SUPPLEMENTAL INFORMATION for Stoichiometry and energetics of poly(amidoamine) dendrimer-phospholipid interactions

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FIGURES



Figure S1: Molecular structure of the phosopholipids and PAMAM dendrimer used in this study. The PAMAM dendrimer contains protonated primary amines at pH 7.4. To reduce the total charge on the dendrimer, the terminal amines may be acetylated, as shown in the inset.¹

Lipid	Net Charge (e)	$\mathbf{T}_{\mathbf{m}}\left(^{\circ}\mathbf{C} ight)$		
DMPG	-1	23		
DMPS	-1	35		
DMPC	0	23		
DMEPC	+1	24		
PAMAM Dendrimer	Mass (kD) ^a	Dendrimer Charge $(e)^{b}$		
G5-Ac(100%)	32	0		
G3	6.9	31		
G5-Ac(40%)	29	70		
G5	27	112		
G6	46	187		
G7	106	450		
G8	260	1090		
G9	428	1880		

Table S1: Lipid and dendrimer properties.

^a Determined by gas phase chromatography with 5% uncertainty.

^b Equal to the number of primary amines per dendrimer at pH 7, as determined by acid-base titrations with 5% uncertainty.²



Figure S2: Power vs. time data and ΔH vs. molar ratio from ITC for PAMAM dendrimers titrated into DMPG SUVs at 50°C in PBS.



Figure S3: Mean diameter of G5-DMPS mixtures as measured with dynamic light scattering (DLS). DMPS was prepared as SUVs and G5 was titrated into the sample to the indicated molar ratio. This data was collected at 50 °C in PBS. Similar data has been obtained for G3, G5, G8 in DMPG (not shown). The flocculation and aggregation at higher dendrimer/lipid ratios yields particle sizes too large for DLS analysis, as visible by eye.

Dynamic light scattering (DLS) was performed on a Nanosizer ZS (Malvern Instruments Ltd.). Samples were prepared in a semi-micro cuvette to a volume of 0.5 mL and varying molar ratios. Samples were in PBS, stirred and 50 °C for > 5 min before measurement, and 50 °C during measurement, to replicate ITC conditions. All samples of dendrimers and lipids were observed to aggregate as the dendrimer/lipid molar ratio increased until the DLS was unable to determine accurately the average particle size due to the aggregates. The molar ratios at which DLS was unable to determine the average particle size were approximately n_L .

PAMAM Dendrimer	n_D^{-1}	n _L ⁻¹	Flattened- dendrimer model _{A,B}			Dendrimer-encased vesicle model ^{B,C}		
	(lipids/dendrimer)							
G3	76	140	70	-	110	1100	-	1200
G5	240	460	110	-	170	1300	-	1500
G6	410	860	170	-	270	1600	-	1900
G7	1200	2300	390	-	610	1900	-	2200
G8	1600	2700	690	-	1100	2300	-	2900
G9	1800	3800	1100	-	1700	3000	-	3800

Table S2: Stoichiometric comparison of ITC results and proposed dendrimer-lipid models

^A Assuming a lipid density of 0.58 nm²/lipid/monolayer and the maximum dendrimer extension on the surface equals that observed on mica under water.³

^B Numbers in bold and italics indicate a general agreement between the model and the ITC-determined stoichiometries.

^C Analogous to an estimate by Mecke et al.⁴ with the minimum dendrimer radius equal to that observed in simulations.

REFERENCES

1. Majoros, I. J.; Keszler, B.; Woehler, S.; Bull, T.; Baker, J. R., Acetylation of poly(amidoamine) dendrimers. *Macromolecules* **2003**, 36, (15), 5526-5529.

2. Majoros, I. J.; Thomas, T. P.; Mehta, C. B.; Baker, J. R., Poly(amidoamine) dendrimerbased multifunctional engineered nanodevice for cancer therapy. *Journal of Medicinal Chemistry* **2005**, 48, (19), 5892-5899.

3. Garidel, P.; Blume, A., 1,2-Dimyristoyl-sn-glycero-3-phosphoglycerol (DMPG) monolayers: influence of temperature, pH, ionic strength and binding of alkaline earth cations. *Chemistry and Physics of Lipids* **2005**, 138, (1-2), 50-59.

4. Mecke, A.; Majoros, I. J.; Patri, A. K.; Baker, J. R.; Holl, M. M. B.; Orr, B. G., Lipid bilayer disruption by polycationic polymers: The roles of size and chemical functional group. *Langmuir* **2005**, 21, (23), 10348-10354.