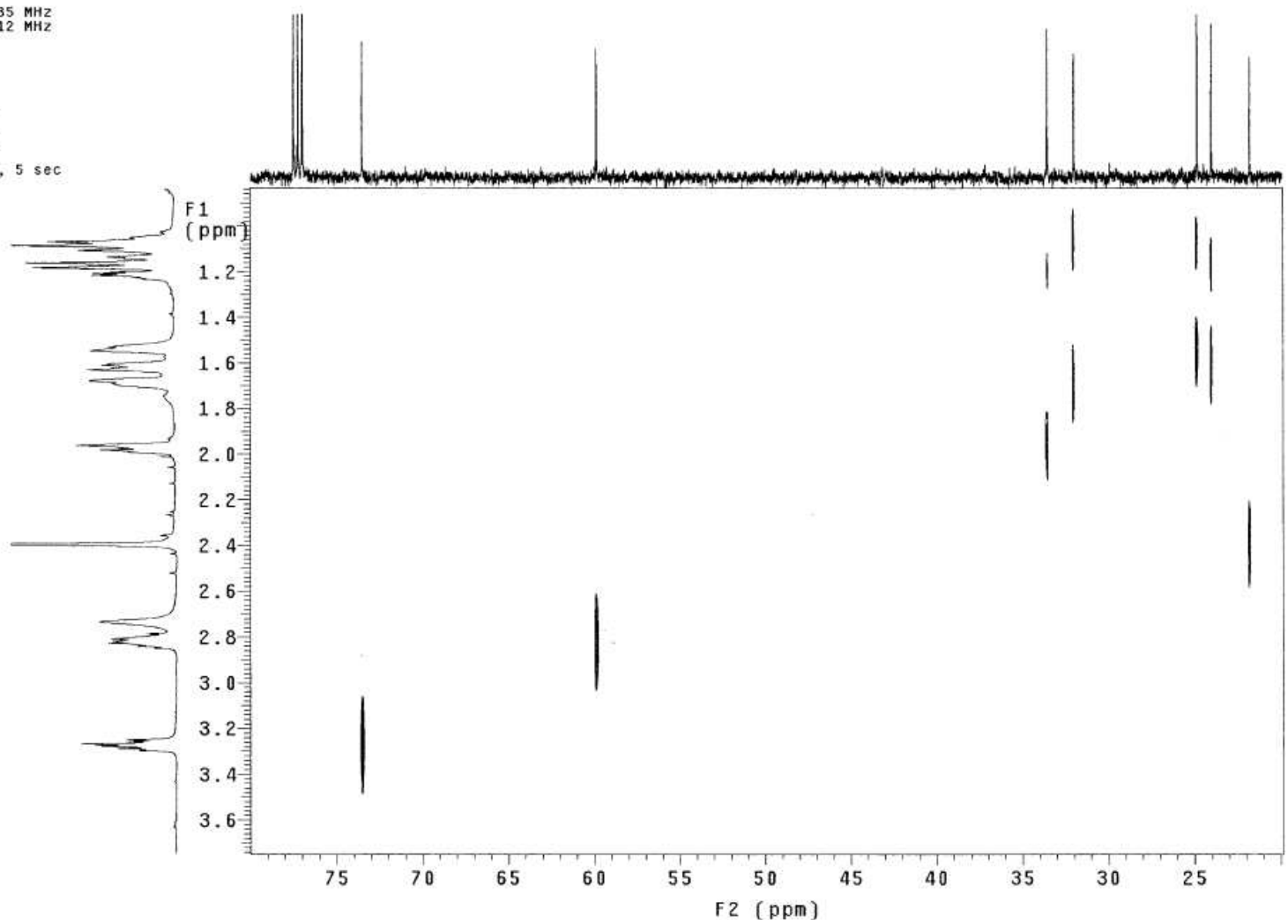
Total time 3 hr, 23 min, 10 sec



TY2-337-B

Pulse Sequence: hetcor  
Solvent: CDC13  
Ambient temperature  
User: 1-14-87  
INDVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE C13, 125.6901685 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
F1 size 4096 x 1024  
Total time 3 hr, 46 min, 5 sec



CH3 carbons



CH2 carbons



CH carbons



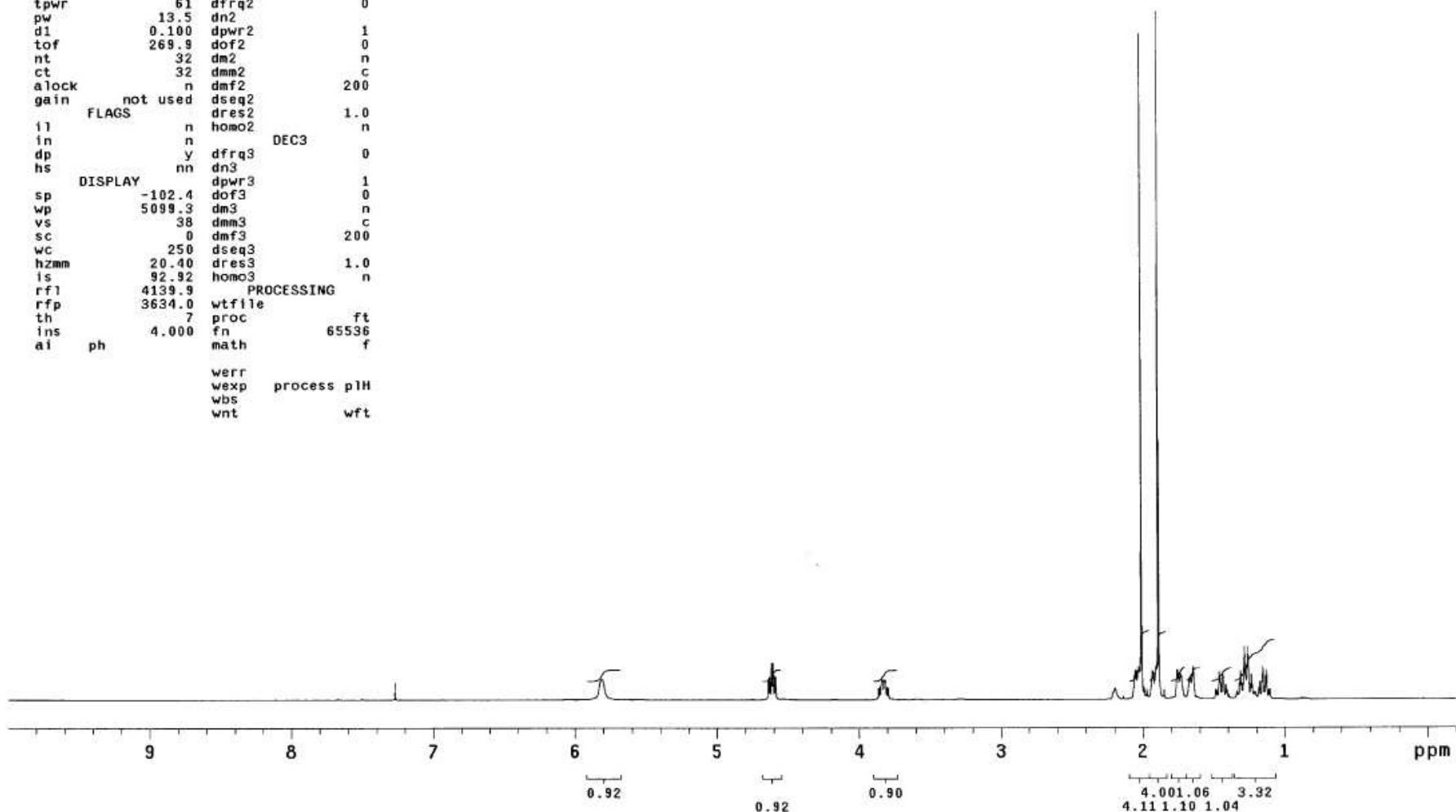
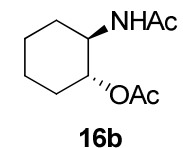
all protonated carbons



TY2-378

expl s2pu1

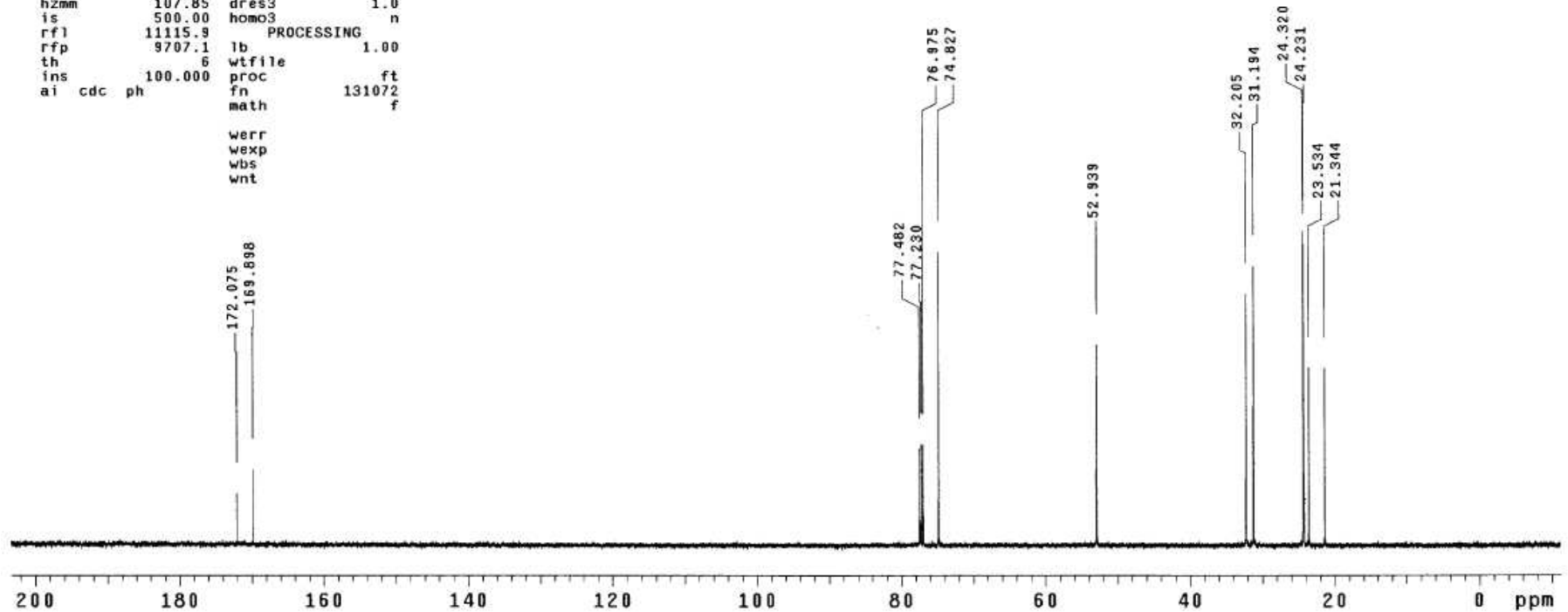
```
SAMPLE          DEC. & VT
date Jun 29 2009 dfrq          499.864
solvent CDC13      dn          H1
file exp         dpwr         30
ACQUISITION    dof          0
sfrq 499.864    dm           nnn
tn    H1        dmm          c
at    5.016     dmf          200
np    65536    dseq
sw    6533.3   dres          1.0
fb    4000     homo
bs    4        DEC2
tpwr  61       dfrq2         0
pw    13.5    dn2
d1    0.100   dpwr2         1
tof   269.9   dof2          0
nt    32      dm2          n
ct    32      dmm2         c
alock n        dmf2          200
gain  not used dseq2
FLAGS n        dres2         1.0
      n        homo2         n
      n        DEC3
      y        dfrq3         0
      nn       dn3
DISPLAY dpwr3         1
sp    -102.4   dof3          0
wp    5099.3   dm3           n
vs    38      dmm3         c
sc    0       dmf3          200
wc    250     dseq3
hzmm  20.40   dres3         1.0
is    92.92   homo3         n
rfl   4139.9  PROCESSING
rfp   3634.0  wtfile
th    7       proc          ft
ins   4.000   fn           65536
al    ph     math          f
                    werr
                    wexp   process plH
                    wbs
                    wnt    wft
```



TY2-378

exp2 s2pu1

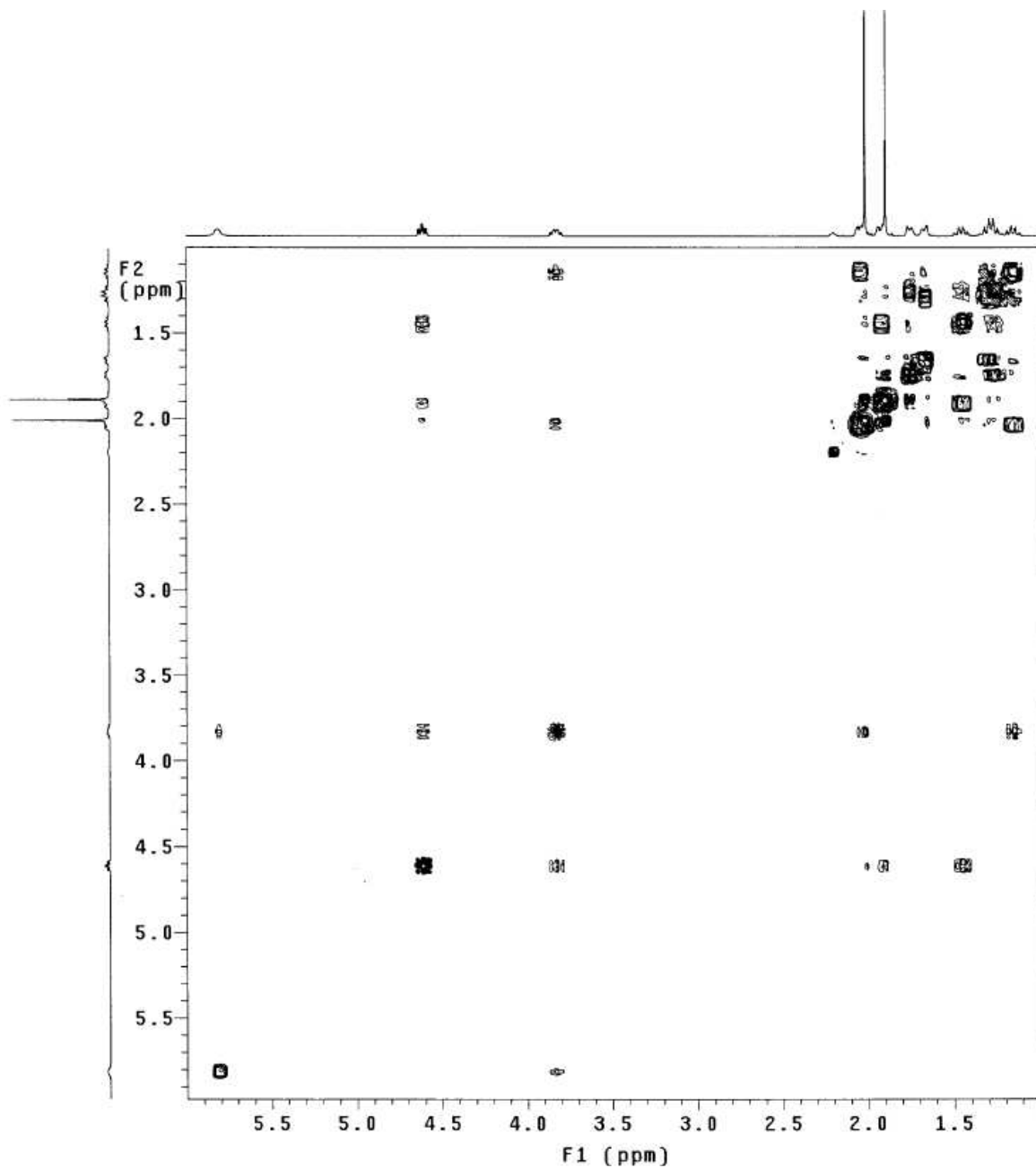
SAMPLE		DEC. & VT	
date	Jun 29 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION			
sfrq	125.702	dof	0
tn	C13	dm	yyy
at	1.215	dmm	w
np	65536	dmf	8787.35
sw	26963.3	dseq	
fb	15000	dres	1.0
bs	4	homo	n
tpwr	52	DEC2	
pw	10.2	dfrq2	0
d1	1.800	dn2	
tof	144.5	dpwr2	1
nt	3000	dof2	0
ct	139	dm2	n
alock	n	dmm2	c
gain	not used	dmf2	10000
FLAGS			
i1	n	dseq2	1.0
in	n	dres2	
dp	y	homo2	n
hs	nn	DEC3	
DISPLAY			
sp	-1408.5	dfrq3	0
wp	26962.9	dn3	
vs	124	dpwr3	1
sc	0	dof3	0
wc	250	dm3	n
hzmm	107.85	dmm3	c
is	500.00	dmf3	10000
rfl	11115.9	dseq3	
rfp	9707.1	dres3	1.0
th	6	homo3	n
ins	100.000	PROCESSING	
ai cdc ph		lb	1.00
		wfile	
		proc	ft
		fn	131072
		math	f



TY2-378

Pulse Sequence: relayh  
Solvent: CDCl3  
Ambient temperature  
INNOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE H1, 499.8611705 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 3 hr, 23 min, 10 sec

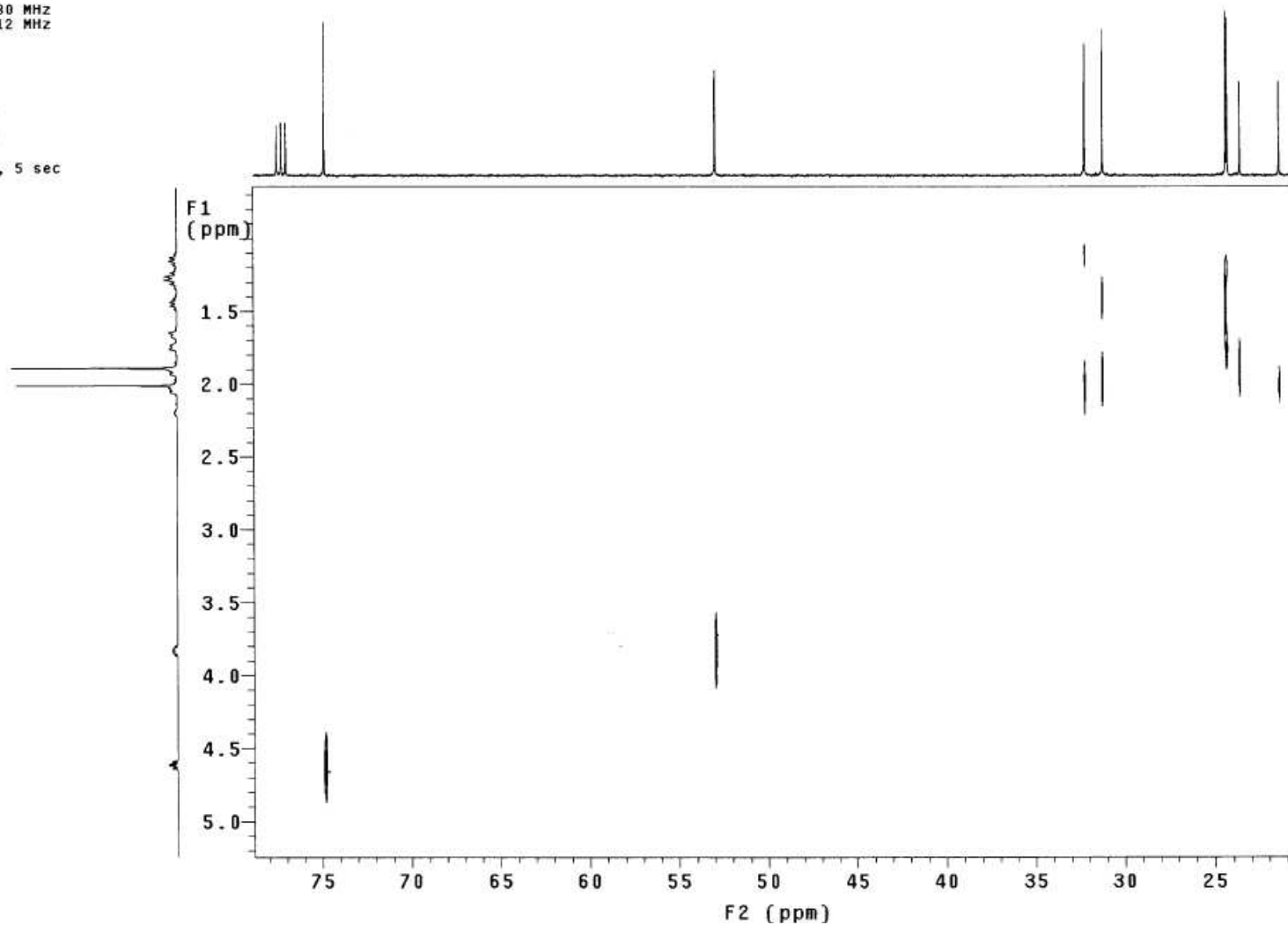


TY2-378

Pulse Sequence: hetcor

Solvent: CDC13  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE C13, 125.6901730 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 3 hr, 46 min, 5 sec





CH3 carbons



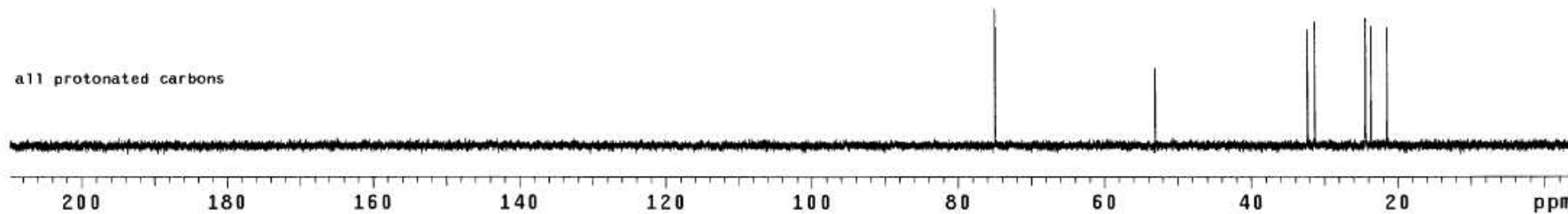
CH2 carbons

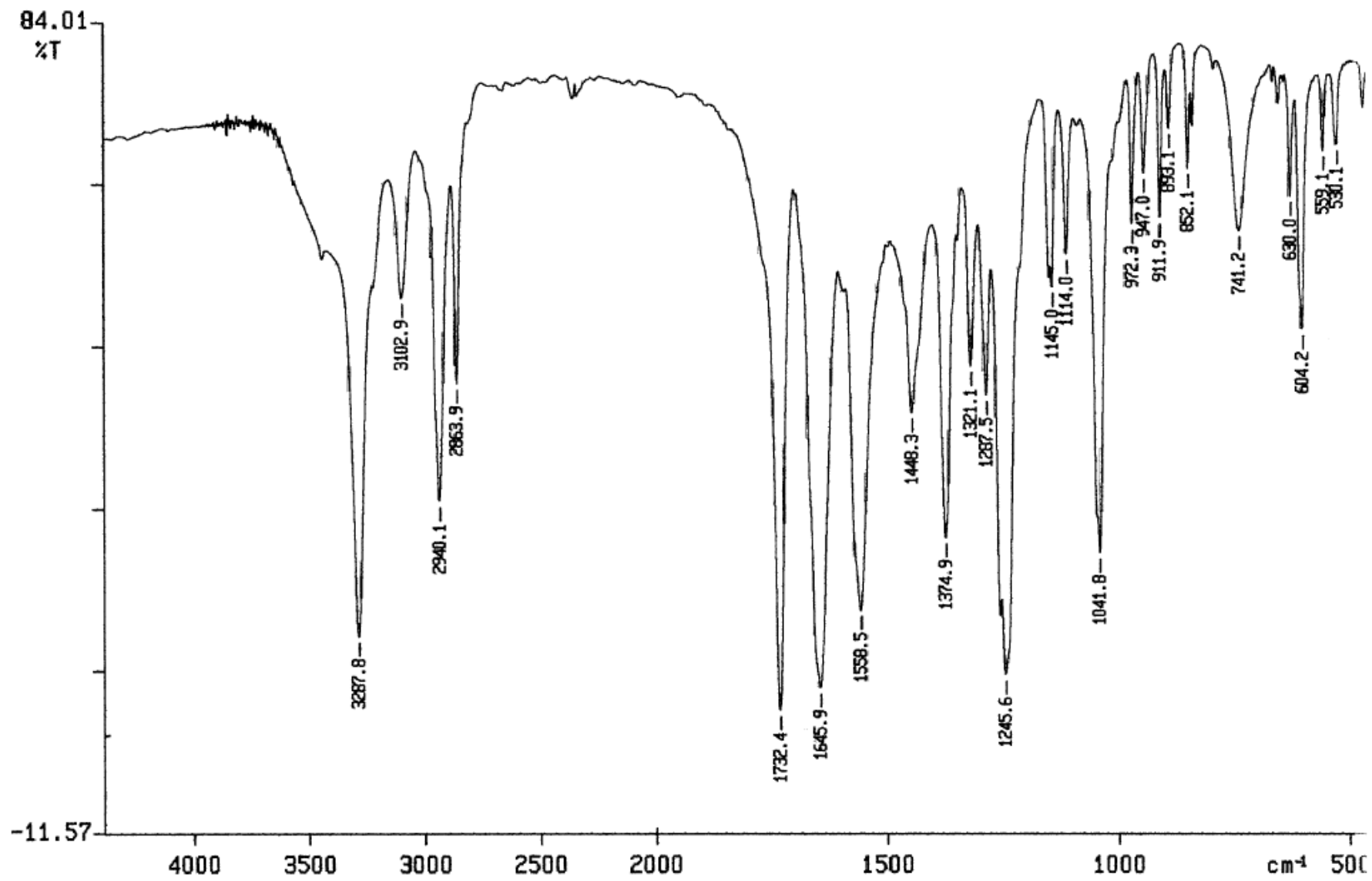


CH carbons



all protonated carbons

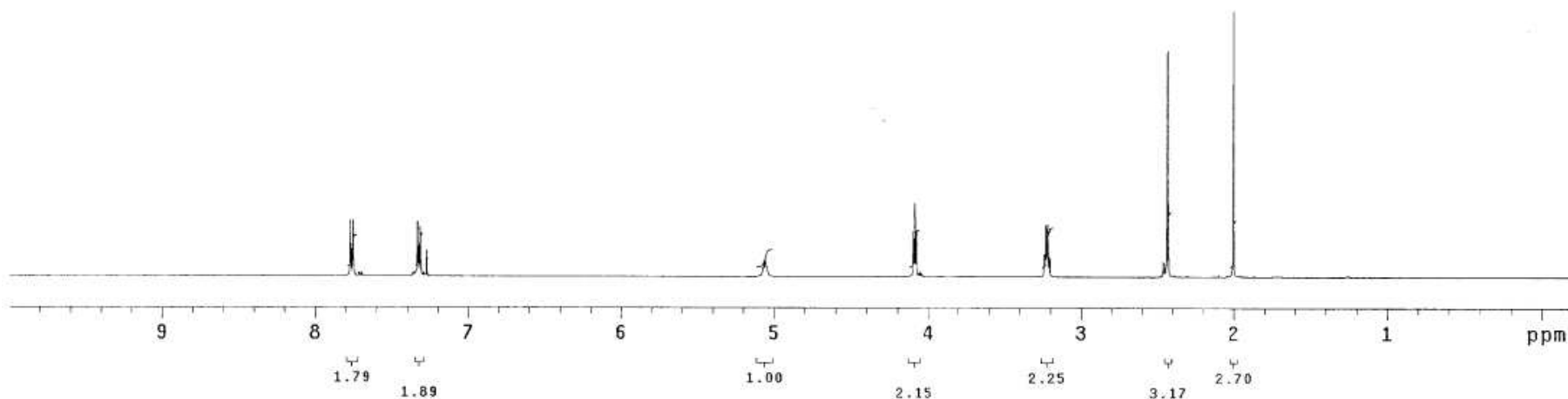
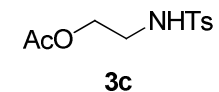




TY2-344

expl s2pu1

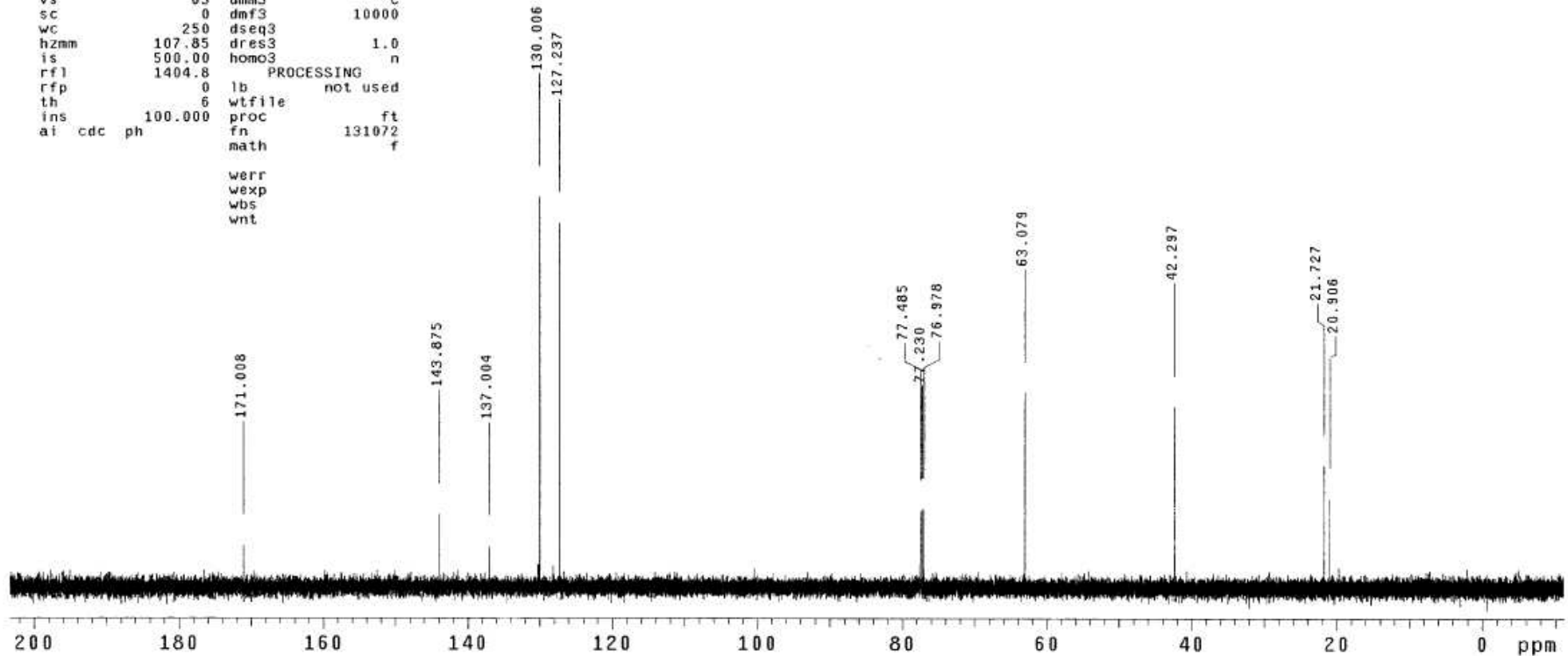
```
SAMPLE          DEC. & VT
date   Apr 23 2009  dfrq      499.864
solvent CDC13      dn        H1
file    exp        dpwr      30
ACQUISITION      dof        0
sfrq    499.864   dm        nnn
tn      H1        dmm        C
at      5.016     dmf        200
np      65536     dseq       1.0
sw      6533.3   dres       1.0
fb      4000     homo       n
bs      4        DEC2
tpwr    61       dfrq2      0
pw      13.5     dn2        1
d1      0.100    dpwr2      0
tof     269.9    dof2       0
nt      4        dm2        n
ct      4        dmm2       C
alock   n        dmf2       200
gain    not used dseq2      1.0
        FLAGS   n        homo2     n
        in      n        DEC3
        dp      y        dfrq3     0
        hs     nn       dn3
        DISPLAY  dpwr3     1
        sp     -102.2  dof3      0
        wp     5099.3  dm3       n
        vs     15     dmm3      C
        sc     0      dmf3      200
        wc     250    dseq3     1.0
        hzmm   20.40  dres3     1.0
        is     117.48 homo3     n
        rfl    4139.7 PROCESSING
        rfp    3634.0 wtfile
        th     7      proc
        ins   1.000  fn        65536
        al    ph     math        f
        werr
        wexp   process pH
        wbs
        wnt    wft
```



TY2-344

exp2 s2pu1

SAMPLE		DEC. & VT	
date	Apr 23 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION		dof	0
sfrq	125.702	dm	yyy
tn	C13	dmm	w
at	1.215	dmf	8787.35
np	65536	dseq	
sw	26963.3	dres	1.0
fb	15000	homo	n
bs	4	DEC2	
tpwr	52	dfrq2	0
pw	10.2	dn2	
d1	1.800	dpwr2	1
tof	144.5	dof2	0
nt	2000	dm2	n
ct	60	dmm2	c
alock	n	dmf2	10000
gain	not used	dseq2	
FLAGS		dres2	1.0
il	n	homo2	n
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY		dpwr3	1
sp	-1404.3	dof3	0
wp	26962.9	dm3	n
vs	63	dmm3	c
sc	0	dmf3	10000
wc	250	dseq3	
h2mm	107.85	dres3	1.0
is	500.00	homo3	n
rfl	1404.8	PROCESSING	
rfp	0	lb	not used
th	6	wtfile	
ins	100.000	proc	ft
ai	cdc ph	fn	131072
		math	f



TY2-344

Pulse Sequence: relayh

Solvent: CDCl3

Ambient temperature

INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec

COSY 90-90

Acq. time 0.157 sec

Width 6533.3 Hz

2D Width 6533.3 Hz

32 repetitions

256 increments

OBSERVE H1, 499.8611703 MHz

DATA PROCESSING

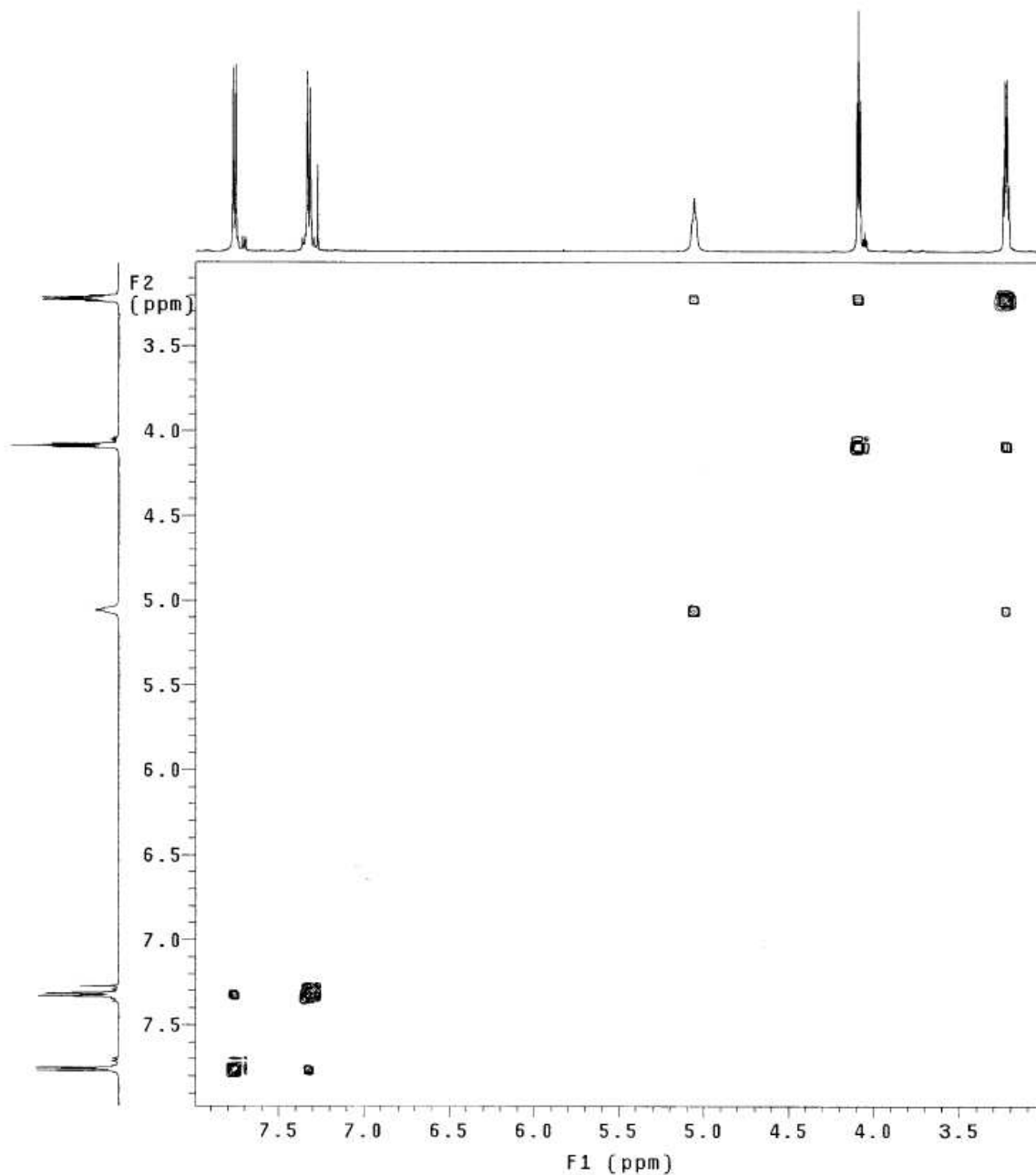
Sine bell 0.078 sec

F1 DATA PROCESSING

Sine bell 0.039 sec

FT size 2048 x 2048

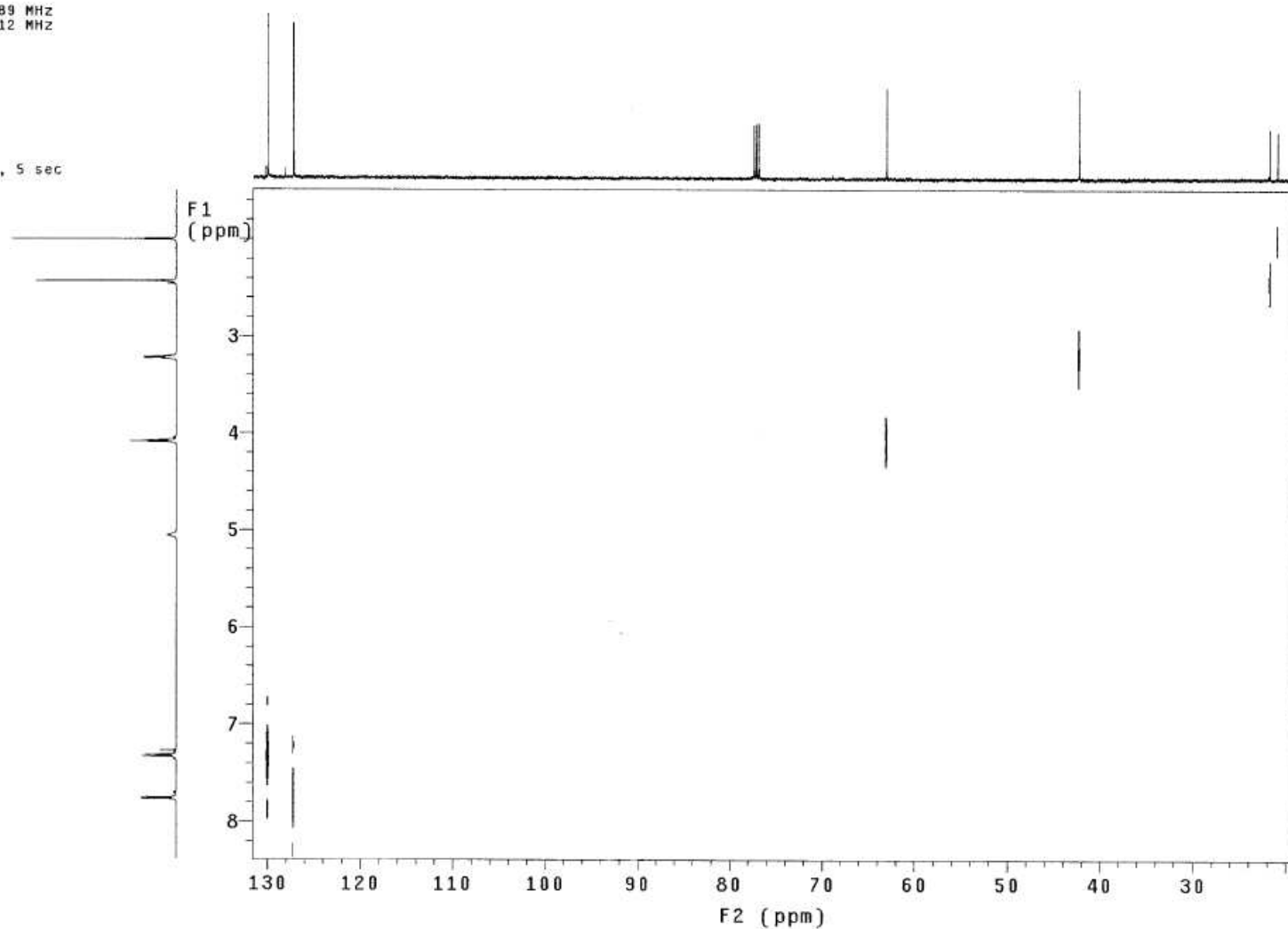
Total time 3 hr, 23 min, 10 sec



TY2-344

Pulse Sequence: hetcor  
Solvent: CDC13  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE C13, 125.6901689 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 3 hr, 46 min, 5 sec



CH3 carbons



CH2 carbons



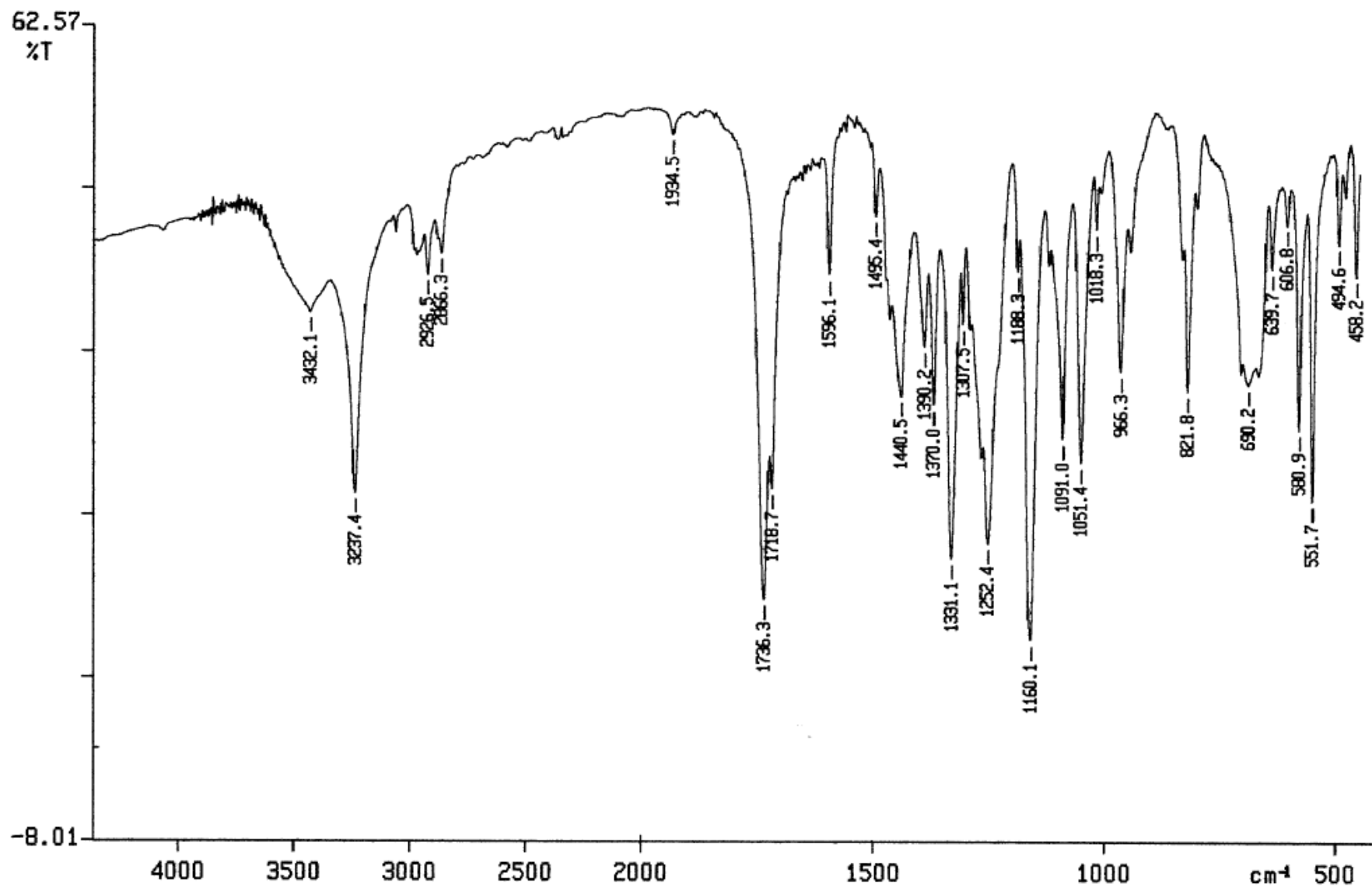
CH carbons



all protonated carbons



200 180 160 140 120 100 80 60 40 20 ppm





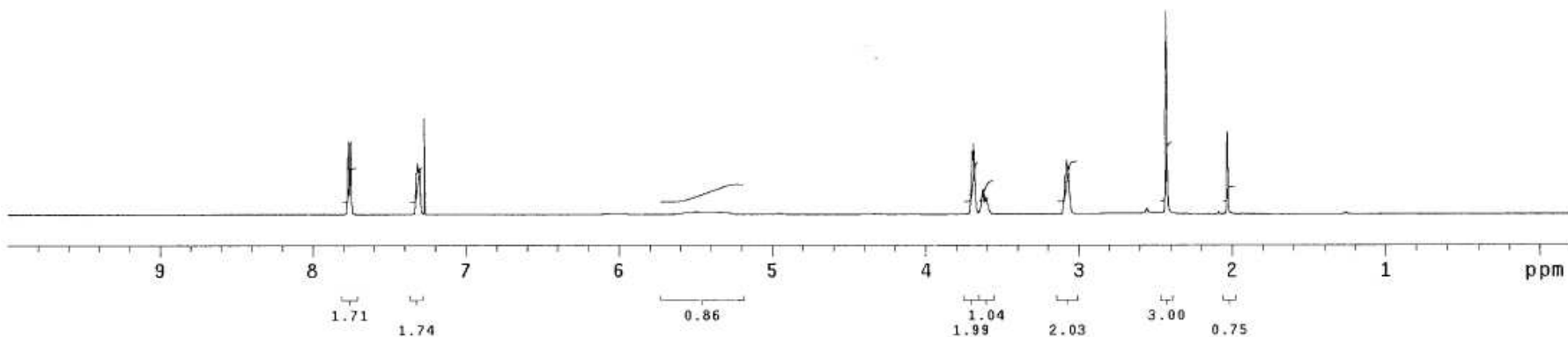
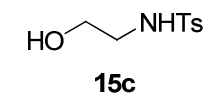
TY2-362-B-c15-22

exp1 s2pu1

```

SAMPLE          DEC. & VT
date Jun 6 2009 dfrq          499.864
solvent CDC13      dn          H1
file exp          dpwr         30
ACQUISITION    dof          0
sfrq 499.864     dm           nnn
tn H1            dmm          c
at 5.016         dmf          200
np 65536         dseq
sw 6533.3        dres          1.0
fb 4000          homo
bs 4             DEC2
tpwr 61          dfrq2         0
pw 13.5          dn2
dl 0.100         dpwr2         1
tof 269.3        dof2          0
nt 16           dm2           n
ct 16           dmm2          c
alock n          dmf2          200
gain not used    dseq2
FLAGS          dres2         1.0
il n            homo2         n
ln n            DEC3
dp y            dfrq3         0
hs nn          dn3
DISPLAY        dpwr3         1
sp -103.0       dof3          0
wp 5099.3       dm3           n
vs 29          dmm3          c
sc 0           dmf3          200
wc 250         dseq3
hzmm 20.40     dres3         1.0
is 136.39     homo3         n
rfl 4140.5     PROCESSING
rfp 3634.0     wtfile
th 50          proc          ft
ins 3.000      fn           65536
ai ph          math          f

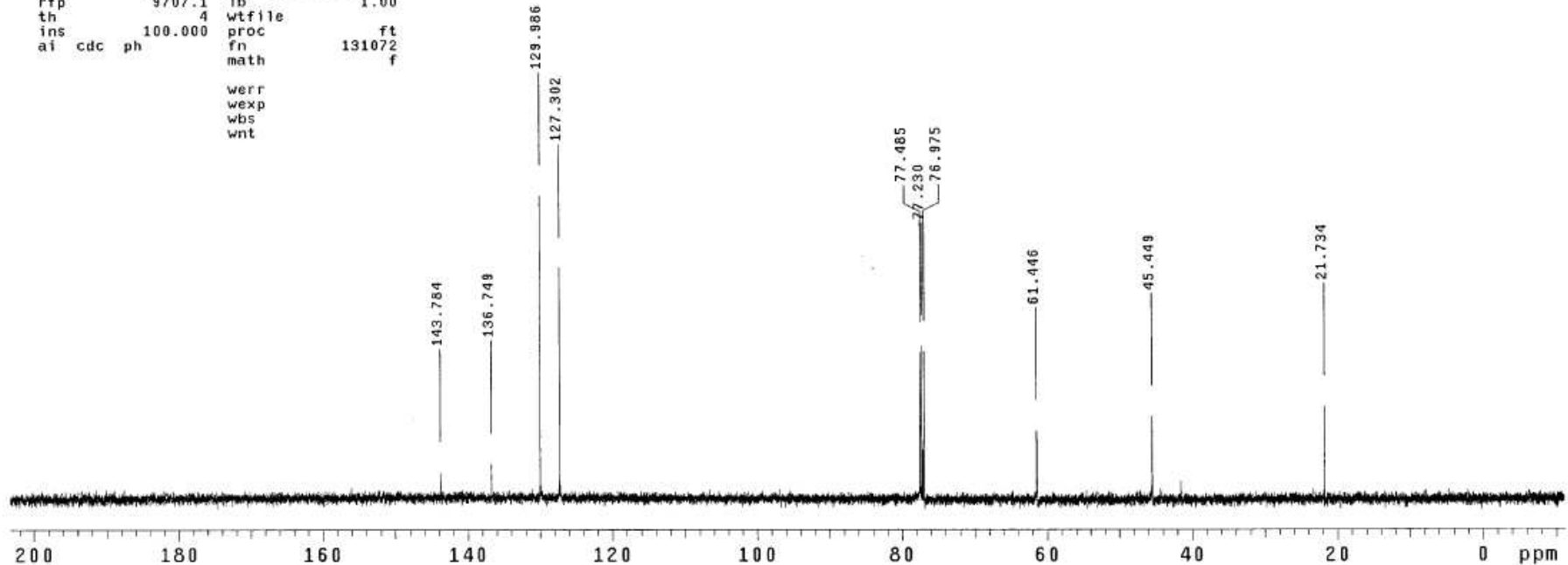
werr
wexp process pH
wbs
wnt wft
```



TY2-362-B-c15-22

exp2 s2pu1

SAMPLE		DEC. & VT	
date	Jun 6 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION			
sfrq	125.702	dof	0
tn	C13	dm	yyv
at	1.215	dmm	w
np	65536	dmf	8787.35
sw	26963.3	dseq	
fb	15000	dres	1.0
bs	4	homo	n
tpwr	52	DEC2	
pw	10.2	dfrq2	0
d1	1.800	dn2	
tof	144.5	dpwr2	1
nt	2000	dof2	0
ct	84	dm2	n
alock	n	dmm2	c
gain	not used	dmf2	10000
FLAGS			
il	n	dseq2	1.0
in	n	dres2	1.0
dp	y	homo2	n
hs	nn	DEC3	
DISPLAY			
sp	-1403.5	dfrq3	0
wp	26962.9	dn3	
vs	207	dpwr3	1
sc	0	dof3	0
vc	250	dm3	n
hzmm	107.85	dmm3	c
is	500.00	dmf3	10000
rfl	11111.0	dseq3	
rfp	9707.1	dres3	1.0
th	4	homo3	n
ins	100.000	PROCESSING	
ai	cdc ph	lb	1.00
		wfille	
		proc	ft
		fn	131072
		math	f

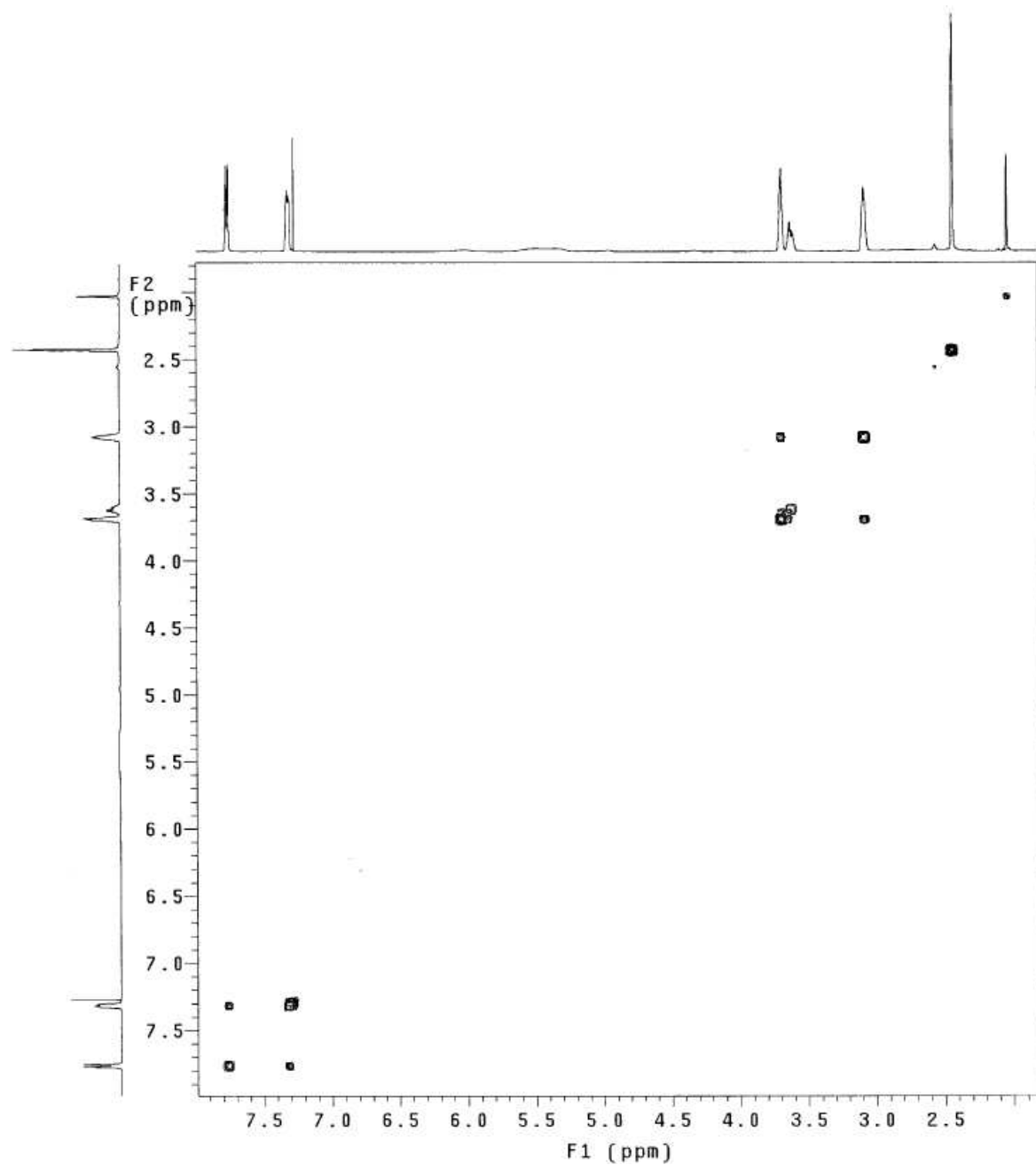


TY2-362-B-c15-22

Pulse Sequence: relayh

Solvent: CDC13  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

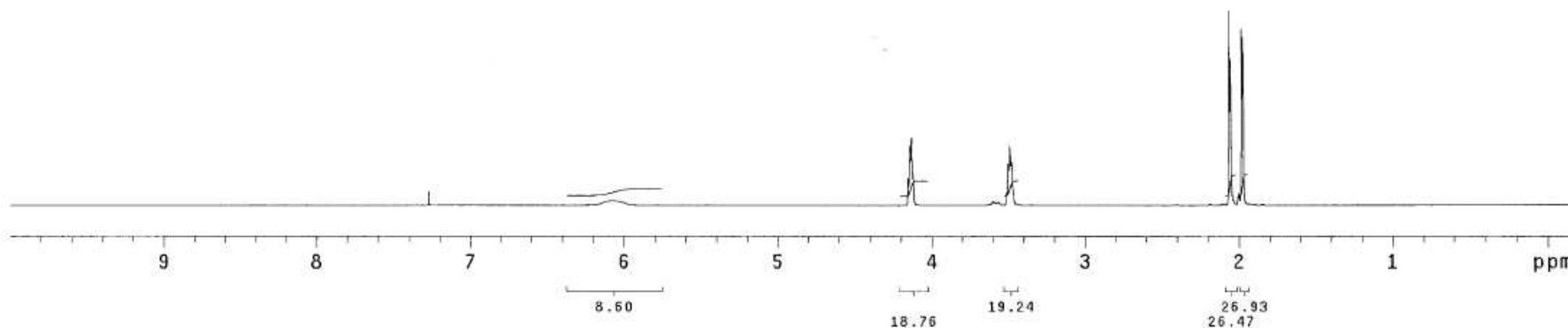
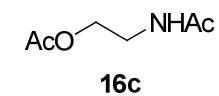
Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE H1, 499.8611711 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 3 hr, 23 min, 10 sec



TY2-362-C-c24-28

expl s2pu1

```
SAMPLE          DEC. & VT
date Jun 6 2009  dfrq          499.864
solvent CDC13    dn           H1
file      exp   dpwr          30
ACQUISITION     dof           0
sfrq      499.864 dm           nnn
tn         H1   dmm           c
at         5.016 dmf          200
np         65536 dseq
sw         6533.3 dres          1.0
fb         4000  homo
bs         4      DEC2
tpwr       61   dfrq2         0
pw         13.5 dn2
d1         0.100 dpwr2         1
tof        269.9 dof2          0
nt         4    dm2           n
ct         4    dmm2          c
alock      n    dmf2          200
gain      not used dseq2
FLAGS      n    dres2          1.0
           n    homo2         n
in         n      DEC3
dp         y    dfrq3         0
hs         nn   dn3
DISPLAY    dpwr3          1
sp        -102.2 dof3          0
wp         5099.3 dm3           n
vs         16   dmm3          c
sc         0    dmf3          200
wc         250  dseq3
hzmm       20.40 dres3          1.0
ls         31.00 homo3         n
rfl        4139.7 PROCESSING
rfp        3634.0 wtfile
th         50   proc          ft
ins       100.000 fn           65536
al ph      math           f
           werr
           wexp process pH
           wbs
           wnt          wft
```



TY2-362-C-c24-28

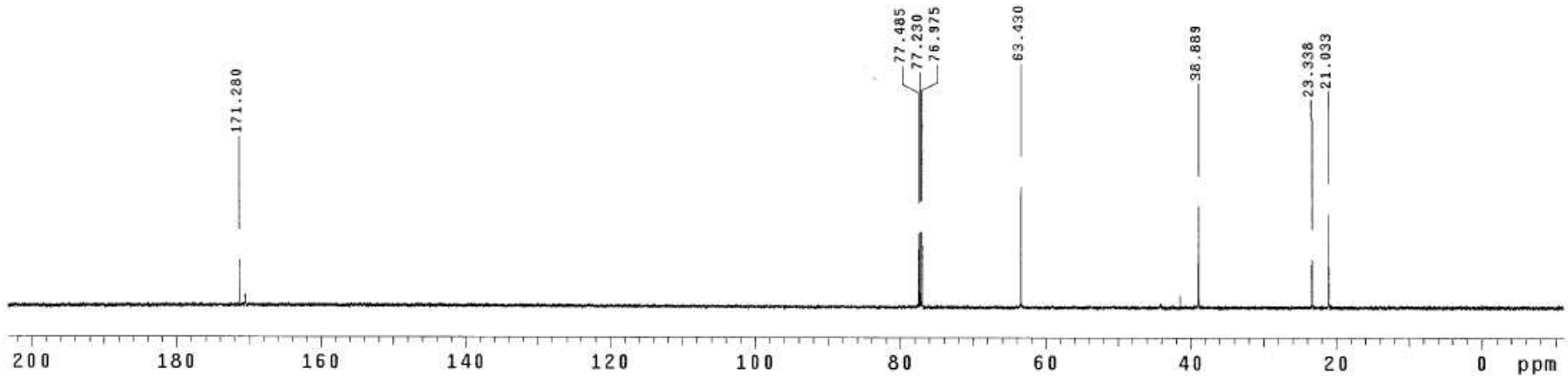
exp2 s2pu1

```

SAMPLE          DEC. & VT
date Jun 6 2009 dfrq 499.864
solvent CDC13   dn      M1
file exp       dpwr   40
ACQUISITION    dof     0
sfrq 125.702   dm      yyy
tn C13         dmm     w
at 1.215       dmf     8787.35
np 65536       dseq
sw 26963.3     dres   1.0
fb 15000       homo   n
bs 4           DEC2
tpwr 52        dfrq2  0
pw 10.2        dn2    0
d1 1.800       dpwr2  1
tof 144.5      dof2   0
nt 300         dm2    n
ct 257         dmm2   c
alock n         dmf2   10000
gain not used  dseq2
FLAGS          dres2   1.0
il n           homo2  n
in n           DEC3
dp y          dfrq3  0
hs nn         dn3    0
DISPLAY       dpwr3   1
sp -1407.2     dof3   0
wp 26962.9    dm3    n
vs 123        dmm3   c
sc 0          dmf3   10000
wc 250        dseq3
hzmm 107.85   dres3  1.0
ls 500.00    homo3  n
rfl 11114.7  PROCESSING
rfp 9707.1   lb      1.00
th 6         wtfile
lms 100.000  proc   ft
al cdc ph   fn     131072
            math  f

```

werr  
wexp  
wbs  
wnt

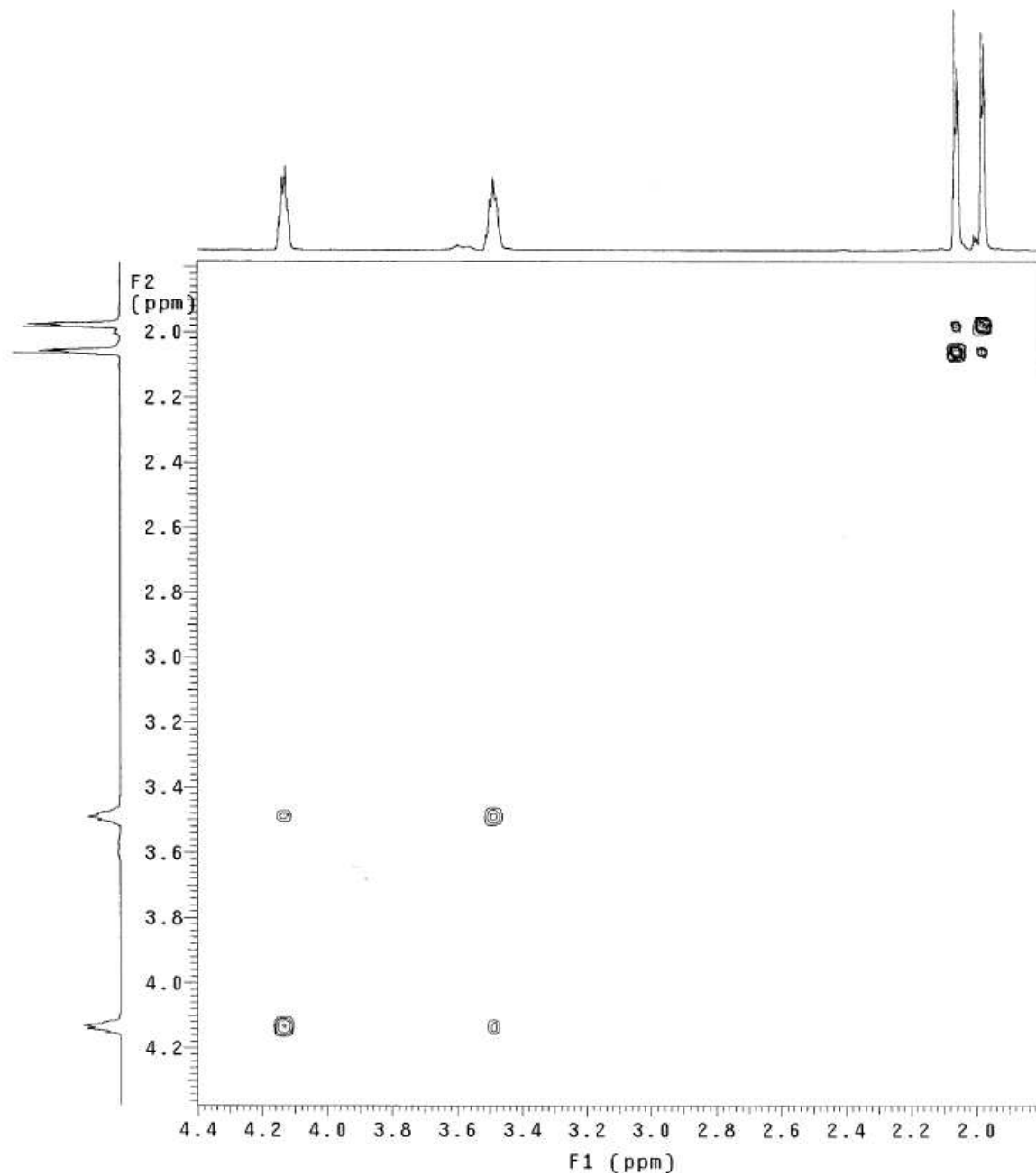


TY2-362-C-c24-28

Pulse Sequence: relayh

Solvent: CDCl<sub>3</sub>  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE H1, 499.8611703 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 3 hr, 23 min, 10 sec



TY2-362-C-c24-28

Pulse Sequence: hetcor

Solvent: CDCl<sub>3</sub>

Ambient temperature

User: 1-14-87

INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec

Acq. time 0.111 sec

Width 18403.5 Hz

2D Width 6533.3 Hz

32 repetitions

256 increments

OBSERVE C13, 125.6901718 MHz

DECOUPLE H1, 499.8639312 MHz

Power 40 dB

on during acquisition

off during delay

WALTZ-16 modulated

DATA PROCESSING

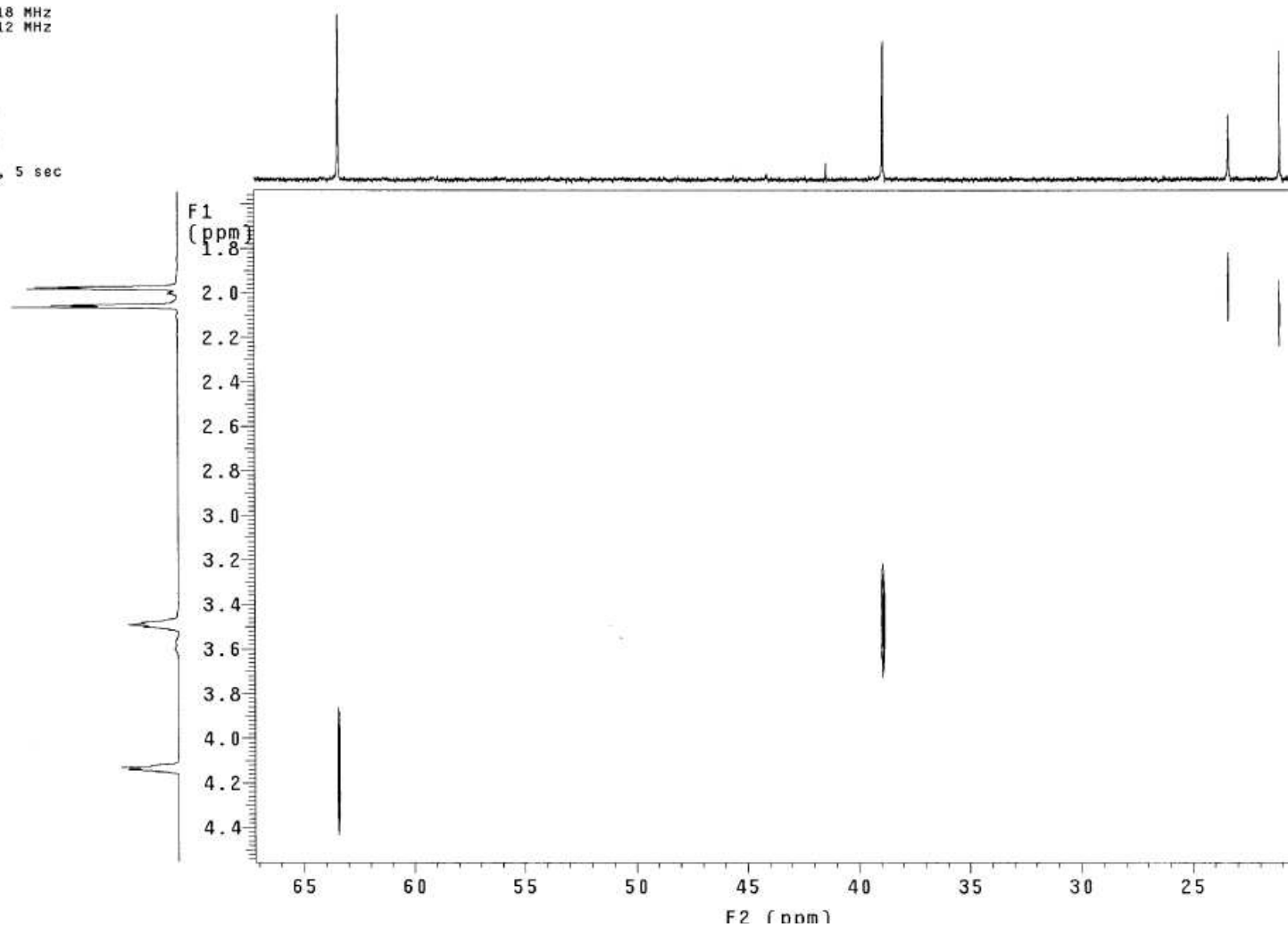
Line broadening 1.0 Hz

F1 DATA PROCESSING

Line broadening 0.3 Hz

FT size 4096 x 1024

Total time 3 hr, 46 min, 5 sec



CH3 carbons



CH2 carbons



CH carbons

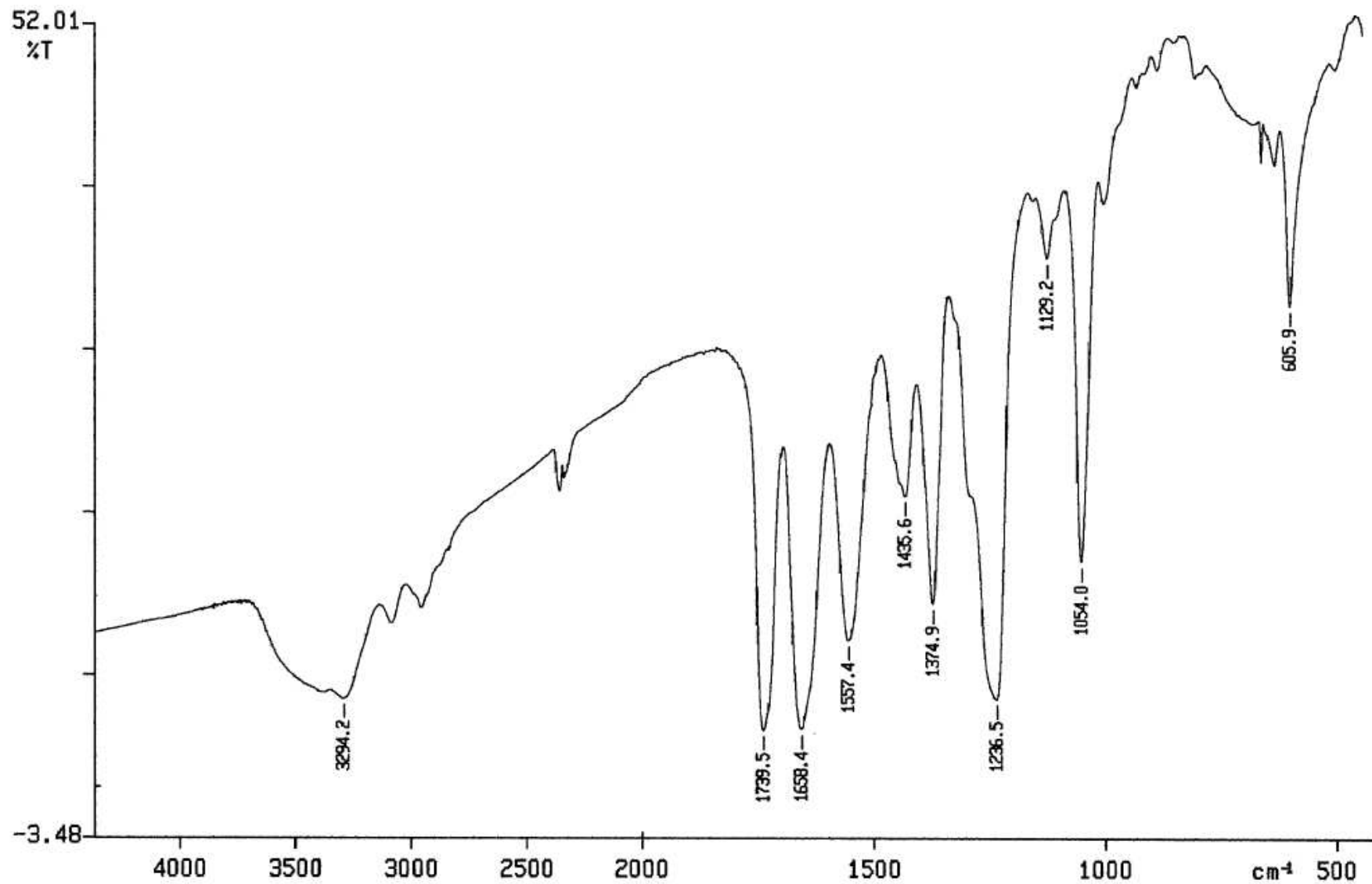


all protonated carbons



200 180 160 140 120 100 80 60 40 20 ppm

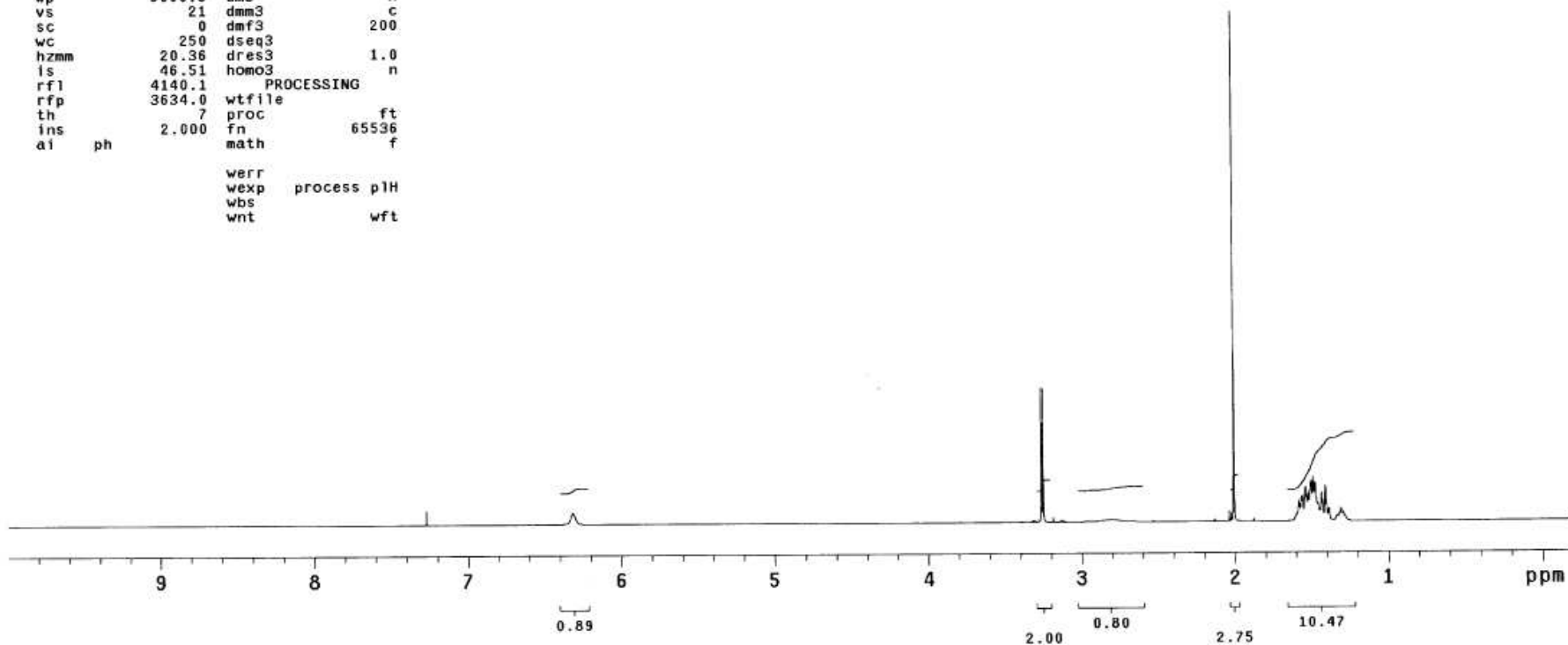
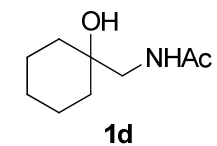




TY2-393

exp2 s2pu1

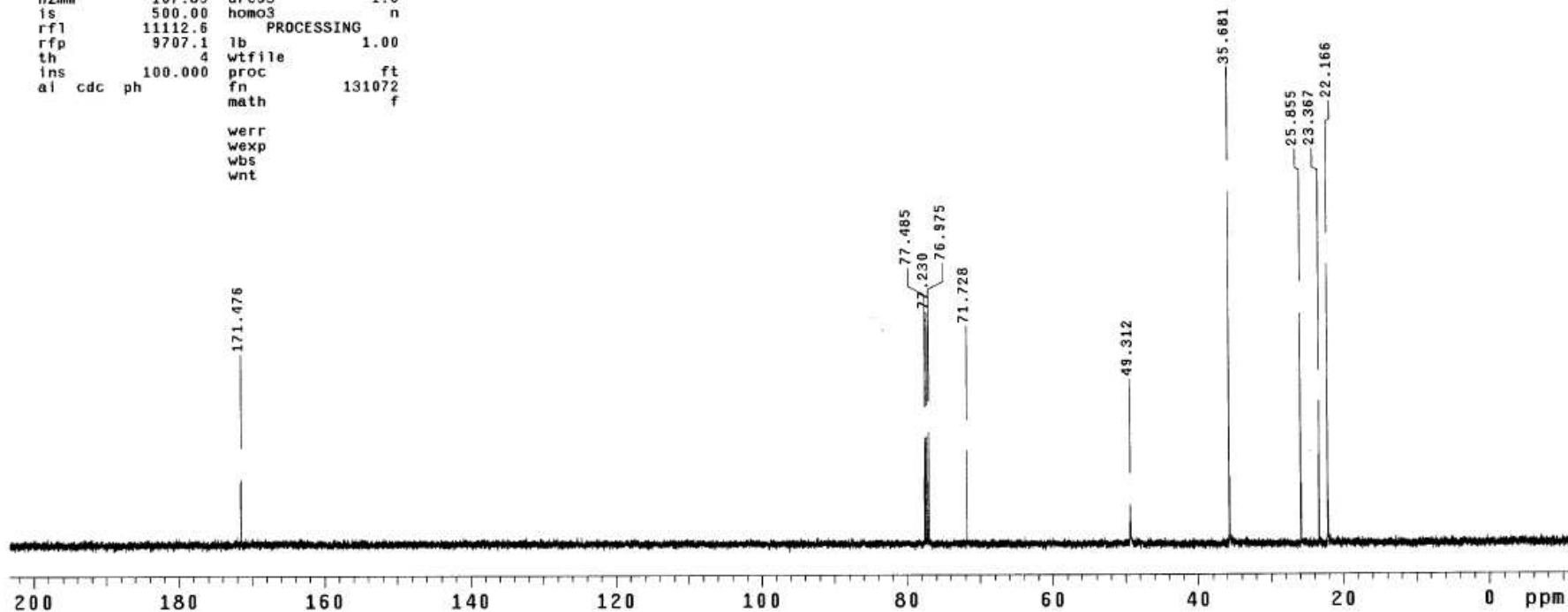
```
SAMPLE          DEC. & VT
date Jul 2 2009  dfrq          499.864
solvent CDC13    dn            H1
file exp        dpwr          30
ACQUISITION     dof            0
sfrq 499.864   dm            nnn
tn    H1       dmm            c
at    5.016    dmf            200
np    65536    dseq           1.0
sw    6533.3   dres           1.0
fb    4000     homo           n
bs    4        DEC2
tpwr  61       dfrq2          0
pw    13.5     dn2            1
d1    0.100    dpwr2          0
tof   269.9    dof2           0
nt    32       dm2            n
ct    32       dmm2           c
alock n        dmf2           200
gain  not used dseq2          1.0
      FLAGS    dres2          n
      n        homo2          n
      in       DEC3
      dp       y        dfrq3          0
      hs      nn       dn3            1
      DISPLAY  dpwr3          0
      sp      -94.0   dof3           0
      wp      5090.8  dm3            n
      vs      21     dmm3           c
      sc      0      dmf3           200
      wc      250    dseq3          1.0
      hzmm    20.36  dres3          1.0
      is      46.51  homo3          n
      rfl     4140.1  PROCESSING
      rfp     3634.0  wfile
      th      7      proc           ft
      ins    2.000   fn            65536
      ai      ph     math           f
      werr
      wexp process pH
      wbs
      wnt wft
```

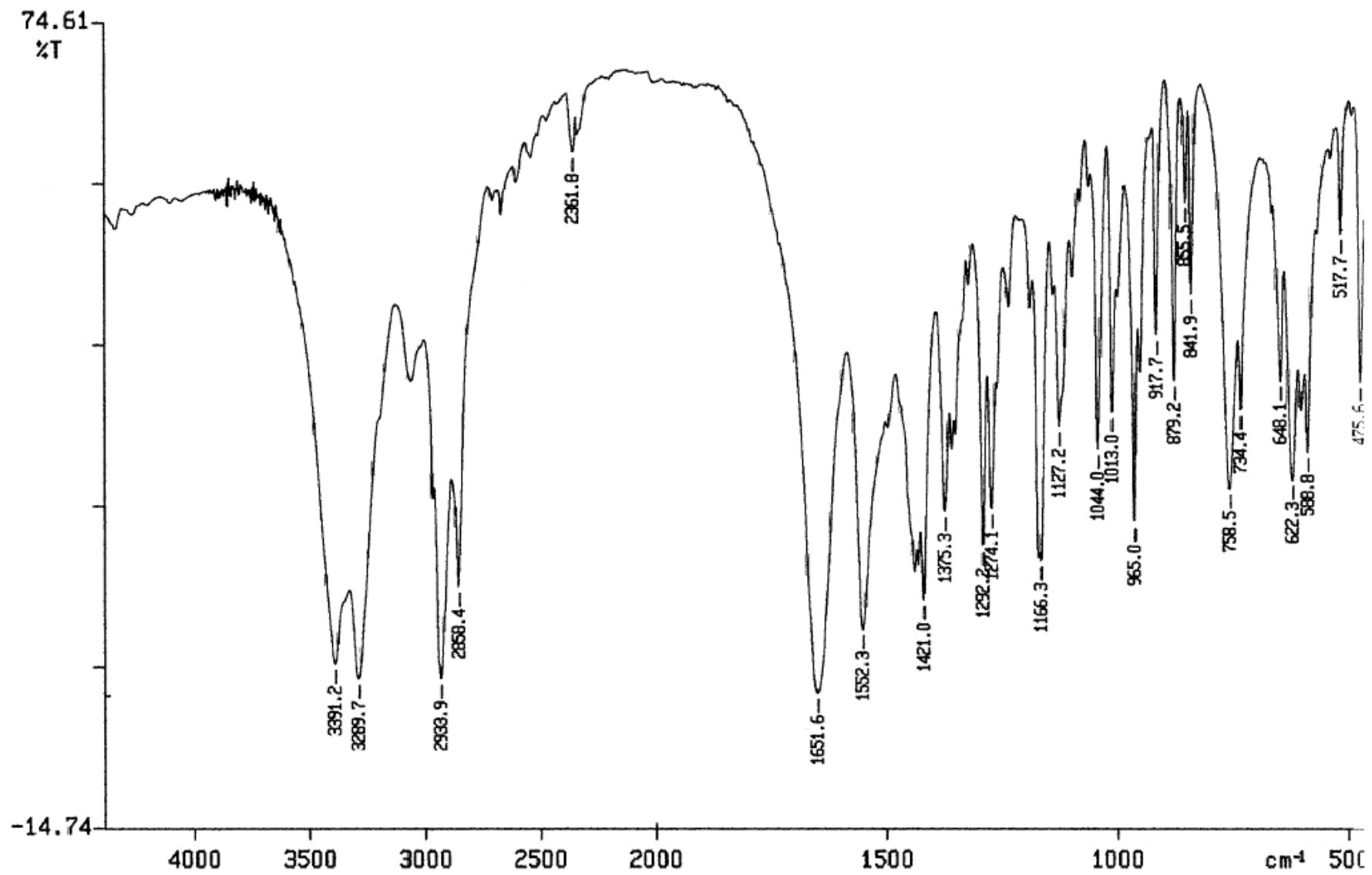


TY2-393

exp3 s2pu1

SAMPLE		DEC. & VT	
date	Jul 2 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION		dof	0
sfrq	125.702	dm	yyY
tn	C13	dmm	w
at	1.215	dmf	8787.35
np	65536	dseq	
sw	26963.3	dres	1.0
fb	15000	homo	n
bs	4	DEC2	
tpwr	52	dfrq2	0
pw	10.2	dn2	
d1	1.800	dpwr2	1
tof	144.5	dof2	0
nt	1000	dm2	n
ct	52	dmm2	c
alock	not used	dmf2	10000
gain		dseq2	
FLAGS		dres2	1.0
il	n	homo2	n
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY		dpwr3	1
sp	-1405.2	dof3	0
wp	26962.9	dm3	n
vs	112	dmm3	c
sc	0	dmf3	10000
wc	250	dseq3	
hzmm	107.85	dres3	1.0
is	500.00	homo3	n
rf1	11112.6	PROCESSING	
rfp	9707.1	lb	1.00
th	4	wfile	
ins	100.000	proc	ft
ai cdc ph		fn	131072
		math	f





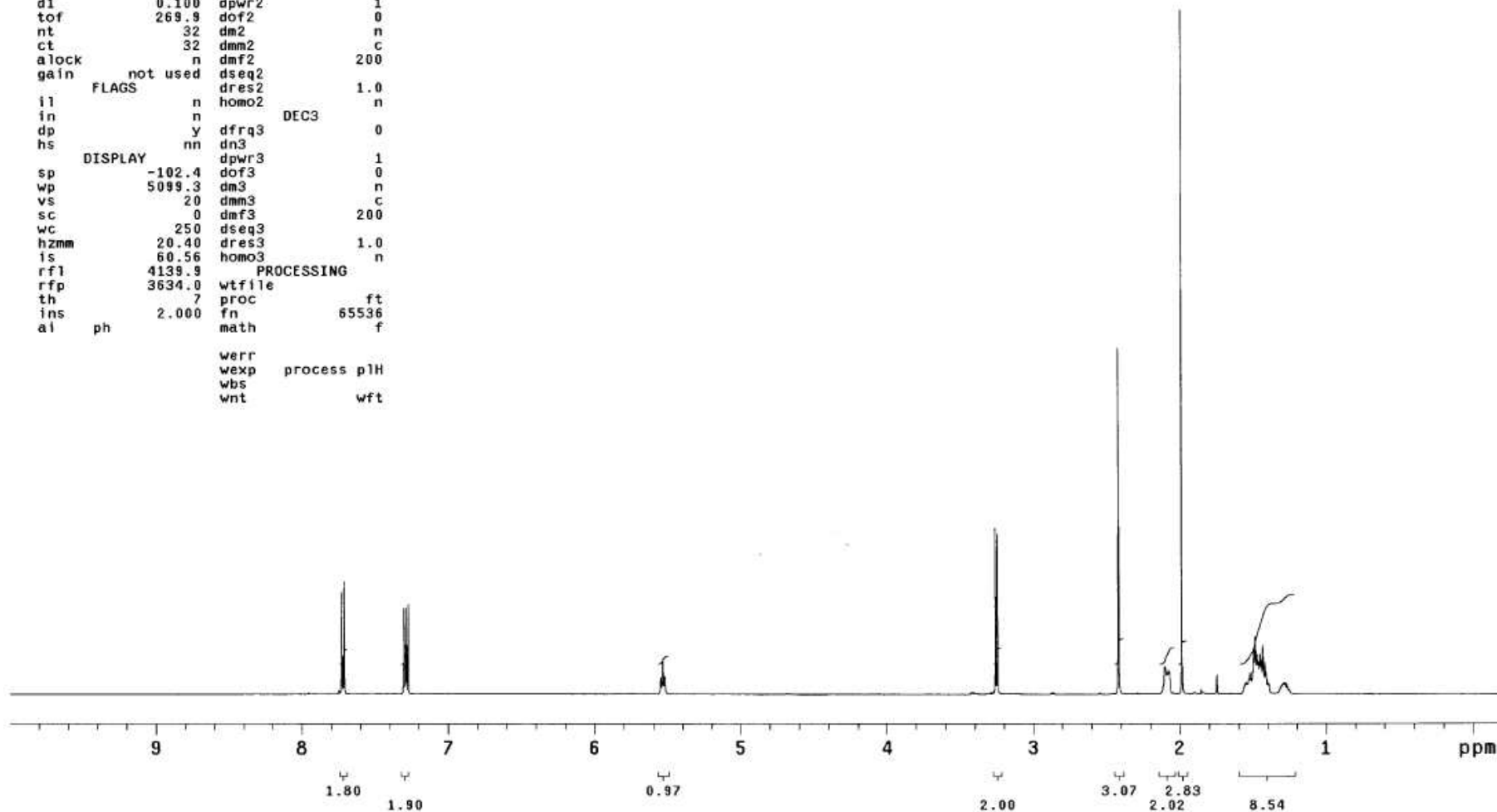
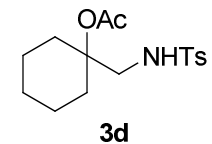
TY2-398-c2

exp2 s2pu1

```

SAMPLE          DEC. & VT
date    Jul 7 2009  dfrq    499.864
solvent  CDC13      dn      H1
file     exp        dpwr    30
ACQUISITION      dof      0
sfrq     499.864   dm      nnn
tn       H1       dmm      c
at       5.016    dmf     200
np       65536    dseq
sw       6533.3   dres    1.0
fb       4000    homo    n
bs       4        DEC2
tpwr     61      dfrq2   0
pw      13.5    dn2
d1       0.100  dpwr2   1
tof      269.9  dof2    0
nt       32     dm2     n
ct       32     dmm2    c
alock    n      dmf2    200
gain     not used dseq2
FLAGS    n      dres2   1.0
in       n      homo2   n
dp       y      dfrq3   0
hs       nn     dn3
DISPLAY  dpwr3   1
sp      -102.4  dof3    0
wp      5099.3  dm3     n
vs      20     dmm3    c
sc      0      dmf3    200
wc      250    dseq3
hzmm    20.40  dres3   1.0
is      60.56  homo3   n
rf1     4139.9 PROCESSING
rfp     3634.0 wtfille
th      7      proc    ft
ins     2.000  fn      65536
al      ph     math    f
werr
wexp   process pH
wbs
wnt    wft

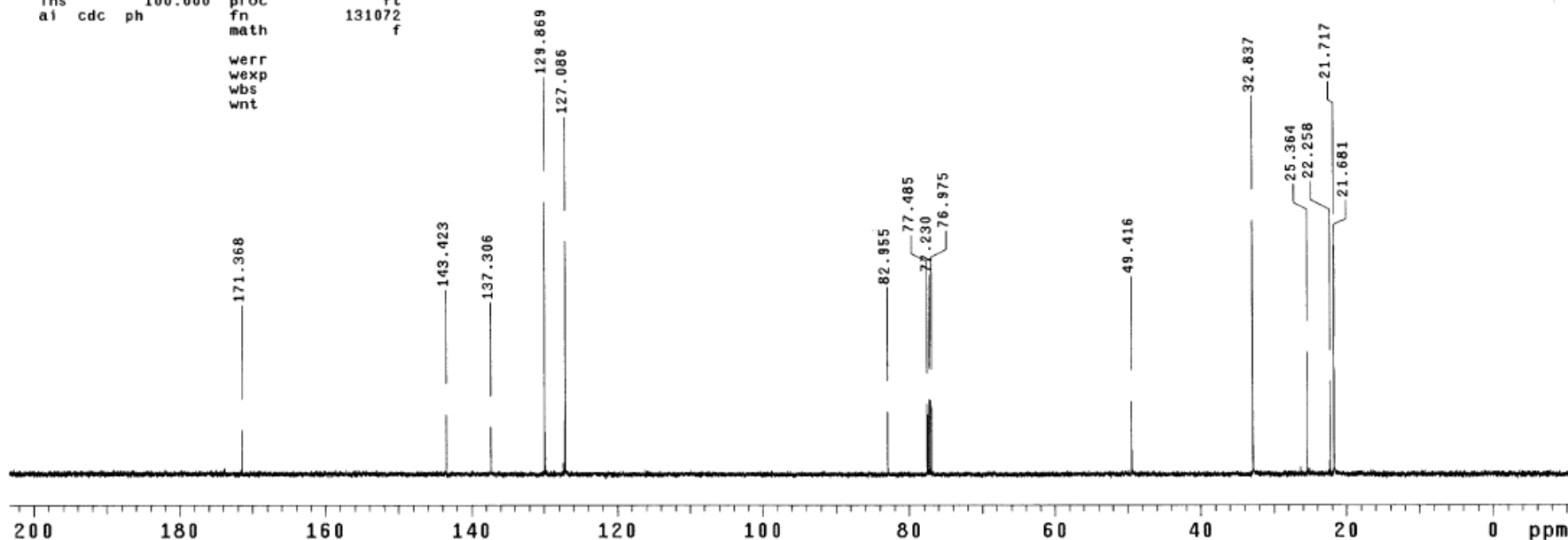
```



TY2-398-c2

exp3 s2pu1

SAMPLE		DEC. & VT	
date	Jul 7 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION		dof	0
sfrq	125.702	dm	yyy
tn	C13	dmm	w
at	1.215	dmf	8787.35
np	65536	dseq	
sw	26963.3	dres	1.0
fb	15000	homo	n
bs	4	DEC2	
tpwr	52	dfrq2	0
pw	10.2	dn2	
d1	1.800	dpwr2	1
tof	144.5	dof2	0
nt	1000	dm2	n
ct	60	dmm2	c
alock	n	dmf2	10000
gain	not used	dseq2	
FLAGS		dres2	1.0
il	n	homo2	n
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY		dpwr3	1
sp	-1406.4	dof3	0
wp	26962.9	dm3	n
vs	70	dmm3	c
sc	0	dmf3	10000
wc	250	dseq3	
hzmm	107.85	dres3	1.0
is	500.00	homo3	n
rf1	11113.9	PROCESSING	
rfp	9707.1	lb	1.00
th	5	wtfile	
ins	100.000	proc	ft
al	cdc ph	fn	131072
		math	f

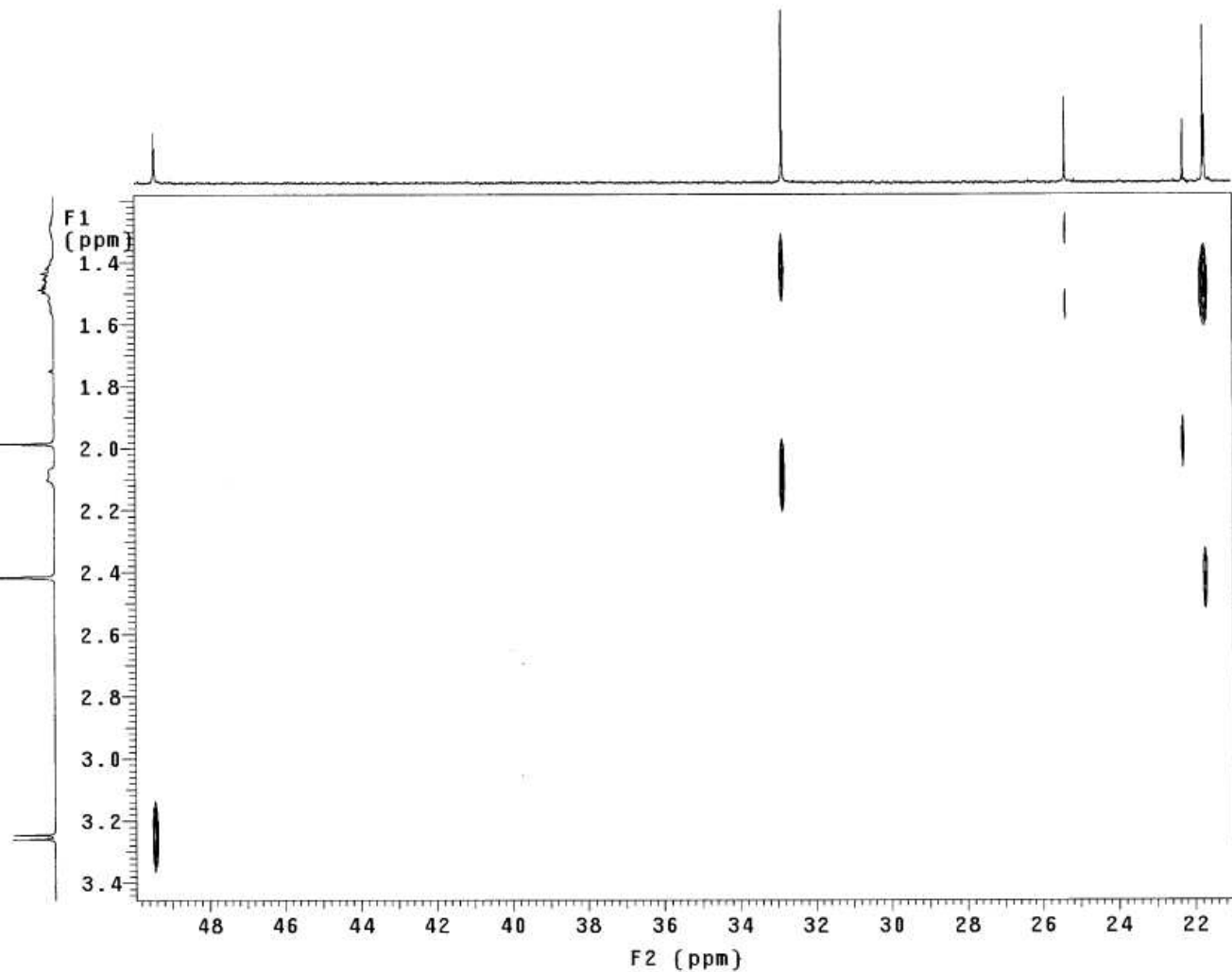


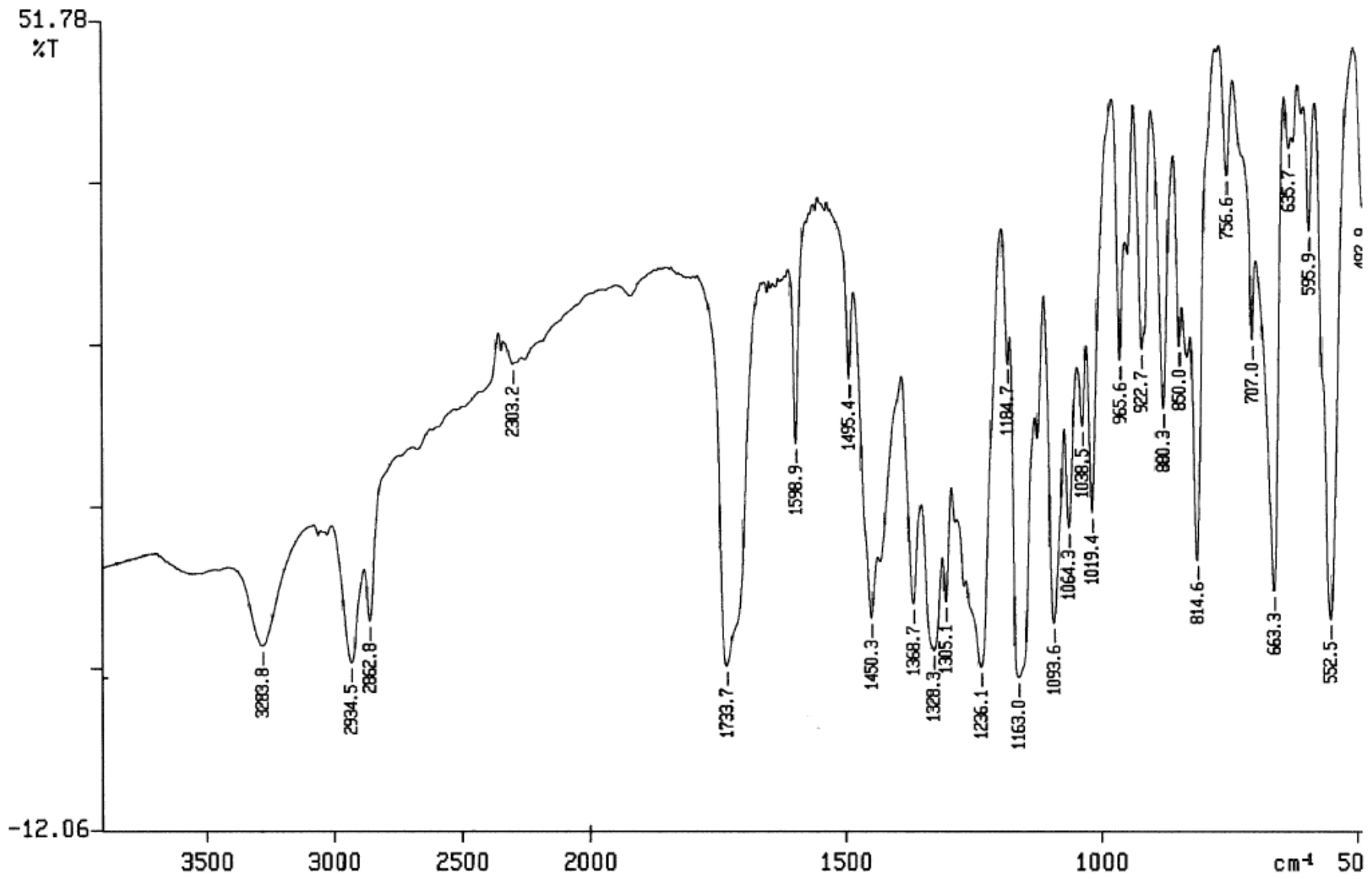
TY2-398-c2

Pulse Sequence: hetcor

Solvent: CDC13  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE C13, 125.6901710 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 3 hr, 46 min, 5 sec



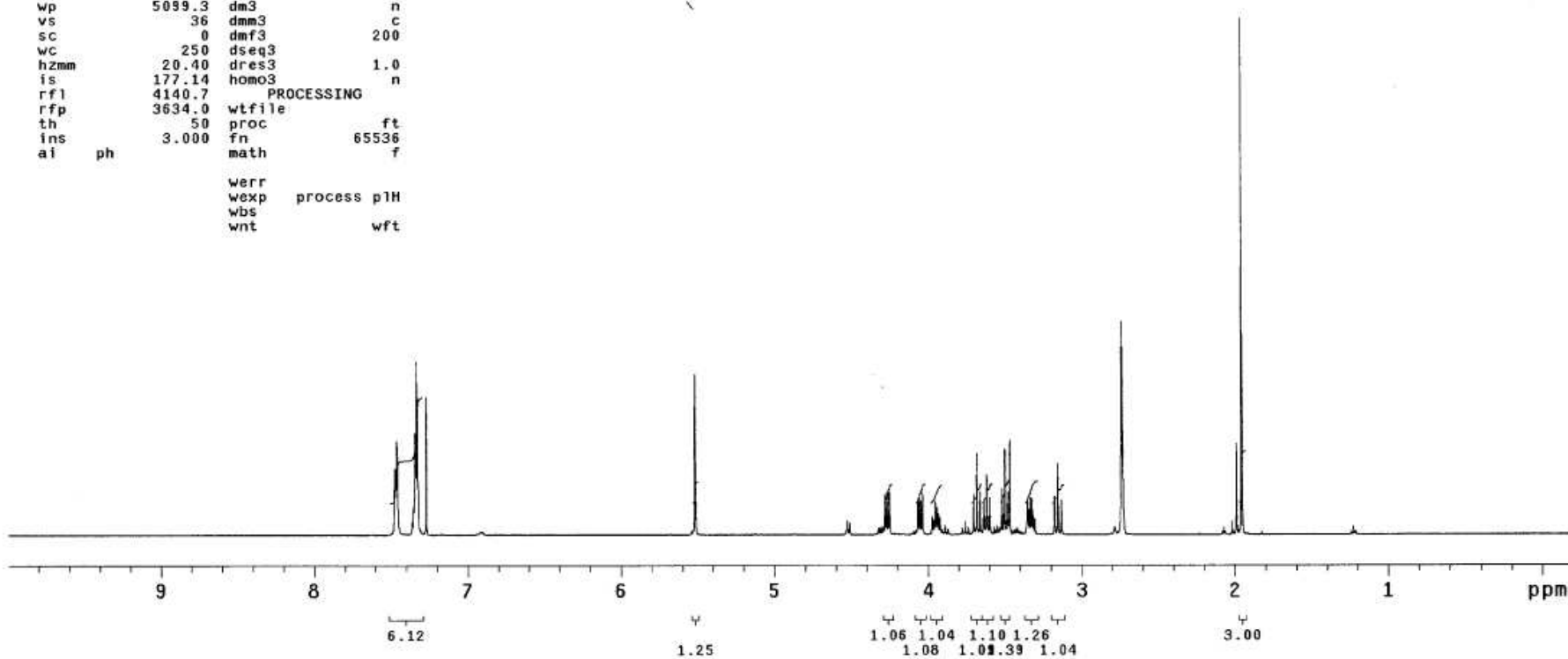
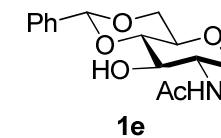




TY2-500

exp2 s2pu1

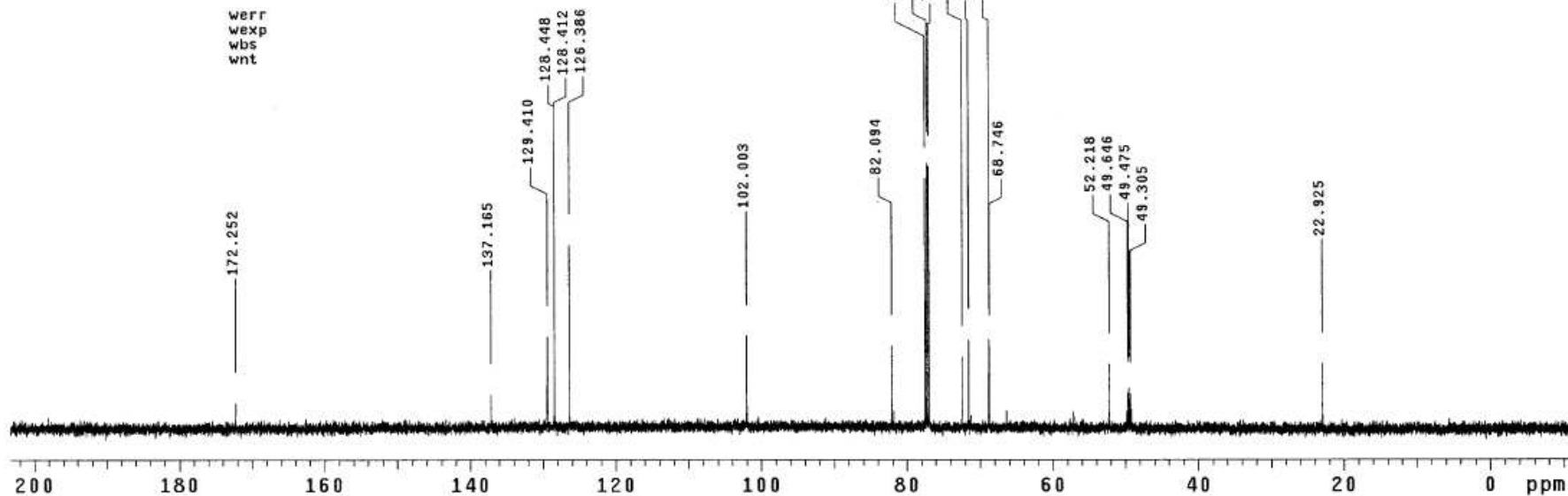
date	Jan 4 2010	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	30
ACQUISITION			
sfrq	499.864	dm	nnn
tn	H1	dmm	c
at	5.016	dmf	200
np	65536	dseq	
sw	6533.3	dres	1.0
fb	4000	homo	n
bs	4	DEC2	
tpwr	61	dfrq2	0
pw	13.5	dn2	
d1	0.100	dpwr2	1
tof	269.9	dof2	0
nt	16	dm2	n
ct	16	dmm2	c
alock	n	dmf2	200
gain	not used	dseq2	
il	FLAGS	dres2	1.0
ln	n	homo2	n
dp	n	DEC3	
hs	nn	dfrq3	0
DISPLAY			
sp	-103.2	dn3	
wp	5099.3	dpwr3	1
vs	36	dof3	0
sc	0	dm3	n
wc	250	dmm3	c
h2mm	20.40	dmf3	200
is	177.14	dseq3	
rfl	4140.7	dres3	1.0
rfp	3634.0	homo3	n
th	50	PROCESSING	
ins	3.000	wtfile	ft
ai	ph	proc	65536
		fn	f
		math	
		werr	
		wexp	process p1H
		wbs	
		wnt	wft



TY2-500

exp3 s2pu1

SAMPLE		DEC. & VT	
date	Jan 4 2010	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION			
sfrq	125.702	dof	0
tn	C13	dm	yyy
at	1.215	dmm	w
np	65536	dof2	8787.35
sw	26963.3	dm2	
fb	15000	dmm2	
bs	4	dof3	1.0
tpwr	52	dmf	n
pw	10.2	homo	DEC2
d1	1.800	dfrq2	0
tof	144.5	dn2	
nt	2000	dpwr2	1
ct	182	dof2	0
alock	n	dm2	n
gain	not used	dmm2	c
FLAGS			
il	n	dmf2	10000
in	n	dseq2	1.0
dp	y	dres2	1.0
hs	nn	homo2	n
DISPLAY			
sp	-1407.2	dfrq3	DEC3
wp	26962.9	dn3	0
vs	313	dpwr3	1
sc	0	dof3	0
wc	250	dm3	n
hzmm	107.85	dmm3	c
is	500.00	dmf3	10000
rfl	11114.7	dseq3	1.0
rpf	9707.1	dres3	1.0
th	3	homo3	n
ins	100.000	PROCESSING	
ai	cdc ph	lb	1.00
		wtfile	
		proc	ft
		fn	131072
		math	f

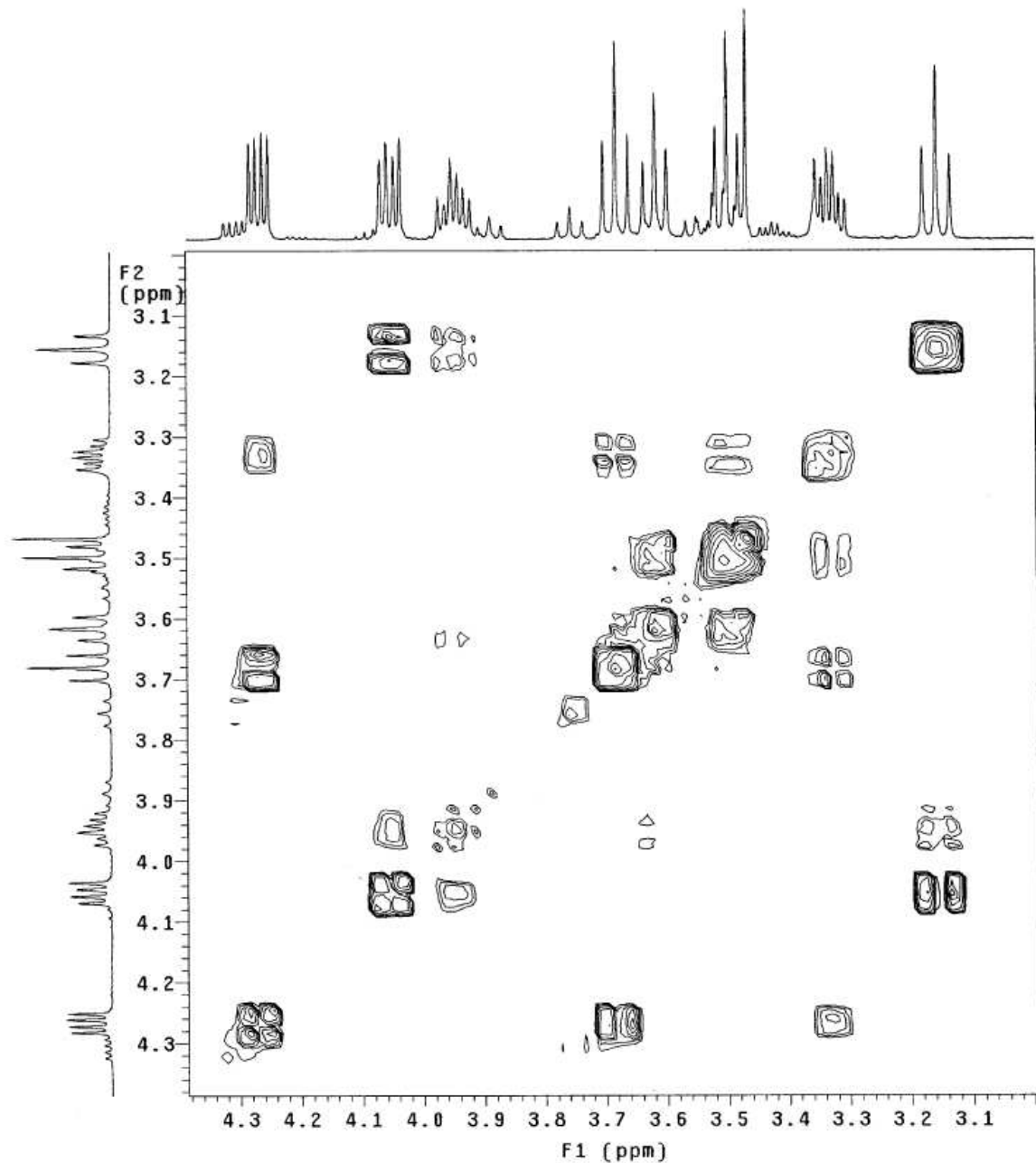


TY2-500

Pulse Sequence: relayh

Solvent: CDCl3  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE H1, 499.8611713 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 3 hr, 23 min, 10 sec

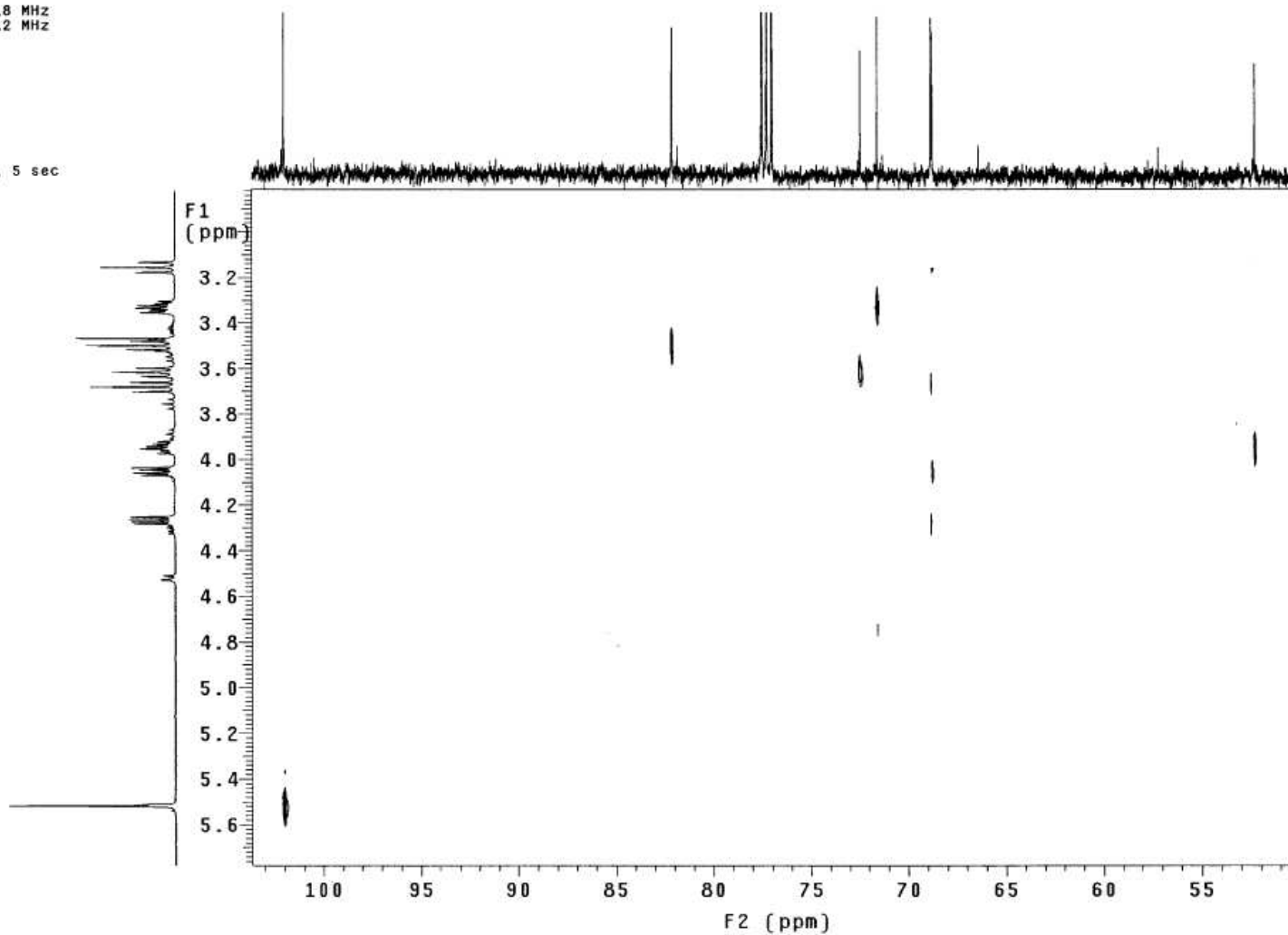


TY2-500

Pulse Sequence: hetcor

Solvent: CDC13  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18409.5 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE C13, 125.6901718 MHz  
DECOUPLE H1, 499.8699312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 3 hr, 46 min, 5 sec



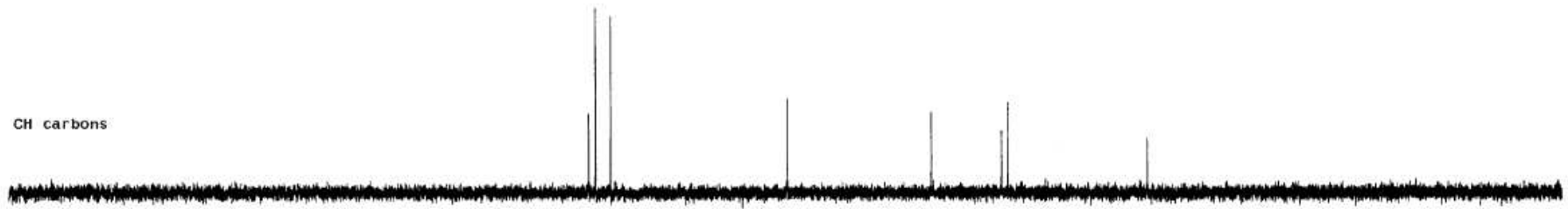
CH3 carbons



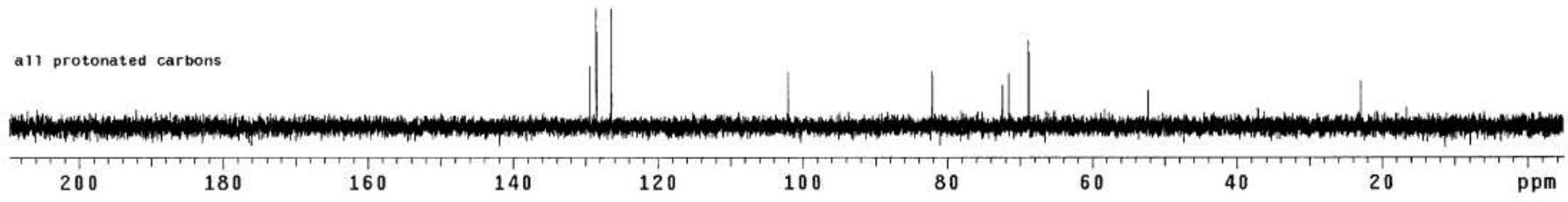
CH2 carbons



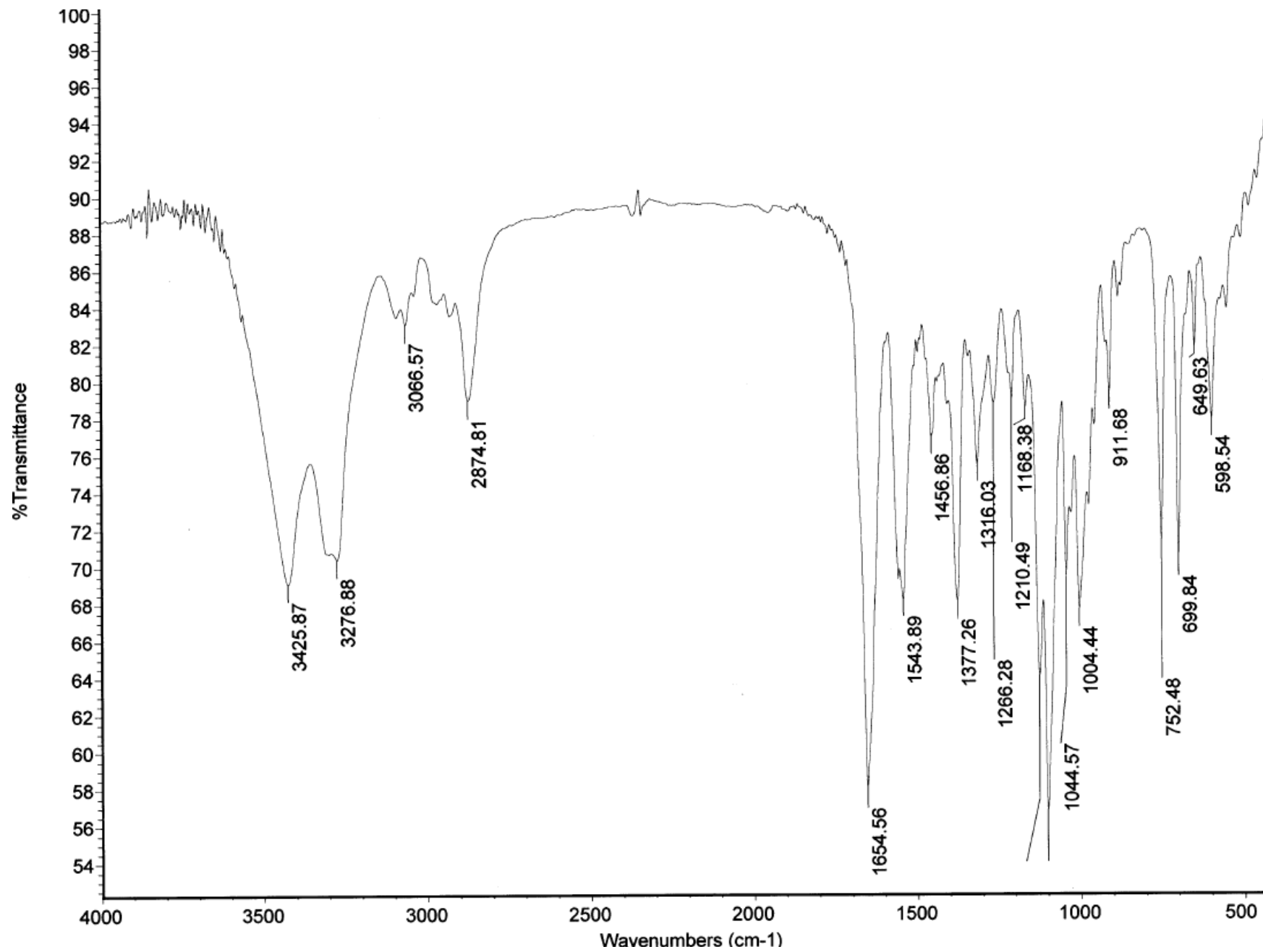
CH carbons



all protonated carbons



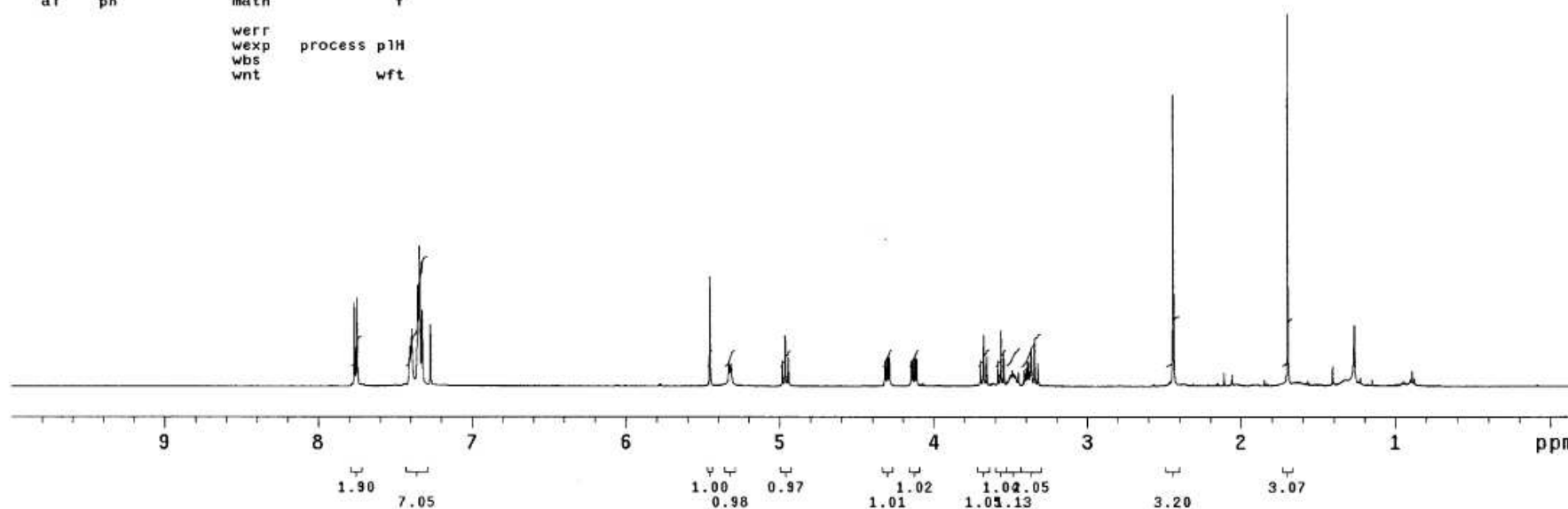
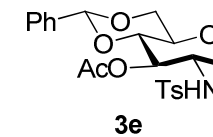
200 180 160 140 120 100 80 60 40 20 ppm



TY2-501

exp2 s2pu1

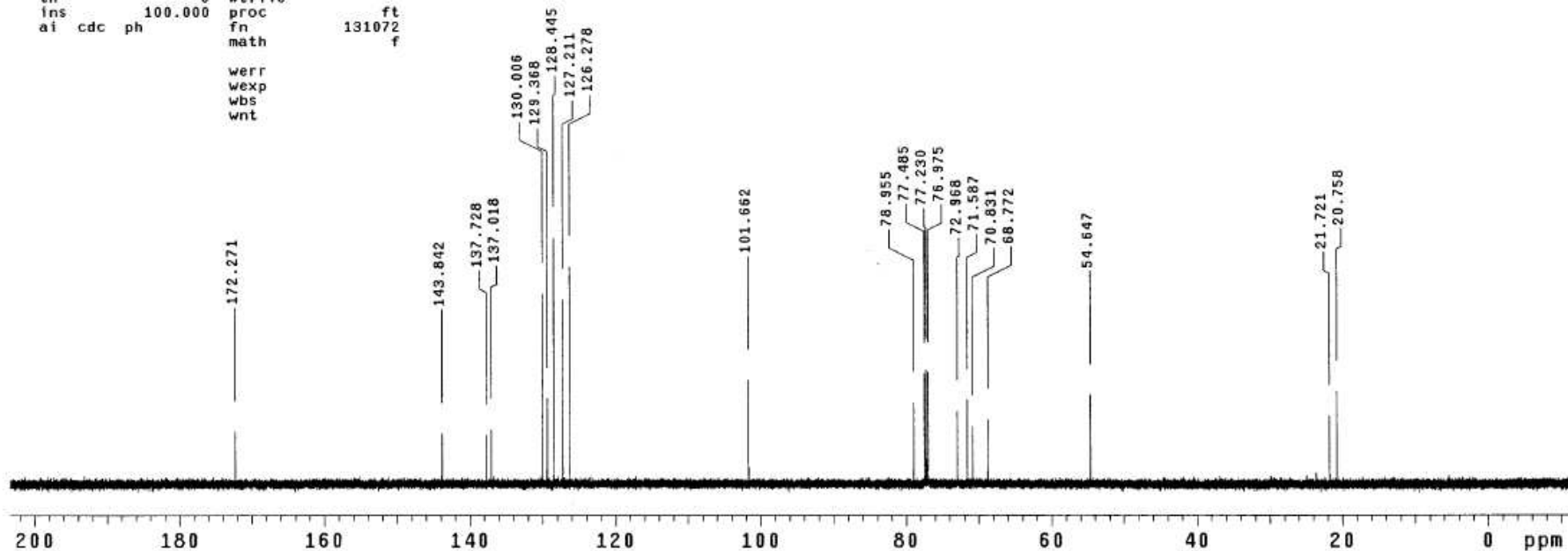
```
SAMPLE          DEC. & VT
date    Dec 23 2009    dfrq          499.864
solvent  CDC13         dn           H1
file     exp          dpwr          30
ACQUISITION
sfrq     499.864      dm           nnn
tn        H1         dmm          c
at        5.016      dmf          200
np        65536      dseq         1.0
sw        6533.3     dres         n
fb        4000       homo
bs         4         DEC2
tpwr      61         dfrq2        0
pw        13.5       dn2
d1        0.100      dpwr2        1
tof       269.9     dof2         0
nt        16        dm2          n
ct        16        dmm2         c
alock     n         dmf2         200
gain      not used   dseq2        1.0
FLAGS     n         homo2        n
il        n         DEC3
in        y         dfrq3        0
dp        nn        dn3
hs        nn        dpwr3        1
DISPLAY
sp       -102.8     dof3         0
wp       5099.3    dm3          n
vs        22      dmm3         c
sc         0      dmf3         200
wc        250     dseq3        1.0
hzmm     20.40    dres3        n
is       127.97   homo3
rfl      4140.3   PROCESSING
rfp      3634.0  wfile
th        7      proc         ft
ins      1.000   fn          65536
ai       ph     math         f
werr
wexp    process pH
wbs
wnt     wft
```



TY2-501

exp3 s2pu1

SAMPLE		DEC. & VT	
date	Dec 23 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION			
sfrq	125.702	dof	0
tn	C13	dm	yy
at	1.215	dmm	w
np	65536	dmf	8787.35
sw	26963.3	dseq	
fb	15000	dres	1.0
bs	4	homo	n
tpwr	52	DEC2	
pw	10.2	dfrq2	0
d1	1.800	dn2	
tof	144.5	dpwr2	1
nt	2000	dof2	0
ct	164	dm2	n
alock	n	dmm2	c
gain	not used	dmf2	10000
FLAGS			
il	n	dseq2	1.0
in	n	dres2	1.0
dp	y	homo2	n
hs	nn	DEC3	
DISPLAY			
sp	-1403.5	dfrq3	0
wp	26962.9	dn3	
vs	85	dpwr3	1
sc	0	dof3	0
wc	250	dm3	n
hzmm	107.85	dmm3	c
is	500.00	dmf3	10000
rfl	11111.0	dseq3	
rfp	9707.1	dres3	1.0
th	6	homo3	n
lms	100.000	PROCESSING	
ai	cdc ph	lb	not used
		wfile	
		proc	ft
		fn	131072
		math	f
		werr	
		wexp	
		wbs	
		wnt	

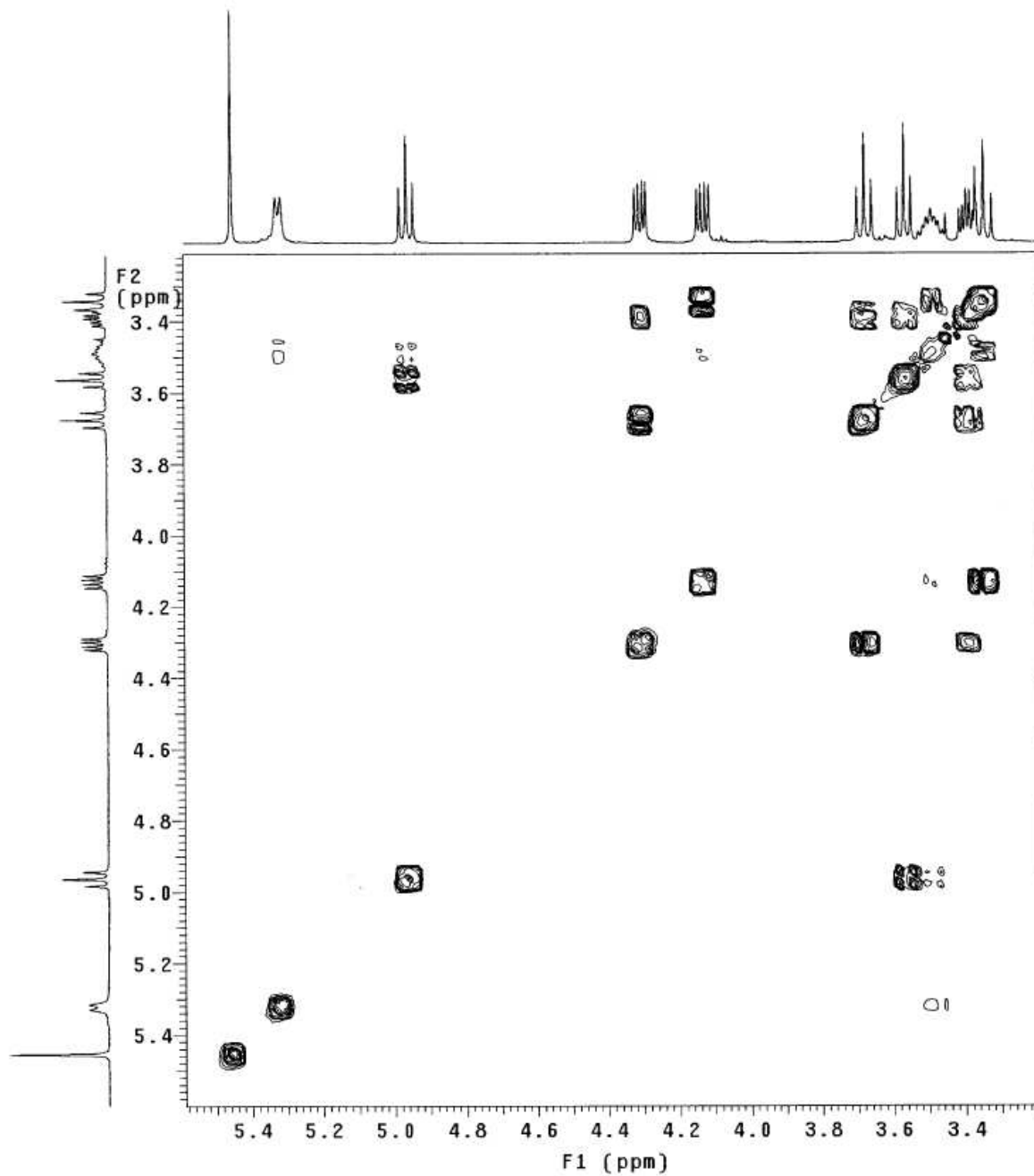




TY2-501

Pulse Sequence: relayh  
Solvent: CDC13  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE H1, 499.8611709 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 3 hr, 23 min, 10 sec

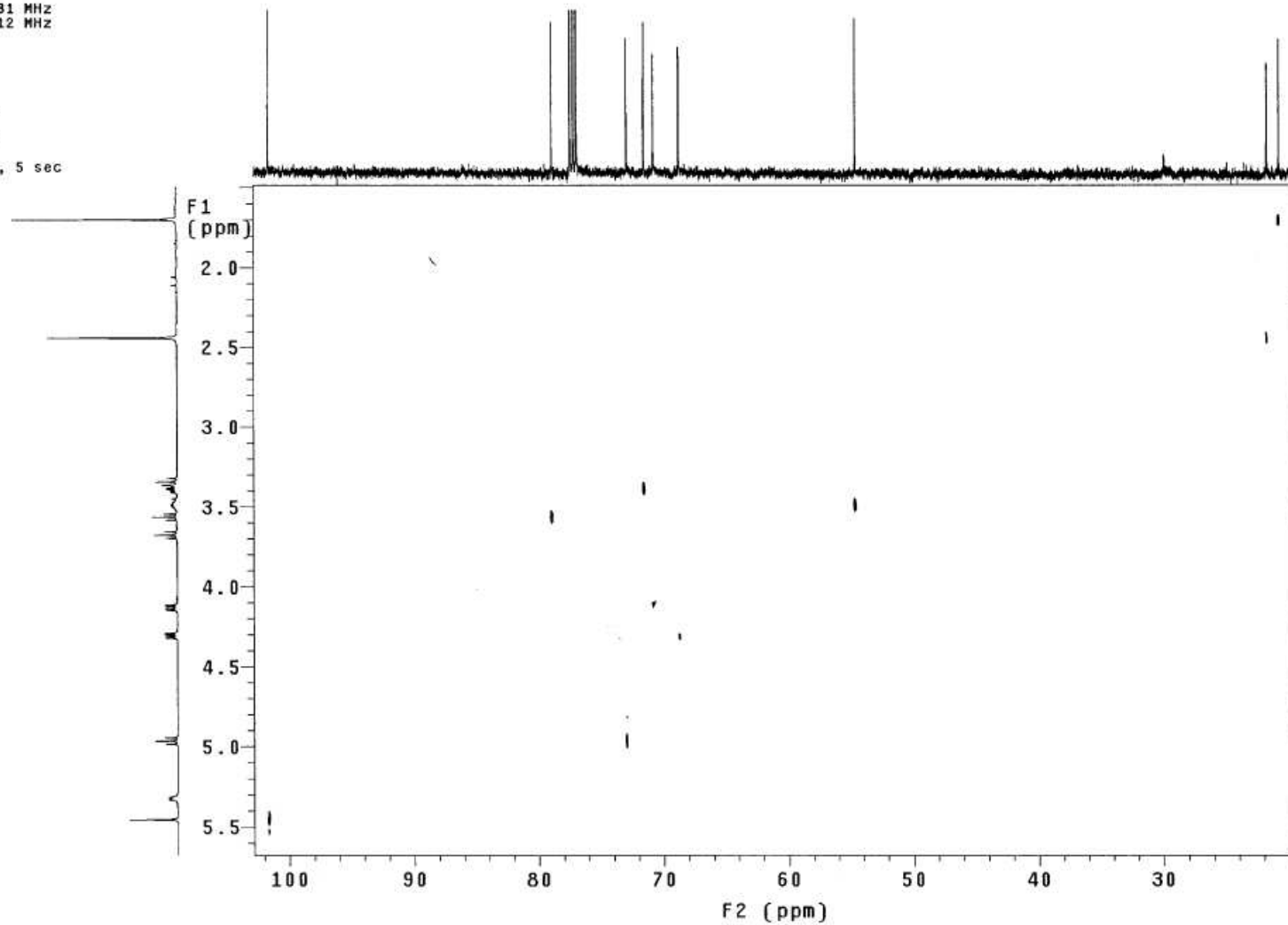


TY2-501

Pulse Sequence: hetcor

Solvent: CDCl3  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE C13, 125.6901681 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 3 hr, 46 min, 5 sec



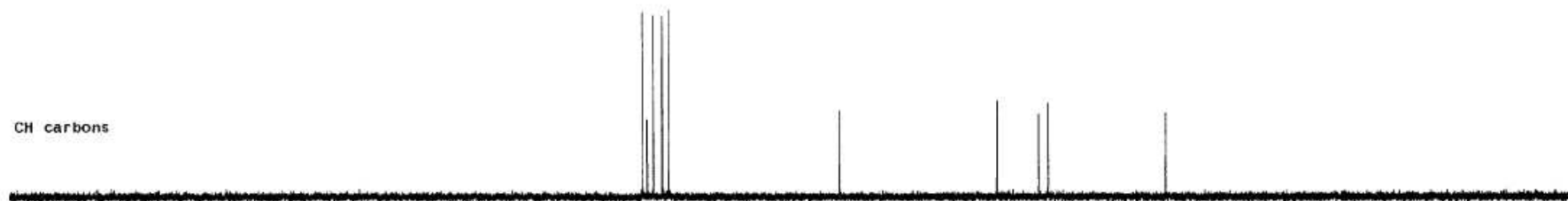
CH3 carbons



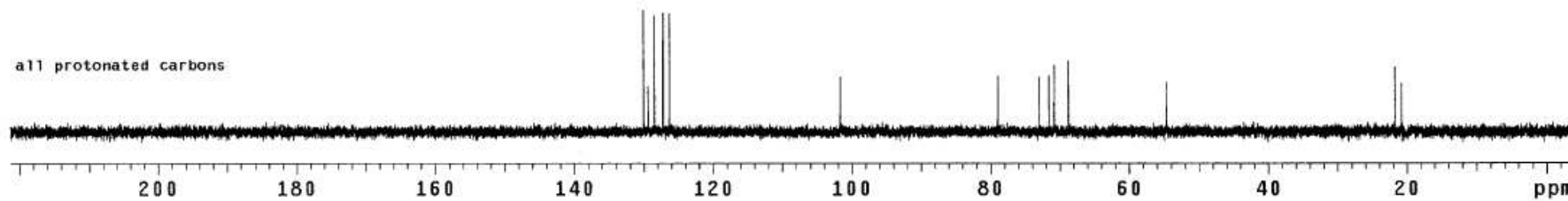
CH2 carbons

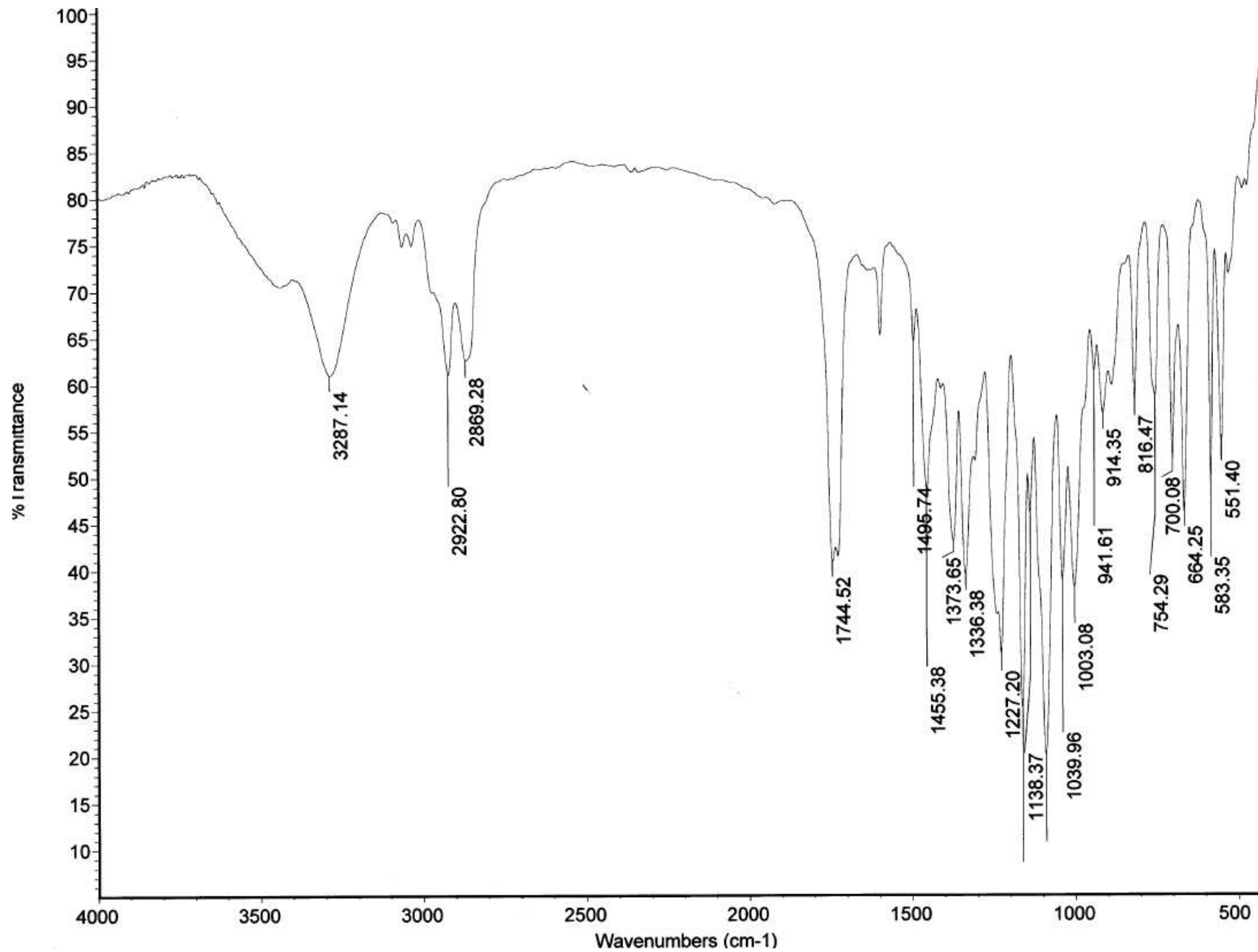


CH carbons



all protonated carbons

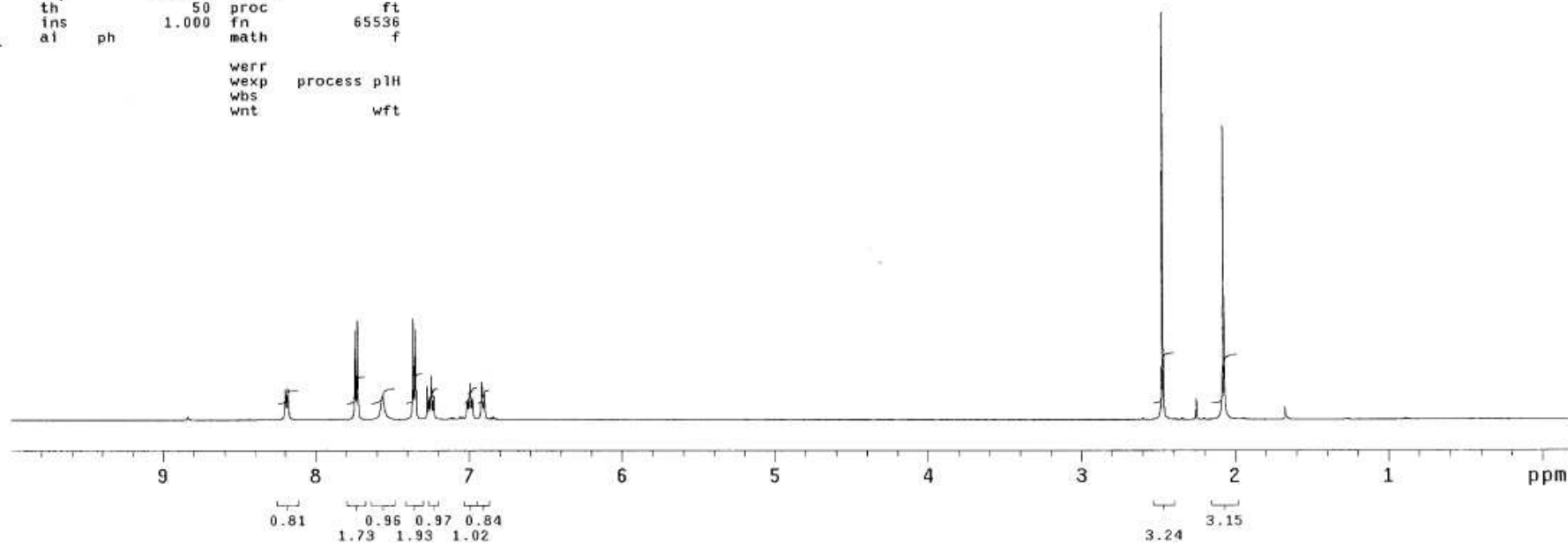
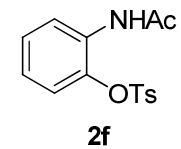




TY2-363

exp1 s2pu1

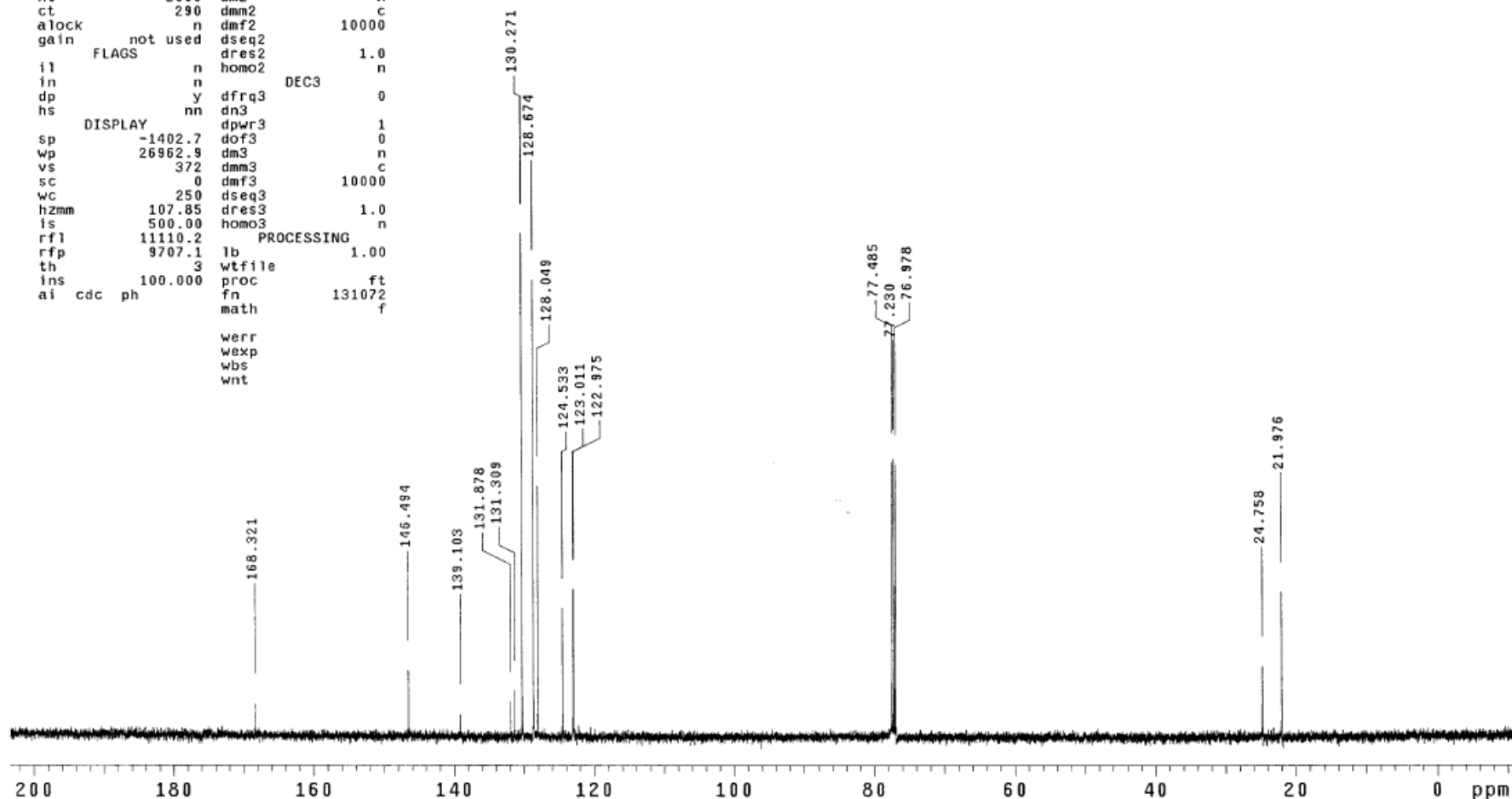
```
SAMPLE          DEC. & VT
date Jun 9 2009  dfrq          499.864
solvent CDC13    dn           H1
file exp        dpwr          30
ACQUISITION    dof           0
sfrq          499.864 dm          nnn
tn            H1    dmm          c
at            5.016 dmf          200
np            65536 dseq         1.0
sw            6533.3 dres        n
fb            4000  homo
bs            4      DEC2
tpwr          61    dfrq2        0
pw            13.5 dn2
dl            0.100 dpwr2        1
tof           269.9 dof2         0
nt            32    dm2          n
ct            32    dmm2         C
alock         n    dmf2          200
gain          not used dseq2       1.0
FLAGS        n    homo2        n
il            n      DEC3
in            n    dfrq3        0
dp            y    dn3
hs            nn   dpwr3          1
DISPLAY      dof3          0
sp            -102.6 dm3          n
wp            5099.3 dmm3         C
vs            34    dmf3          200
sc            0    dseq3         1.0
wc            250  dres3        n
hzmm          20.40 homo3
is            123.23
rf1           4140.1 PROCESSING
rfp           3634.0 wtf file
th            50    proc
ins           1.000 fn           65536
ai ph         math          f
werr
wexp process pH
wbs
wnt wft
```



TY2-363

exp2 s2pu1

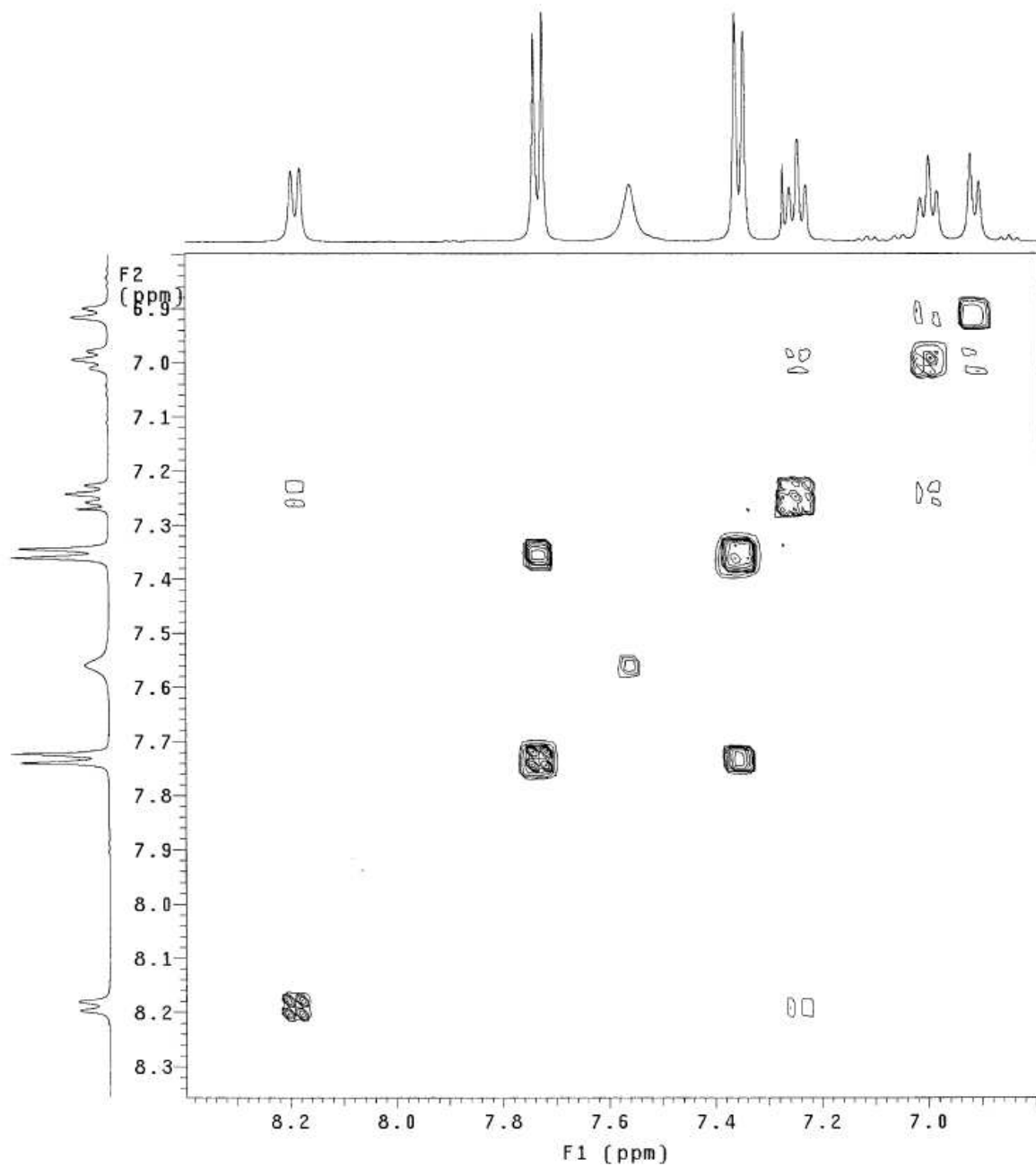
SAMPLE		DEC. & VT	
date	Jun 9 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION		dof	0
sfrq	125.702	dm	yyy
tn	C13	dmm	w
at	1.215	dmf	8787.35
np	65536	dseq	
sw	26963.3	dres	1.0
fb	15000	homo	n
bs	4	DEC2	
tpwr	52	dfrq2	0
pw	10.2	dn2	
d1	1.800	dpwr2	1
tof	144.5	dof2	0
nt	2000	dm2	n
ct	290	dmm2	c
alock	not used	dmf2	10000
gain	not used	dseq2	
FLAGS		dres2	1.0
il	n	homo2	n
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY		dpwr3	1
sp	-1402.7	dof3	0
wp	26962.9	dm3	n
vs	372	dmm3	c
sc	0	dmf3	10000
wc	250	dseq3	
hzmm	107.85	dres3	1.0
is	500.00	homo3	n
rfl	11110.2	PROCESSING	
rfp	9707.1	lb	1.00
th	3	wtfile	
ins	100.000	proc	ft
ai	cdc ph	fn	131072
		math	f



TY2-363

Pulse Sequence: relayh  
Solvent: CDC13  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

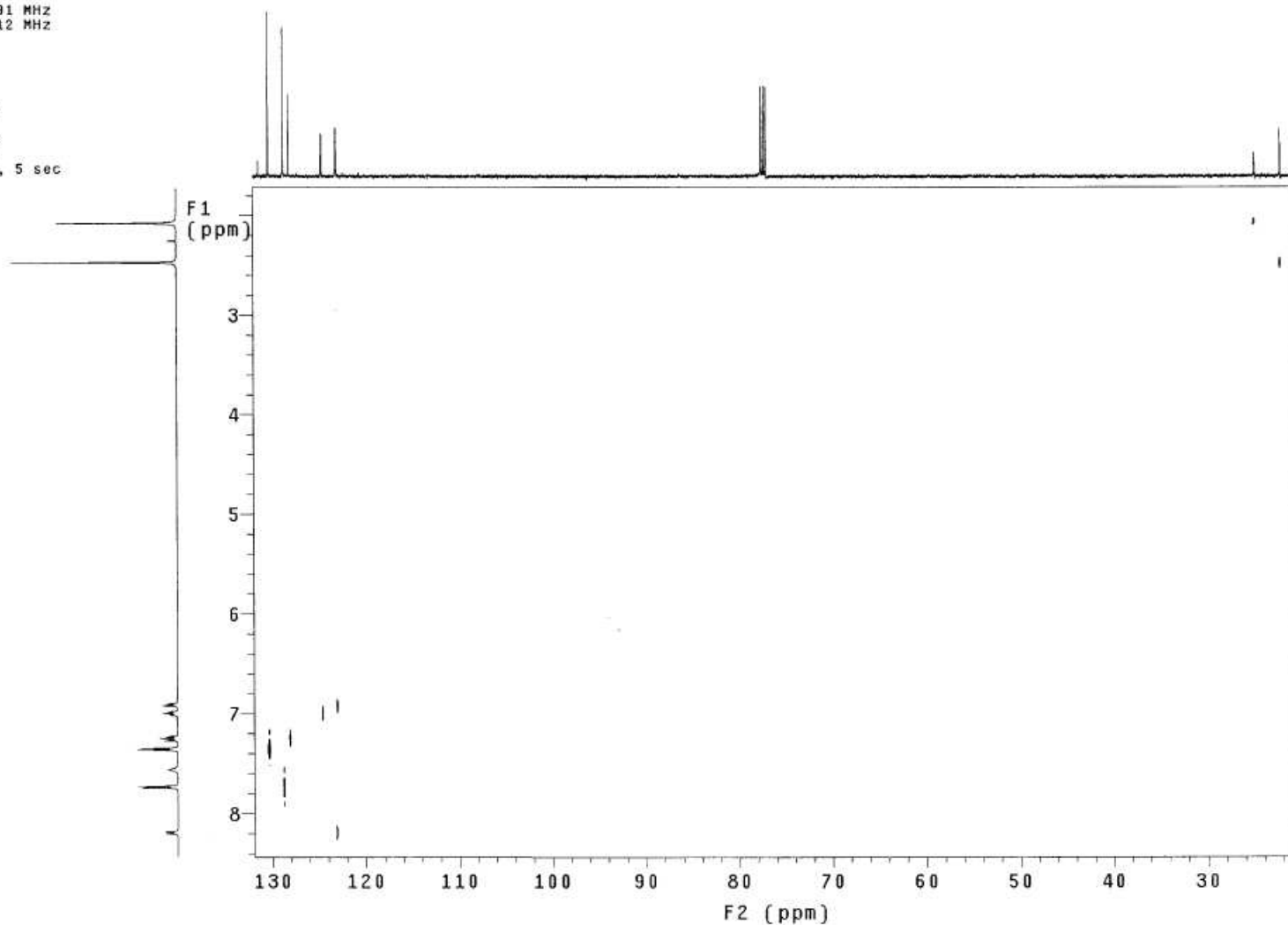
Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
32 repetitions  
258 increments  
OBSERVE H1, 499.8611707 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 3 hr, 24 min, 46 sec



TY2-363

Pulse Sequence: hetcor  
Solvent: CDC13  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE C13, 125.6901591 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 3 hr, 46 min, 5 sec



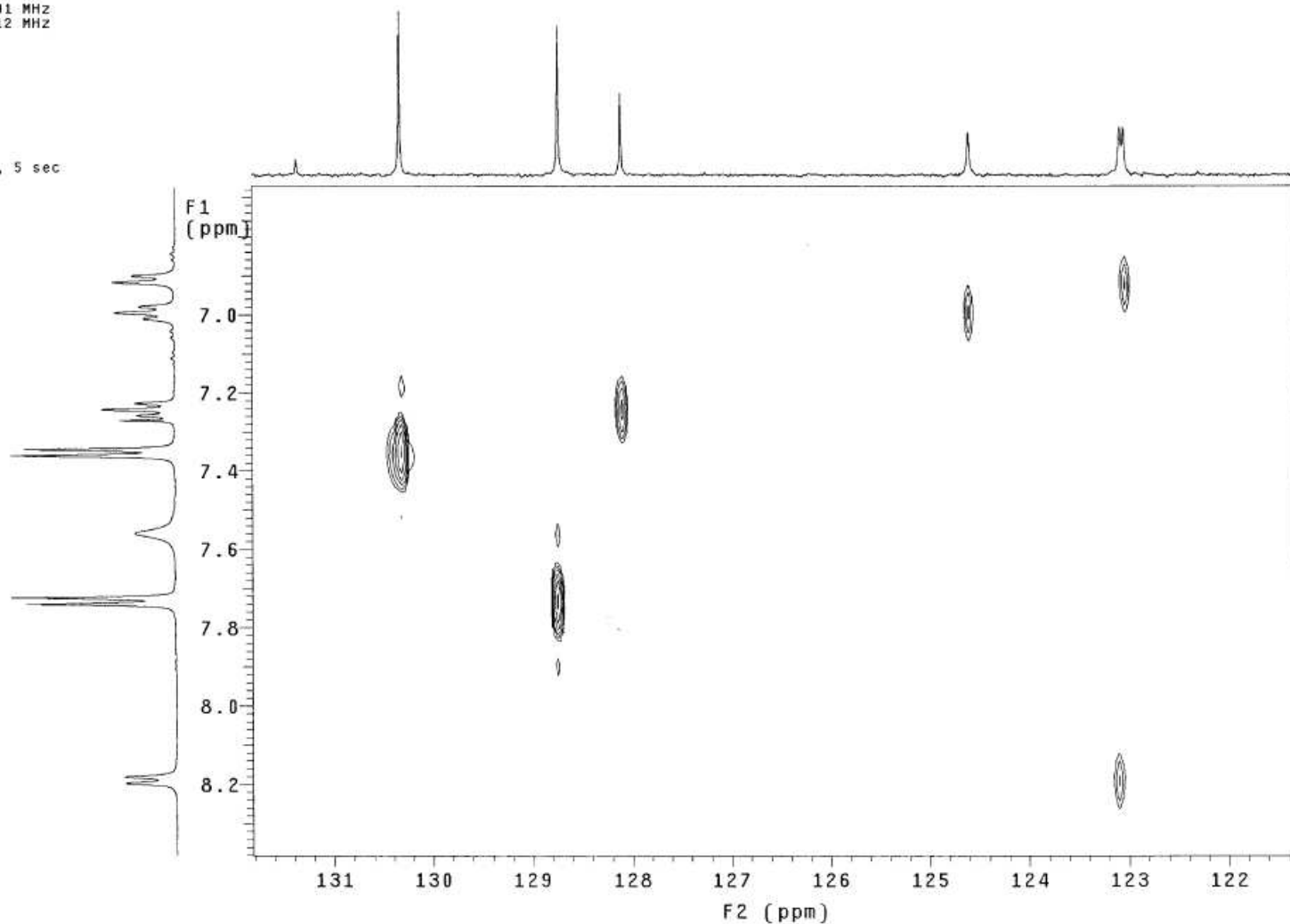


TY2-363

Pulse Sequence: hetcor

Solvent: CDC13  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE C13, 125.6901591 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 3 hr, 46 min, 5 sec



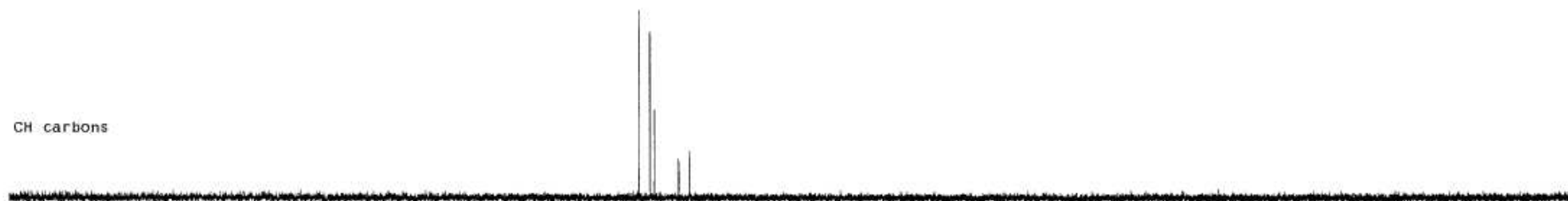
CH3 carbons



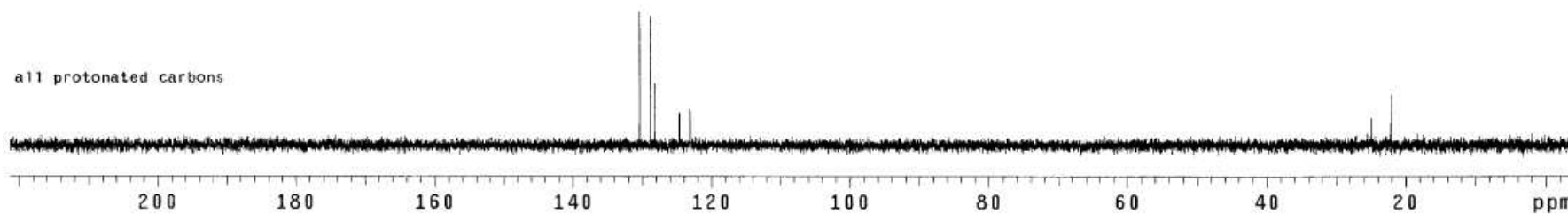
CH2 carbons

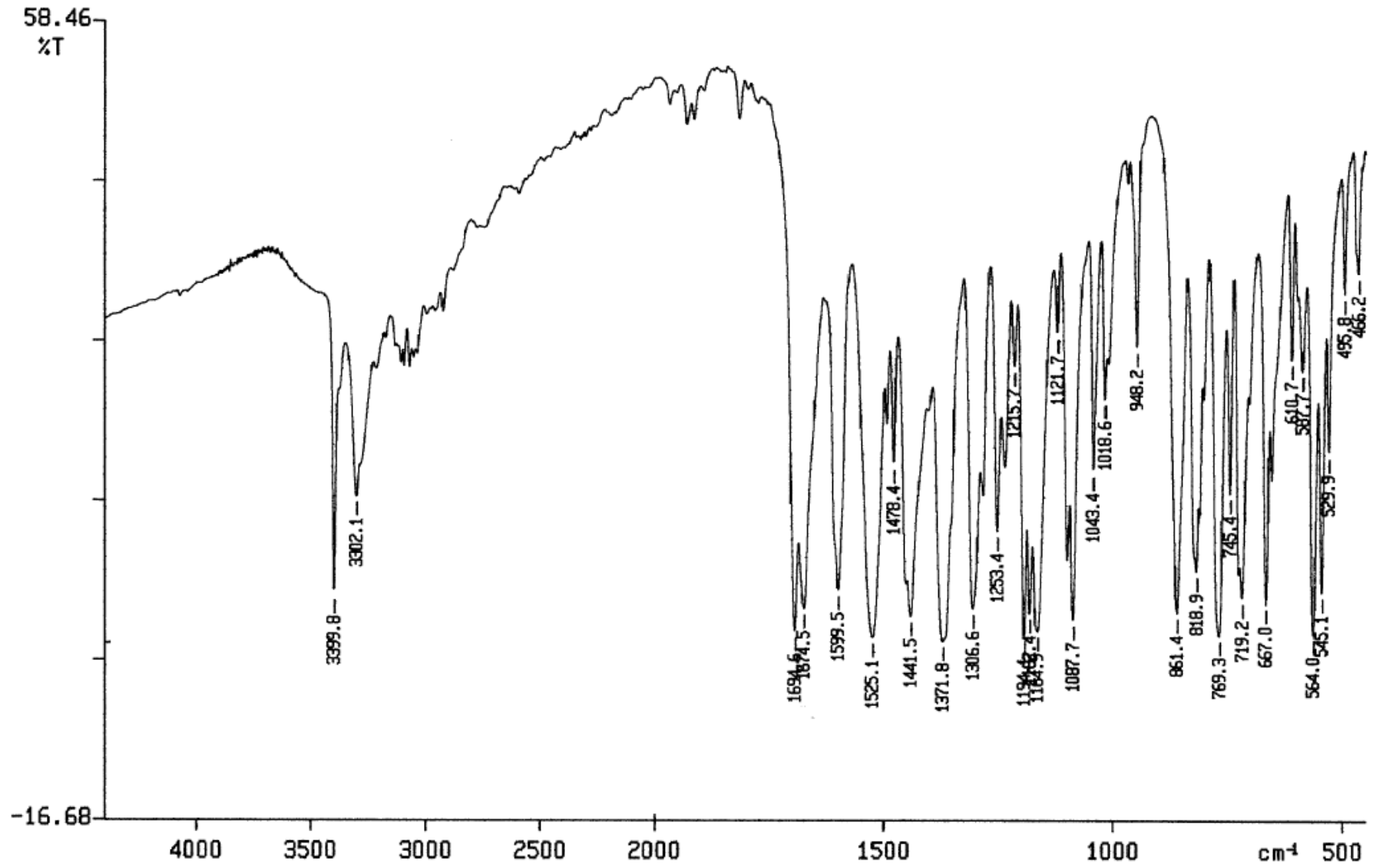


CH carbons



all protonated carbons

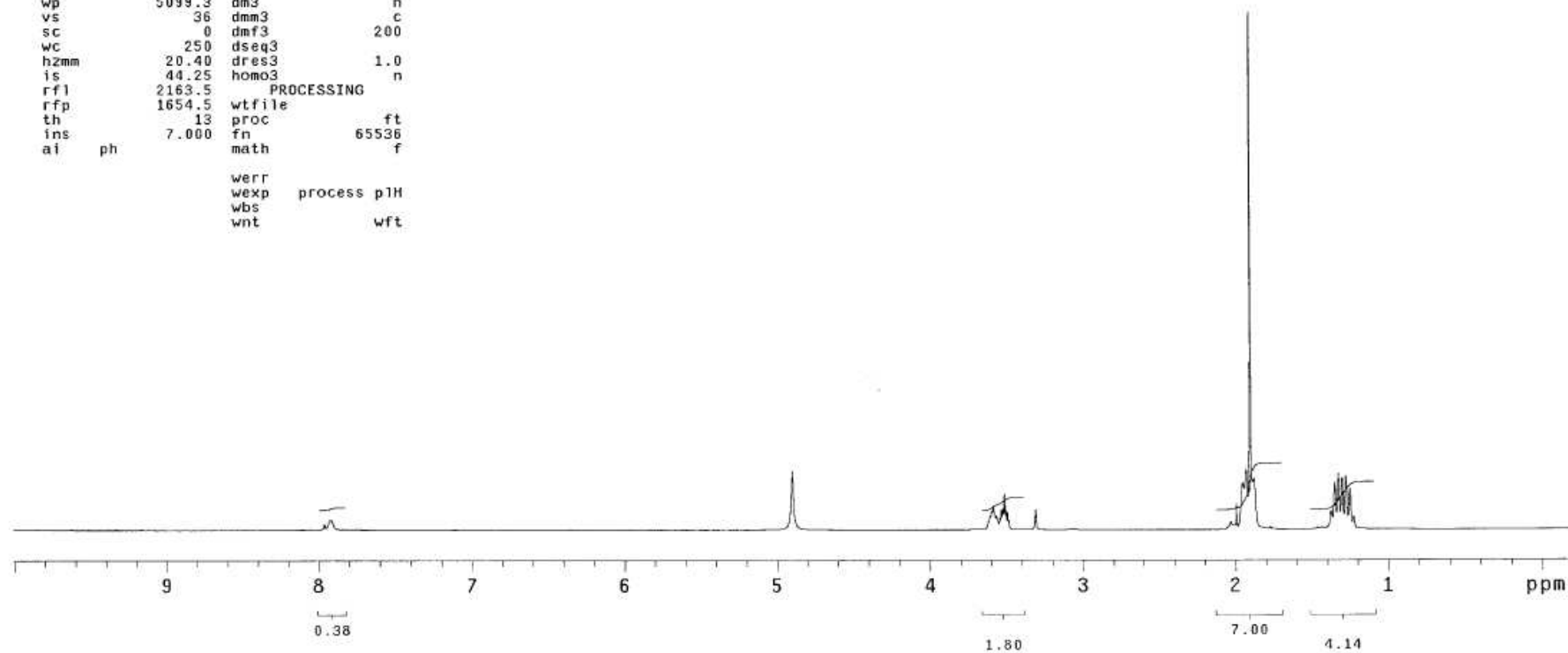
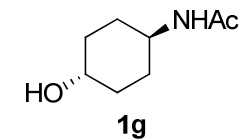




TY2-376

exp1 s2pu1

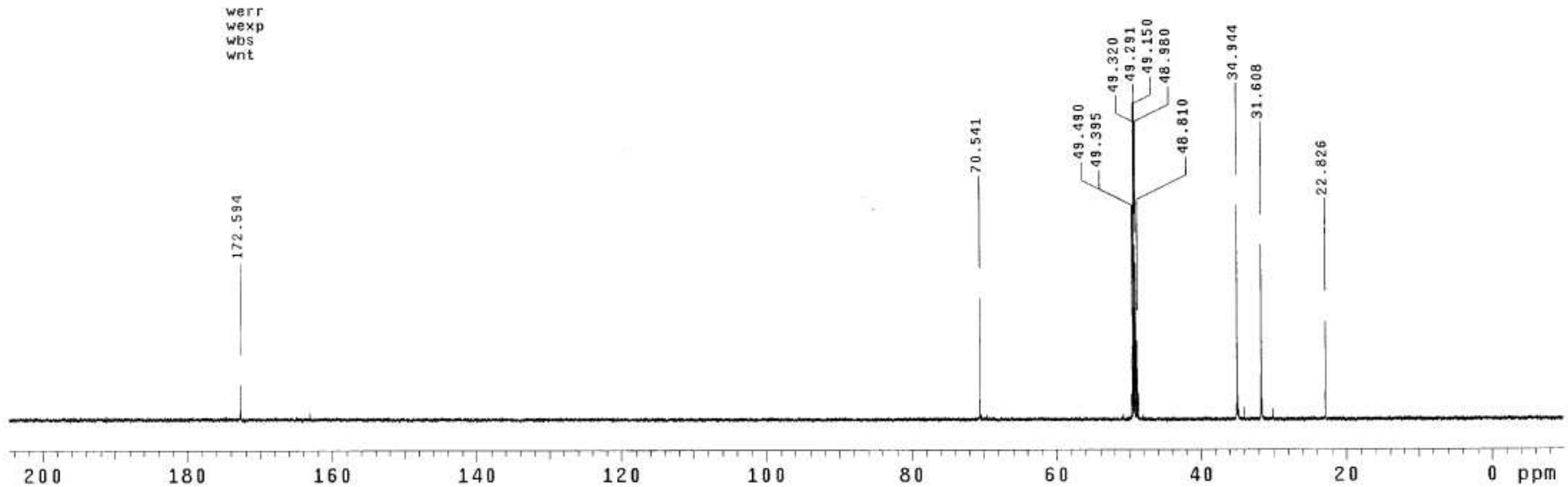
```
SAMPLE          DEC. & VT
date Jun 21 2009 dfrq          499.866
solvent CD300      dn          H1
file      exp     dpwr         30
ACQUISITION    dof          0
sfrq          499.866 dn          nnn
tn            H1    dmm          c
at            5.016 dmf         200
np            65536 dseq
sw            6533.3 dres         1.0
fb            4000 homo
bs            4      DEC2
tpwr          61    dfrq2        0
pw            13.5 dn2
d1            0.100 dpwr2        1
tof           269.9 dof2         0
nt            16    dm2          n
ct            16    dmm2         c
alock         n     dmf2         200
gain          not used dseq2
FLAGS          n     dres2         1.0
              n     homo2         n
in            n     DEC3
dp            y     dfrq3        0
hs            nn    dn3
DISPLAY       dpwr3        1
sp            -96.8 dof3         0
wp            5099.3 dm3          n
vs            36    dmm3         c
sc            0     dmf3         200
wc            250  dseq3
hzmm          20.40 dres3         1.0
is            44.25 homo3         n
rfl           2163.5 PROCESSING
rfp           1654.5 wtf file
th            13    proc          ft
ins           7.000 fn            65536
ai            ph    math          f
              werr
              wexp process pH
              wbs
              wnt wft
```

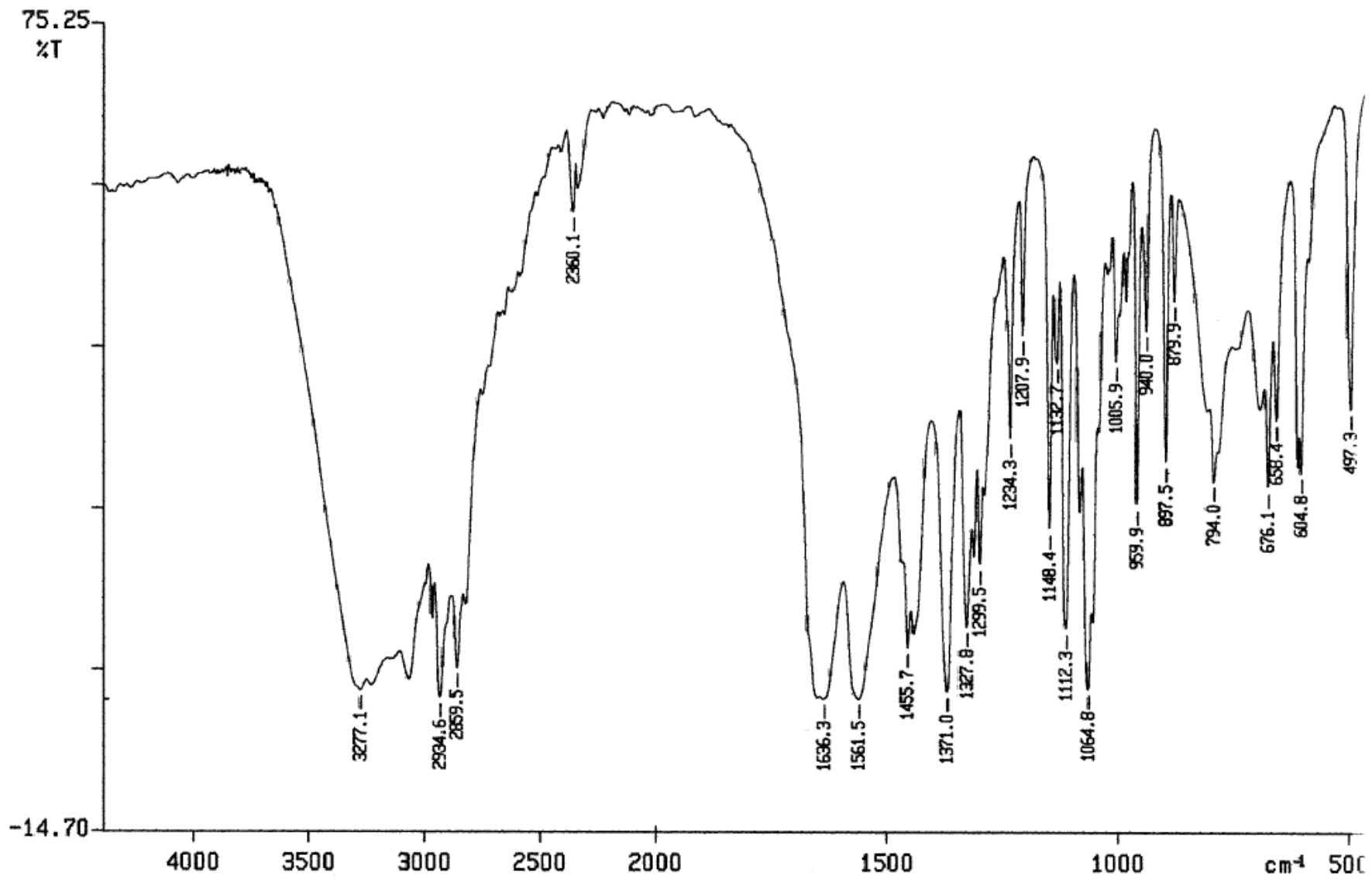


TY2-376

exp2 s2pu1

```
SAMPLE          DEC. & VT
date Jun 21 2009 dfrq      499.866
solvent cd3od      dn       H1
file exp          dpwr     40
ACQUISITION     dof       0
sfrq 125.703     dm        yyy
tn C13           dmm       w
at 1.215         dmf      8787.35
np 65536         dseq
sw 26963.3       dres     1.0
fb 15000         homo    n
bs 4             DEC2
tpwr 52          dfrq2   0
pw 10.2          dn2
dl 1.800         dpwr2   1
tof 144.5        dof2    0
nt 1000          dm2     n
ct 58           dmm2    c
alock n          dmf2    10000
gain not used    dseq2
FLAGS           dres2   1.0
jl n            homo2   n
in n            DEC3
dp y            dfrq3   0
hs nn           dn3
DISPLAY        dpwr3   1
sp -1234.2       dof3    0
wp 26962.9      dm3     n
vs 61           dmm3    c
sc 0            dmf3    10000
wc 250          dseq3
hzmm 107.85     dres3   1.0
ls 500.00       homo3   n
rfl 7412.3      PROCESSING
rfp 6177.7      lb       1.00
th 5            wtfile
ins 100.000     proc
al cdc ph      fn       131072
math          f
```

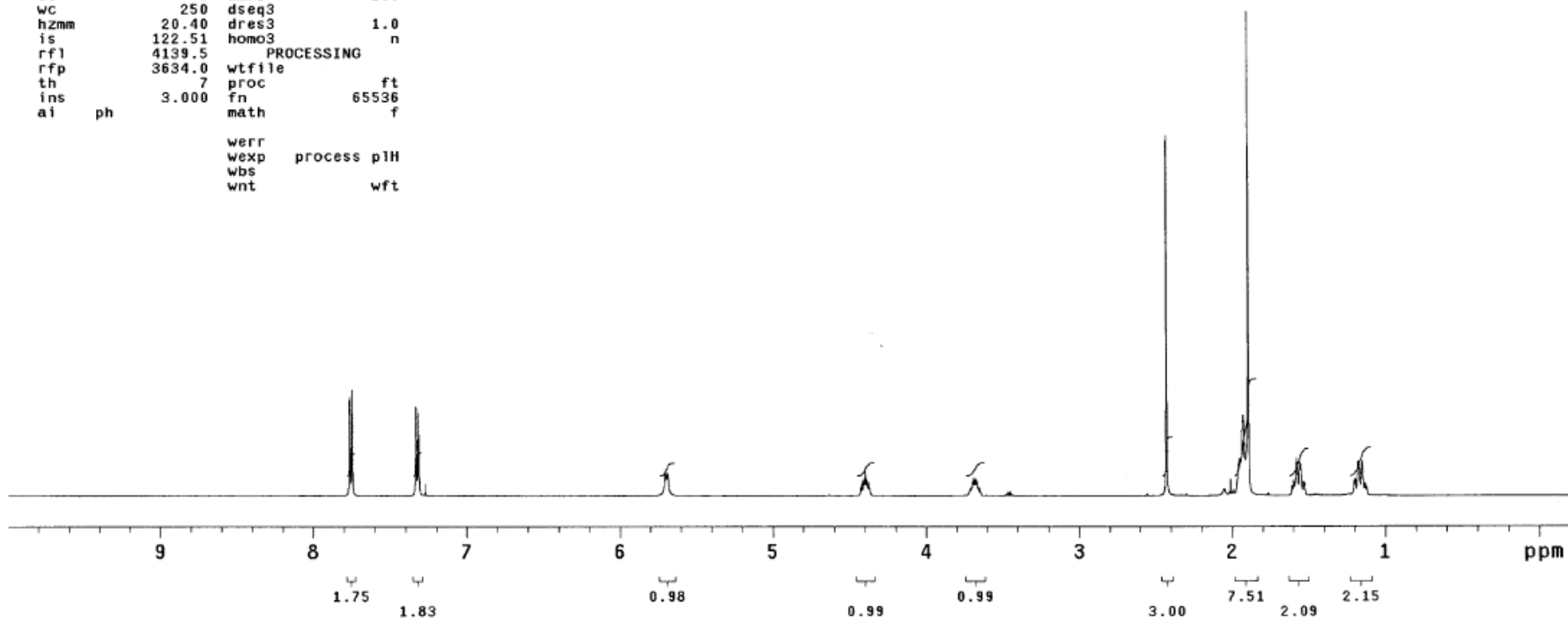
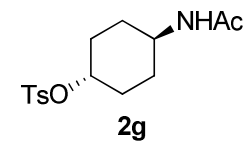




TY2-381-c5-24-recrystallization

exp2 s2pu1

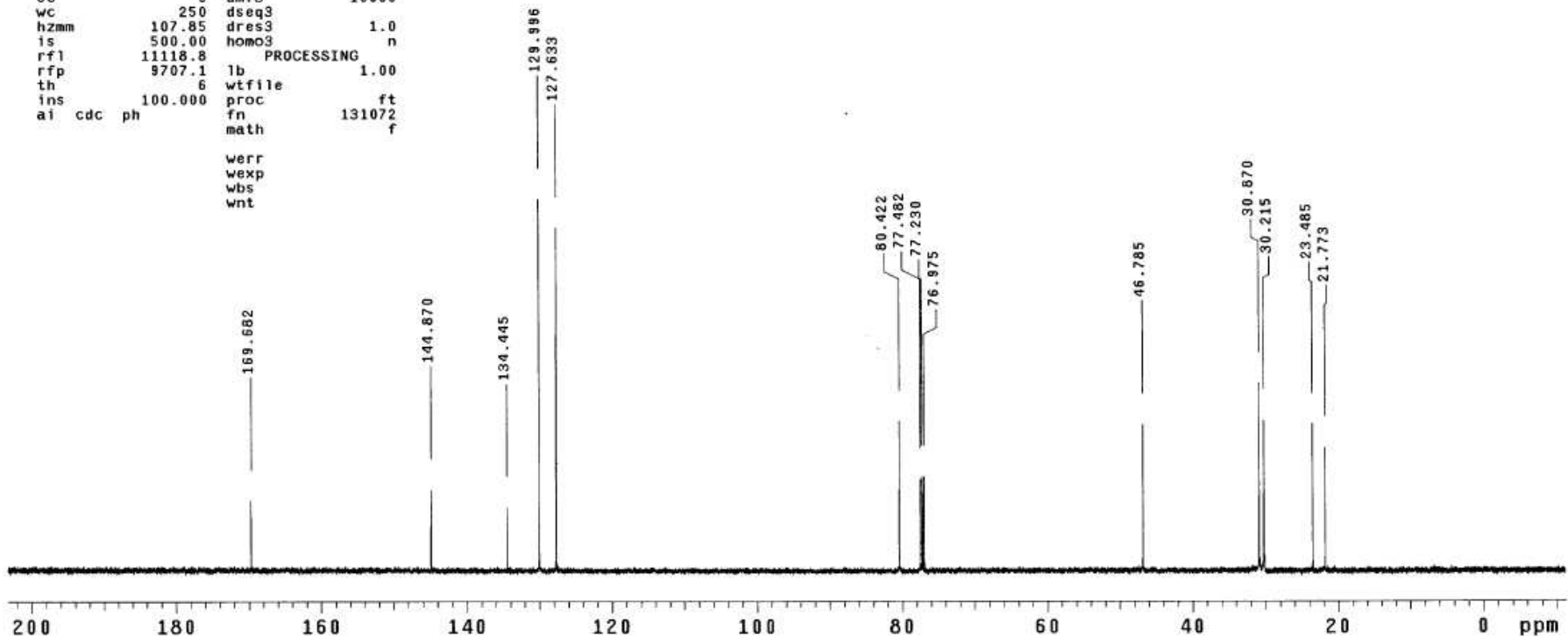
```
SAMPLE          DEC. & VT
date Jul 7 2009 dfrq      499.864
solvent CDC13      dn      H1
file      exp      dpwr     30
ACQUISITION    dof      0
sfrq      499.864 dm      nnn
tn        H1      dmm      c
at        5.016  dmf      200
np        65536  dseq
sw        6533.3 dres     1.0
fb        4000   homo
bs        4      DEC2
tpwr      61     dfrq2    0
pw        13.5  dn2
d1        0.100 dpwr2    1
tof       269.9 dof2     0
nt        32    dm2      n
ct        32    dmm2     c
alock     n     dmf2     200
gain      not used dseq2
il        FLAGS  n homo2   1.0
in        n     DEC3
dp        y     dfrq3    0
hs        nn    dn3
DISPLAY      dpwr3    1
sp        -102.0 dof3    0
wp        5099.3 dm3     n
vs        33    dmm3     c
sc        0     dmf3     200
wc        250   dseq3
hzmm      20.40 dres3    1.0
is        122.51 homo3   n
rfl       4139.5 PROCESSING
rfp       3634.0 wtfile
th        7     proc
ins       3.000 fn      65536
ai        ph    math
          werr
          wexp process pH
          wbs
          wnt wft
```



TY2-381-c5-24-recrystallization

exp3 s2pu1

date	Jul 7 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION			
sfrq	125.702	dm	yyy
tn	C13	dmm	w
at	1.215	dmf	8787.35
np	65536	dseq	
sw	26963.3	dres	1.0
fb	15000	homo	n
bs	4	DEC2	
tpwr	52	dfrq2	0
pw	10.2	dn2	
d1	1.800	dpwr2	1
tof	144.5	dof2	0
nt	1000	dm2	n
ct	67	dmm2	c
alock	n	dmf2	10000
gain	not used	dseq2	
FLAGS		dres2	1.0
l1	n	homo2	n
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY		dpwr3	1
sp	-1411.3	dof3	0
wp	26962.9	dm3	n
vs	98	dmm3	c
sc	0	dmf3	10000
wc	250	dseq3	
hzmm	107.85	dres3	1.0
ls	500.00	homo3	n
rfl	11118.8	PROCESSING	
rtp	9707.1	lb	1.00
th	6	wfile	
ins	100.000	proc	ft
ai	cdc ph	fn	131072
		math	f



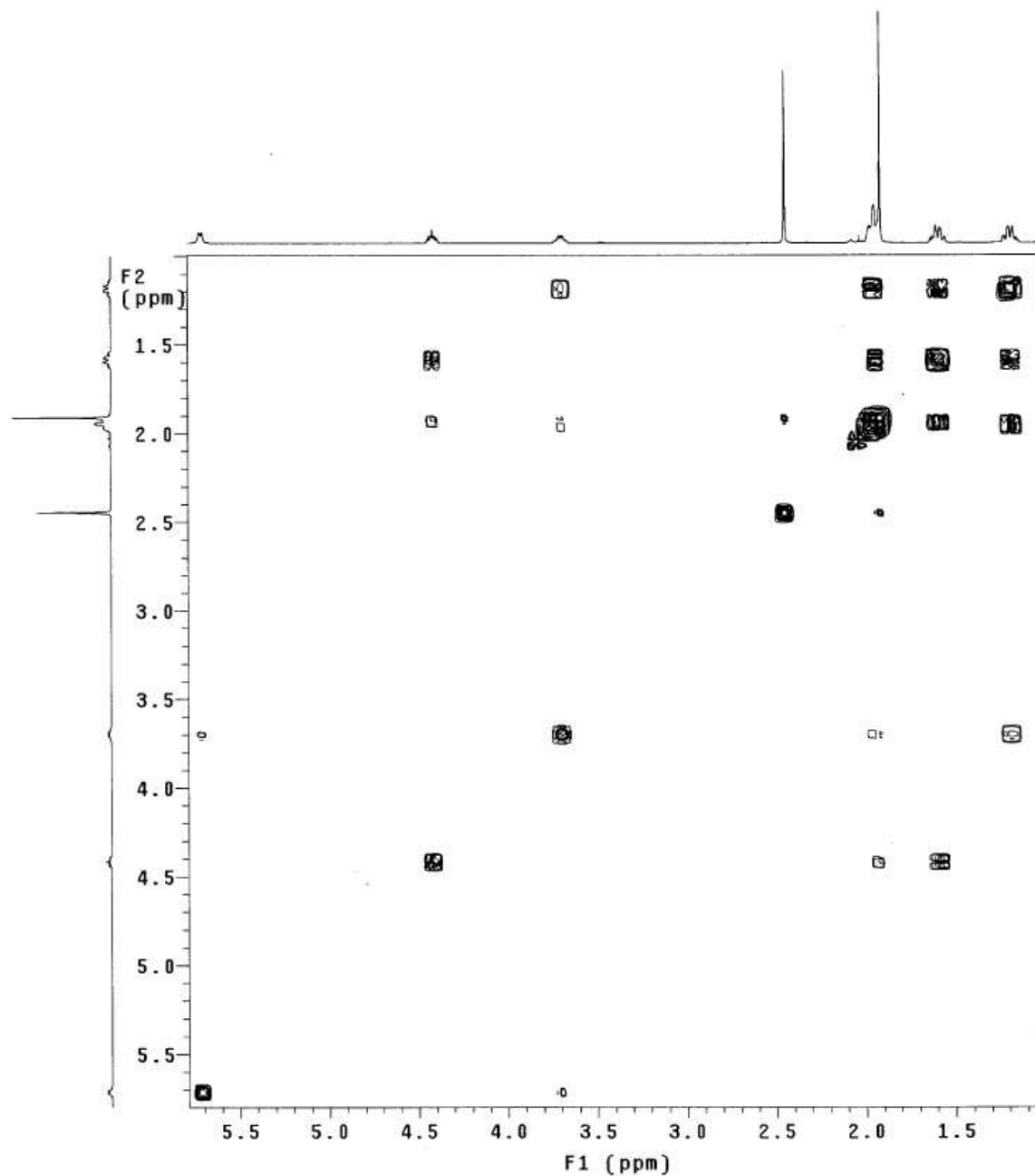


TY2-381-c5-24-recrystallization

Pulse Sequence: relayh

Solvent: CDCl<sub>3</sub>  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE H1, 499.8611619 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 3 hr, 23 min, 10 sec

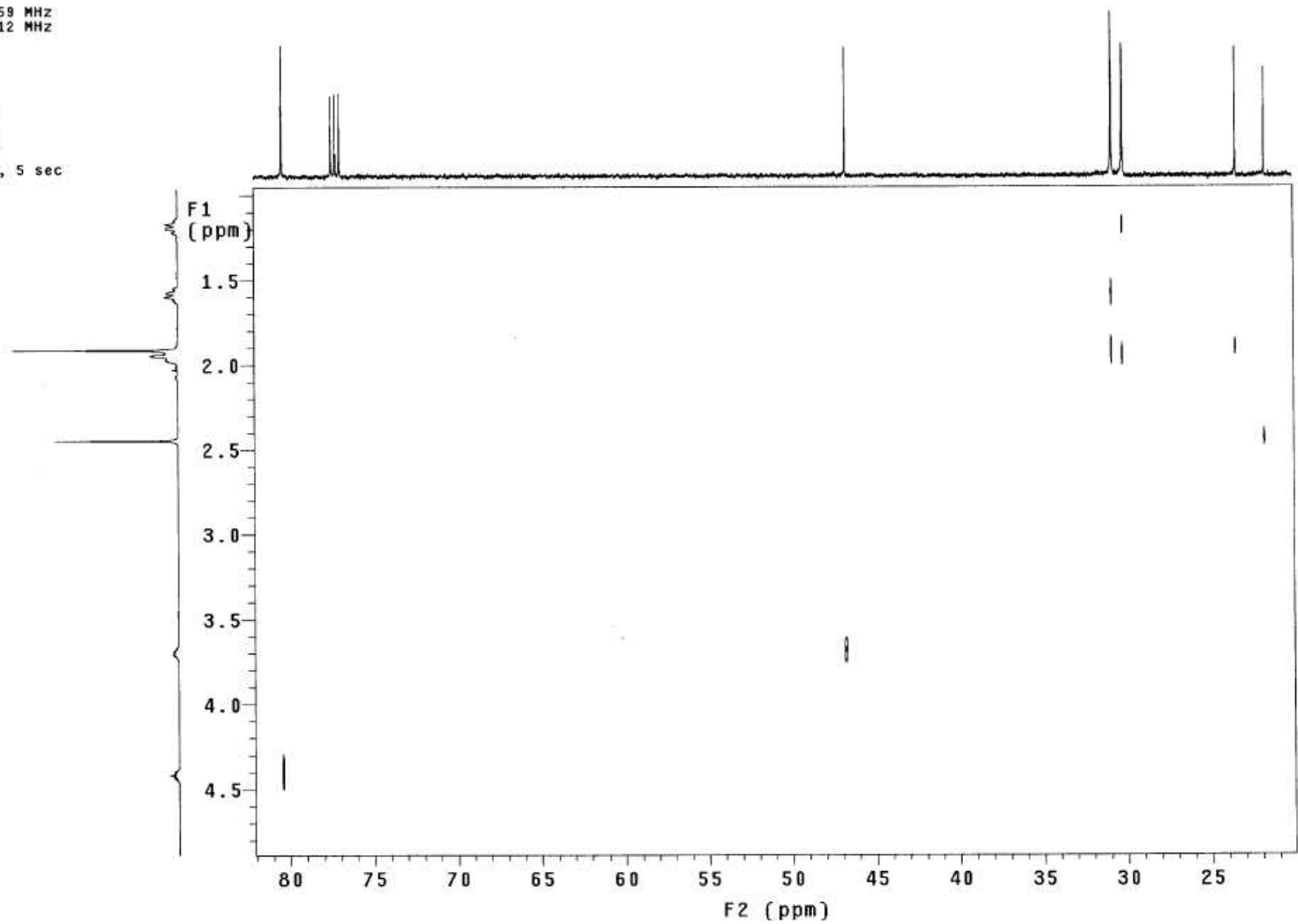


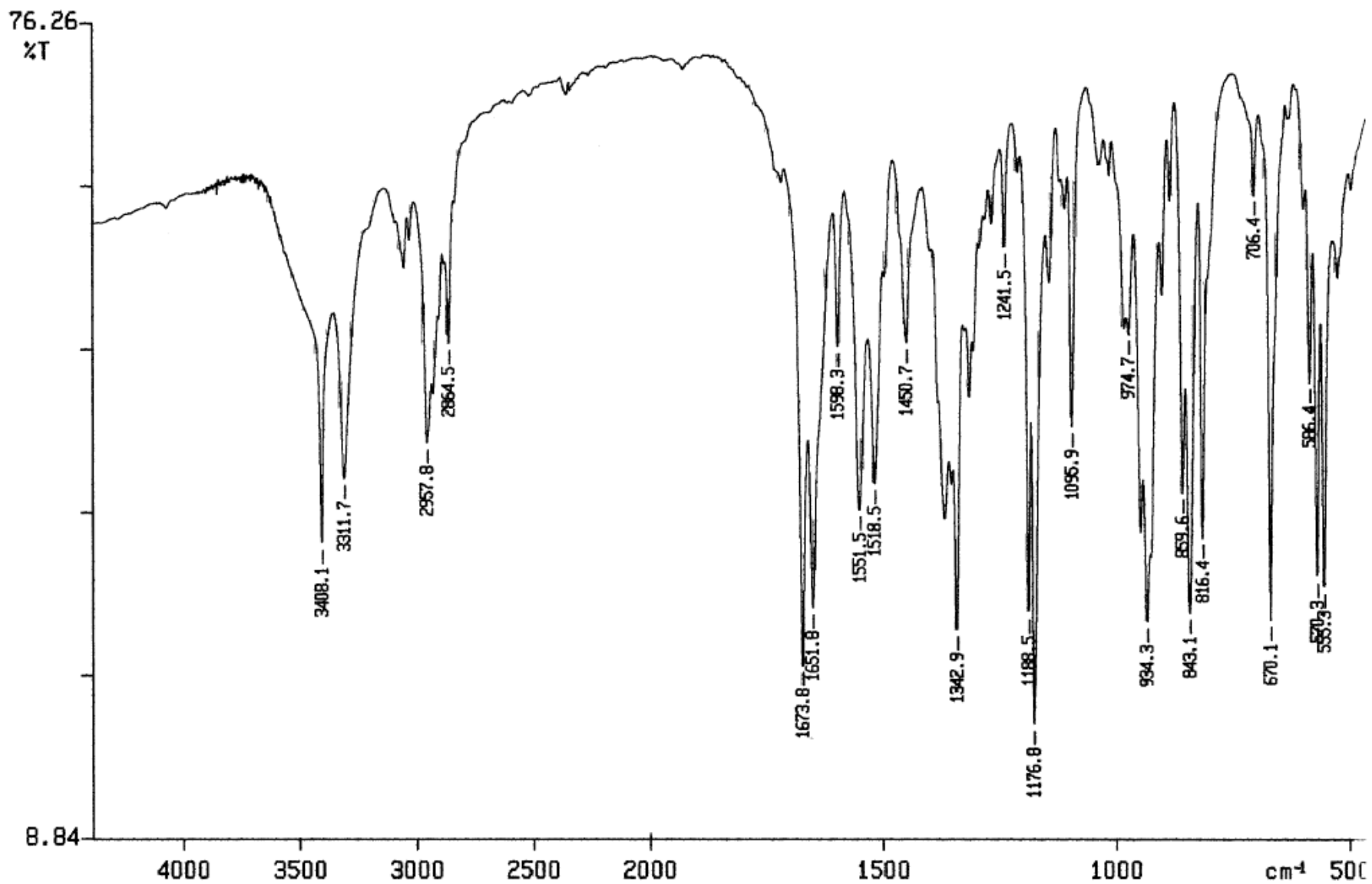
TY2-381-c5-24-recrystallization

Pulse Sequence: hetcor

Solvent: CDCl<sub>3</sub>  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE C13, 125.6901759 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 3 hr, 46 min, 5 sec

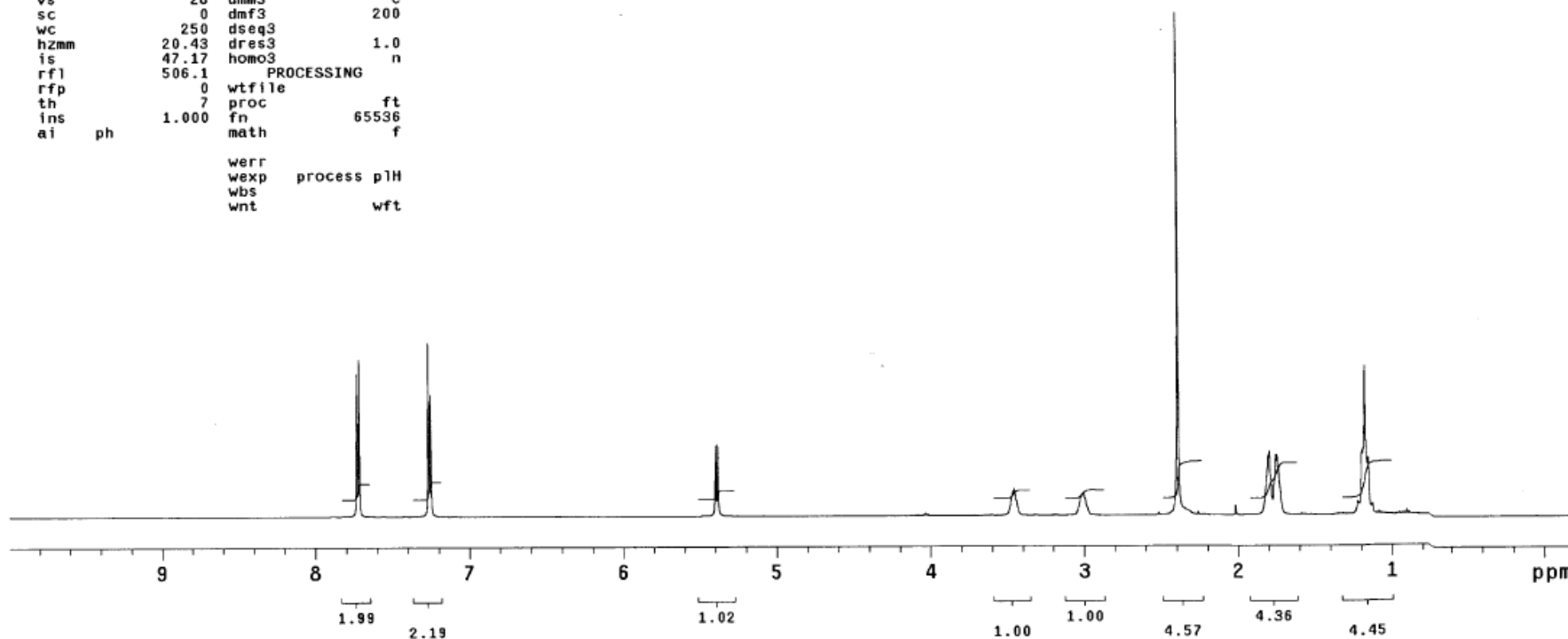
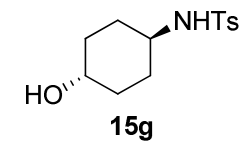




TY2-411

exp2 s2pu1

```
SAMPLE          DEC. & VT
date Jul 20 2009 dfrq          499.864
solvent CDC13      dn           H1
file exp         dpwr          30
ACQUISITION
sfrq 499.864      dm           nnn
tn    H1          dmm          c
at    5.016       dmf          200
np    65536       dseq
sw    6533.3      dres          1.0
fb    4000        homo
bs    4           DEC2
tpwr  61          dfrq2         0
pw    13.5        dn2
d1    0.100       dpwr2         1
tof   269.9      dof2          0
nt    16          dm2           n
ct    16          dmm2          c
alock n           dmf2          200
gain  not used   dseq2
FLAGS          dres2          1.0
              homo2          n
il            n           DEC3
in            n           dfrq3         0
dp            y           dn3
hs            nn          dpwr3         1
DISPLAY
sp    -111.2      dof3          0
wp    5107.9     dm3           n
vs    28         dmm3          c
sc    0          dmf3          200
wc    250        dseq3
hzmm  20.43      dres3         1.0
is    47.17      homo3         n
rfl    506.1     PROCESSING
rfp    0          wfile
th     7          proc          ft
ins    1.000     fn           65536
al     ph        math          f
              werr
              wexp   process pH
              wbs
              wnt    wft
```

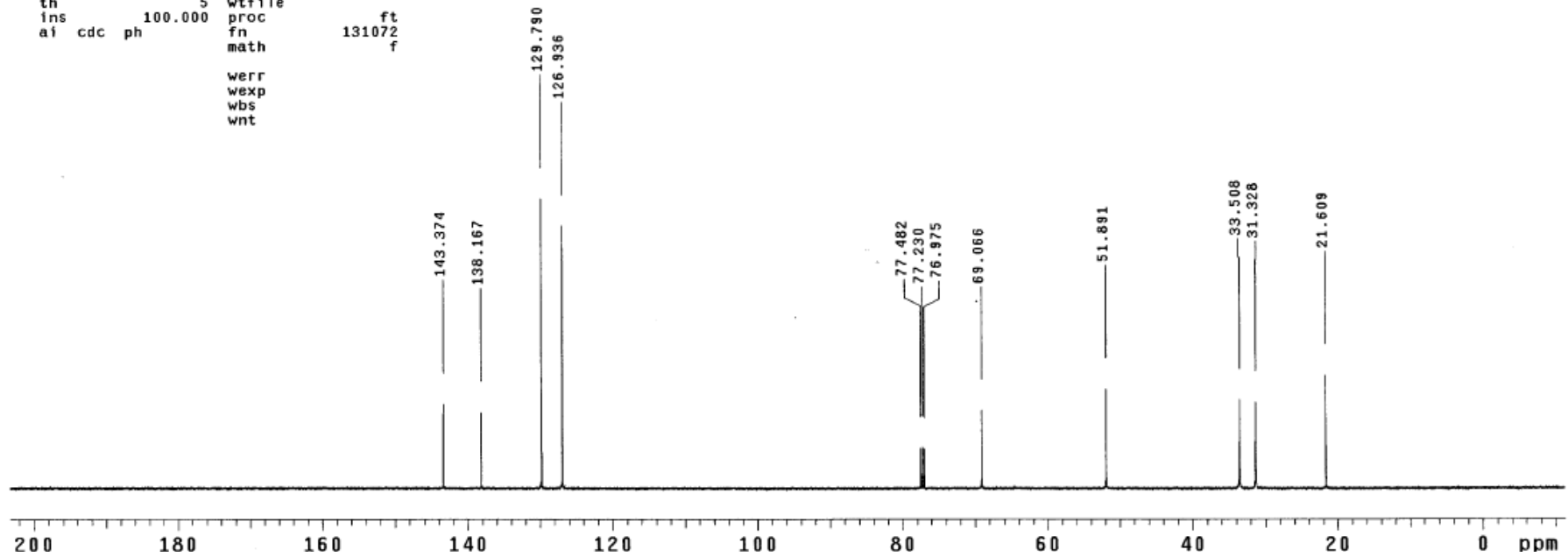


TY2-411

exp3 s2pu1

SAMPLE		DEC. & VT	
date	Jul 20 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION		dof	0
sfrq	125.702	dm	yyy
tn	C13	dmm	w
at	1.215	dmf	8787.35
np	65536	dseq	
sw	26963.3	dres	1.0
fb	15000	homo	n
bs	4	DEC2	
tpwr	52	dfrq2	0
pw	10.2	dn2	
d1	1.800	dpwr2	1
tof	144.5	dof2	0
nt	256	dm2	n
ct	67	dmm2	c
alock	n	dmf2	10000
gain	not used	dseq2	
il	FLAGS	dres2	1.0
in	n	homo2	n
dp	y	dfrq3	DEC3
hs	nn	dn3	0
DISPLAY		dpwr3	1
sp	-1417.1	dof3	0
wp	26962.9	dm3	n
vs	41	dmm3	c
sc	0	dmf3	10000
wc	250	dseq3	
hzmm	107.85	dres3	1.0
is	500.00	homo3	n
		PROCESSING	
rfl	11124.6	lb	1.00
rfp	9707.1	wtfile	
th	5	proc	ft
ins	100.000	fn	131072
ai	cdc ph	math	f

werr  
wexp  
wbs  
wnt

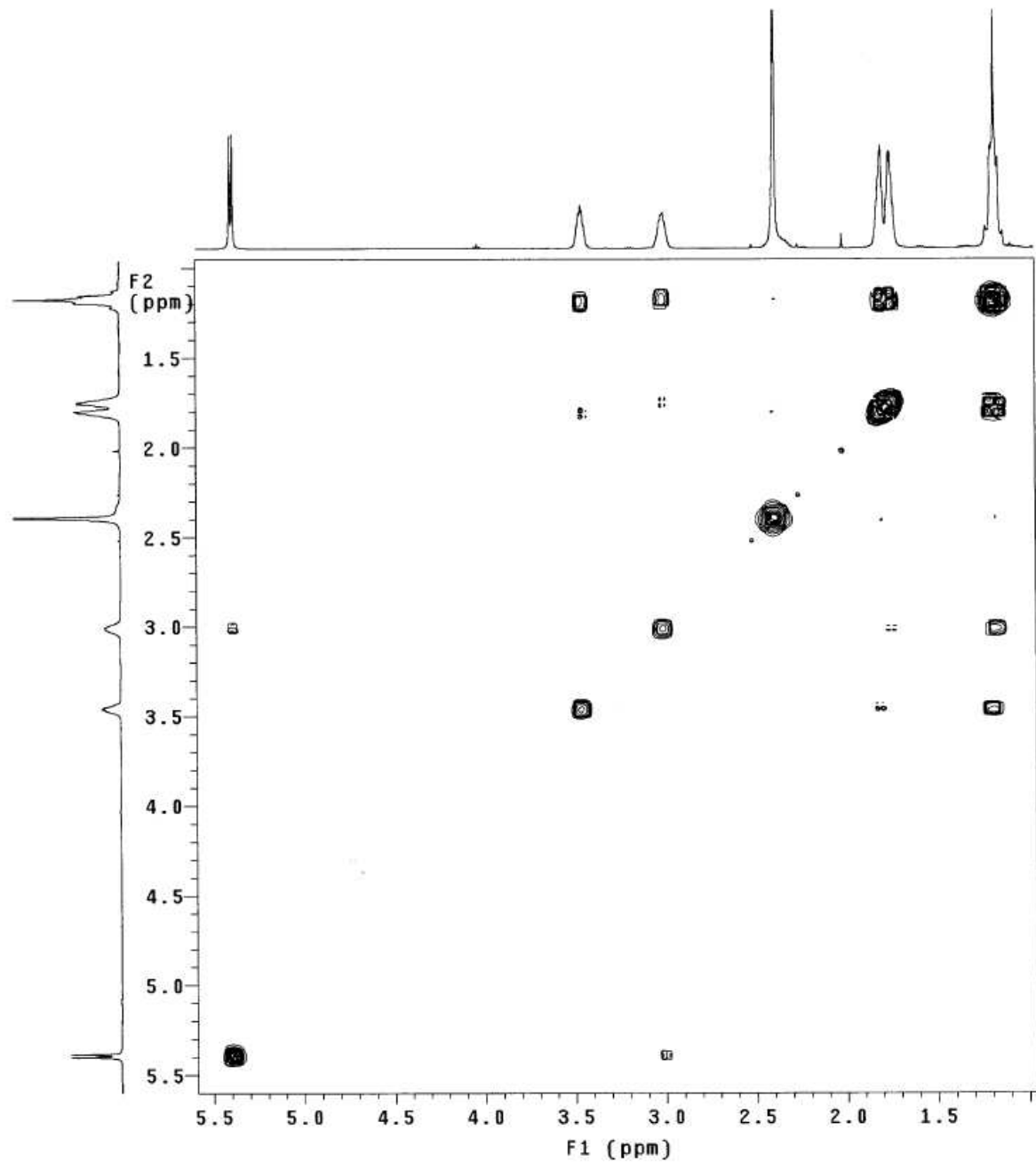


TY2-411

Pulse Sequence: relayh

Solvent: CDC13  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec  
CDSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
16 repetitions  
256 increments  
OBSERVE H1, 499.8611707 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 1 hr, 41 min, 40 sec

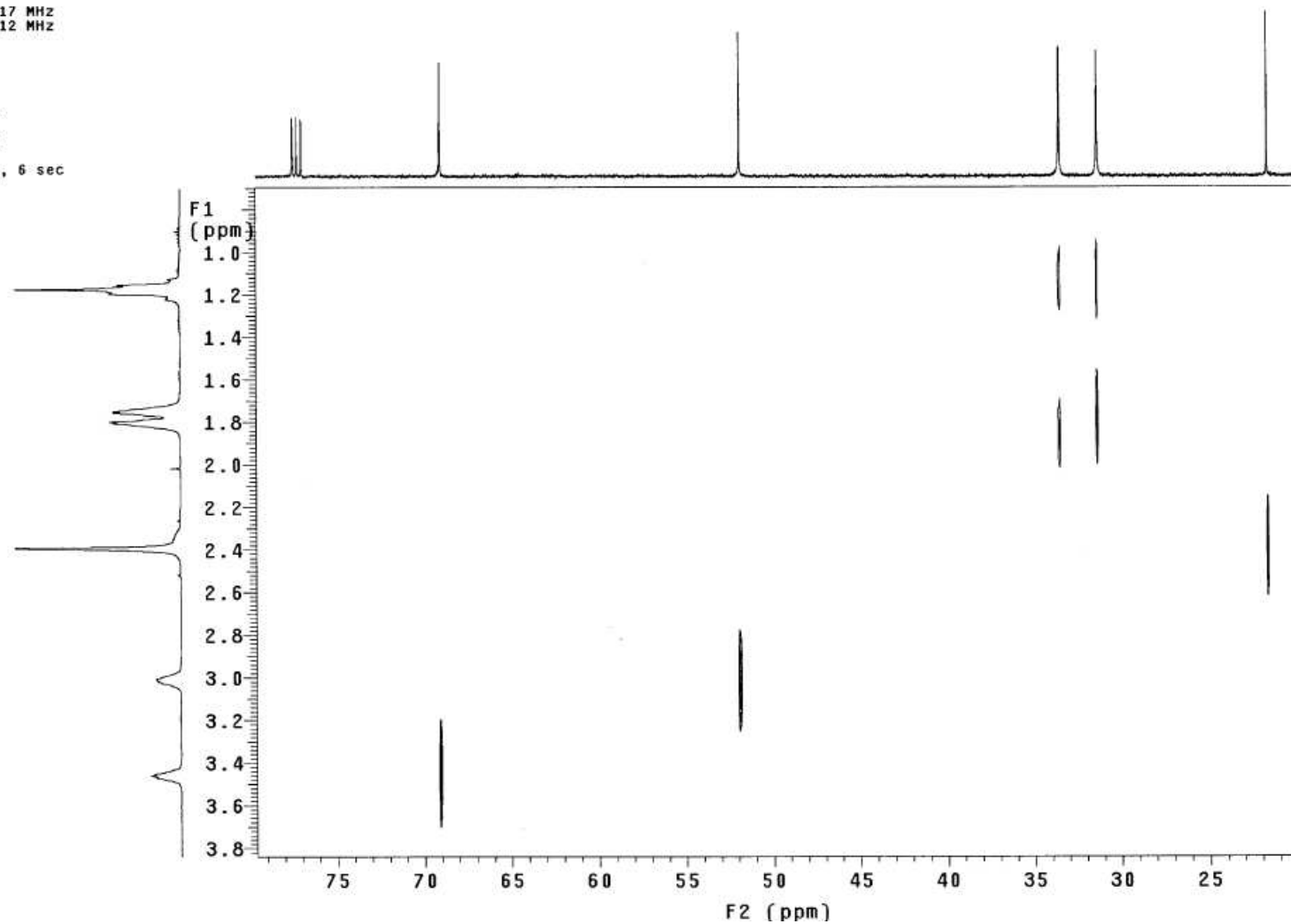


TY2-411

Pulse Sequence: hetcor

Solvent: CDCl<sub>3</sub>  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
16 repetitions  
256 increments  
OBSERVE C13, 125.6901817 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 1 hr, 53 min, 6 sec



CH3 carbons



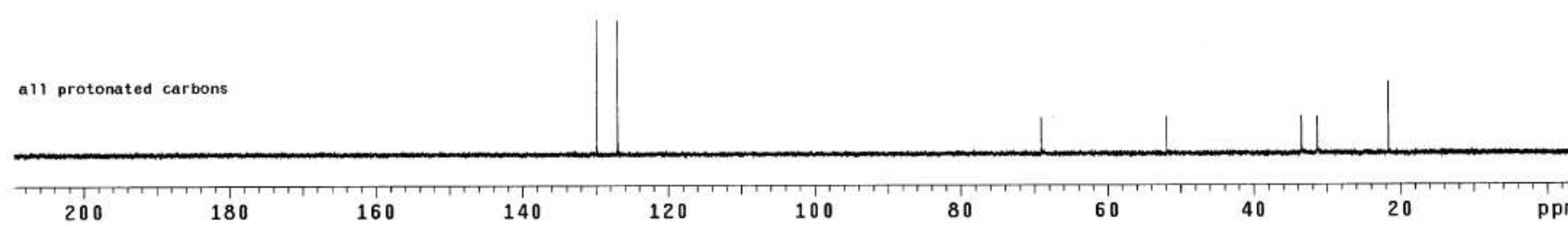
CH2 carbons



CH carbons



all protonated carbons



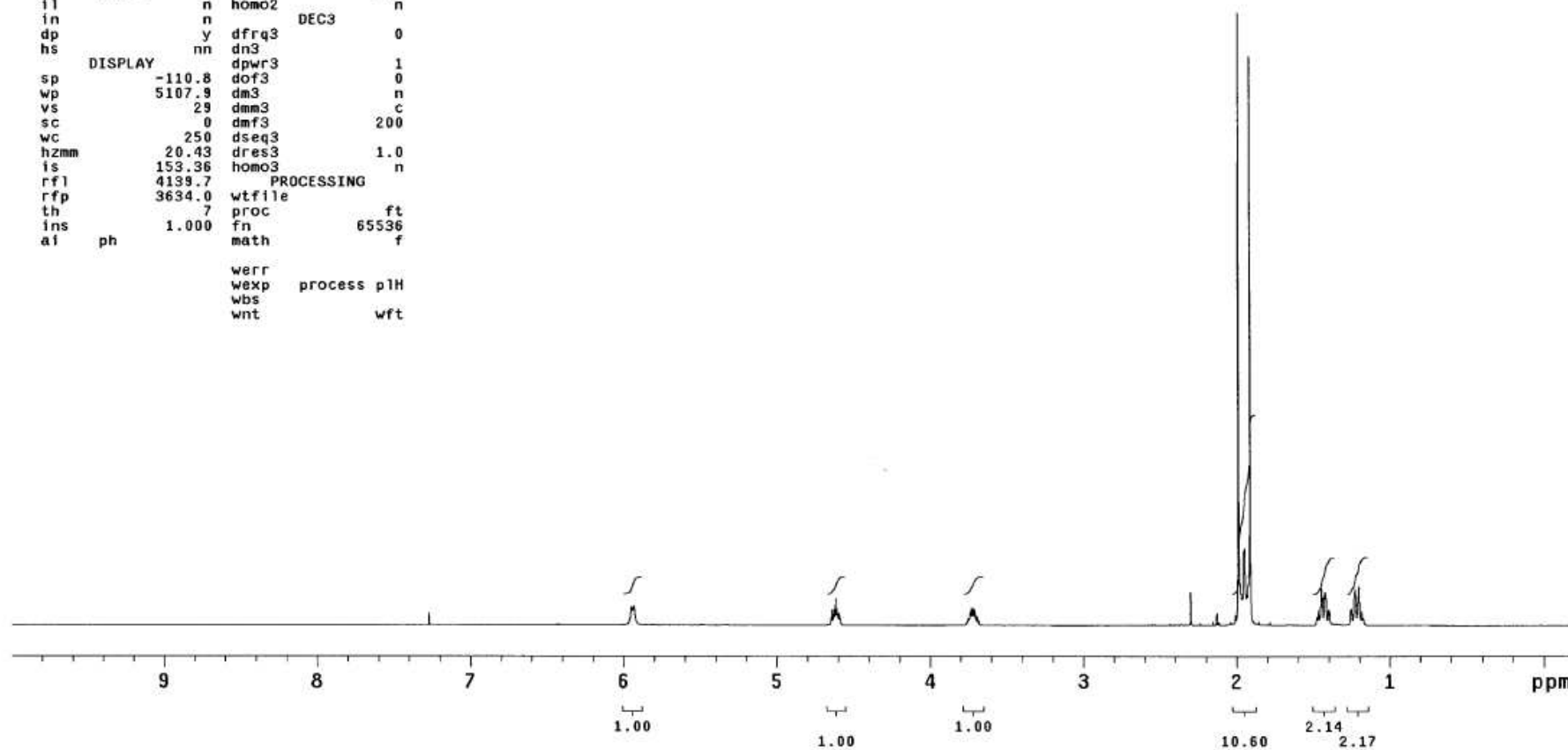
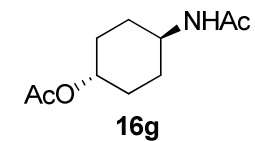
200 180 160 140 120 100 80 60 40 20 ppm



TY2-415

exp2 s2pu1

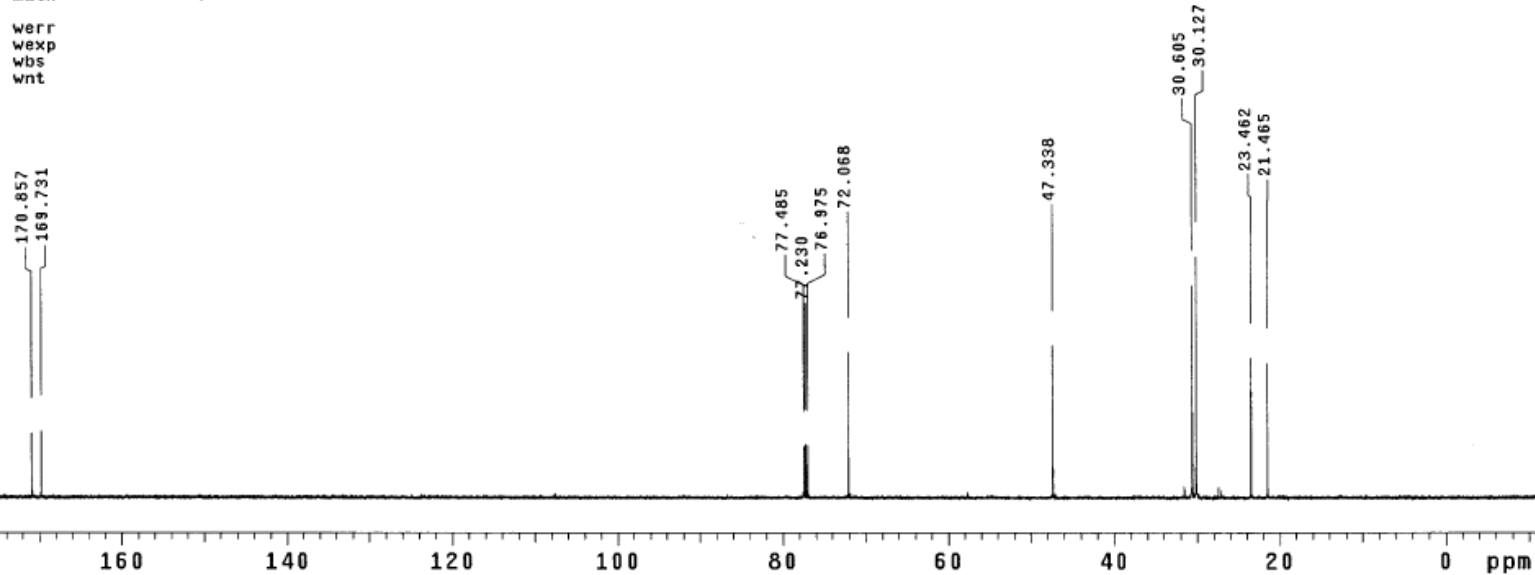
SAMPLE		DEC. & VT	
date	Jul 21 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	499.864	dm	nnn
tn	H1	dmm	c
at	5.016	dmf	200
np	65536	dseq	
sw	6533.3	dres	1.0
fb	4000	homo	n
bs	4	DEC2	
tpwr	61	dfrq2	0
pw	13.5	dn2	
d1	0.100	dpwr2	1
tof	269.9	dof2	0
nt	16	dm2	n
ct	16	dmm2	c
alock	n	dmf2	200
gain	not used	dseq2	
FLAGS		dres2	1.0
il	n	homo2	n
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY		dpwr3	1
sp	-110.8	dof3	0
wp	5107.9	dm3	n
vs	29	dmm3	c
sc	0	dmf3	200
wc	250	dseq3	
hzmm	20.43	dres3	1.0
is	153.36	homo3	n
rfl	4139.7	PROCESSING	
rfp	3634.0	wffile	
th	7	proc	ft
ins	1.000	fn	65536
af	ph	math	f
		werr	
		wexp	process pH
		wbs	
		wnt	wft



TY2-415

exp3 s2pu1

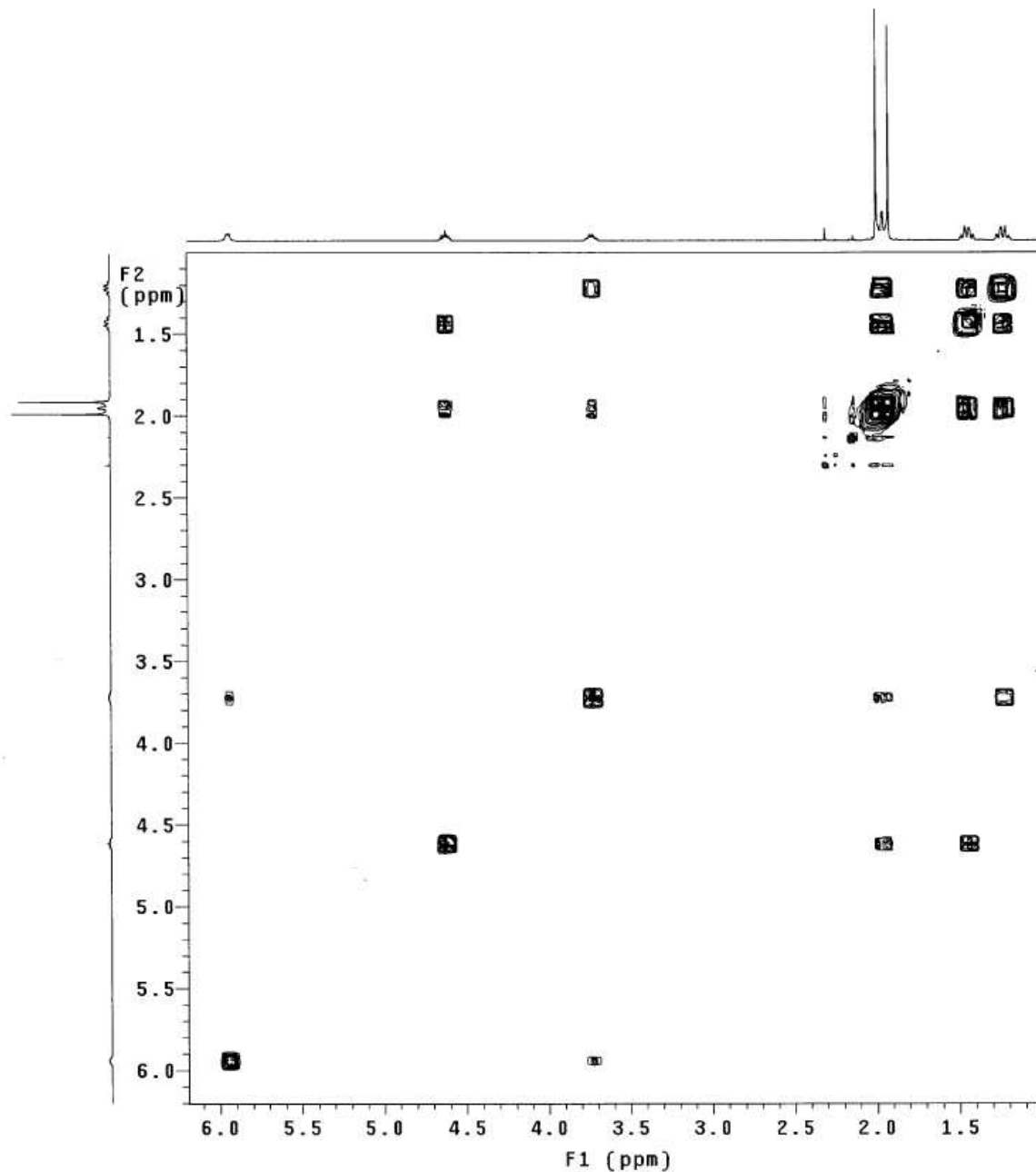
SAMPLE		DEC. & VT	
date	Jul 21 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION		dof	
sfrq	125.702	dm	yyy
tn	C13	dmm	w
at	1.215	dmf	8787.35
np	65536	dseq	
sw	26963.3	dres	1.0
fb	15000	homo	n
bs	4	DEC2	
tpwr	52	dfrq2	0
pw	10.2	dn2	
d1	1.800	dpwr2	1
tof	144.5	dof2	0
nt	512	dm2	n
ct	69	dmm2	c
alock	n	dmf2	10000
gain	not used	dseq2	
FLAGS		dres2	
il	n	homo2	n
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY		dpwr3	
sp	-1411.8	dof3	0
wp	26962.9	dm3	n
vs	47	dmm3	c
sc	0	dmf3	10000
wc	250	dseq3	
hzmm	107.85	dres3	1.0
is	500.00	homo3	n
rfl	11119.2	PROCESSING	
rfp	9707.1	lb	1.00
th	6	wffile	
ins	100.000	proc	ft
ai	cdc ph	fn	131072
		math	f



TY2-415

Pulse Sequence: relayh  
Solvent: CDC13  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
16 repetitions  
256 increments  
OBSERVE H1, 499.8611703 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 1 hr, 41 min, 40 sec

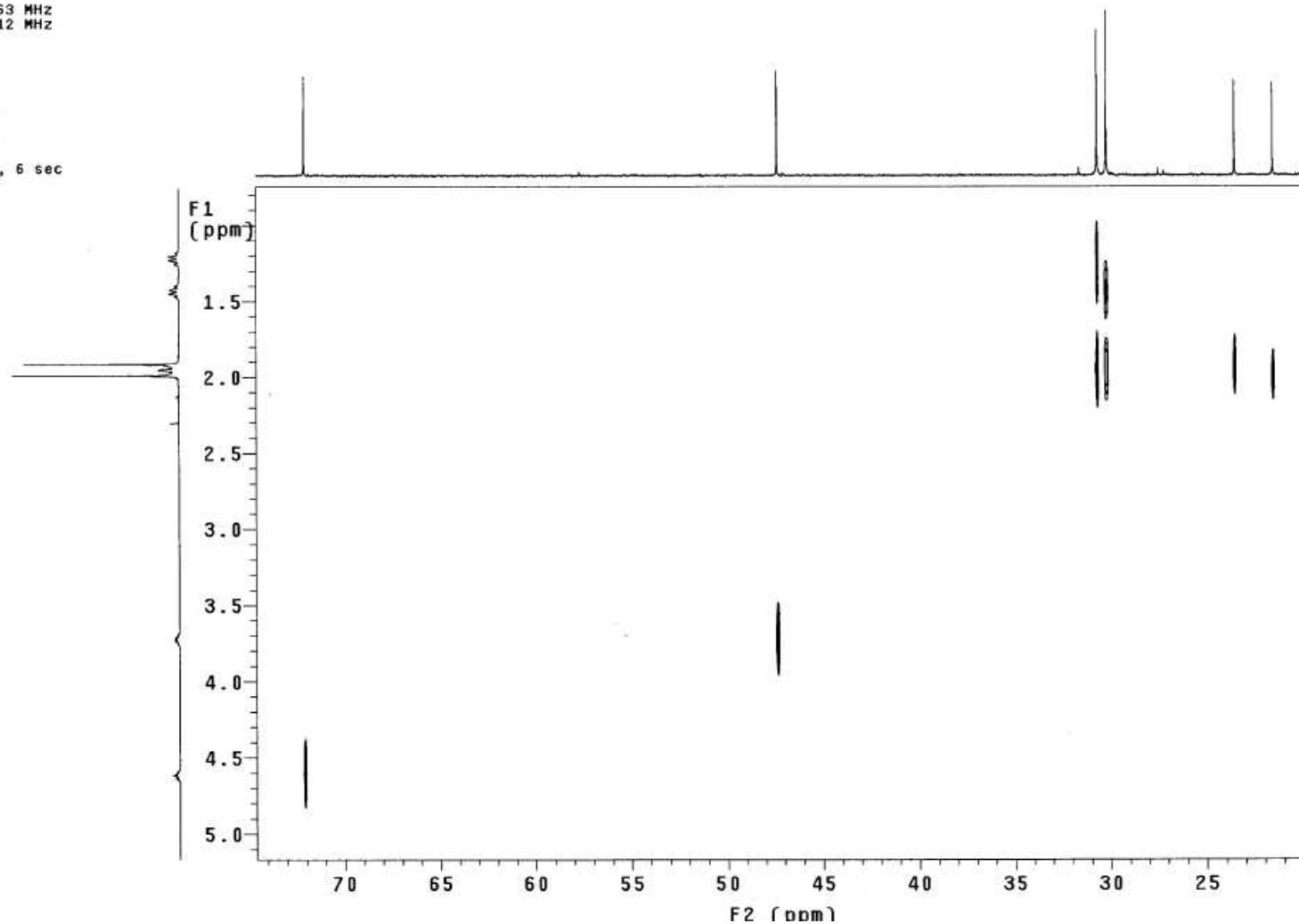


TY2-415

Pulse Sequence: hetcor

Solvent: CDC13  
Ambient temperature  
User: 1-14-87  
INNOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
16 repetitions  
256 increments  
OBSERVE C13, 125.6901763 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 1 hr, 53 min, 6 sec



CH3 carbons



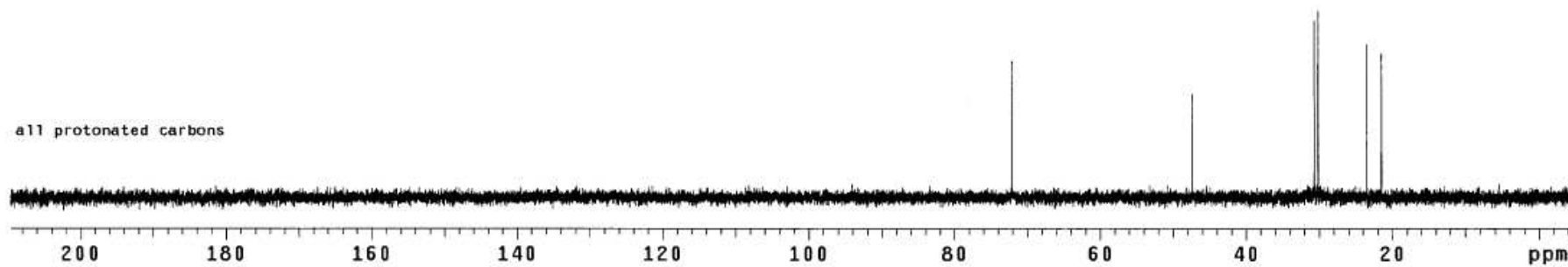
CH2 carbons

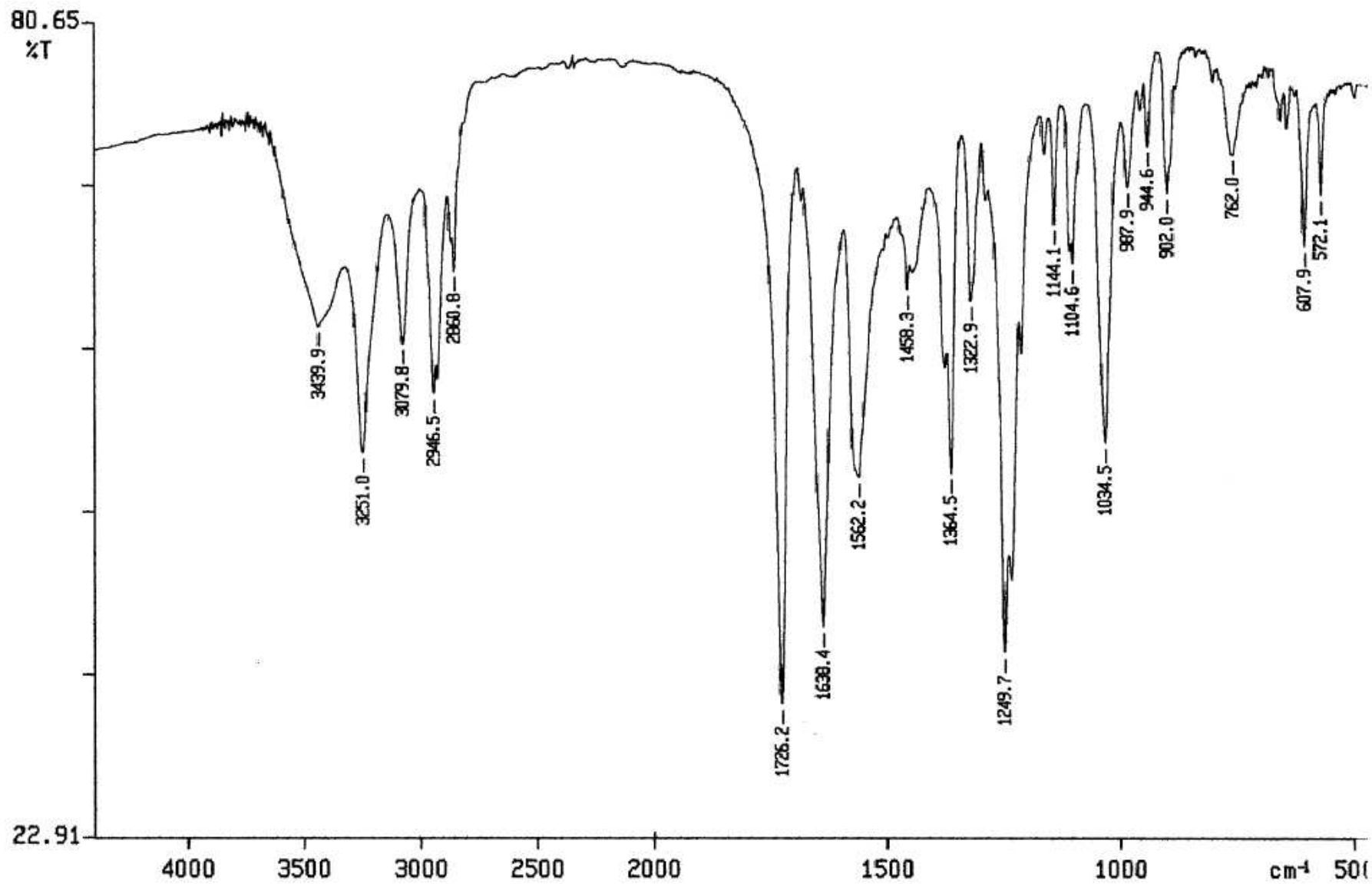


CH carbons



all protonated carbons

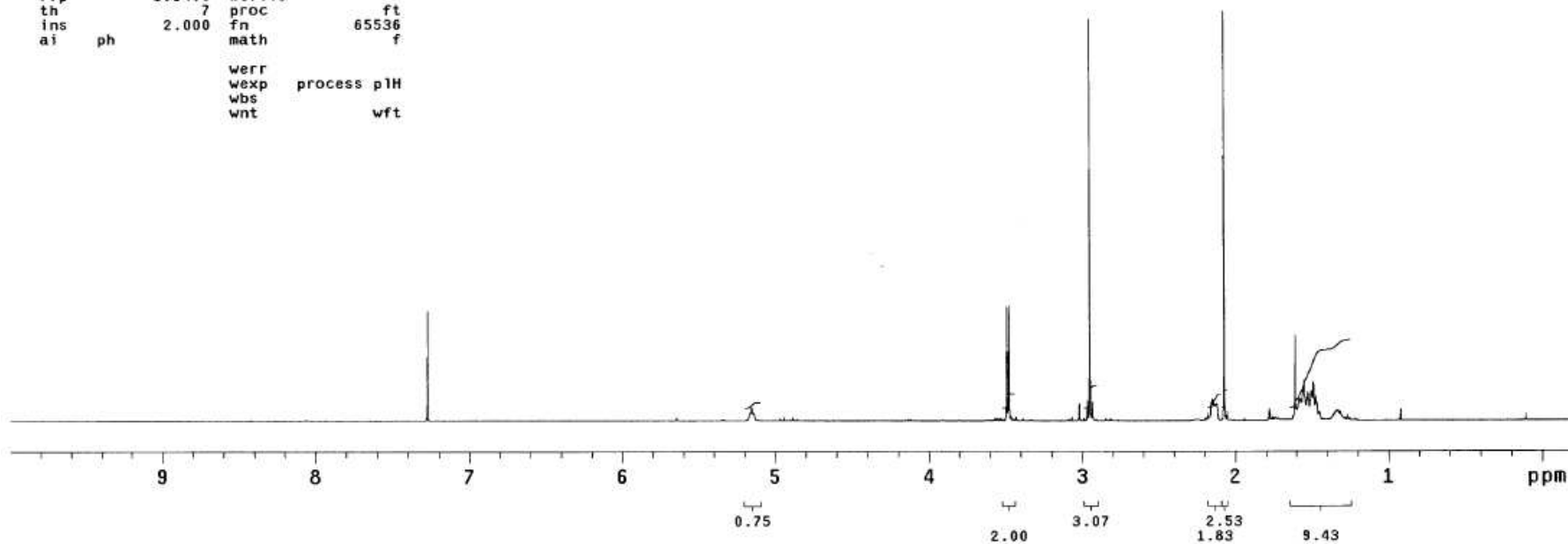
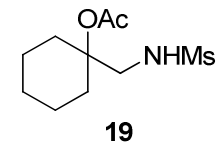




TY2-416

exp2 s2pu1

```
SAMPLE          DEC. & VT
date Jul 18 2009 dfrq          499.864
solvent CDC13      dn          H1
file      exp      dpwr         30
ACQUISITION     dof          0
sfrq      499.864 dm           nnn
tn         H1      dmm          c
at         5.016 dmf           200
np         65536 dseq          1.0
sw         6533.3 dres         1.0
fb         4000  homo          n
bs         4      DEC2
tpwr       61      dfrq2        0
pw         13.5 dn2
d1         0.100 dpwr2         1
tof        269.9 dof2         0
nt         16      dm2          n
ct         16      dmm2         c
alock      n      dmf2         200
gain      not used dseq2
FLAGS      n      dres2         1.0
           n      homo2         n
           n      DEC3
           y      dfrq3         0
           nn     dn3
DISPLAY    dpwr3         1
sp         -102.2 dof3         0
wp         5099.3 dm3          n
vs         21      dmm3         c
sc         0      dmf3         200
wc         250    dseq3         1.0
hzmm       20.40 dres3         1.0
is         69.57 homo3         n
rf1        4139.7 PROCESSING
rfp        3634.0 wtfile
th         7      proc
ins        2.000 fn           65536
ai         ph      math
           werr
           wexp process pH
           wbs
           wnt wft
```

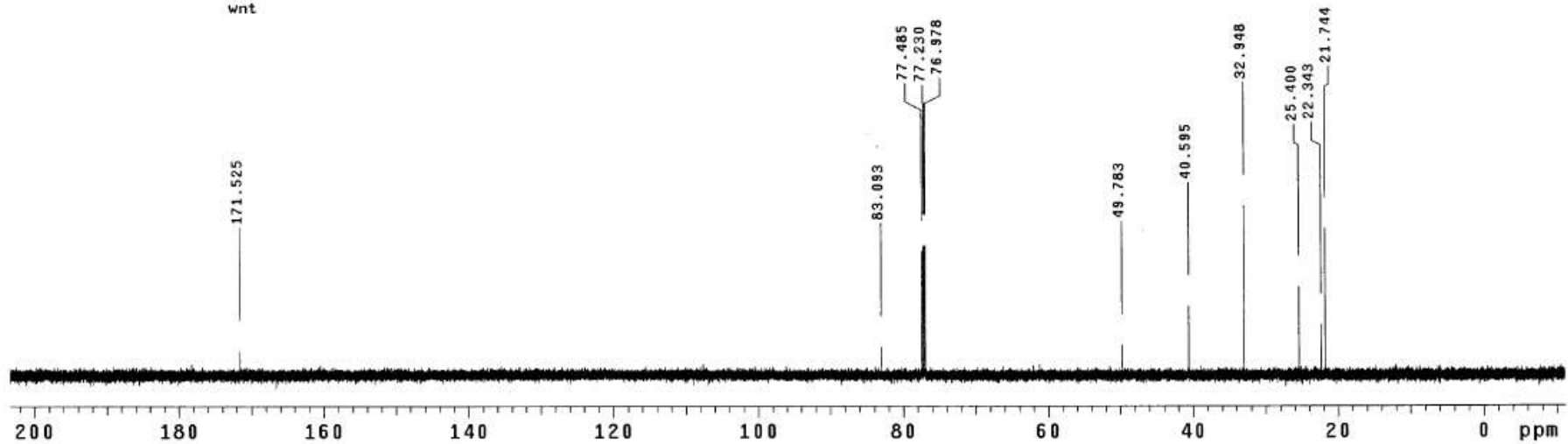


TY2-416

exp3 s2pu1

SAMPLE		DEC. & VT	
date	Jul 18 2009	dfrq	499.864
solvent	CDC13	dn	H1
file	exp	dpwr	40
ACQUISITION		dof	0
sfrq	125.702	dm	yyy
tn	C13	dmm	w
at	1.215	dmf	8787.35
np	65536	dseq	
sw	26963.3	dres	1.0
fb	15000	homo	n
bs	4	DEC2	
tpwr	52	dfrq2	0
pw	10.2	dn2	
d1	1.800	dpwr2	1
tof	144.5	dof2	0
nt	1000	dm2	n
ct	122	dmm2	c
alock	n	dmf2	10000
gain	not used	dseq2	
FLAGS		dres2	1.0
il	n	homo2	n
in	n	DEC3	
dp	y	dfrq3	0
hs	nn	dn3	
DISPLAY		dpwr3	1
sp	-1401.5	dof3	0
wp	26962.9	dm3	n
vs	95	dmm3	c
sc	0	dmf3	10000
wc	250	dseq3	
hzmm	107.85	dres3	1.0
is	500.00	homo3	n
rf1	11108.9	PROCESSING	
rfl	9707.1	lb	not used
th	3	wtfile	
ins	100.000	proc	ft
ai	cdc ph	fn	131072
		math	f

werr  
wexp  
wbs  
wnt

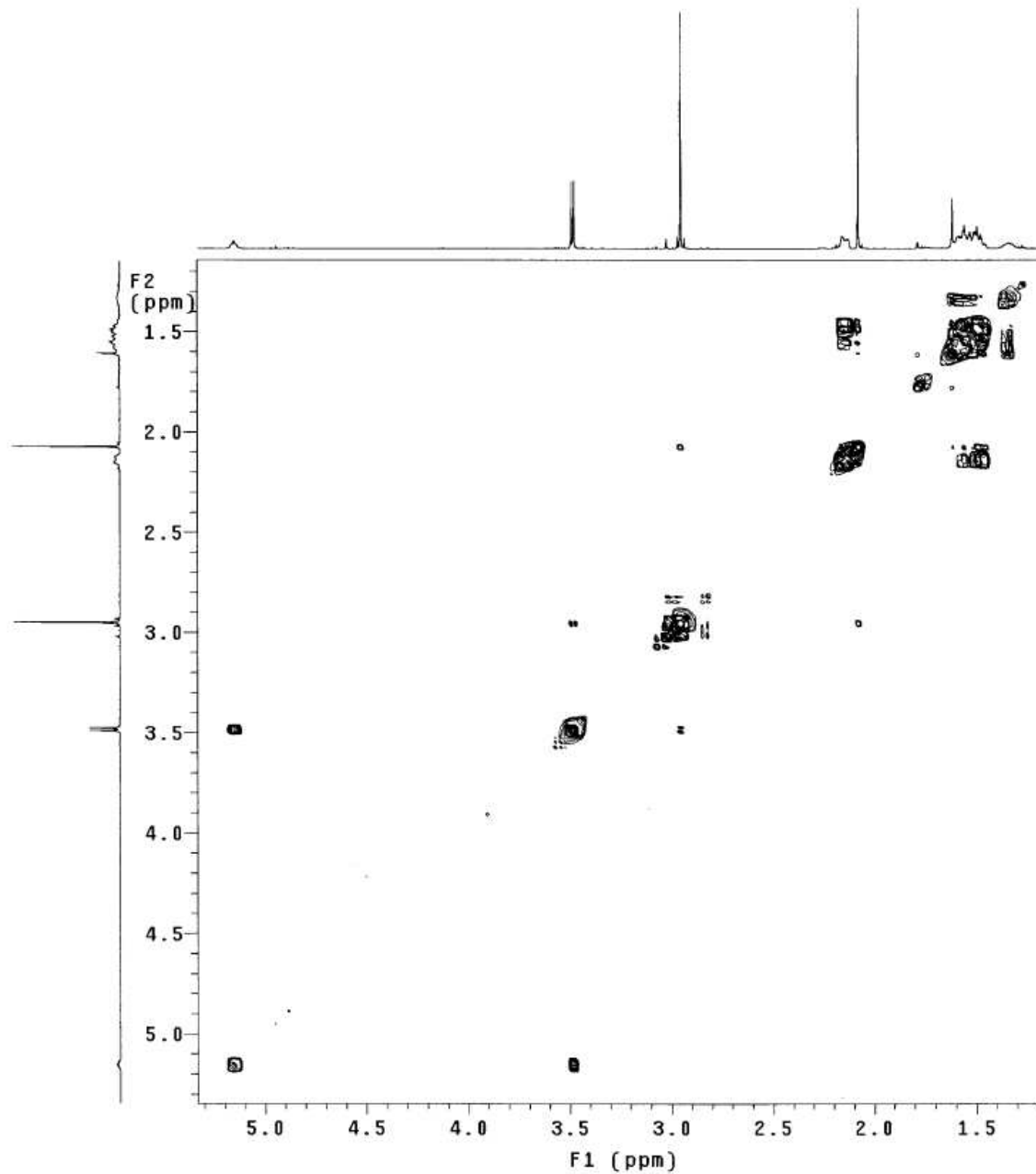




TY2-416

Pulse Sequence: relayh  
Solvent: CDC13  
Ambient temperature  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.157 sec  
Width 6533.3 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE H1, 499.8611703 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.039 sec  
FT size 2048 x 2048  
Total time 3 hr, 23 min, 10 sec

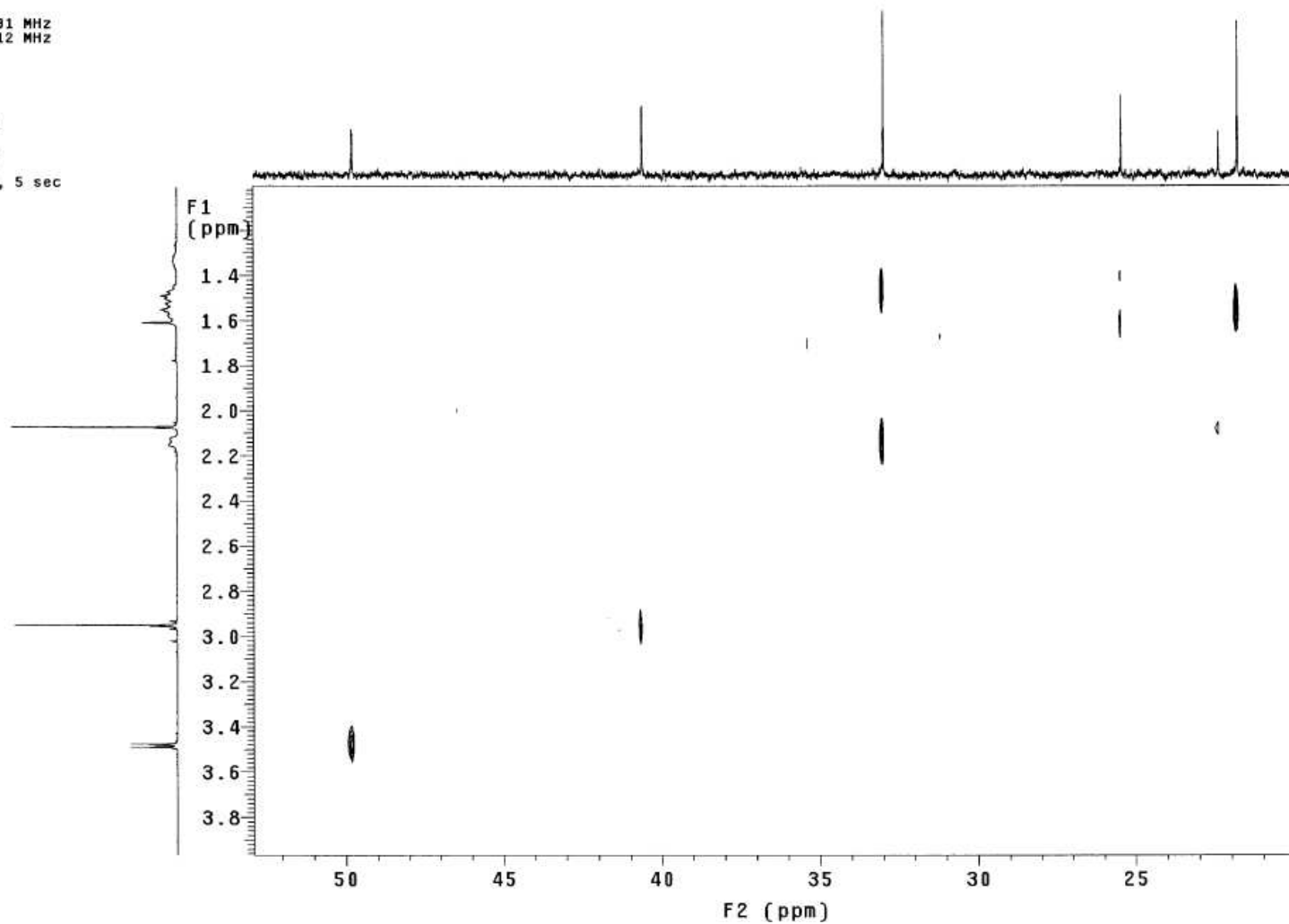


TY2-416

Pulse Sequence: hetcor

Solvent: CDCl<sub>3</sub>  
Ambient temperature  
User: 1-14-87  
File: TY2-416-CH  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 6533.3 Hz  
32 repetitions  
256 increments  
OBSERVE C13, 125.6901591 MHz  
DECOUPLE H1, 499.8639312 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 3 hr, 46 min, 5 sec



CH3 carbons



CH2 carbons



CH carbons



all protonated carbons

