

1
2 **SUPPLEMENTAL DATA**

3
4 **MATERIALS & METHODS.**

5
6 ***Skeletal muscle incubations and [³H]-2-deoxyglucose uptake.*** Muscle incubation experiments
7 were performed as previously described (1). Briefly, mice were fasted overnight and killed by
8 cervical dislocation. The soleus muscles were removed and placed in Krebs-Ringer-Bicarbonate
9 (KRB) solution containing (in mM): 117 NaCl, 4.7 KCl, 2.5 CaCl₂•2H₂O, 1.2 KH₂PO₄, 1.2
10 MgSO₄•7H₂O, 24.6 NaHCO₃) supplemented with 2 mM pyruvic acid.

11 For the basal state, muscles were incubated in KRB, 2 mM pyruvic acid, DMSO (0.2%), and
12 EtOH (0.2%) for 20 min. All other muscles were incubated in KRB, 2 mM pyruvic acid, DMSO
13 (0.2%), EtOH (0.2%), and 50 mU/ml insulin (Eli Lilly) for 20 min to stimulate the movement of
14 GLUT4 to the plasma membrane. After 20 min, muscles were transferred to vials containing
15 KRB supplemented with 1.5 μCi/ml [³H]-2-deoxyglucose, 1 mM deoxyglucose, 0.45 μCi/ml
16 [¹⁴C]-mannitol, and 7 mM cold-mannitol. For the basal state, muscles were incubated in the
17 radioactive KRB buffer. The insulin-stimulated muscles were divided into one of four groups as
18 follows: 1] 50 mU/ml insulin (insulin only); 2] wortmannin (100 nM, Sigma-Aldrich); 3]
19 cytochalasin-B (10 μM, Sigma-Aldrich); or 4] KN-62 (10 μM, Sigma-Aldrich) for 10 min.
20 Appropriate amounts of DMSO and/or EtOH were added to the radioactive buffer to maintain
21 concentrations at 0.2% for all conditions. Muscles were frozen in liquid N₂, solubilized in 1 M
22 NaOH at 80°C and neutralized with 1 M HCl. Samples were centrifuged at 11,000 x g for 1 min.
23 Aliquots were removed for scintillation counting of the [³H] and [¹⁴C] labels, and [³H]-
24 deoxyglucose uptake calculated.

25
26 ***Immunoblot analysis.*** Immunoblots were performed using standard procedures as described in
27 (1). Primary antibodies were obtained from sources as follows: phospho-ACC1/2 (Ser^{79/212})
28 from Fisher Scientific; ACC (streptavidin-HRP) from Pierce Biotechnology.

29
30
31 **REFERENCES.**

- 32 1. **Witczak CA, Fujii N, Hirshman MF and Goodyear LJ.** CaMKK{alpha} Regulates
33 Skeletal Muscle Glucose Uptake Independent of AMPK and Akt Activation. *Diabetes* 56:
34 1403-1409, 2007.

35
36
37 **FIGURE LEGENDS.**

38 **Supplemental FIG S1.** The CaMKII inhibitory peptide, AC3-I, exhibits near identical homology
39 with the inhibitory domain of CaMKII. Sequence of the CaMKII inhibitory and control peptides, and
40 their alignment with the activating phosphorylation site (i.e. Thr287) of the three main isoforms of
41 CaMKII in mouse skeletal muscle, CaMKIIβ_M, CaMKIIγ, and CaMKIIδ.

42
43 **Supplemental FIG S2.** KN-62 does not compete for the glucose binding site on glucose
44 transporters. Mouse soleus muscles with stimulated with insulin (50 mU/ml, 20 min) to induce
45 translocation of GLUT4 to the plasma membrane, and then muscles were incubated with either the
46 phosphoinositide-3-kinase inhibitor, wortmannin (100 nM), the glucose transporter inhibitor,

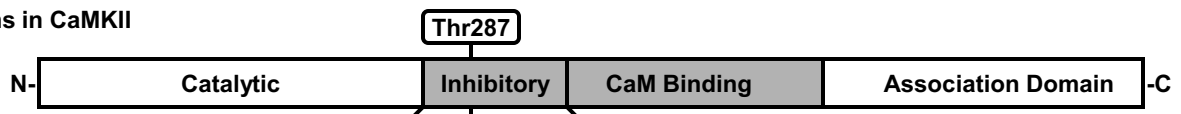
47 cytochalasin B (10 μ M) or the Ca²⁺/calmodulin-competitive inhibitor, KN-62 (10 μ M). KN-62 does not
48 inhibit muscle glucose uptake via the same mechanism as cytochalasin B, which competes with glucose
49 for the glucose binding site on glucose transporters. Statistical significance was defined as P<0.05
50 where * vs. basal, and # vs. insulin only. N = 4-6 muscles/group.

51
52 **Supplemental FIG S3. The GFP-CaMKII inhibitory peptide enhances contraction-induced**
53 **increases in acetyl-CoA carboxylase (ACC) phosphorylation.** Mouse tibialis anterior muscles were
54 transfected with DNA vectors containing either the GFP-CaMKII inhibitory peptide or the GFP-control
55 peptide. After 1 wk, mice were anesthetized and then stimulated by contraction for 15 min. Muscles
56 were harvested to assess signaling proteins by immunoblot analysis. The contraction-induced
57 phosphorylation of ACC1/2 (Ser^{79/212}) was enhanced in muscles expressing the GFP-CaMKII inhibitory
58 peptide compared to the control peptide. There was no difference in ACC1/2 protein expression.
59 Statistical significance was defined as P<0.05 where * vs. sham, and # vs. control peptide. (N=6-8
60 muscles/group)

61

Witczak et al., *AJP-Endo* – Supplemental Figure S1

Domains in CaMKII



CaMKII β_M - 281- HRQEIVECLKKF - 294

CaMKII γ - 281- HRQEIVECLRKF - 294

CaMKII δ - 281- HRQEIVDCLKKF - 294

KKAL HRQEAVDCL **GFP** Inhibitory peptide

KKAL HAQERVDCL **GFP** Control peptide

