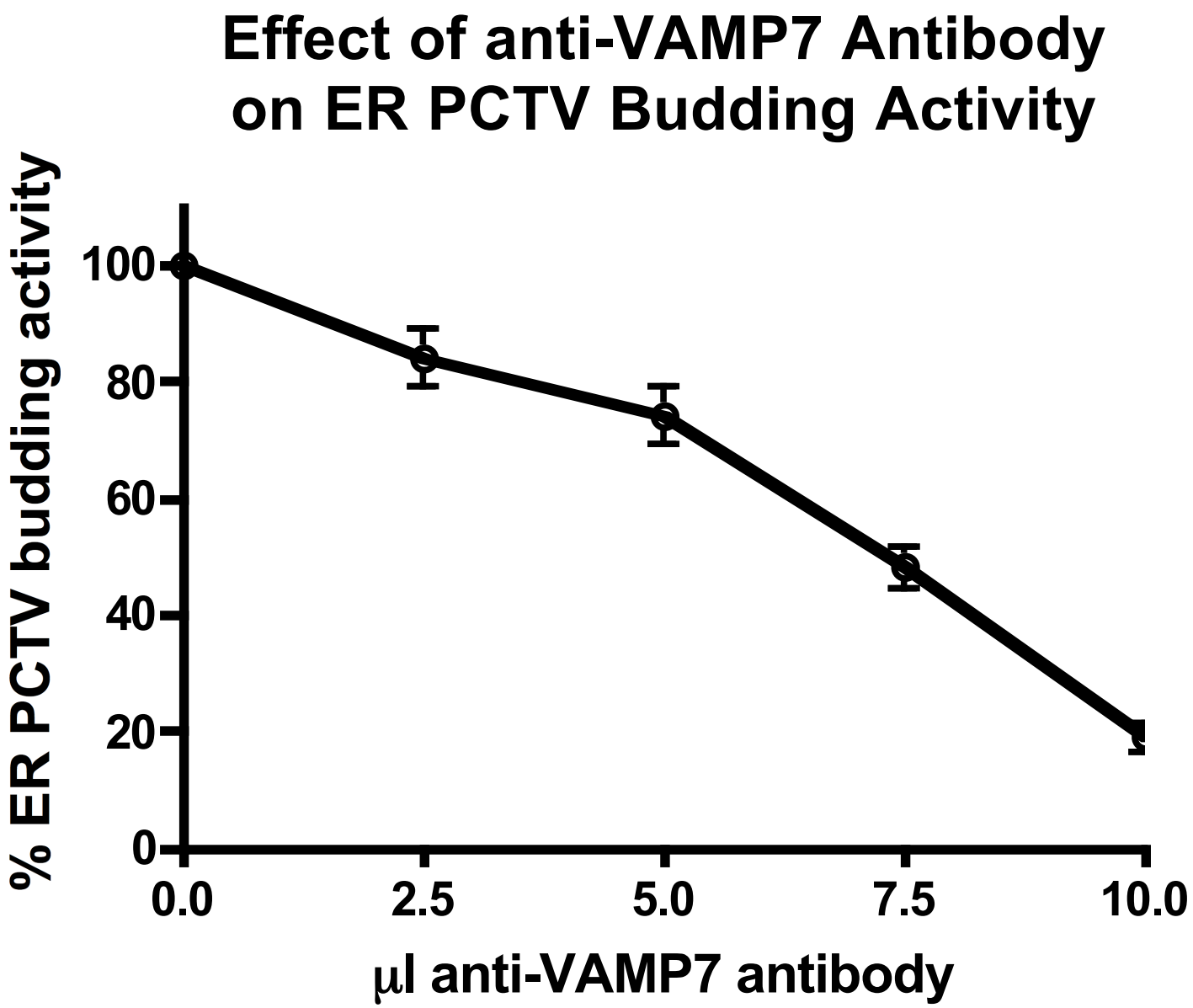
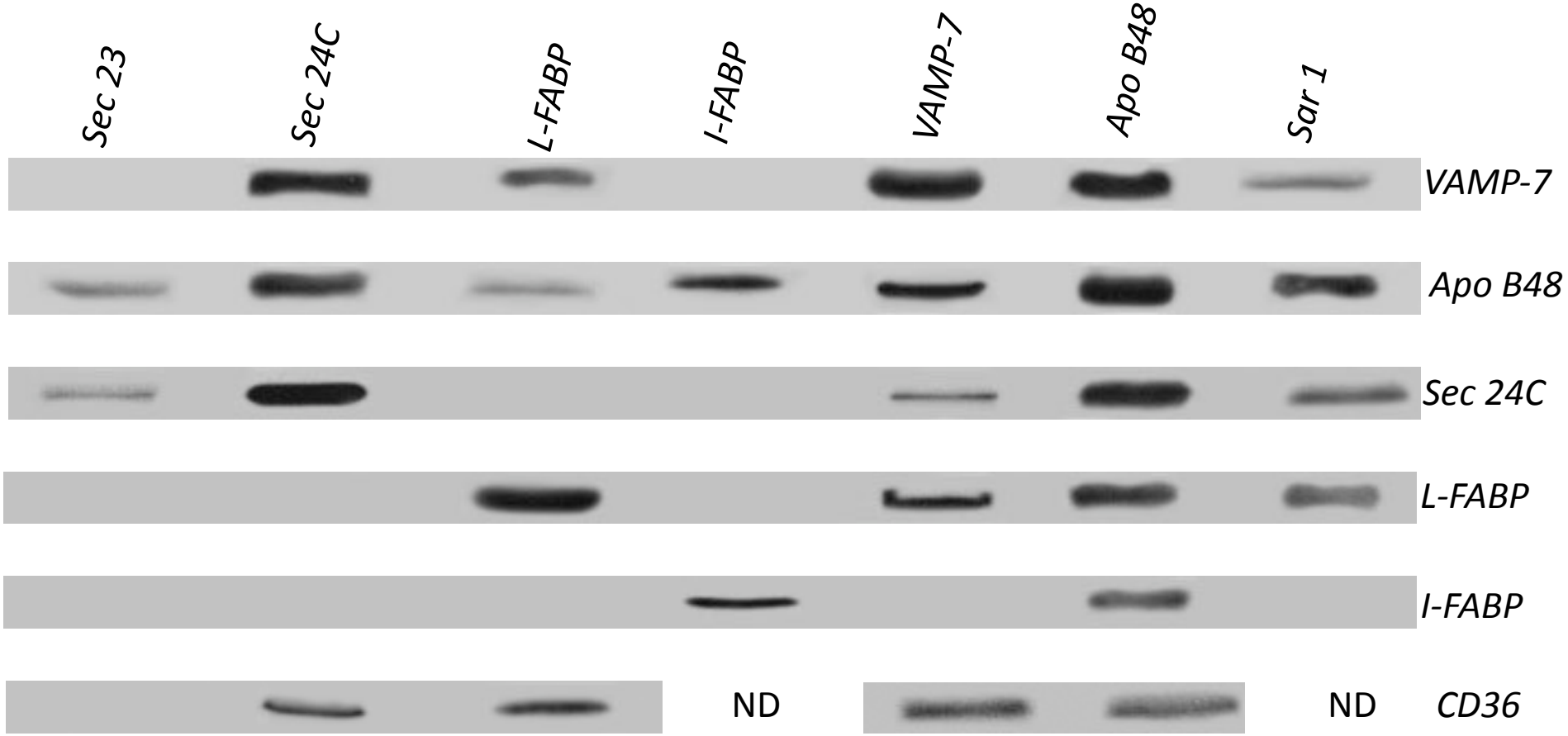
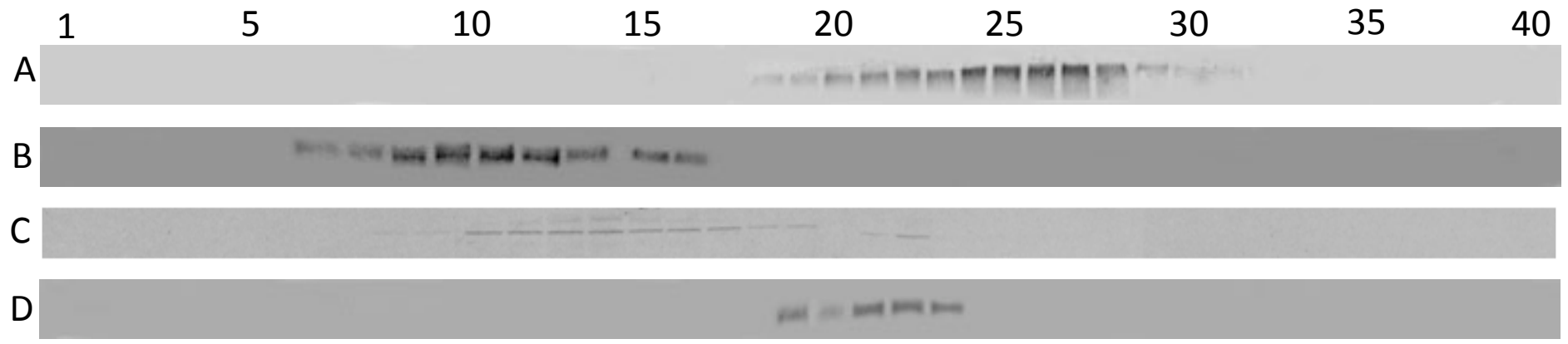

Effect of anti-VAMP7 Antibody
on ER PCTV Budding Activity
% ER PCTV budding activity
100
80
60
40
20
0
0.0
2.5
5.0
7.5
10.0
μl anti-VAMP7 antibody

Fig. S1

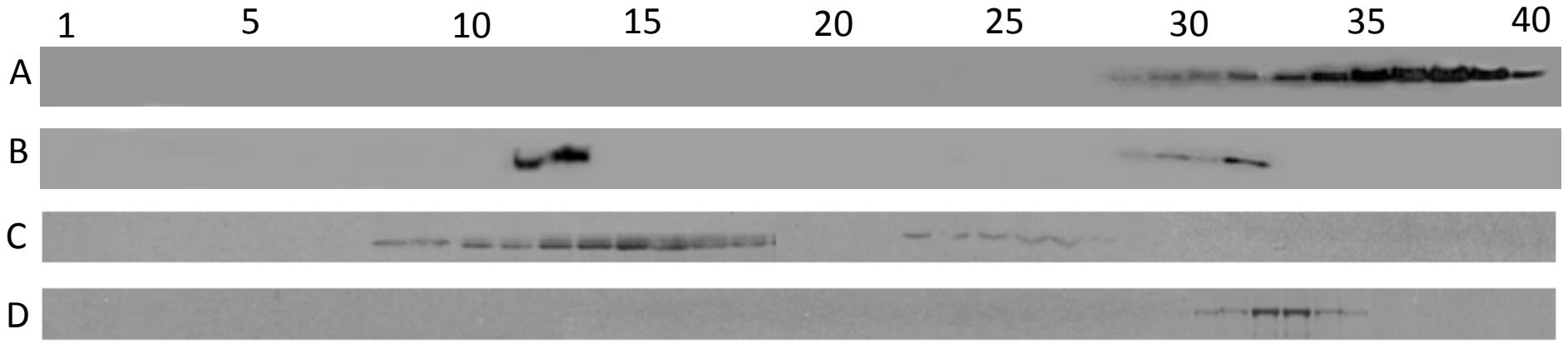


The immunoblots shown represent the original data from which Table 1 was derived. It follows exactly the same format as Table 1. Antibodies to the proteins shown on the right hand side were used for immuno-precipitation studies and the proteins associated with the initiating immuno-precipitated protein are shown at the top.

Fig. S2



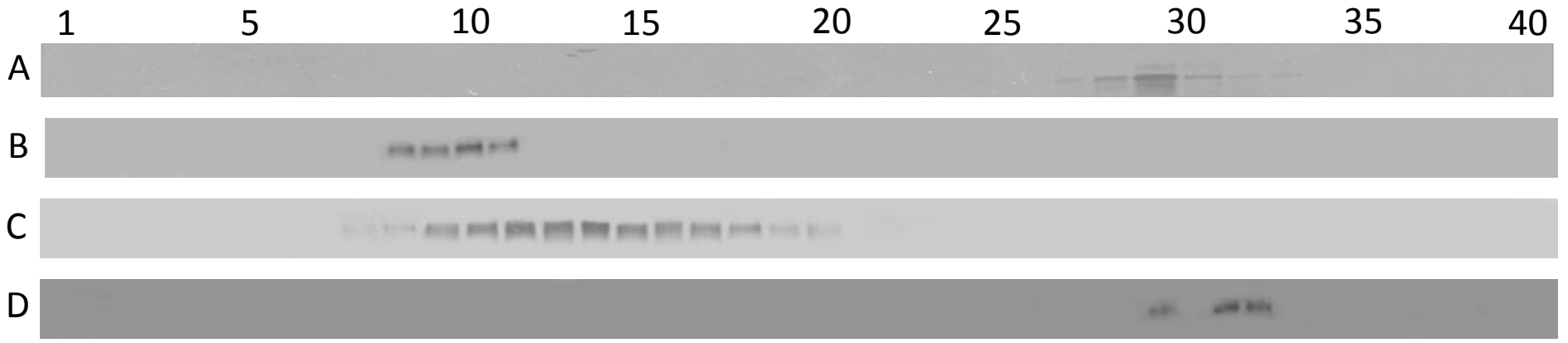
ApoB48



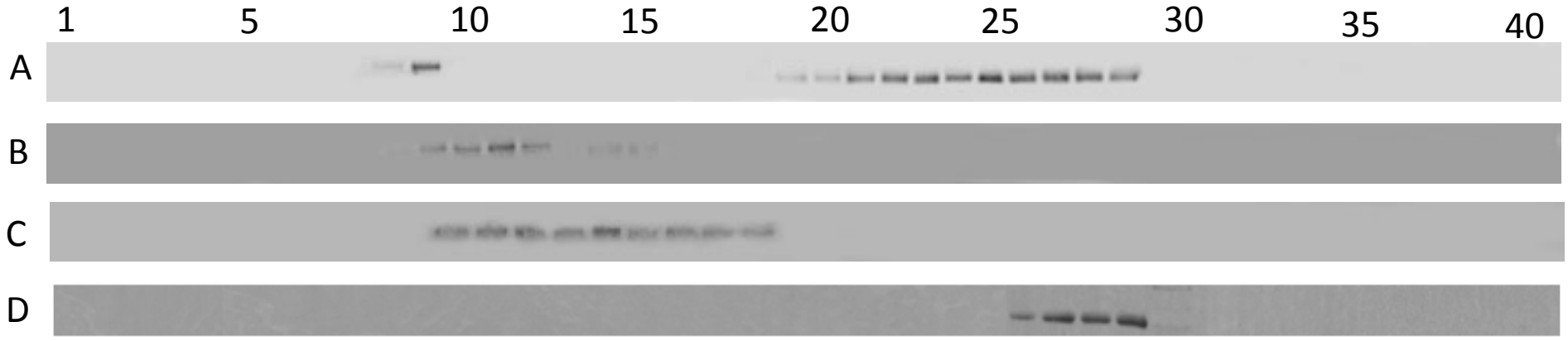
L-FABP

Immunoblots showing the distribution of apoB48 and L-FABP across the Sephacryl S400 column as indicated. These data are the original data from which the data shown in Figs. 1 and 2 were derived.

Fig. S3



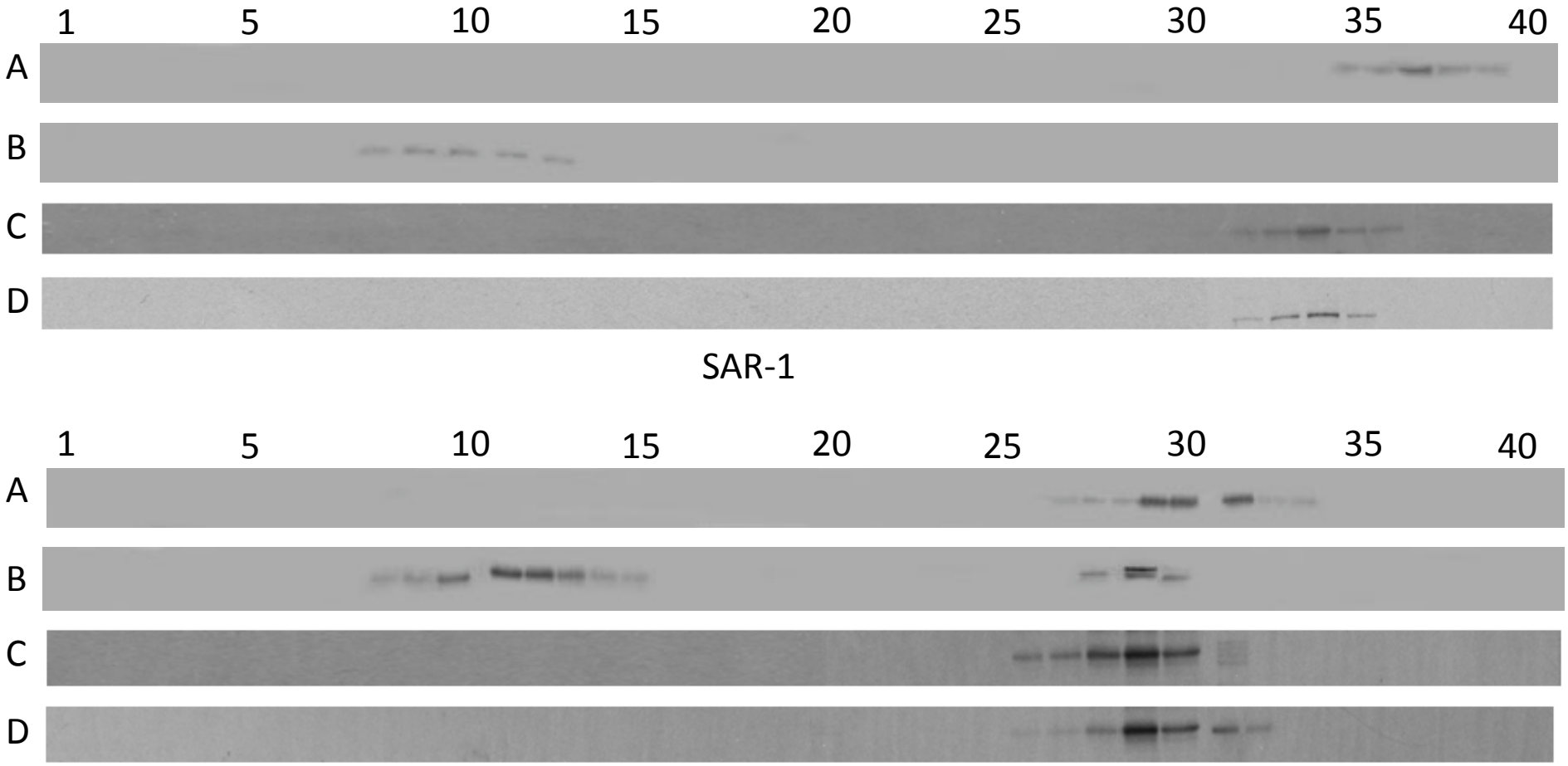
CD36



VAMP7

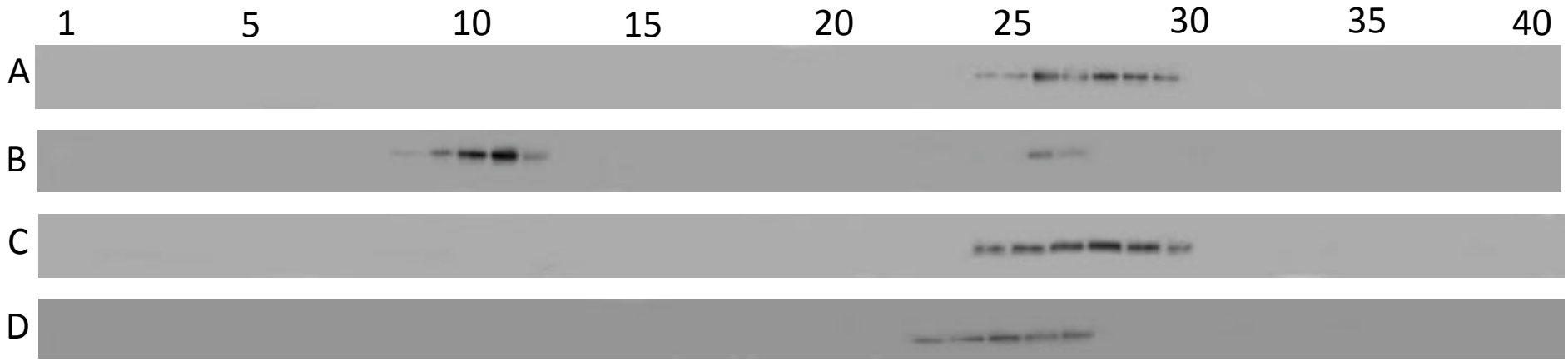
Immunoblots showing the distribution of apoB48 and L-FABP across the Sephacryl S400 column as indicated. These data are the original data from which the data shown in Figs. 3 and 4 were derived.

Fig. S4



Immunoblots showing the distribution of apoB48 and L-FABP across the Sephacryl S400 column as indicated. These data are the original data from which the data shown in Figs. 7 and 8 were derived.

Fig. S5



Sec 31

Immunoblots showing the distribution of apoB48 and L-FABP across the Sephacryl S400 column as indicated. These data are the original data from which the data shown in Fig. 9. was derived.

Fig. S6

SAR1

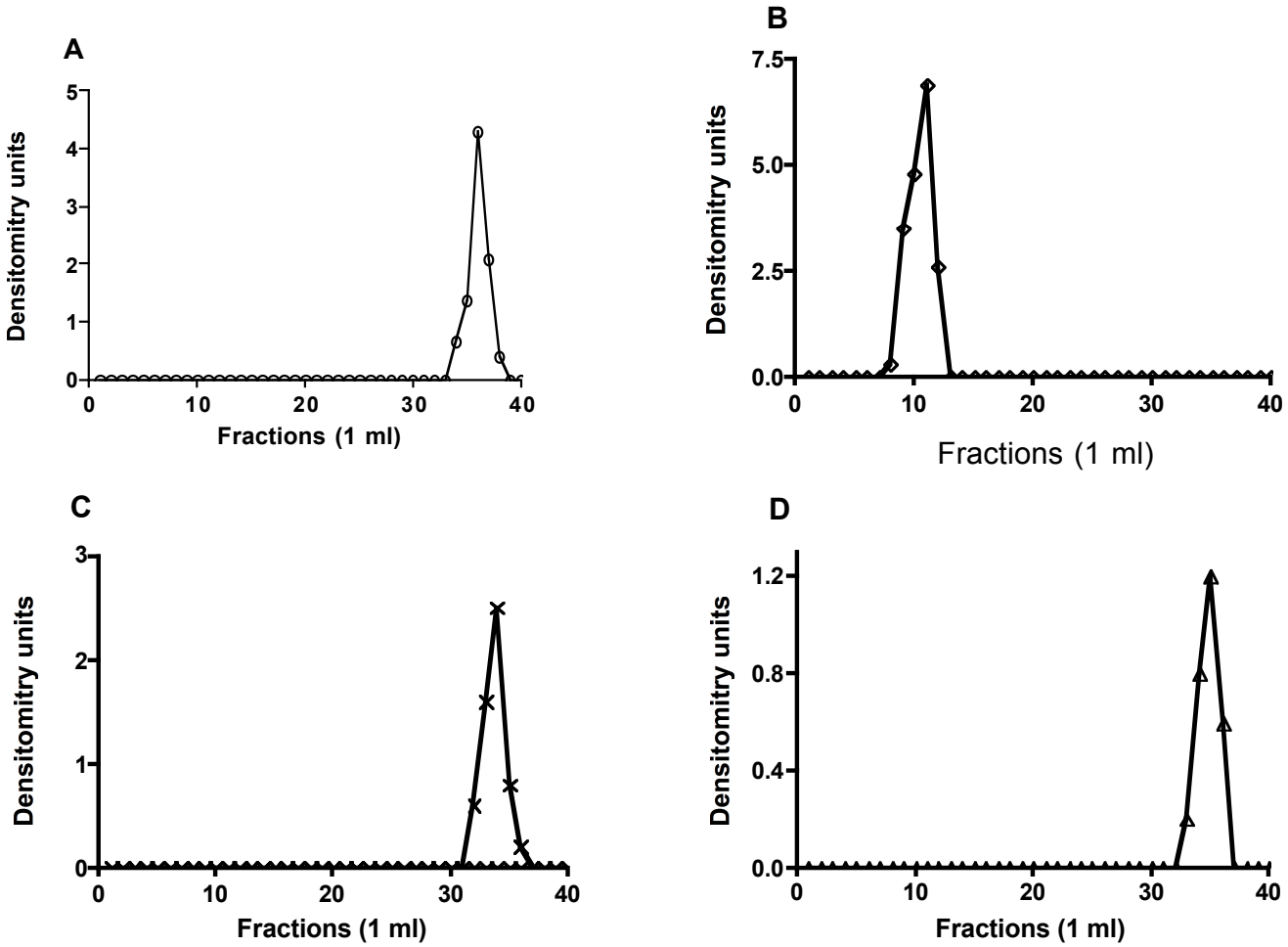

A
Densitomitry units
0
1
2
3
4
5
0
10
20
30
40
Fractions (1 ml)
B
Densitomitry units
0.0
2.5
5.0
7.5
0
10
20
30
40
Fractions (1 ml)
C
Densitomitry units
0
1
2
3
0
10
20
30
40
Fractions (1 ml)
D
Densitomitry units
0.0
0.4
0.8
1.2
0
10
20
30
40
Fractions (1 ml)

Fig. S7

Sec 24C

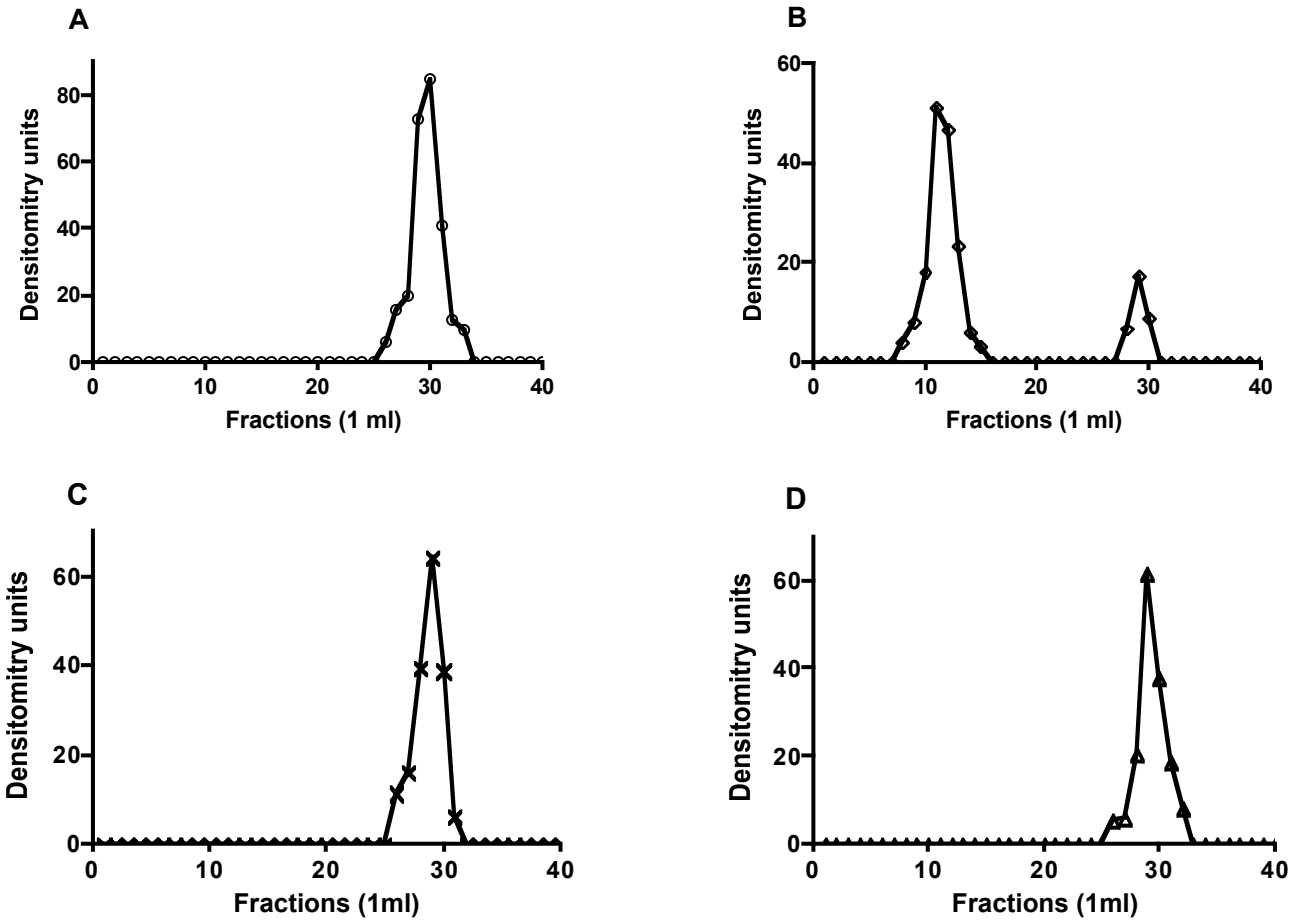


Fig. S8

Sec31

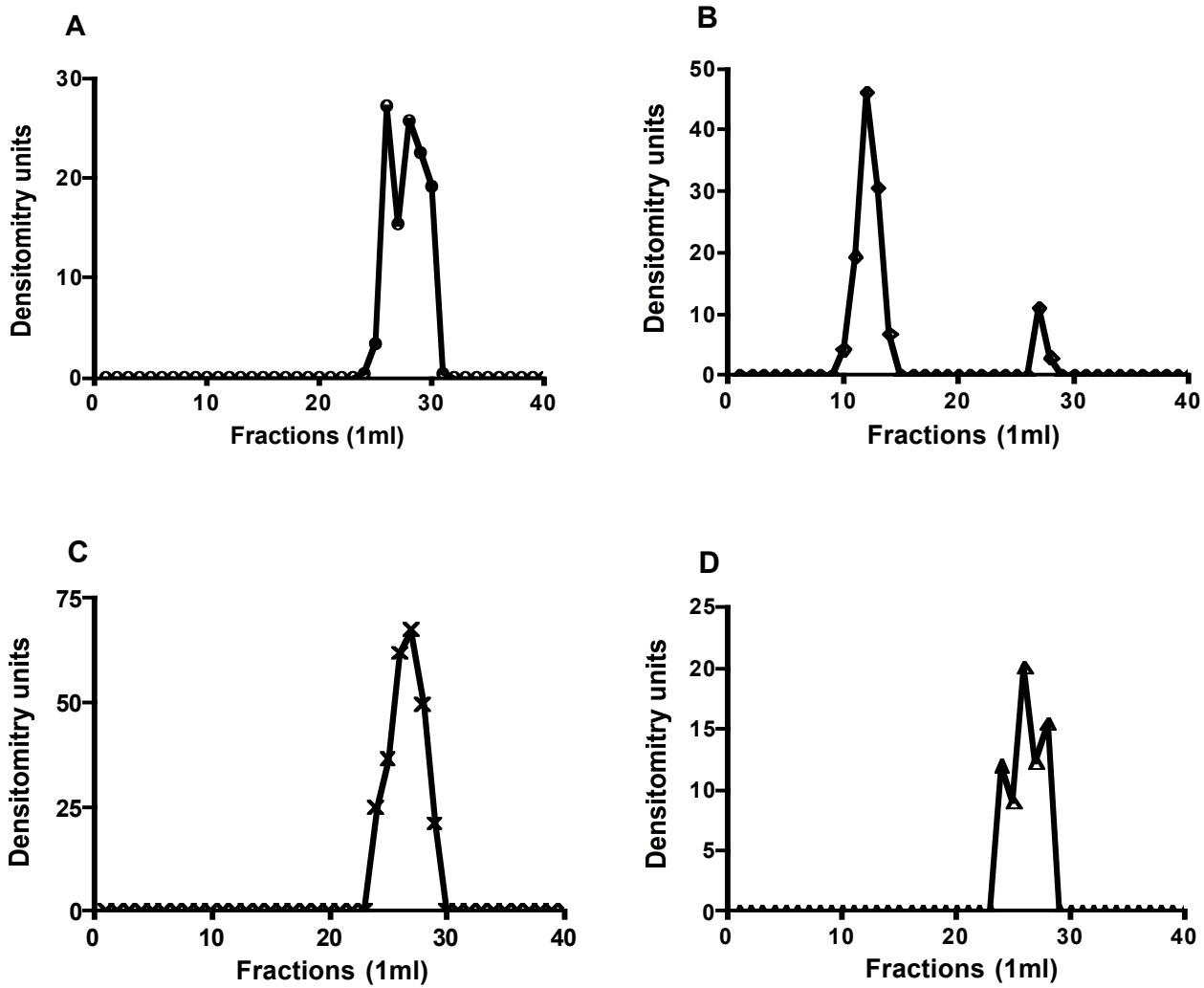

A
Densitomitry units
30
20
10
0
0
10
20
30
40
Fractions (1ml)
B
Densitomitry units
50
40
30
20
10
0
0
10
20
30
40
Fractions (1ml)
C
Densitomitry units
75
50
25
0
0
10
20
30
40
Fractions (1ml)
D
Densitomitry units
25
20
15
10
5
0
0
10
20
30
40
Fractions (1ml)

Fig. S9

Legend to Supplementary Figures:

Figure S1: Increasing amounts of anti-VAMP7 antibody progressively reduces PCTV budding activity from intestinal ER. Progressively more anti-VAMP7 antibody was incubated with intestinal ^{3}H-TAG loaded ER as indicated (Methods). The excess antibodies were removed by washing and a PCTV budding assay performed on the resulting ER. ER budding activity when no antibody was added was set to 100%. The data are the mean ± SEM, N=4.

Figure S7: Distribution of ER-Sar1 across a Sephacryl S-400 HR column under differing conditions. Native ER membranes (1mg prot) were incubated with: no cytosol (A), cytosol plus ATP (B), rL-FABP (C), PKCζ depleted cytosol and urea washed ER (D). The treated ER was solubilized, chromatographed, and the eluate immunoblotted for Sar1. The band densities were determined as in Figure 1 and reported in arbitrary units on the ordinate.

Figure S8: Distribution of ER-Sec24C across a Sephacryl S-400 HR column under differing conditions. Native ER membranes (1mg prot) were incubated with: no cytosol (A), cytosol plus ATP (B), rL-FABP (C), PKCζ depleted cytosol and urea washed ER (D). The treated ER was solubilized, chromatographed, and the eluate immunoblotted for Sec24C. The band densities were determined as in Figure 1 and reported in arbitrary units on the ordinate.

Figure S9: Distribution of ER-Sec31 across a Sephacryl S-400 HR column under differing conditions. Native ER membranes (1mg prot) were incubated with: no cytosol (A), cytosol plus ATP (B), rL-FABP (C), PKCζ depleted cytosol and urea washed ER (D). The treated ER was solubilized, chromatographed, and the eluate immunoblotted for Sec31. The band densities were determined as in Figure 1 and reported in arbitrary units on the ordinate.