A Ruthenium-catalyzed, atom-economical synthesis of nitrogen heterocycles

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Supporting Information

Materials and Methods:

All reactions were carried out under an atmosphere of nitrogen or argon in oven-dried glassware with magnetic stirring, unless otherwise indicated. Reaction solvents were dried using J. C. Meyer's Solvent Purification System passing through activated alumina prior to use. All other reagents were purchase from commercial sources and used without further purification, unless otherwise indicated.

Flash Chromatography was performed using SiliCycle SilicaFlash F 60 40-60 μ m 60 Å silica gel. Analytical thin-layer chromatography was performed with 0.25 mm coated commercial silica gel plates (E. Merck, DC-Glasfolien, Kieselgel 60 F254) and visualized with UV light and potassium permanganate stain. Melting points were obtained on a Thomas-Hoover apparatus in open capillary tubes and are uncorrected. Proton nuclear magnetic resonance (¹H-NMR) data were acquired on a Varian Gemini 300 (300 MHz), Mercury 400 (400 MHz), Varian 400 (400 MHz), or on a Varian Unity Inova-500 (500 MHz) spectrometer. Carbon-13 nuclear magnetic resonance (¹³C-NMR) data were acquired at 100 MHz on a Mercury 400 or at 125 MHz on a Varian Unity Inova 500 spectrometer. Chemical shifts are reported in delta (δ) units, part per million (ppm) relative to deuterochloroform (7.26 ppm for ¹H NMR and 77.23 ppm for ¹³C). Infrared (IR) data were recorded as films on potassium bromide (KBr) pellets on a Thermo Scientific Nicolet IR 100 FT-IR spectrometer. High resolution mass spectra were obtained from Stanford University on a Micromass Q-Tof API US Mass Spectrometer using positive electrospray ionization (+ESI). Elemental analyses were conducted by M-H-W Laboratories, Phoenix, Az.

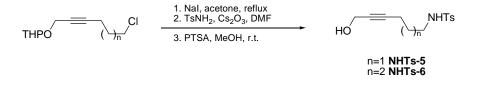
Abbreviations: Me = methyl Et = ethyl Ph = phenyl Ind = indenyl EA = ethyl acetate PE = petroleum ether

> Page S1-S15: Experimental procedures Page 16-69: Copies of spectra

Experimental procedures

<u>Substrates</u>

General procedure



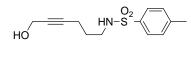
In a 25 mL flask, the chloroalkane¹ (430 mg, 2 mmol, 1 eq) is dissolved in 10 mL of acetone and NaI (1.05 g, 7 mmol, 3.5 eq) are added. The ensuing suspension is heated to reflux overnight. 15 mL of water are added to the reaction mixture which is then extracted by 3x 15 mL of ether. The combined yellow organic layers are washed with 5% Na₂S₂O3 solution, washed with brine, dried (MgSO₄) and concentrated. The light yellow oil thus obtained is pure enough for the subsequent step.

In a flame-dried 25 mL flask, the iodide is dissolved in 8 mL dry DMF. Then, $TsNH_2$ (2.4 mmol, 1.2 eq) is added followed by cesium carbonate (2.4 mmol, 1.2 eq) and the mixture is heated to 60°C for 8 h. After cooling, 5 mL of saturated aqueous NH₄Cl are added, followed by 10 mL of ether. The aqueous layer is extracted with 3x10 mL of ether and the combined organic layers are washed with 2x10 mL of brine. The ensuing organic extracts are then dried (MgSO₄) and concentrated. The resulting residue can be purified by chromatography (silica, PE/EA 4:1) or used crude in the deprotection (*vide* example towards **5a**).

¹H-NMR (400 MHz, CDCl₃): δ = 7.74 (dm, *J* = 8.0 Hz, 2H), 7.29 (dm, *J* = 7.5 Hz, 2H), 4.87-4.84 (m, 1H), 4.75 (t, *J* = 3.4 Hz, 1H), 4.25-4.10 (m, 2H), 3.84-3.78 (m, 1H), 3.53-3.48 (m, 1H), 3.03 (q, *J* = 6.4 Hz, 2H), 2.42 (s, 3H), 2.26-2.21 (m, 2H), 1.82-1.48 (m, 8H). ¹³C – NMR (100 MHz, CDCl₃): δ = 143.3, 136.9, 129.7, 127.0, 96.8, 84.8, 62.0, 54.5, 42.2, 30.2, 28.2, 25.3, 21.5, 19.0, 16.1.

The protected tosylamine is dissolved in enough methanol to make a 0.1 M solution. Then, 10 mol% PTSA is added and stirring is continued at room temperature for 3 h. At this stage, saturated aqueous NaHCO₃ is added, followed by ether. Extraction of the aqueous layer with 3 portions of ether, followed by drying (MgSO₄) and concentration affords a residue, which can be purified by short-path chromatography (silica, PE/EA 2:1) to afford typically 60-70% overall yields of **5a-6a** substrates.

¹ Haynes, R. K.; Lambert, D. E.; Parafiniuk, K. A.; Schober, P. A.; Turner, S. G. Austr. J. Chem. **1987**, 40, 273



5a

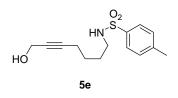
5a

IR (CHCl₃): 3435 (br), 2943, 2123, 1641, 1432, 1322, 1157 cm⁻¹.

¹H-NMR (400 MHz, CDCl₃): δ = 7.74 (dm, *J* = 8.4 Hz, 2H), 7.30 (dm, *J* = 7.5 Hz, 2H), 5.06 (t, *J* = 6 Hz, 1H), 4.19 (t, *J* = 2 Hz, 2H), 3.05 (q, *J* = 6.8 Hz, 2H), 2.42 (s, 3H), 2.27-2.23 (m, 2H), 1.66 (quin, *J* = 6.8 Hz, 2H).

¹³C – NMR (100 MHz, CDCl₃): δ = 143.7, 137.0, 130.0, 127.3, 84.9, 80.0, 51.4, 42.4, 28.3, 21.8, 16.3.

HRMS: Calculated for C₁₃H₁₇NO₃NaS: 290.0827. Found: 290.0828.



<u>5e</u>

IR (CHCl₃): 3433 (br), 2947, 2120, 1645, 1433, 1322, 1158 cm⁻¹.

¹H-NMR (400 MHz, CDCl₃): δ = 7.76 (dm, *J* = 7.4 Hz, 2H), 7.31 (dm, *J* = 7.5 Hz, 2H), 4.95 (br s, 1H), 4.22 (s, 2H), 2.95 (q, *J* = 5.7 Hz, 2H), 2.43 (s, 3H), 2.21-2.17 (m, 2H), 1.61-1.58 (m, 2H), 1.52-1.49 (m, 2 H).

¹³C – NMR (100 MHz, CDCl₃): δ = 143.4, 136.7, 129.7, 127.0, 85.5, 79.1, 51.2, 42.6, 28.5, 25.2, 21.5, 18.2.

HRMS: Calculated for C₁₄H₁₉NO₃NaS: 304.0983. Found: 304.0981.

Preparation of N-tosylalkynyl amines

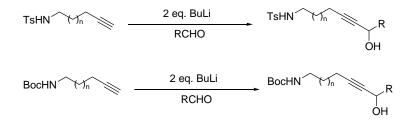
HO \mathcal{V}_n 1. MsCl, NEt₃ TsHN \mathcal{V}_n 2. TsNH₂, Cs₂CO₃ $n = 1 \frac{NM I-97}{n}$ $n = 2 \frac{NM II-11}{NM I-11}$

Sample procedure

In a flame-dried 100 mL flask kept under Argon, pent-4-ynol (10 mmol, 1 eq) is dissolved in 40 mL of dichloromethane. To this solution, triethylamine (30 mmol, 3 eq) is added and the resulting mixture is cooled to 0° C. Then, methanesulfonyl chloride (14 mmol, 1.4 eq) is added dropwise, causing the reaction mixture to turn from light yellow to a turbid orange mixture. After stirring for 3 hours, 15 mL of water are added. Following phase separation, the aqueous layer is extracted with 3x15 mL of dichloromethane and the combined organic extracts are washed with 2x20 mL of brine. The ensuing organic extracts are then dried (MgSO₄) and concentrated. The crude orange oil thus obtained is directly dissolved in 20 mL DMF. Then,

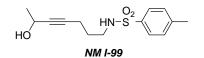
TsNH₂ (16 mmol, 1.6 eq) and Cs₂CO₃ (12 mmol, 1.2 eq) are sequentially added and the mixture is heated to 120° C. After completion (TLC monitoring), the reaction mixture is cooled down to room temperature and 15 mL saturated aqueous NH₄Cl are added, followed by 10 mL of ether. The aqueous layer is extracted with 3x10 mL of ether and the combined organic layers are washed with 2x10 mL of brine. The ensuing organic extracts are then dried (MgSO₄) and concentrated. The resulting residue is purified by chromatography (silica, PE/EA 4:1 to 1:2) to afford typically 45-55% yields of product. NMR data match those previously reported for these compounds (cf. *J. Am. Chem. Soc.* **2007**, *129*, 3794).

Addition to aldehydes



General procedure

In a flame-dried 25 mL flask kept under Argon, the alkynyl amine (1 mmol, 1 eq) is dissolved in 4 mL of THF and cooled down to -78 °C. To this solution is added dropwise *n*-BuLi (2.2 mmol, 2.2 eq) and stirring is continued at the same temperature for 1 hour. At this point, the corresponding aldehyde is added neat (5 mmol, 5 eq) and the mixture is gradually warmed to room temperature before quenching with 15 mL saturated aqueous NH_4Cl , followed by 10 mL of ether. Extractive workup followed by chromatography then affords the pure amino alcohols.



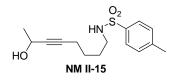
<u>5b</u>

IR (CHCl₃): 3430, 3291, 2936, 1657, 1597, 1431, 1329, 1157 cm⁻¹.

¹H-NMR (500 MHz, CDCl₃): δ = 7.76 (d, *J* = 8.4 Hz, 2H), 7.31 (dm, *J* = 8.4 Hz, 2H), 5.11 (t, *J* = 6.4 Hz, 1H), 4.46 (tq, *J* = 6.4, 2.0 Hz, 1H), 3.06 (q, *J* = 6.8 Hz, 2H), 2.43 (s, 3H), 2.08-2.04 (m, 2H), 2.04 (br s, 1H), 1.66 (quin, *J* = 6.8 Hz, 2H), 1.39 (d, *J* = 6.4 Hz, 3H).

 13 C – NMR (100 MHz, CDCl₃): δ = 143.7, 137.1, 130.0, 127.4, 84.0, 83.1, 58.6, 42.5, 28.3, 24.8, 21.8, 16.2.

HRMS: Calculated for C₁₄H₁₉NO₃NaS: 304.0983. Found: 304.0981.



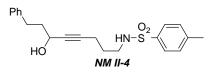
<u>6f</u>

IR (CHCl₃): 3430, 3201, 2974, 2936, 1657, 1598, 1431, 1329, 1157cm⁻¹.

¹H-NMR (400 MHz, CDCl₃): $\delta = 7.75$ (d, J = 8.0 Hz, 2H), 7.30 (d, J = 8.0 Hz, 2H), 5.19 (m, 1H), 4.47 (qm, J = 6.8, 1H), 2.92 (q, J = 6.7 Hz, 2H), 2.43 (s, 3H), 2.16 (dt, J = 6.8, 1.8 Hz, 2H), 2.05 (d, J = 7.2 Hz, 1H), 1.59 (quin, J = 6.8 Hz, 2H), 1.49 (quin, J = 6.8 Hz, 2H), 1.39 (d, J = 6.4 Hz, 3H), 1.28-1.19 (m, 2H).

¹³C – NMR (100 MHz, CDCl₃): δ = 143.3, 137.0, 129.6, 127.0, 83.5, 82.9, 58.3, 42.5, 28.4, 25.2, 24.5, 21.4, 18.0.

HRMS: Calculated for C₁₅H₂₁NO₃NaS: 318.1140. Found: 318.1141.



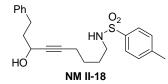
<u>5c</u>

IR (CHCl₃): 3422, 2936, 2872, 2366, 2334, 1649, 1496, 1451, 1330, 1157 cm⁻¹.

¹H-NMR (500 MHz, CDCl₃): $\delta = 7.75$ (dm, J = 8.5 Hz, 2H), 7.29-7.26 (m, 4H), 7.20-7.17 (m, 3H), 4.99 (t, J = 6.25 Hz, 1H), 4.31 (tm, J = 6.5 Hz, 1H), 3.06 (q, J = 6.5 Hz, 2H), 2.74 (t, J = 8.0 Hz, 2H), 2.40 (s, 3H), 2.36 (d, J = 4.5 Hz, 1H), 2.26 (dt, J = 6.8, 2.0 Hz, 2H), 1.99-1.92 (m, 2H), 1.67 (quin, J = 6.9 Hz, 2H).

¹³C – NMR (125 MHz, CDCl₃): δ = 143.4, 141.3, 136.8, 129.7, 128.4, 128.4, 127.0, 125.9, 84.0, 82.4, 61.8, 42.1, 39.4, 31.4, 28.1, 21.5, 16.0.

HRMS: Calculated for C₂₁H₂₅NO₃NaS: 394.1453. Found: 394.1451.



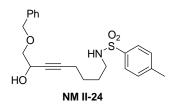
<u>5g</u>

IR (CHCl₃): 3418, 2937, 2348, 2233, 1638, 1597, 1469, 1306, 1157 cm⁻¹.

¹H-NMR (400 MHz, CDCl₃): $\delta = 7.74$ (dm, J = 8.3 Hz, 2H), 7.32-7.26 (m, 4H), 7.22-7.18 (m, 3H), 4.70 (br s, 1H), 4.33 (tm, J = 6.5 Hz, 1H), 2.96 (q, J = 6.7 Hz, 2H), 2.76 (tm, J = 7.9 Hz, 3H), 2.42 (s, 3H), 2.20 (dt, J = 6.7, 1.8 Hz, 2H), 2.03-1.92 (m, 2H), 1.63-1.57 (m, 2H), 1.54-1.47 (m, 2H).

¹³C – NMR (125 MHz, CDCl₃): δ = 143.4, 141.4, 136.8, 129.7, 129.0, 128.4, 127.0, 125.9, 84.9, 81.8, 61.8, 42.6, 39.5, 31.4, 28.6, 25.8, 21.5, 18.2.

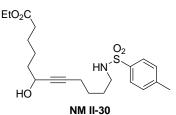
HRMS: Calculated for C₂₂H₂₇NO₃NaS: 408.1609. Found: 408.1609.



<u>5h</u>

 $\overline{\text{IR}}$ (CHCl₃): 3430, 2928, 2859, 2234, 2090, 1650, 1458, 1341, 1162, 1094 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ = 7.73 (dm, *J* = 8.3 Hz, 2H), 7.38-7.28 (m, 7H), 4.86 (br s, 1H), 4.61-4.54 (m, 1H), 4.57 (d, *J* = 2.8 Hz, 2H), 3.60-3.48 (AB m, 2H), 2.90 (q, *J* = 6.6 Hz, 2H), 2.77 (br s, 1H), 2.41 (s, 3H), 2.15 (dt, *J* = 6.4, 1.8 Hz, 2H), 2.03-1.92 (m, 2H), 1.58-1.43 (m, 4H).

¹³C – NMR (100 MHz, CDCl₃): δ = 143.3, 137.6, 136.8, 129.6, 128.4, 127.8, 127.7, 127.0, 94.3, 85.5, 78.4, 73.8, 73.2, 73.2, 71.3, 61.8, 61.7, 42.5, 28.4, 25.1, 21.4, 18.1. HRMS: Calculated for C₁₄H₁₉NO₃NaS: 424.1559. Found: 424.1556.



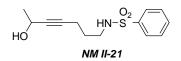
<u>5d</u>

IR (CHCl₃): 3423, 2941, 2864, 1725, 1711, 1660, 1636, 1446, 1340, 1168 cm⁻¹.

¹H-NMR (400 MHz, CDCl₃): δ = 7.74 (dm, *J* = 8.3 Hz, 2H), 7.30 (dm, *J* = 8.0 Hz, 2H), 4.97 (t, *J* = 6.3 Hz, 1H), 4.30 (t, *J* = 6.5 Hz, 1H), 4.12 (q, *J* = 7.1 Hz, 2H), 3.05 (q, *J* = 6.6 Hz, 2H), 2.42 (s, 3H), 2.30 (t, *J* = 7.4, 2H), 2.25 (dt, *J* = 6.8, 1.9 Hz, 1H), 1.70-1.45 (m, 8 H), 1.24 (t, *J* = 7.1 Hz, 3H).

 ^{13}C – NMR (100 MHz, CDCl₃): δ = 173.8, 143.4, 136.9, 129.7, 127.0, 94.4, 83.8, 82.5, 62.2, 60.3, 42.1, 37.5, 34.1, 28.1, 24.6, 24.5, 21.5, 15.9, 14.2.

HRMS: Calculated for C₂₀H₂₉NO₅NaS: 418.1664. Found: 418.1660.

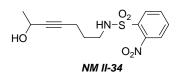


7i

IR (CHCl₃): 3432, 2935, 2858, 2372, 1661, 1140, 1028 cm⁻¹.

¹H-NMR (400 MHz, CDCl₃): δ = 7.89-7.86 (m, 2H), 7.60-7.49 (m, 3H), 5.10 (br s, 1H), 4.46 (qm, *J* = 6.4 Hz, 1H), 3.07 (q, *J* = 6.4 Hz, 2H), 2.32 (br s, 1H), 2.23 (dt, *J* = 6.8, 2.0 Hz, 2H), 1.66 (quin, *J* = 6.4 Hz, 2H), 1.38 (d, *J* = 6.4 Hz, 3H).

¹³C – NMR (100 MHz, CDCl₃): δ = 139.8, 132.6, 129.1, 127.0, 83.7, 82.8, 58.3, 42.2, 28.0, 24.5, 15.9. HRMS: Calculated for C₁₃H₁₇NO₃NaS: 290.0827. Found: 290.0822.



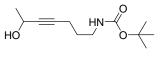
<u>7j</u>

IR (CHCl₃): 3392, 2921, 1640, 1543, 1416, 1332, 1163 cm⁻¹.

¹H-NMR (400 MHz, CDCl₃): $\delta = 8.14-8.11$ (m, 1H), 7.87-7.84 (m, 1H), 7.77-7.71 (m, 2H), 5.55 (t, J = 6.3 Hz, 1H), 4.47 (qm, J = 6.0 Hz, 1H), 3.22 (q, J = 6.4 Hz, 2H), 2.28 (dt, J = 6.6, 2.0 Hz, 2H), 2.21 (br s, 1H), 1.73 (quin, J = 6.8 Hz, 2H), 1.39 (d, J = 6.8 Hz, 3H).

¹³C – NMR (100 MHz, CDCl₃): δ = 147.9, 133.6, 133.5, 132.8, 131.0, 125.4, 83.8, 82.4, 58.4, 42.8, 28.1, 24.5, 15.9.

HRMS: Calculated for C₁₃H₁₆N₂O₅NaS: 335.1137. Found: 335.1137.

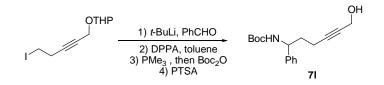




<u>7k</u>

IR (CHCl₃): 3390, 2982, 2926, 2350, 2082, 1691, 1640, 1522, 1377, 1254, 1165 cm⁻¹. ¹H-NMR (500 MHz, CDCl₃): δ = 4.74 (br s, 1H), 4.47 (qm, *J* = 6.5 Hz, 1H), 3.22 (q, *J* = 6.4 Hz, 2H), 2.56 (br s, 1H), 2.24 (dt, *J* = 6.7, 2.0 Hz, 2H) 1.66 (quin, *J* = 7.0 Hz, 2H), 1.42 (s, 9H), 1.39 (d, *J* = 6.5 Hz, 3H).

¹³C – NMR (100 MHz, CDCl₃): δ = 155.9, 83.3, 83.2, 79.3, 58.3, 39.6, 28.5, 228.4, 24.5, 16.1. HRMS: Calculated for C₁₂H₂₁NO₃Na: 250.1419. Found: 250.1421.



The iodide² (1.47 g, 5.00 mmol) was dissolved in freshly distilled ether (50 mL) in a 250-ml flask. The solution was cooled to -78° C and *t*-BuLI (1.7 M in pentane, 6.47 mL, 11.0 mmol) was added. The mixture was stirred at the same temperature for 30 min and freshly distilled benzaldehyde (1.02 mL, 1.06 g, 10.0 mmol) was added via syringe. The mixture was allowed to

² Nishibayashi, Y.; Milton, M. D.; Inada, Y.; Yoshikawa, M.; Wakiji, I.; Hidai, M.; Uemura, S. *Chem. Eur. J.* **2005**, *11*, 1433

warm to room temperature and was quenched at -78°C with acetic acid (1.2 eq); upon warming back to room temperature water was added. Following aqueous workup and chromatographic purification, the intermediate alcohol (930 mg, 67% yield) was carried forward without complete characterization.

¹H NMR (300 MHz, CDCI3): δ = 7.36-7.25 (m, 5H), 4.82 (dd, *J* = 7.9, 5.1 Hz, 2H), 4.30 (dt, *J* = 15.3, 2.2 Hz, 1H), 4.20 (dd, J = 15.2, 2.1 Hz, 1H), 3.88-3.80 (m, 1H), 3.56-3.49 (m, 1H), 2.41-2.27 (m, 2H), 2.02-1.51 (m, 9H)

The previous alcohol (685 mg, 2.50 mmol) was dissolved in freshly distilled toluene (3 mL) and the solution was cooled in an ice bath. Diphenylphosphorazidate (648 μ L, 3.00 mmol) and DBU (449 μ L, 3.00 mmol) were added and the mixture was stirred at 0 °C for 2 h. The mixture was allowed to warm to room temperature and stirring was continued for 48 h. The reaction was worked up with water (10 mL) and ether (3x10 mL) and the combined extracts were dried over magnesium sulfate. Removal of the solvent under reduced pressure and flash chromatography on silica gel (10:1 PE: EA) gave 663 mg of clear oil (89% yield). This intermediate was carried forward without complete characterization.

IR(film): 2944, 2870, 2096, 1455, 1346, 1247, 1202, 1118, 1024, 903, 872, 762 cm⁻¹. ¹H NMR (300 MHz, CDCI3): δ = 7.42-7.30 (m, 5H), 4.82 (t, *J* = 3.2 Hz, 1H), 4.62 (dd, *J* = 8.2, 6.4 Hz, 1H), 4.32 (dt, *J* =15.3, 2.2 Hz, 1H), 4.22 (dt, *J* =15.3, 2.1 Hz, 1H), 3.89-3.82 (m, 1H), 3.58-3.51 (m, 1H), 2.38-2.1.9 (m, 2H), 2.07-1.52 (m, 8H). ¹³C NMR (75 MHz, CDCI3) δ = 139.1, 128.9 (2), 128.4, 126.9 (2), 96.7, 84.7, 77.0, 64.8, 61.9, 54.4, 34.8, 30.1, 25.2, 18.9, 15.7.

To the above azide (599 mg, 2.00 mmol) in THF (10 mL) and water (5 mL) was added trimethyl phosphine (2.40 mL, 1.0M, 2.40 mmol). After stirring 4 h, triethylamine (1 mL) and di-t-butyl dicarbonate (524 mg, 2.40 mmol) were added and the mixture was stirred overnight. The solvent was removed under vacuum and the residue was taken up in methanol. Catalytic PTSA was added and the mixture was stirred 2 h at room temperature. Removal of the solvent under vacuum and flash chromatography on silica (1:1 hexanes: ethyl acetate) gave 480 mg white solid (83% yield).

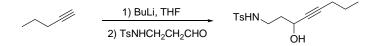
IR(film): 3331, 2978, 2932, 2225,1689, 1526, 1367, 1251, 1168, 1026, 701 cm⁻¹.

¹H NMR (300 MHz, CDCl₃): δ = 7.37-7.26 (m, 5H), 4.91 (br s, 1H), 4.82 (br s, 1H), 4.23 (s, 2H), 2.24 (d, J = 7.1 Hz, 2H), 1.98 (d, J = 6.6 Hz, 1H), 1.61 (s, 1H), 1.42 (s, 9H).

¹³C NMR (75 MHz, CDCl₃) δ = 155.2, 141.9, 128.6 (2), 127.3, 126.3 (2), 84.8, 79.7 (2), 53.9, 51.0, 35.3, 28.3 (3), 15.8.

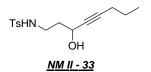
HRMS: Calculated for C₁₃H₁₅NO₃ (M⁺ - C₄H₈): 233.1052. Found 233.1057.

Piperidinone substrates



Sample procedure: 8a (NM II-33)

In a flame-dried 25 mL flask, 1-hexyne (0.26 mL, 2.6 mmol, 3eq) are dissolved in 8 mL dry THF and cooled to -78 °C. To this solution is added BuLi (1.1 mL of a 2.5M hexanes solution, 2.7 mmol, 3.1eq) and stirring is continued at this temperature for 1 hour. Then, the N-tosylaldehyde (200 mg, 0.88 mmol, 1eq) dissolved in 4 mL dry THF is added dropwise. After addition is completed, the reaction is slowly warmed up to room temperature. After reaching room temperature, 5 mL of saturated aqueous NH₄Cl are added, followed by 5 mL of ether. Following conventional extractive workup, purification by chromatography (silica, PE/AcEt 4:1 to 1:1) affords 143 mg (55% yield) of the aminoalcohol.



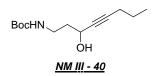
<u>8a</u>

IR (CHCl₃): 3417, 3257, 2957, 2127, 1654, 1437, 1328, 1169 cm⁻¹.

¹H-NMR (400 MHz, CDCl₃): $\delta = 7.74$ (d, J = 8.4 Hz, 2H), 7.29 (dd, J = 8.0, 0.4 Hz, 7H), 5.26 (t, J = 6.0 Hz, 1H), 4.48 (br s, 1H), 3.13 (apparent dsext, J = 6.4 Hz, 2H), 2.43 (s, 3H), 2.33 (d, J = 4.0 Hz, 1H), 2.12 (tm, J = 7.2 Hz, 2H), 1.91-1.76 (m, 2H), 1.52-1.43 (m, 2H), 0.93 (t, J = 7.4 Hz, 3H).

¹³C – NMR (100 MHz, CDCl₃): δ = 143.6, 137.0, 129.9, 127.4, 86.8, 80.3, 61.4, 40.3, 36.8, 22.1, 21.8, 20.8, 13.7.

HRMS: Calculated for C₁₅H₂₁NO₃NaS: 318.1140. Found: 318.1138.



<u>8b</u>

IR (CHCl₃): 3411, 2968, 2358, 2121, 1653, 1522 cm⁻¹.

¹H-NMR (400 MHz, CDCl₃): $\delta = 5.04$ (br s, 1H), 4.40 (br s, 1H), 3.46-3.41 (m, 1H), 3.35 (apparent sext, J = 6.5 Hz, 1H), 3.23-3.14 (m, 1H), 2.12 (dt, J = 7.2, 2.0 Hz, 2H), 1.79 (q, J = 6.2 Hz, 2H), 1.47 (sext, J = 7.2 Hz, 2H), 1.38 (s, 9H), 0.92 (t, J = 7.2 Hz, 3H).

¹³C – NMR (100 MHz, CDCl₃): δ = 156.6, 85.3, 80.7, 79.3, 60.0, 38.0, 36.8, 28.3, 28.2, 21.9, 20.5, 13.4.

HRMS: Calculated for C₁₃H₂₃NO₃Na: 264.1576. Found: 264.1577.

Tandem Redox isomerisation/Cyclisation

General procedure

The substrate is dissolved in enough dry THF to make a 0.1 M solution, in a flame-dried flask fitted with reflux condenser. To this solution is then added the ruthenium catalyst (3 mol%) and CSA (5 mol%), and the resulting reddish mixture is stirred at room temperature from 5 minutes.

At this stage, indium triflate (3 mol%) is added and the solution is brought to reflux in a preheated oil bath. After 1 h stirring, the mixture is filtered through a plug of florisil, rinsing with ether. The filtrate is concentrated and purified by chromatography (silica gel, PE/EA 2:1 to 1:1).

NB: In the case of piperidine formation, 10 mol% CSA were used instead.

<u>NM I-60</u>



<u>6a</u>

IR: 3267, 2957, 2924, 2360, 1720 (sh), 1597, 1451, 1341, 1158, 1092 cm⁻¹.

¹H-NMR (500 MHz, CDCl₃): δ = 9.80 (s, 1H), 7.72 (dm, *J* = 8.0 Hz, 2H), 7.31 (dm, *J* = 7.5 Hz, 2H), 4.00 (sept, *J* = 4.5 Hz, 1H) 3.46-3.41 (m, 1H), 3.20-3.09 (m, 2H), 2.74 (qd, *J* = 5.4, 1.5 Hz, 1H), 2.43 (s, 3H), 1.86-1.73 (m, 2H), 1.58-1.47 (m, 2H).

¹³C – NMR (125 MHz, CDCl₃): δ = 200.7, 143.7, 133.8, 129.8, 127.6, 54.8, 50.9, 49.0, 32.0, 23.9, 21.5.

HRMS: Calculated for C₁₃H₁₇NO₃NaS: 290.0827. Found: 290.0826.

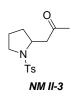


CAS: 934472-39-6

<u>6e</u>

¹H-NMR (500 MHz, CDCl₃): δ = 9.68 (t, *J* = 2.0 Hz, 1H), 7.71 (dm, *J* = 8.0 Hz, 2H), 7.31 (d, *J* = 7.5 Hz, 2H), 4.68-4.64 (m, 1H), 3.80 (dm, *J* = 12.0 Hz, 1H), 2.94 (dt, *J* = 12.7, 2.5 Hz, 1H), 2.67-2.65 (m, 2H), 2.42 (s, 3H), 1.57-1.42 (m, 4H), 1.32-1.24 (m, 2H). ¹³C – NMR (125 MHz, CDCl₃): δ = 200.1, 143.3, 137.9, 129.8, 127.0, 47.6, 43.9, 41.0, 27.8, 24.3, 21.5, 18.3.

HRMS: Calculated for C₁₄H₁₉NO₃NaS: 304.0983. Found: 304.0983.

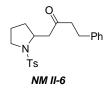


IR: 3435, 2957, 2862, 2145, 1729, 1654, 1603, 1339, 1156 cm⁻¹.

¹H-NMR (500 MHz, CDCl₃): $\delta = 7.70$ (d, J = 8.5 Hz, 2H), 7.31 (dd, J = 8.0, 0.5 Hz, 2H), 3.90 (sept, J = 3.5 Hz, 1H), 3.45-3.40 (m, 1H), 3.23 (dd, J = 17.8, 3.2 Hz, 1H), 3.06 (ddd, J = 10.0, 7.5, 7.5 Hz, 1H), 2.42 (s, 3H), 2.16 (s, 3H), 1.83-1.67 (m, 2H), 1.54-1.43 (m, 2H).

¹³C – NMR (125 MHz, CDCl₃): δ = 207.2, 143.5, 133.7, 129.7, 127.6, 55.8, 50.6, 49.1, 32.0, 30.5, 23.7, 21.5.

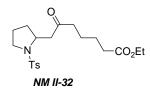
HRMS: Calculated for C₁₄H₁₉NO₃NaS: 304.0983. Found: 304.0985.



<u>6c</u>

IR (CHCl₃): 3423, 3024, 2936, 2859, 1709, 1604, 1492, 1457, 1334, 1158, 1087 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ = 7.69 (dt, *J* = 8.4, 2.0 Hz, 2H), 7.33-7.26 (m, 4H), 7.21-7.18 (m, 3H), 3.95-3.89 (m, 1H), 3.43-3.38 (m, 1H), 3.20 (dd, *J* = 17.4, 3.2 Hz, 1H), 3.09-3.03 (m, 1H), 2.92-2.85 (m, 2H), 2.85-2.68 (m, 2H), 2.43 (s, 3H), 1.80-1.65 (m, 2H), 1.51-1.40 (m, 2H). ¹³C - NMR (100 MHz, CDCl₃): δ = 208.4, 143.5, 140.8, 133.7, 129.7, 128.5, 128.3, 127.6, 126.1, 55.8, 49.9, 49.1, 44.7, 32.0, 29.6, 23.7, 21.5.

HRMS: Calculated for C₂₁H₂₅NO₃NaS: 394.1453. Found: 394.1453.

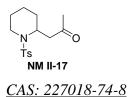


<u>6d</u>

IR (CHCl₃): 3407, 2933, 2873, 2358, 1735, 1712, 1663, 1599, 1456, 1380, 1344, 1178 cm⁻¹. ¹H-NMR (500 MHz, CDCl₃): δ = 7.70 (d, *J* = 8.0 Hz, 2H), 7.31 (d, *J* = 8.0 Hz, 2H), 4.10 (q, *J* = 7.0 Hz, 2H), 3.94-3.89 (m, 1H), 3.43 (ddd, *J* = 10.5, 5.5, 5.5 Hz, 1H), 3.20 (dd, *J* = 17.5, 3.0 Hz, 1H), 3.08 (ddd, *J* = 10.0, 7.0, 7.0 Hz, 1H), 2.63 (dd, *J* = 18.0, 9.5 Hz, 1H), 2.52-2.40 (m, 1H), 2.43 (s, 3H), 2.31 (t, *J* = 6.5 Hz, 2H), 1.83-1.69 (m, 2H), 1.62-1.61 (m, 5H), 1.54-1.44 (m, 2H), 1.25 (t, *J* = 7.5 Hz, 3H).

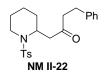
¹³C – NMR (100 MHz, CDCl₃): δ = 208.9, 173.4, 143.5, 133.7, 129.7, 127.6, 60.3, 55.9, 49.7, 49.1, 42.9, 34.0, 32.1, 24.4, 23.8, 23.0, 21.5, 14.2.

HRMS: Calculated for C₂₀H₂₉NO₅NaS: 418.1664. Found: 418.1658.



<u>6f</u>

¹H-NMR (400 MHz, CDCl₃): δ = 7.71 (dm, *J* = 8.3 Hz, 2H), 7.30-7.27 (m, 2H), 4.52 (dd, *J* = 8.8, 4.3 Hz, 1H), 3.78 (dm, *J* = 13.8 Hz, 1H), 2.94 (dt, *J* = 13.7, 2.4 Hz, 1H), 2.79 (dd, *J* = 16.3, 9.3 Hz, 1H), 2.60 (dd, *J* = 16.3, 4.6 Hz, 1H), 2.42 (s, 3H), 2.13 (s, 3H), 1.55-1.22 (m, 6H). ¹³C – NMR (100 MHz, CDCl₃): δ = 206.0, 143.1, 138.0, 129.7, 127.0, 48.6, 43.9, 41.2, 30.2, 27.6, 24.5, 21.5, 18.3.



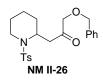
<u>6g</u>

IR (CHCl₃): 2934, 2853, 2362, 1715, 1597, 1498, 1454, 1342, 1149, 1093 cm⁻¹.

¹H-NMR (400 MHz, CDCl₃): δ = 7.69 (dt, *J* = 8.4, 1.8 Hz, 2H), 7.30-7.25 (m, 4H), 7.21-7.16 (m, 3H), 4.52-4.48 (m, 1H), 3.76 (dm, *J* = 13.6 Hz, 1H), 2.94-2.84 (m, 3H), 2.75-2.68 (m, 3H), 2.54 (dd, *J* = 8.0 Hz, 4.6 Hz, 2H), 2.41 (s, 3H), 1.53-1.23 (m, 6H).

¹³C – NMR (100 MHz, CDCl₃): δ = 207.3, 143.1, 140.8, 138.0, 129.7, 128.5, 128.3, 127.0, 126.1, 48.7, 44.6, 43.2, 41.2, 29.6, 27.7, 24.5, 21.5, 18.3.

HRMS: Calculated for C₂₂H₂₇NO₃NaS: 408.1609. Found: 408.1612.



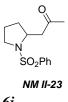
<u>6h</u>

IR (CHCl₃): 3422, 2934, 2863, 2126, 1722, 1652, 1338, 1165 cm⁻¹.

¹H-NMR (500 MHz, CDCl₃): δ = 7.68 (dt, *J* = 8.5, 2.0 Hz, 2H), 7.39-7.30 (m, 5H), 7.27-7.25 (m, 2H), 4.59-4.54 (m, 1H), 4.57 (d, *J* = 2.0 Hz, 2H), 4.01 (ddd, *J* = 16.5, 16.4 Hz, 2H), 3.81-3.78 (m, 1H), 2.95 (dt, *J* = 13.0, 2.5 Hz, 1H), 2.84 (dd, *J* = 16.0, 8.3 Hz, 1H), 2.65 (dd, *J* = 16.0, 5.0 Hz, 1H), 2.40 (s, 3H), 1.53-1.39 (m, 5H), 1.33-1.25 (m, 1H).

¹³C – NMR (125 MHz, CDCl₃): δ = 206.4, 143.1, 138.0, 137.0, 129.7, 128.5, 128.0, 127.9, 127.0, 75.1, 73.4, 48.6, 41.1, 39.4, 27.9, 24.5, 21.5, 18.3.

HRMS: Calculated for C₂₂H₂₇NO₄NaS: 424.1559. Found: 424.1559.



<u>6i</u>

IR (CHCl₃): 3407, 2976, 2885, 1715, 1454, 1411, 1345, 1163, 1102, 1047 cm⁻¹.

¹H-NMR (500 MHz, CDCl₃): δ = 7.83-7.81 (m, 2H), 7.61-7.58 (m, 1H), 7.54-7.51 (m, 2H), 3.95-3.90 (m, 1H), 3.46-3.42 (m, 1H), 3.23 (dd, *J* = 17.75, 3.25 Hz, 1H), 3.08 (ddd, *J* = 14.0, 7.5, 7.0 Hz, 1H), 2.67 (dd, *J* = 17.5, 9.5 Hz, 1H), 2.16 (s, 3H), 1.82-1.69 (m, 2H), 1.55-1.42 (m, 2H).

¹³C–NMR (125 MHz, CDCl₃): δ = 207.1, 136.6, 132.7, 129.1, 127.5, 55.8, 50.5, 49.1, 32.0, 30.5, 23.7.

HRMS: Calculated for C₁₃H₁₇NO₃NaS: 290.0827. Found: 290.0829.



<u>6j</u>

IR: 3579, 3415, 2918, 2121, 1712, 1649, 1549, 1379, 1172, 1115 cm⁻¹.

¹H-NMR (500 MHz, CDCl₃): δ = 7.98 (dd, *J* = 7.5, 2.5 Hz, 1H), 7.73-7.67 (m, 1H), 7.58 (dd, *J* = 7.5, 2.5 Hz, 2H), 4.27-4.22 (m, 1H), 3.50-3.46 (m, 1H), 3.43-3.38 (m, 1H), 3.18 (dd, *J* = 17.75, 3.25 Hz, 1H), 2.64 (dd, *J* = 17.75, 9.75 Hz, 1H), 2.16 (s, 3H), 2.10-2.02 (m, 1H), 1.91-1.83 (m, 1H), 1.80-1.73 (m, 1H), 1.70-1.64 (m, 1H).

¹³C – NMR (125 MHz, CDCl₃): δ = 206.7, 148.6, 133.7, 131.4, 131.1, 130.7, 123.9, 56.0, 49.6, 49.1, 32.2, 30.5, 23.8.

HRMS: Calculated for C₁₃H₁₆N₂O₅NaS: 335.0678. Found: 335.0677.



CAS: 86953-89-1

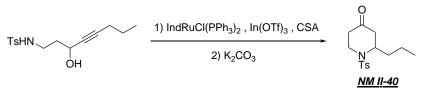
<u>6k</u>

Data is in agreement with the published reference (*J.Org.Chem.* **1983**, *48*, 4058). IR (CHCl₃): 3582, 3452, 2964, 2873, 2347, 1690, 1651, 1450, 1391, 1170 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): $\delta = 4.13$ (br s, 1H), 3.30-3.29 (br m, 2H), 3.01 (dd, *J* = 16.0 Hz, 1H), 2.43-2.35 (m, 1H), 2.14 (s, 3H), 2.05 (dddd, *J* = 12.4, 8.0, 8.0, 7.9 Hz, 1H), 1.80 (quint, *J* = 8.0 Jz, 2H), 1.65-1.60 (m, 1H), 1.44 (s, 9H).



NMR data is given for the major diastereomer. IR(film): 2976, 1723, 1689, 1391, 1367, 1169, 1120, 701. ¹H NMR (300 MHz, CDCl₃): $\delta = 9.83$ (t, J = 2.0 Hz, 1H), 7.35-7.09 (m, 5H), 4.86 (d, J = 8.0Hz), 4.66-4.60 (m, IH), 3.12 (ddd, J = 16.2, 4.6, 1.9 Hz, 1H), 2.54 (ddd, J = 16.3, 8.2, 2.0 Hz), 2.39-2.16 (m, 2H), 1.81-1.75 (m, 1H), 1.65-1.58 (m, 1H), 1.13 (s, 9H). ¹³C NMR (75 MHz, CDCl₃) $\delta = 200.7$, 154.0, 144.7, 128.2 (2), 126.6, 125.1 (2), 79.7, 61.6, 53.1, 48.4, 32.6, 28.4, 28.0 (3). HRMS: Calculated for C₁₆H₂₃NO₂ (M⁺ - CO): 261.1729. Found 261.1726.

Piperidinones



9a (NM II-40)

The aminoalcohol (50 mg, 0.17 mmol, 1eq) is subjected to the standard procedure for redox isomerisation. After 3h, near complete conversion to the amino-enone is observed.

Aminoenone: ¹H-NMR (500 MHz, CDCl₃): δ = 7.73 (d, *J* = 8.5, 2H), 7.30 (d, *J* = 8.0 Hz, 2H), 6.80 (ddd, *J* = 14.0, 7.0 Hz, 1H), 6.02 (d, *J* = 16.0, 1.3 Hz, 1H), 5.19 (t, *J* = 6.5 Hz, 1H), 3.18 (q, *J* = 6.0 Hz, 2H), 2.78 (t, *J* = 5.8 Hz, 2H), 2.42 (s, 3H), 2.18 (dq, *J* = 7.0, 1.5 Hz, 2H), 1.47 (sext, *J* = 7.25 Hz, 2H), 0.93 (t, *J* = 7.25 Hz, 3H).

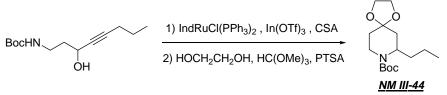
At this stage, the mixture is cooled down to room temperature. Then, 1 mL MeOH is added to the reaction mixture, followed by K_2CO_3 (24 mg, 0.17 mmol, 1 eq) and stirring is continued at room temperature for 12 hours. After filtration over a pad of Florisil, 5 mL of saturated aqueous NH₄Cl and 10 mL of ether are added. Following conventional extractive workup, purification by chromatography (silica, PE/AcEt 4:1 to 1:1) affords 26 mg of the piperidinone (52% yield).



IR (CHCl₃): 3414, 2955, 2362, 2338, 1715, 1601, 1353, 1165, 1093 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ = 7.77 (dt, *J* = 8.4, 2.0 Hz, 2H), 7.31 (dm, *J* = 8.0 Hz, 2H), 4.43-4.37 (m, 1H), 4.13 (tq, *J* = 7.6, 2.0 Hz, 1H), 3.30-3.23 (m, 1H), 2.50 (dd, *J* = 13.6, 6.6 Hz,

1H), 2.44 (s, 3H), 2.41-2.34 (m, 1H), 2.21 (dm, J = 8.0 Hz, 1H), 1.42-1.22 (m, 6H), 0.84 (t, J =7.6 Hz, 3H). ¹³C–NMR (100 MHz, CDCl₃): $\delta = 206.7$, 143.8, 137.5, 129.9, 127.0, 54.3, 45.3, 40.3, 40.0, 34.3, 21.5, 19.0, 13.5.

HRMS: Calculated for C₁₅H₂₁NO₃NaS: 318.1140. Found: 318.1141.



9b (NM III-44)

The aminoalcohol (45 mg, 0.186 mmol, 1eq) is subjected to the standard procedure for redox isomerisation. After 2h, complete conversion to the amino-enone is observed.

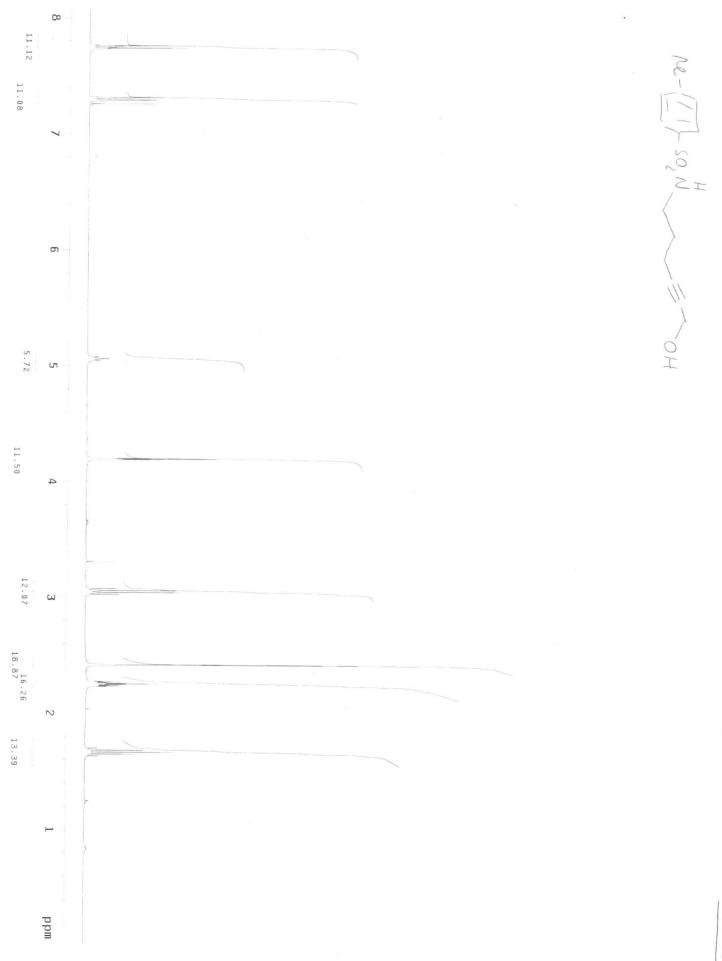
At this stage, the mixture is cooled down to room temperature. Then, PTSA (35 mg, 0.186 mmol, 1 eq), followed by ethylene glycol (20 µL, 0.37 mmol, 2 eq) and trimethyl orthoformate (40 µl, 0.37 mmol, 2 eq) stirring is continued at room temperature for 2 hours. After filtration over a pad of Florisil, 5 mL of saturated aqueous NaHCO₃ and 10 mL of ether are added. Following conventional extractive workup, purification by chromatography (silica, PE/AcEt 4:1 to 1:1) affords 32 mg of the protected piperidinone (62% yield).



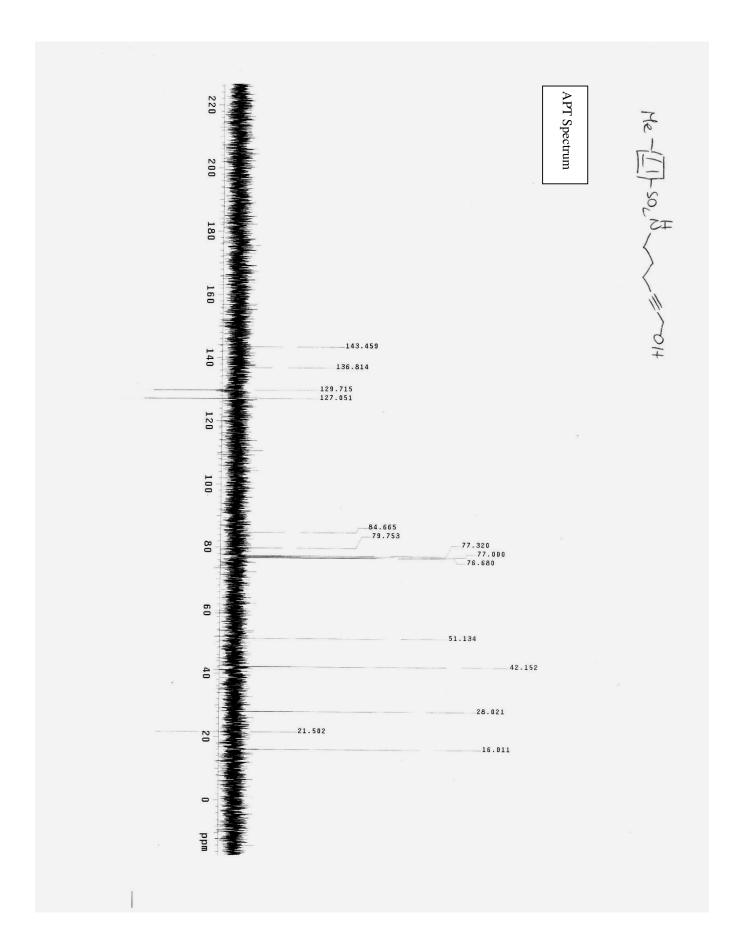
IR (CHCl₃): 34073416, 2960, 2922, 2871, 1687, 1414, 1370, 1250, 1174, 1116 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): $\delta = 4.34-4.29$ (m, 1H), 4.09-3.97 (m, 5H), 3.02-2.95 (m, 1H), 1.86-1.76 (m, 2H), 1.67-1.58 (m, 4H), 1.45 (s, 9H), 1.31-1.20 (m, 2H), 0.91 (t, *J* = 7.2 Hz, 3H). ¹³C–NMR (100 MHz, CDCl₃): $\delta = 154.8$, 107.4, 79.4, 64.6, 63.7, 50.4, 36.9, 34.7, 33.3, 19.8, 14.0.

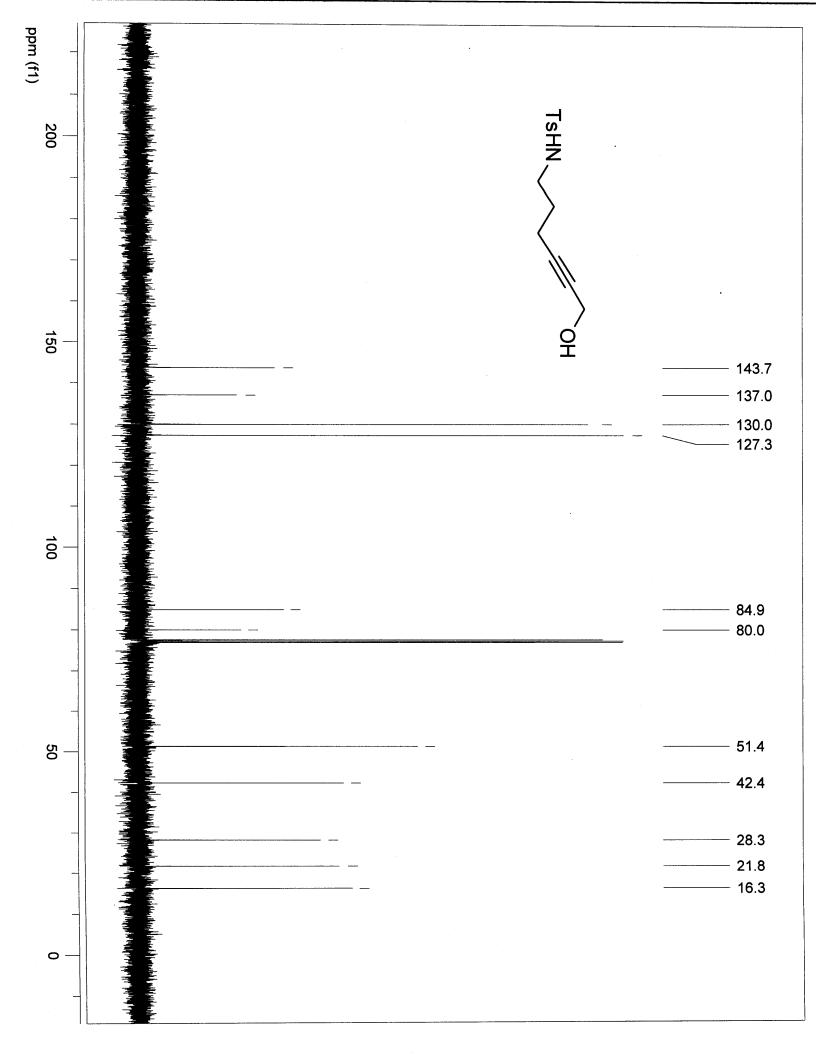
HRMS: Calculated for C₁₅H₂₇NO₄Na: 308.1838. Found: 308.1838.

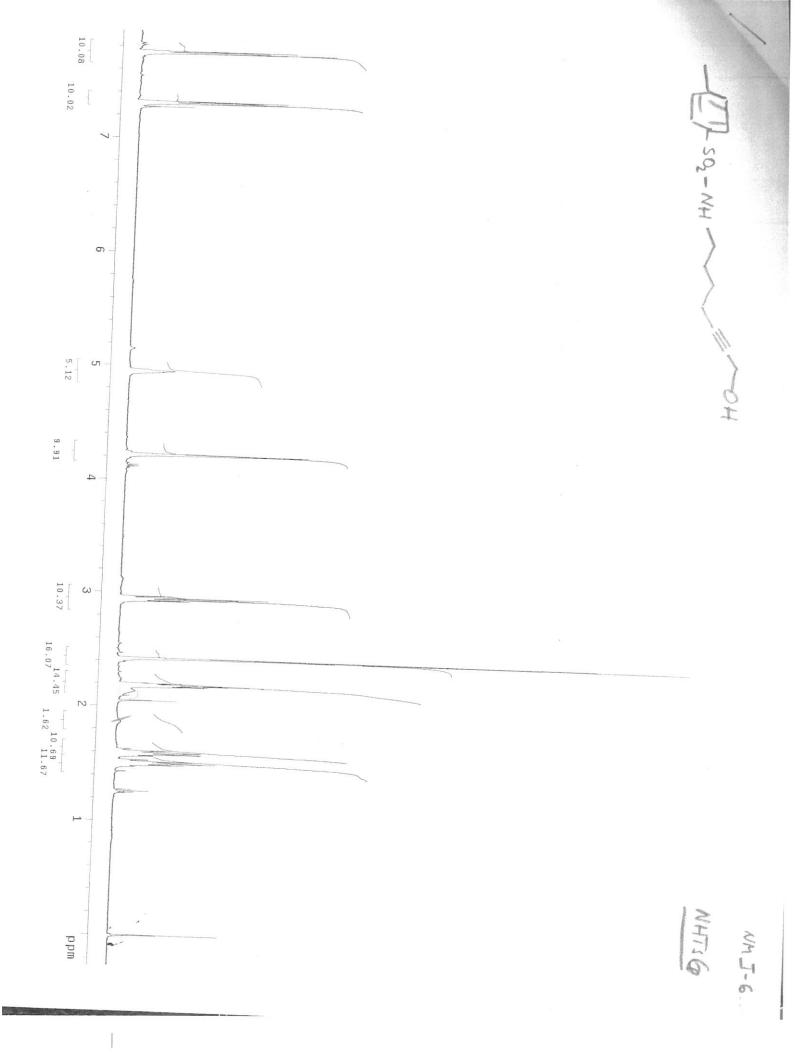
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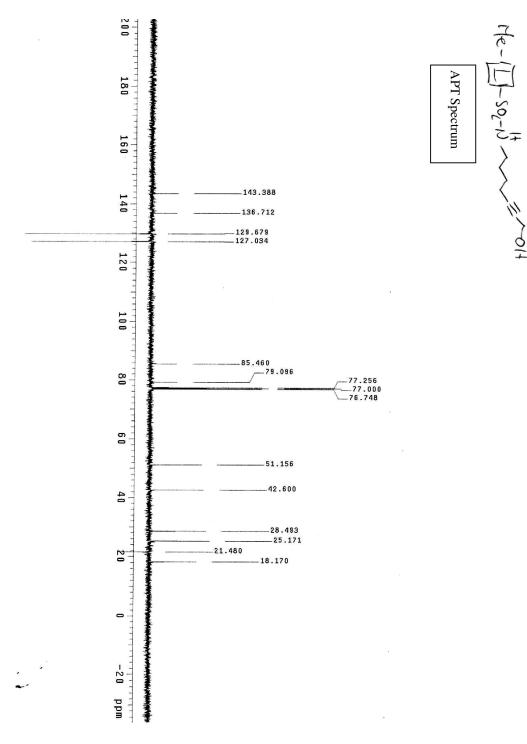


NMI I- 48

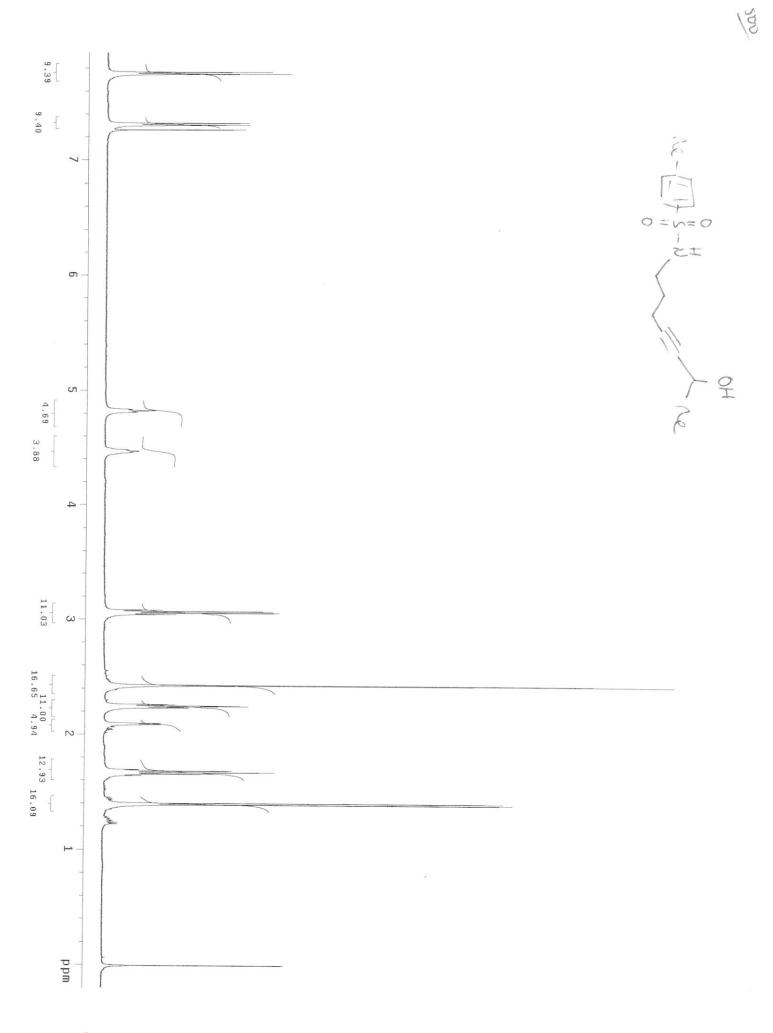


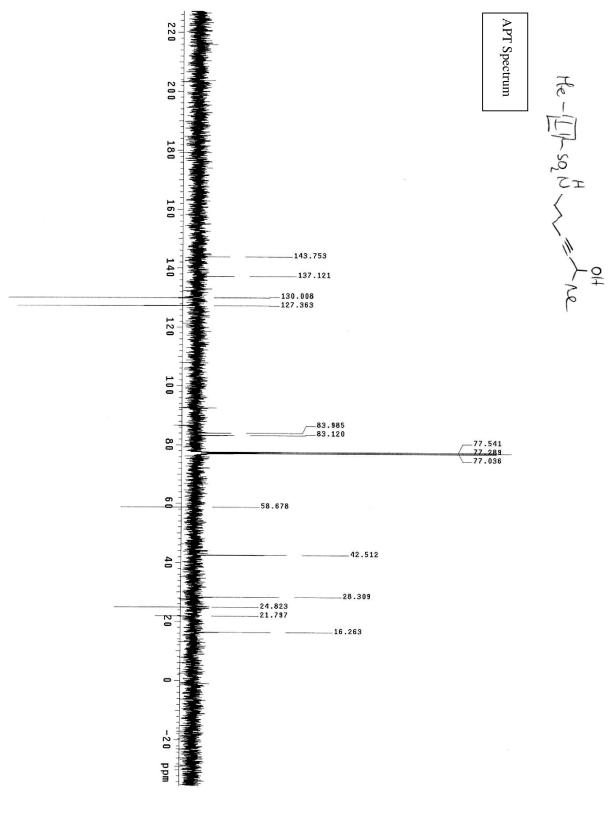


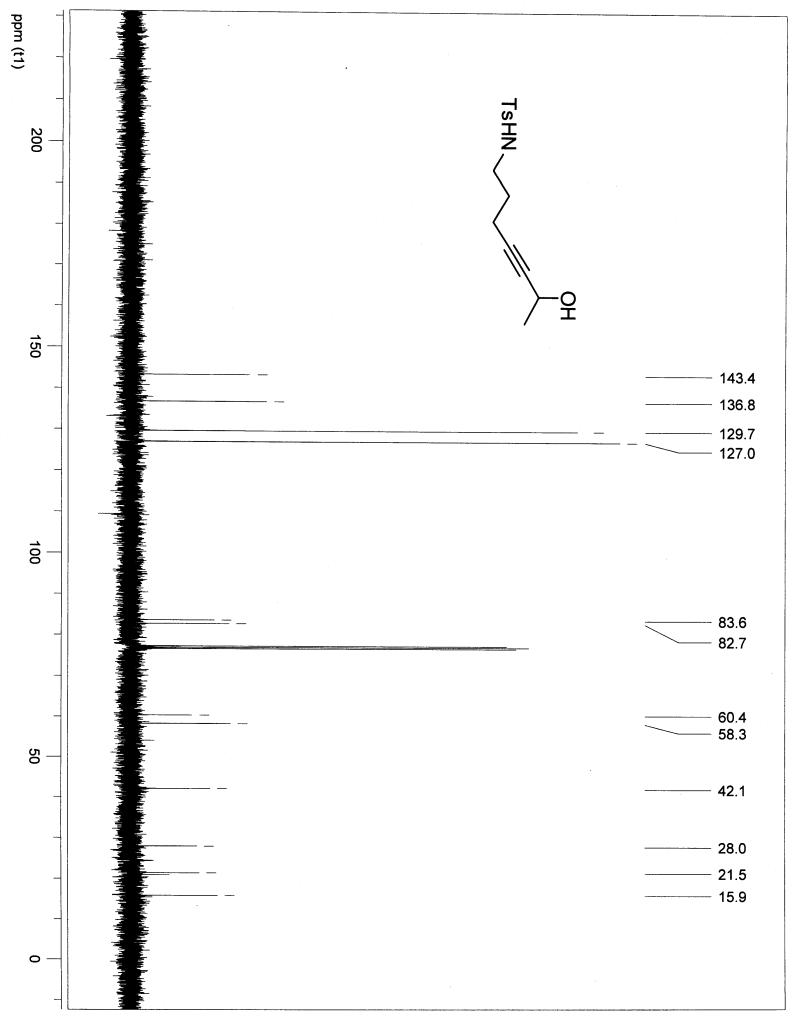


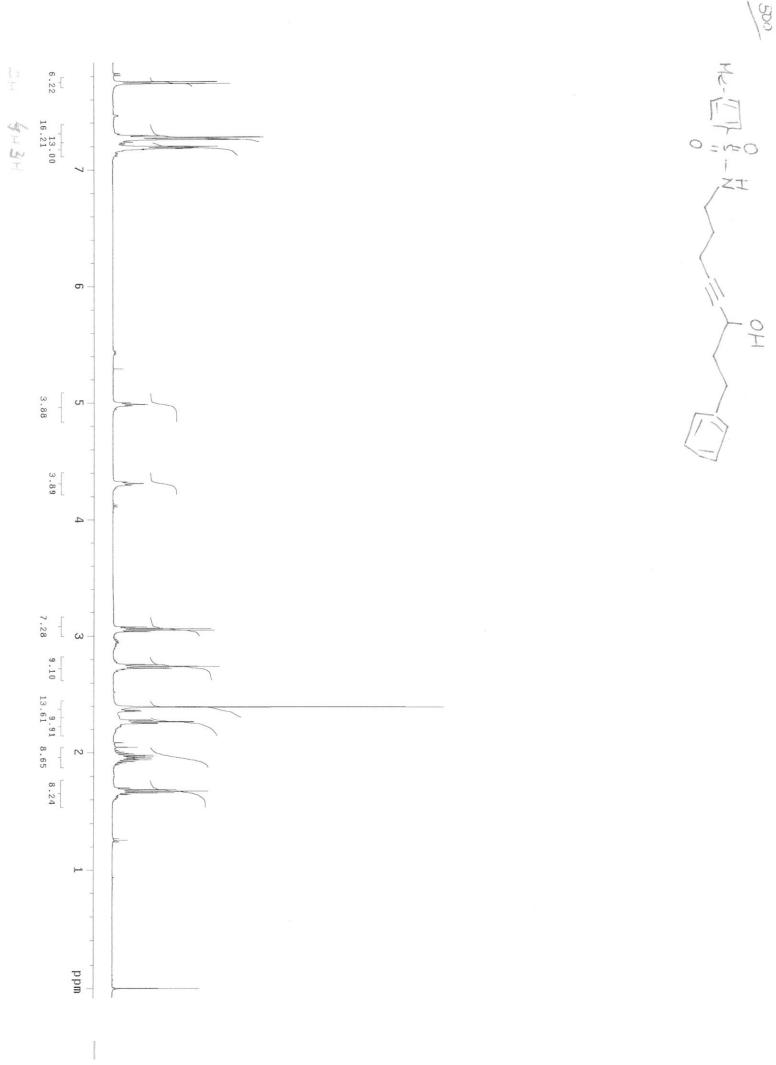


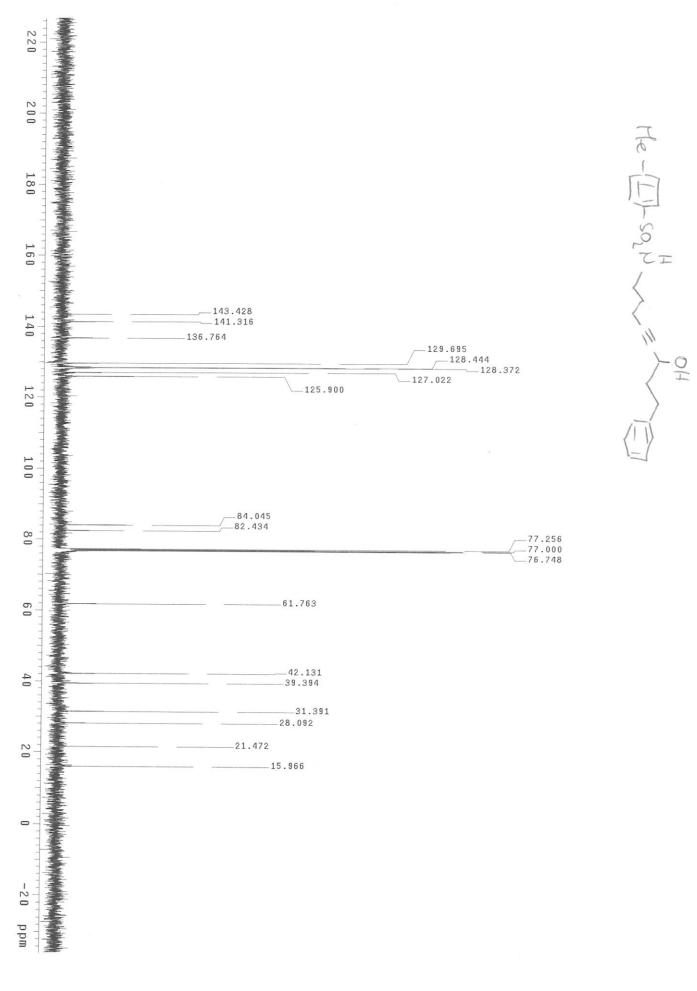
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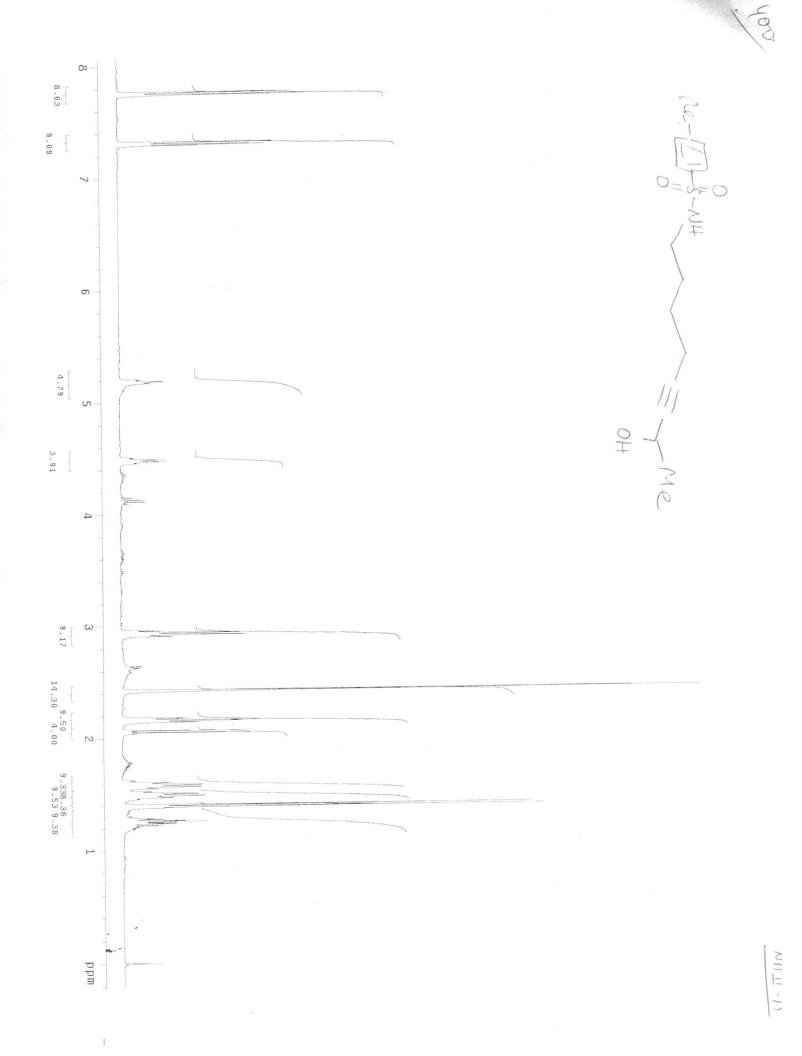


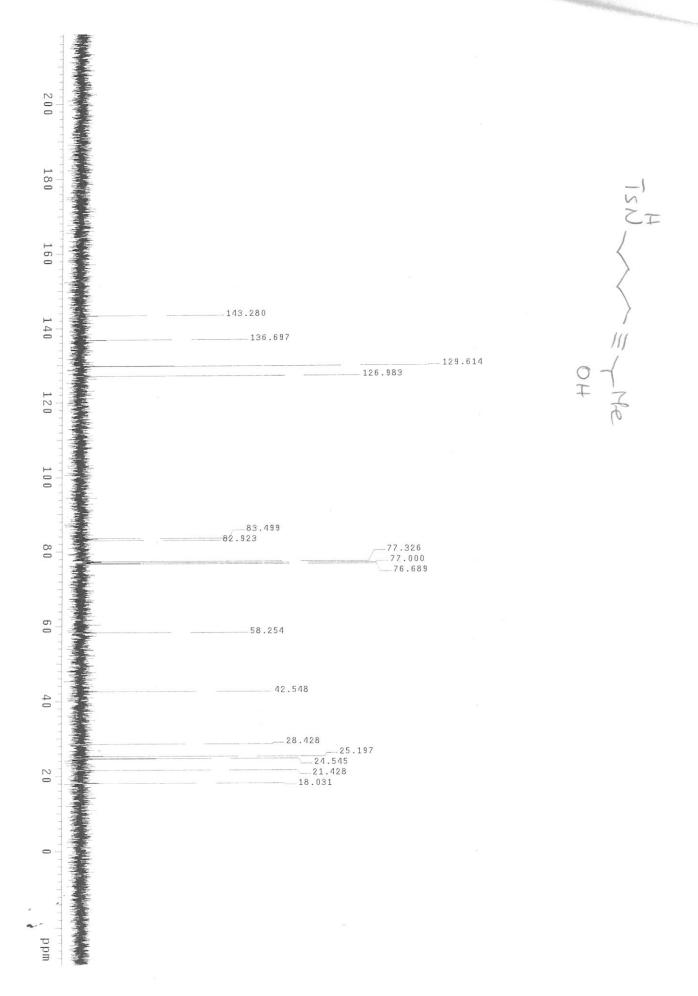


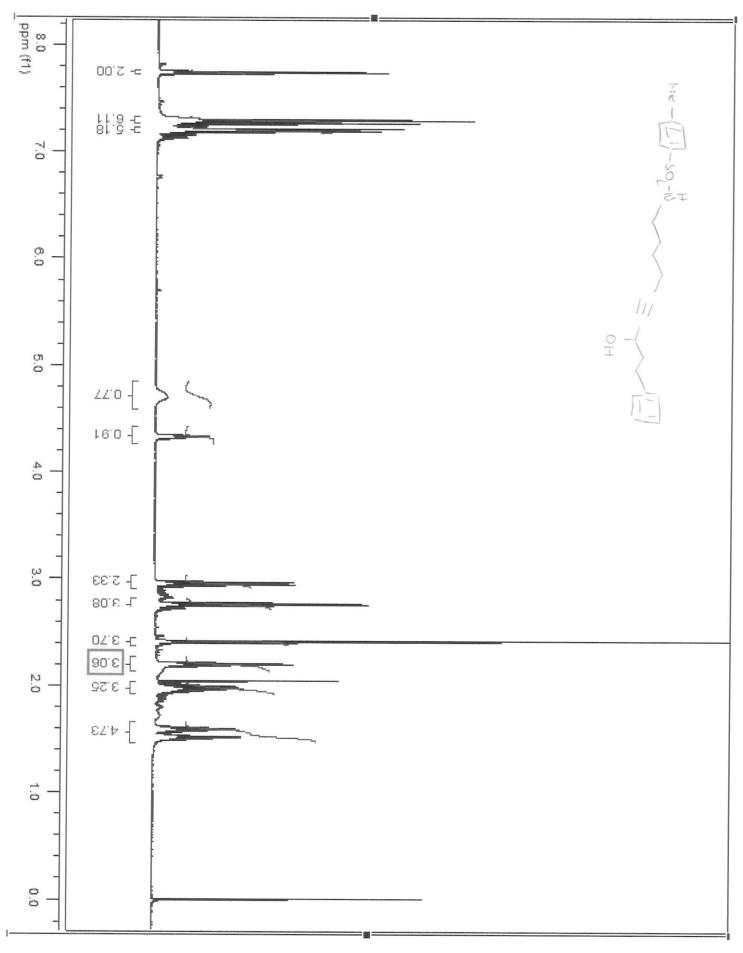


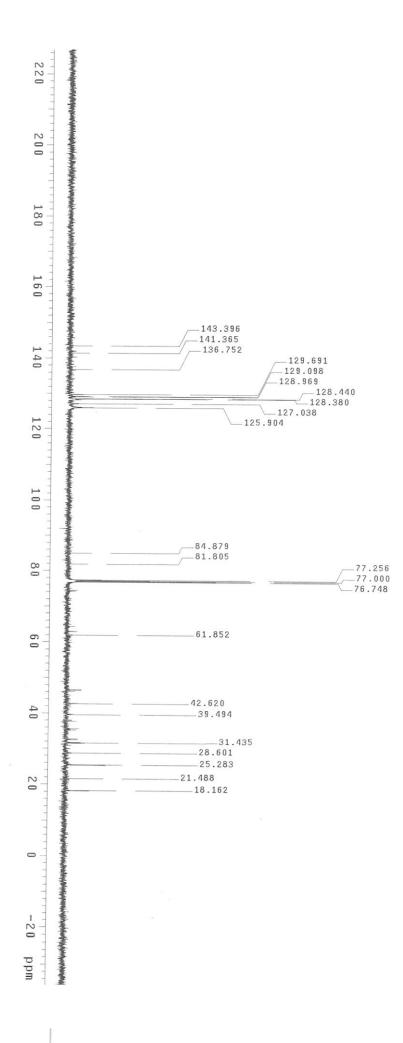




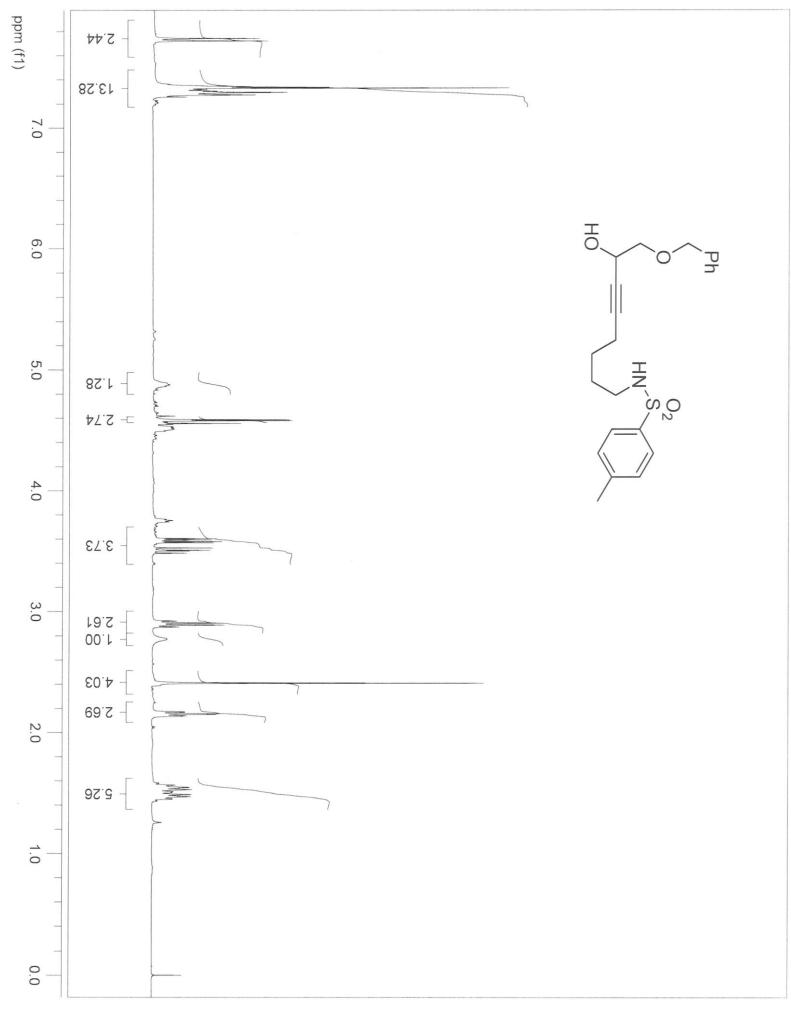


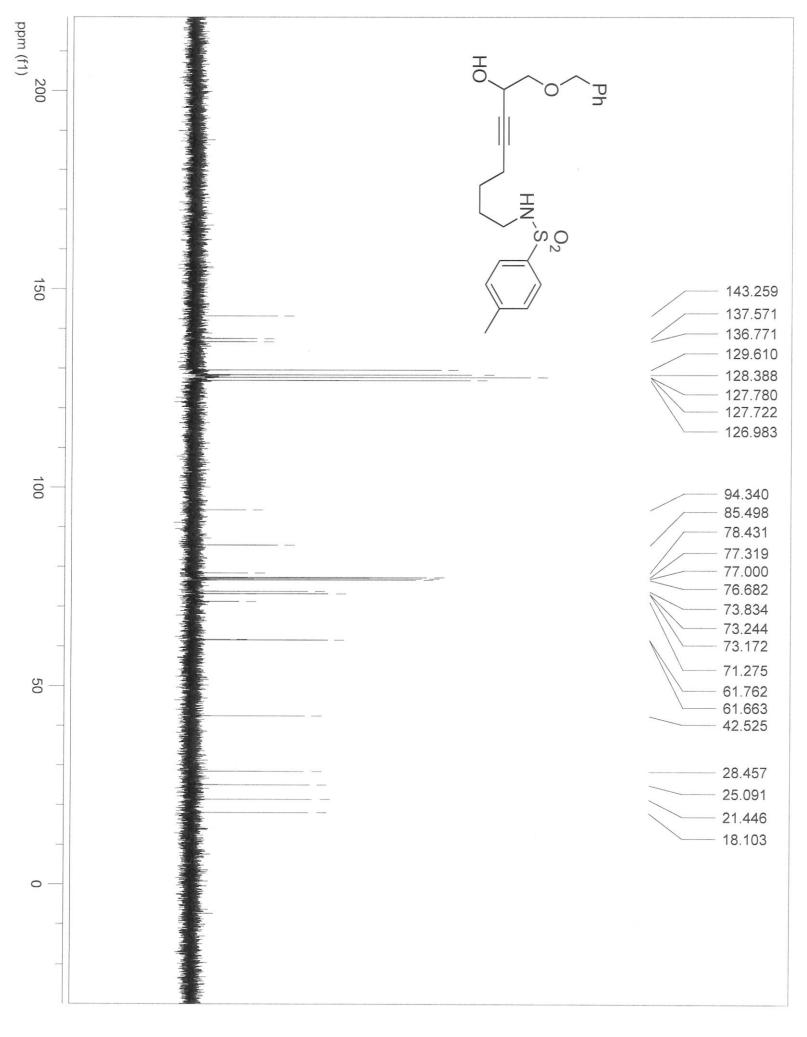




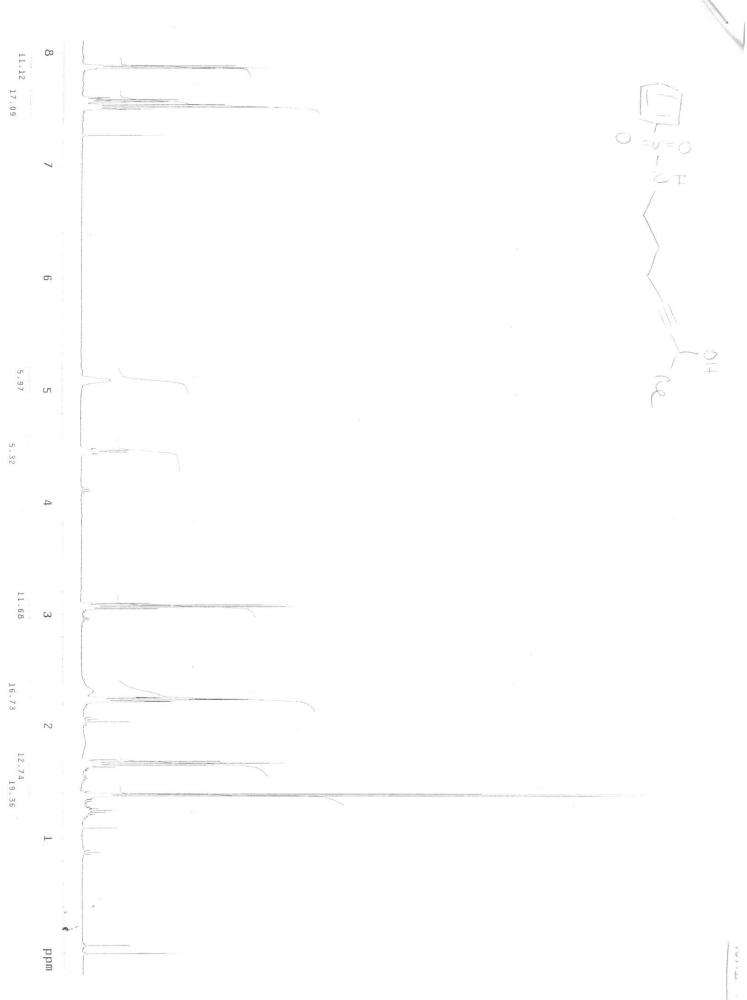


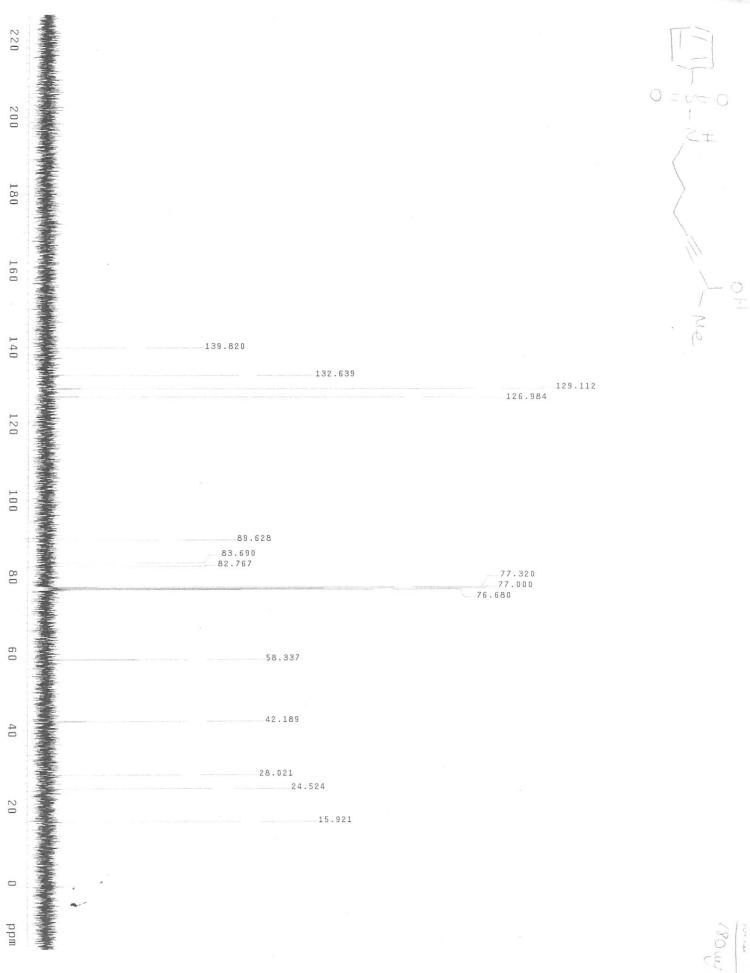




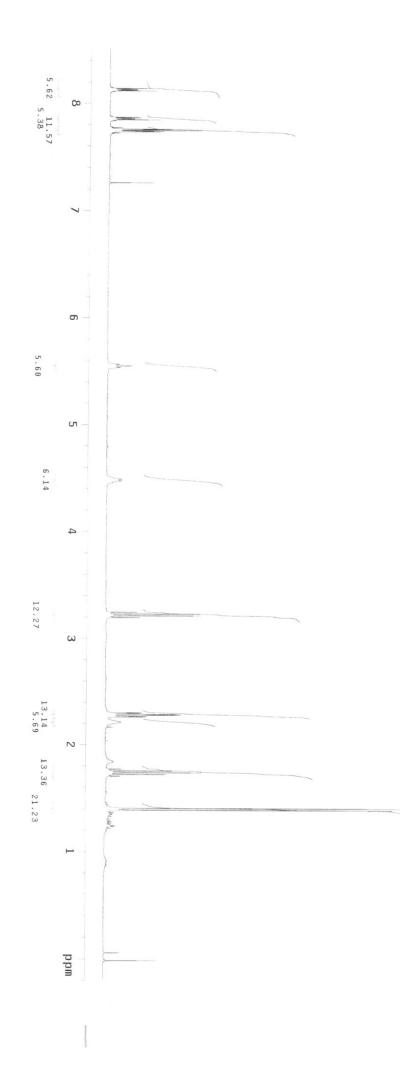








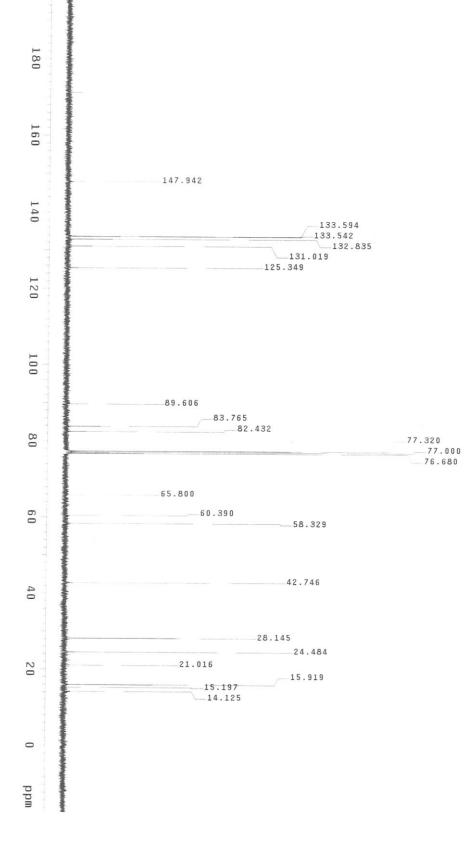
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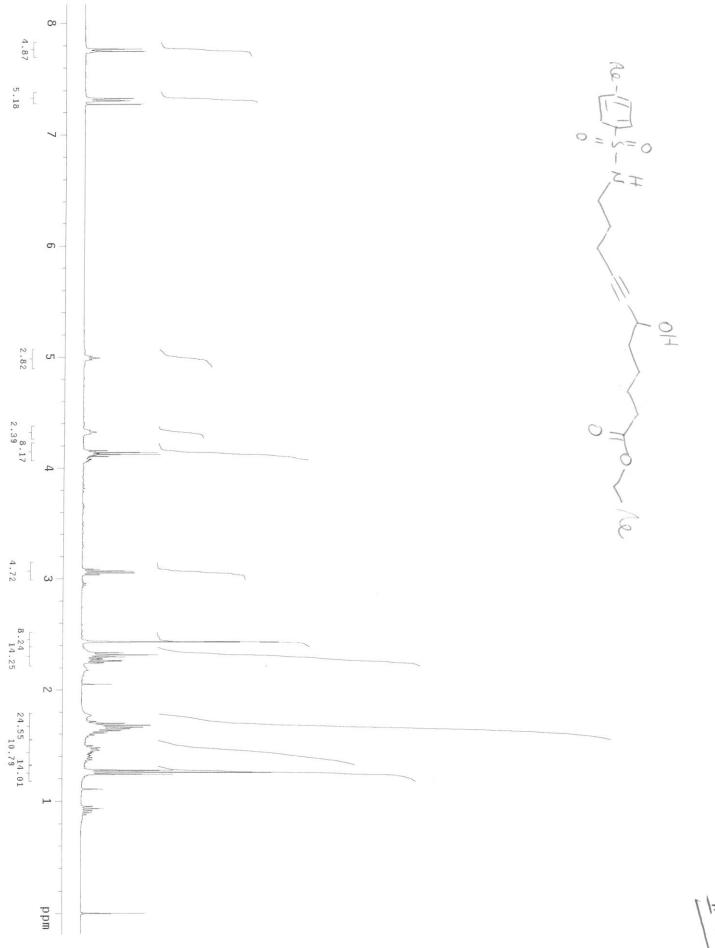


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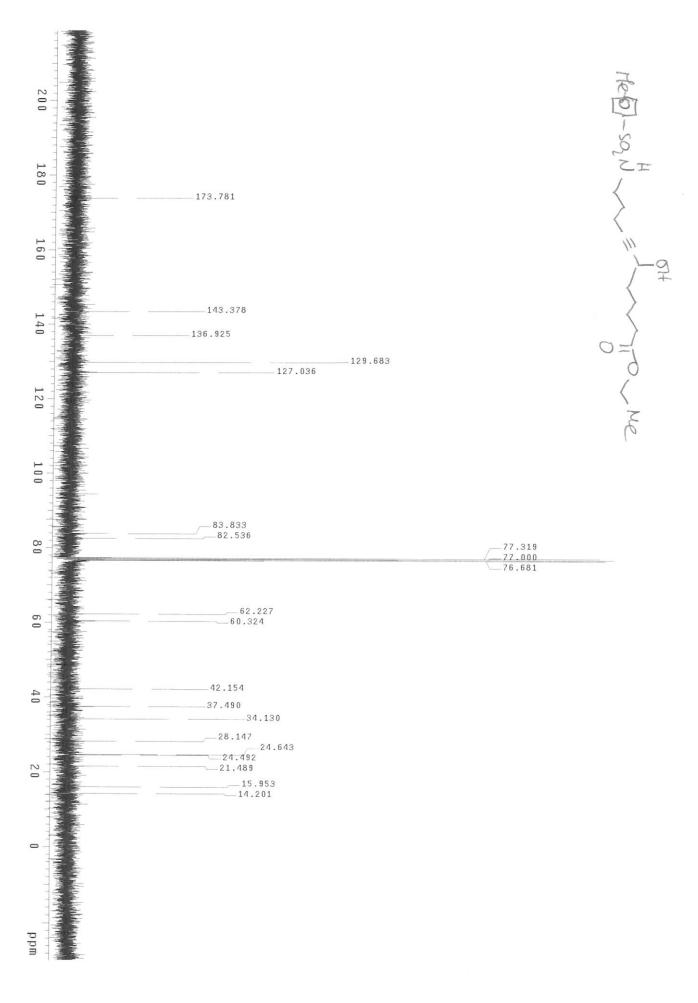
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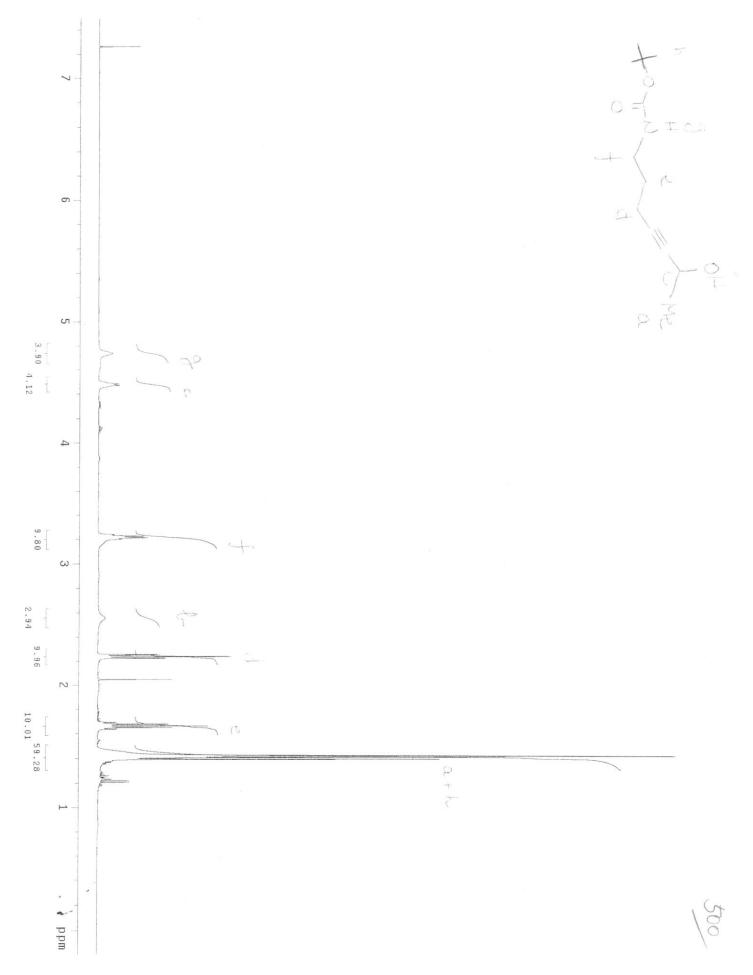


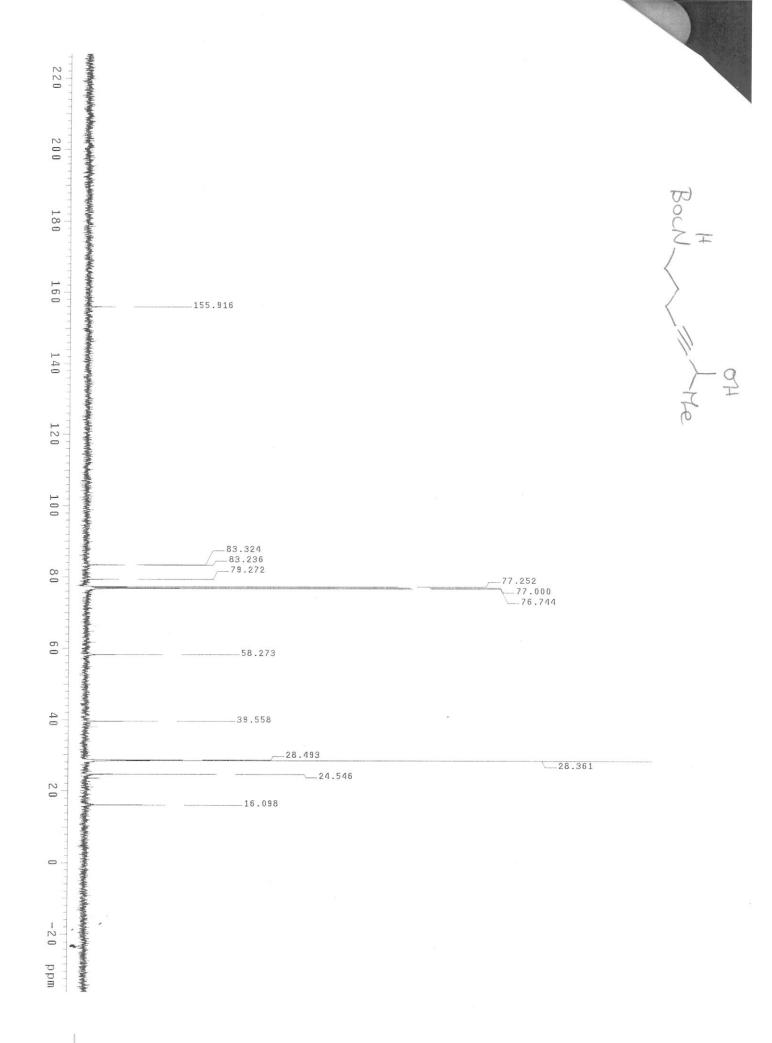


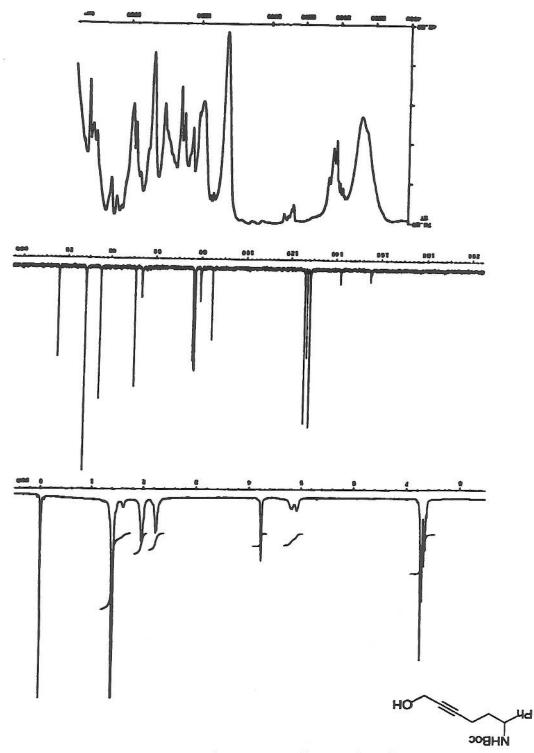


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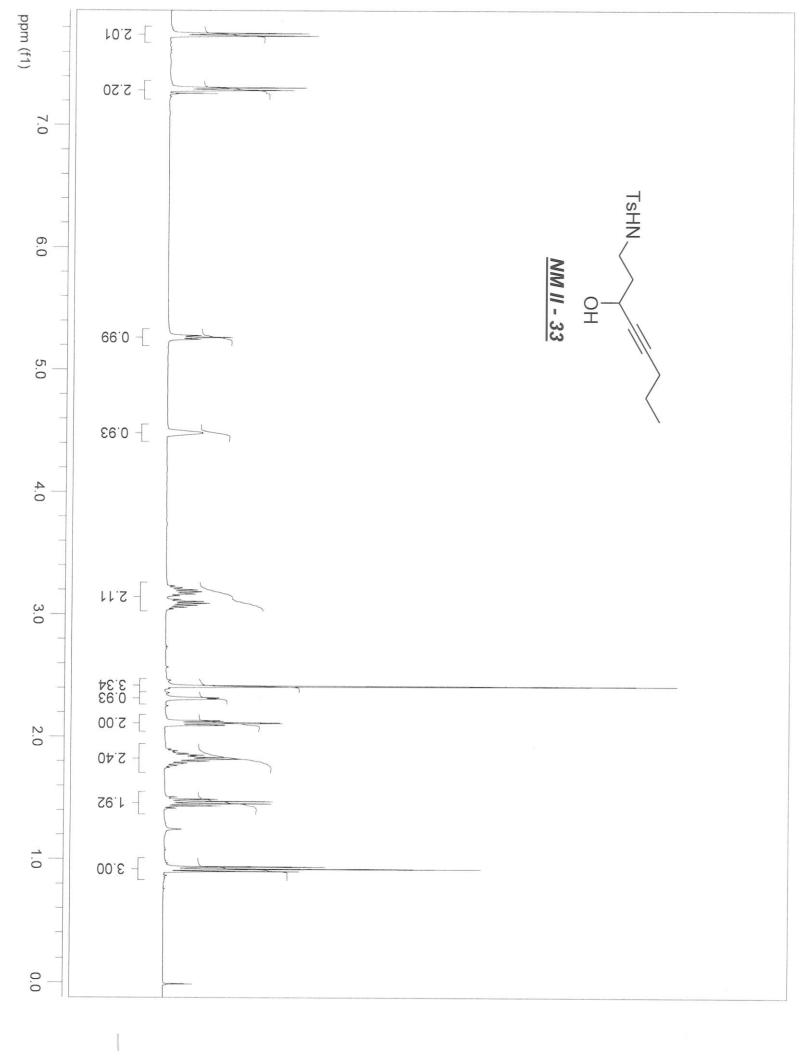


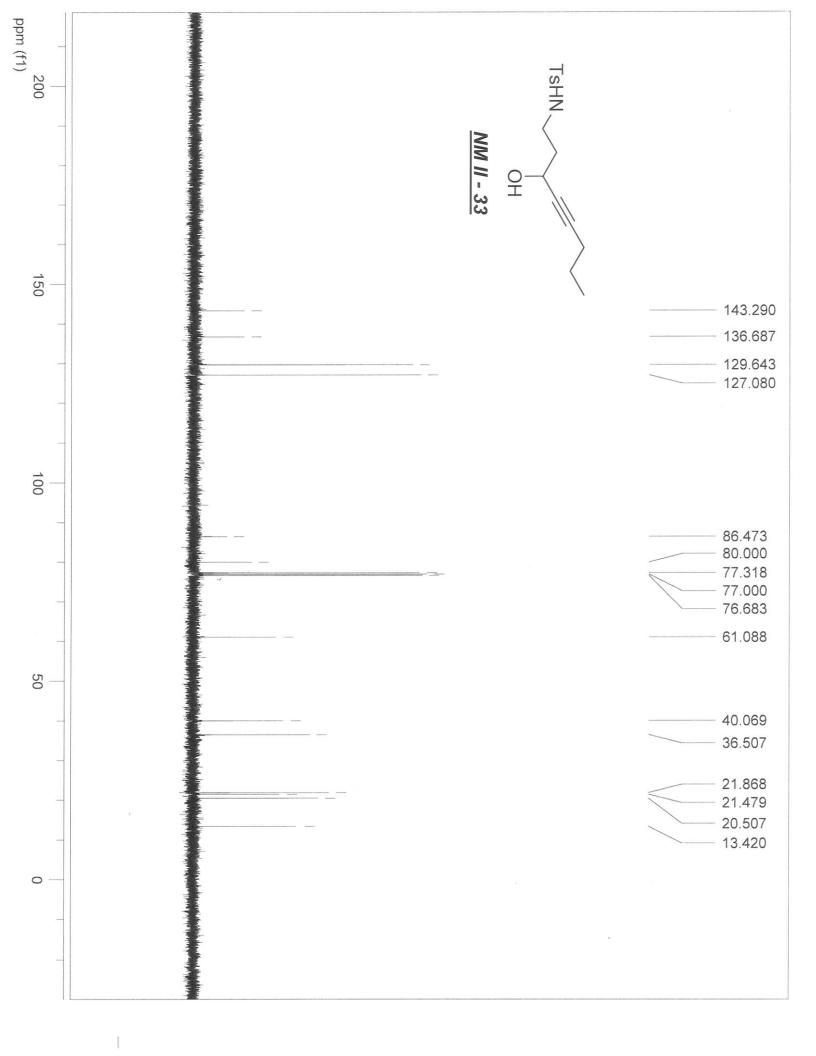


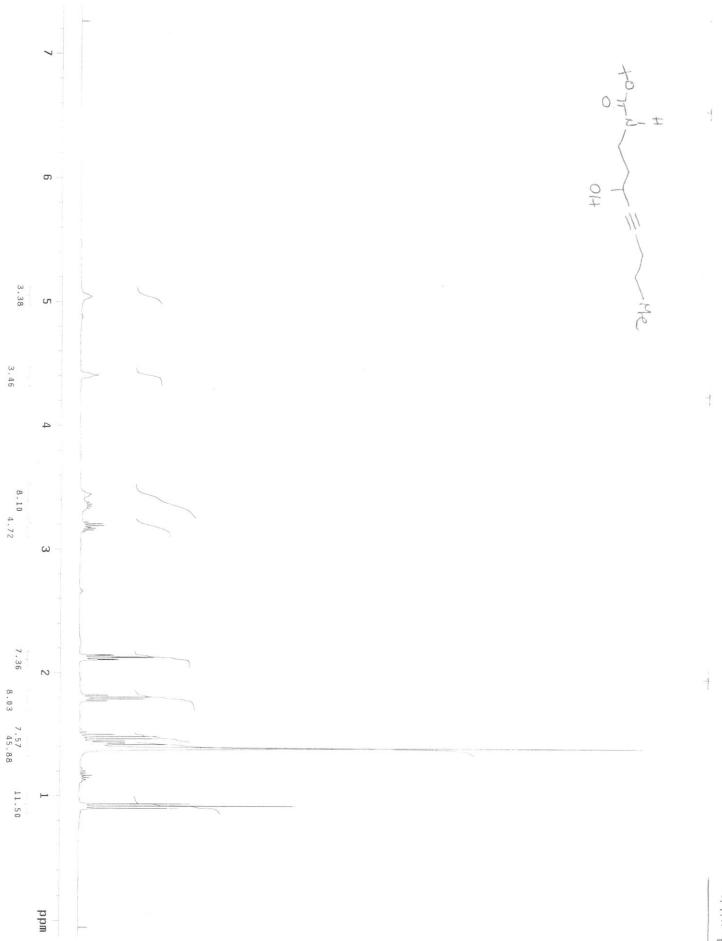




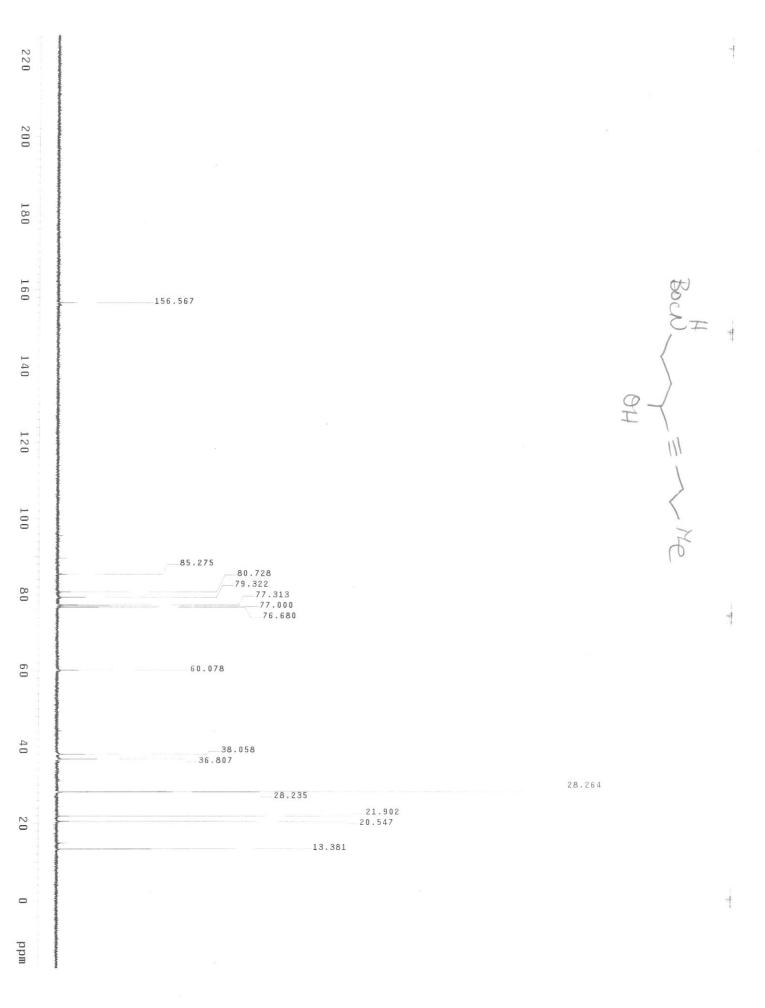


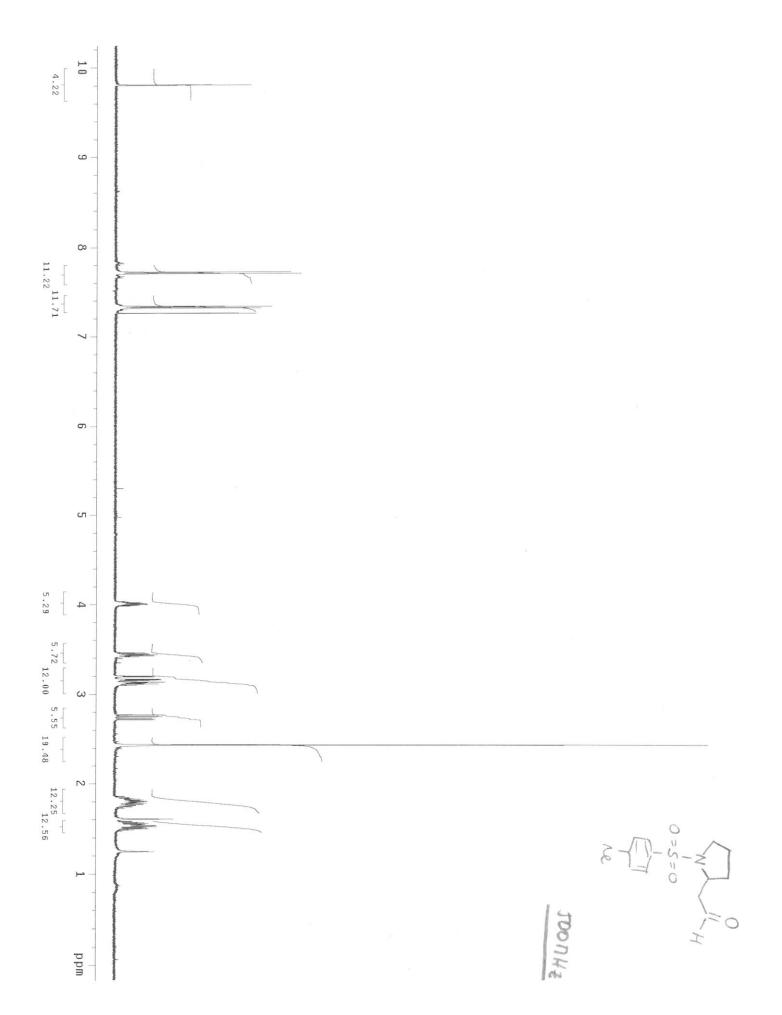


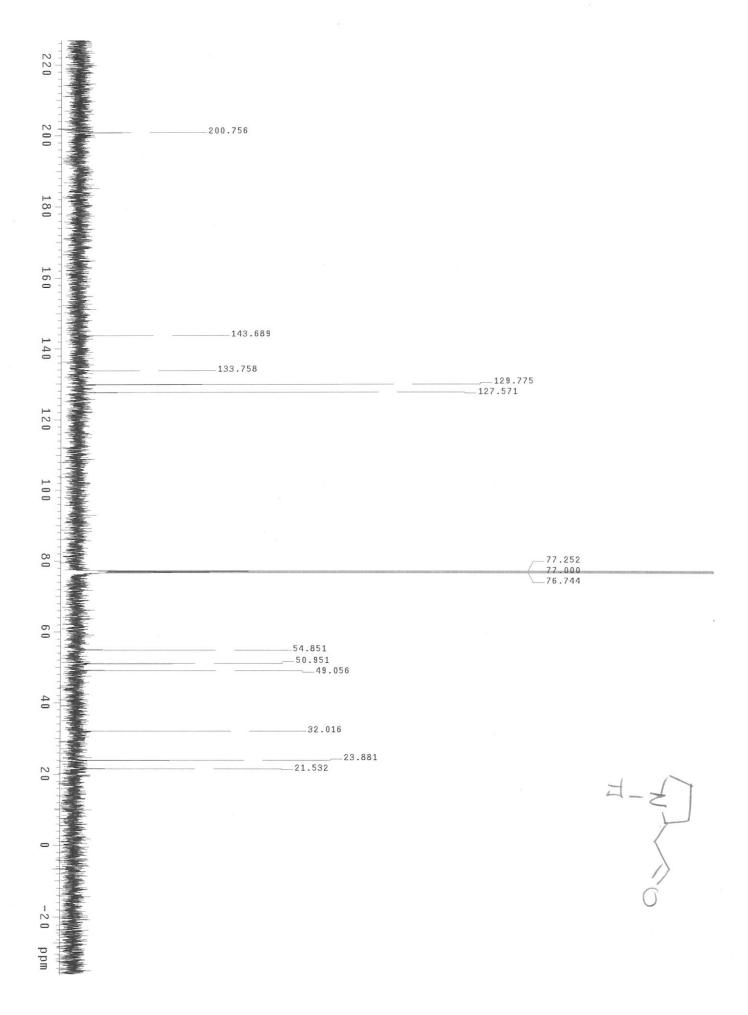


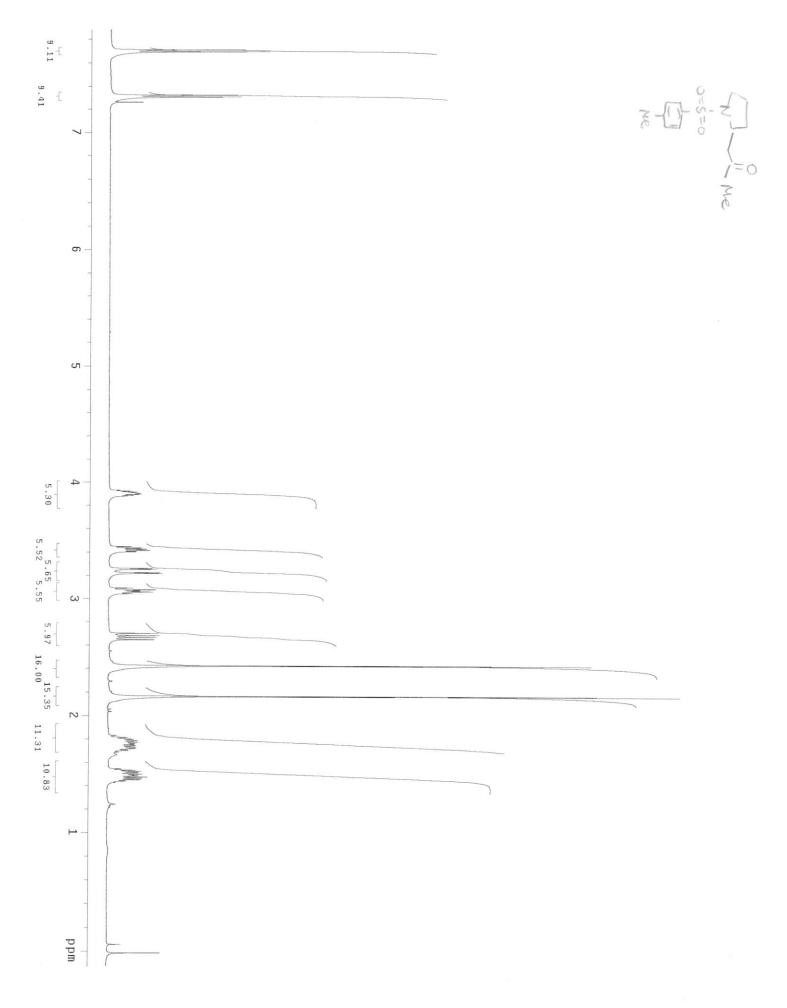


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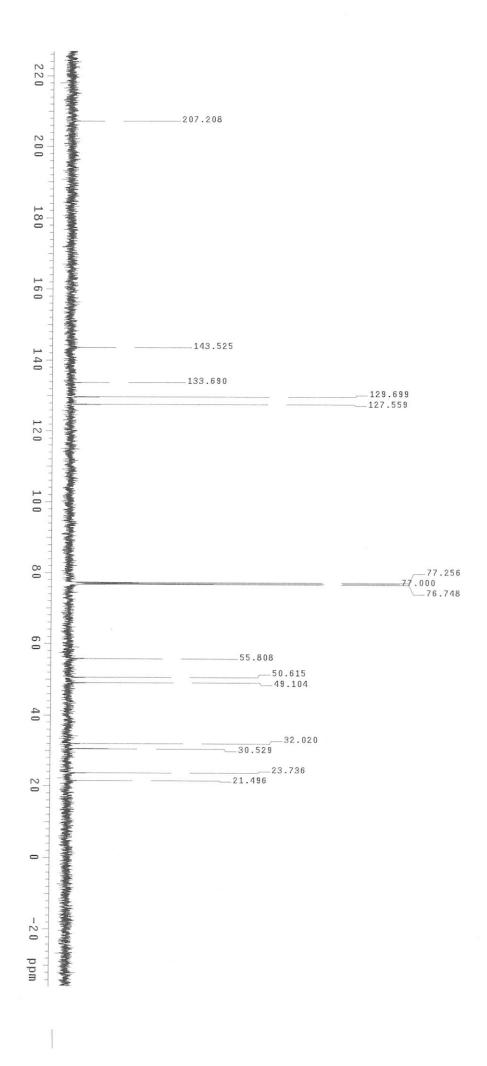


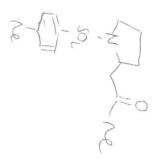




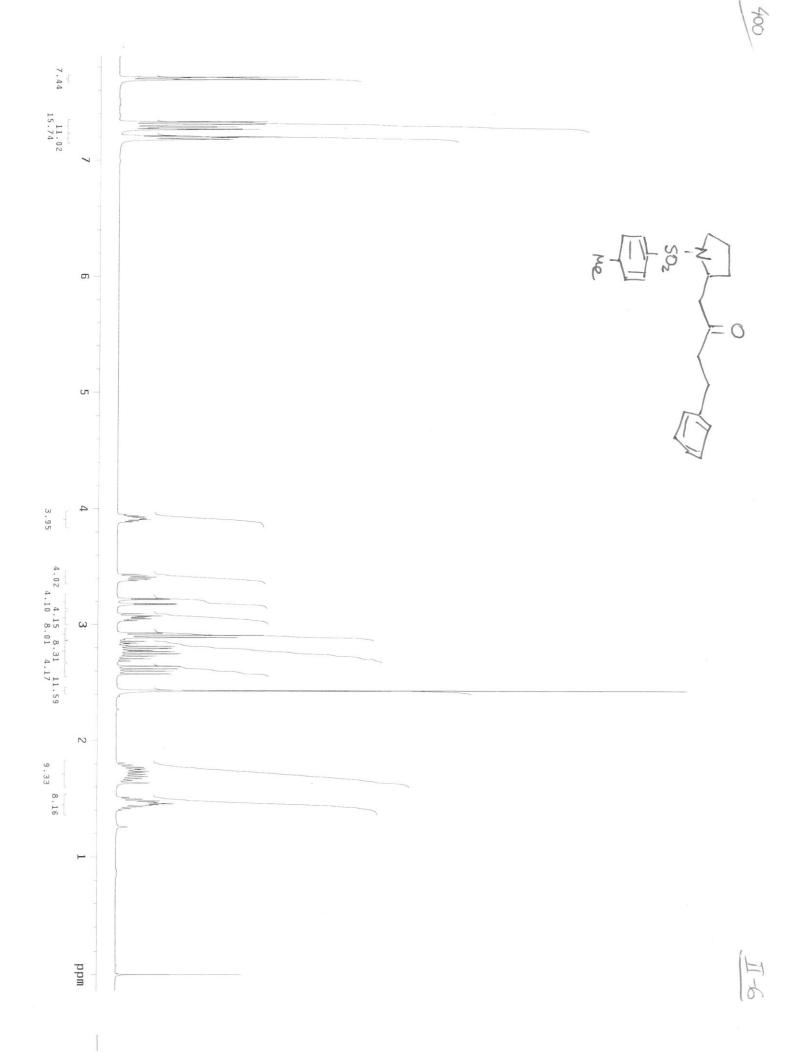


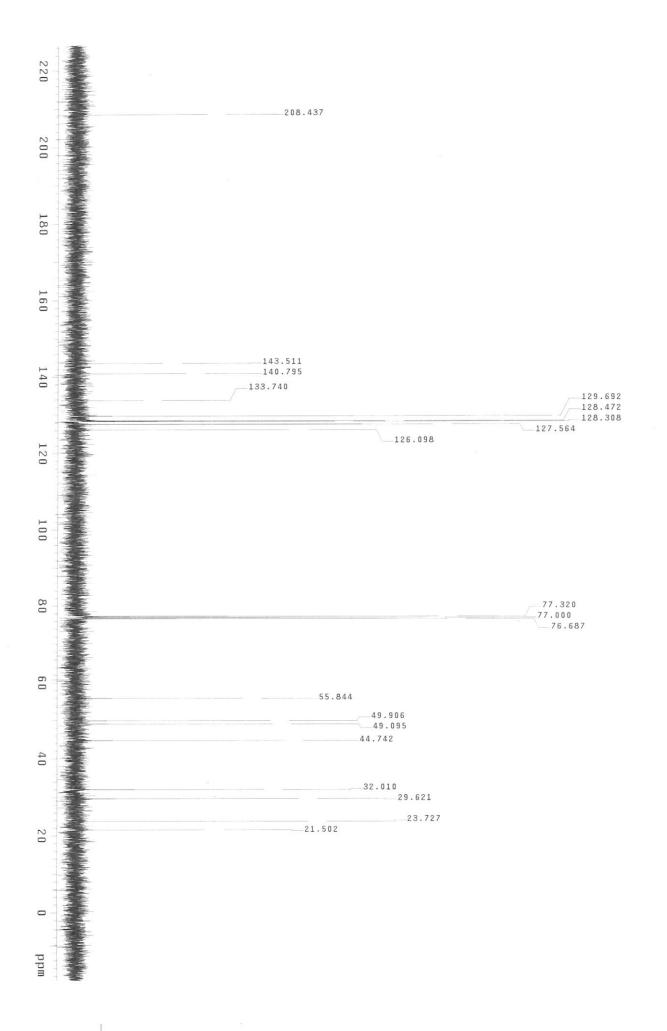
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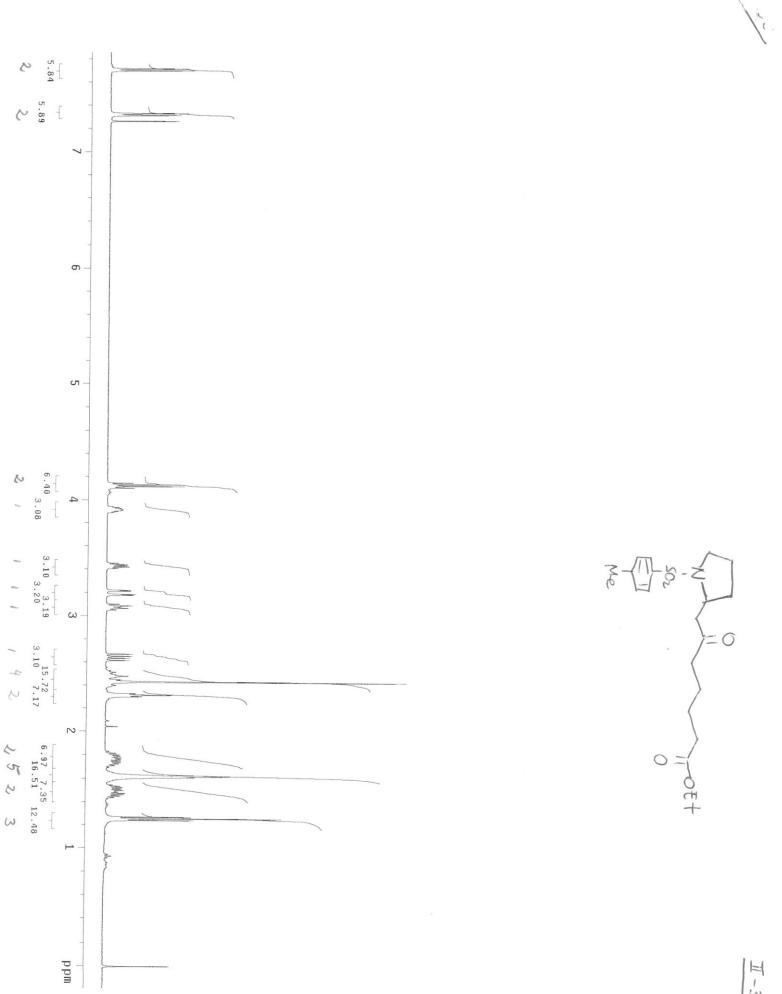


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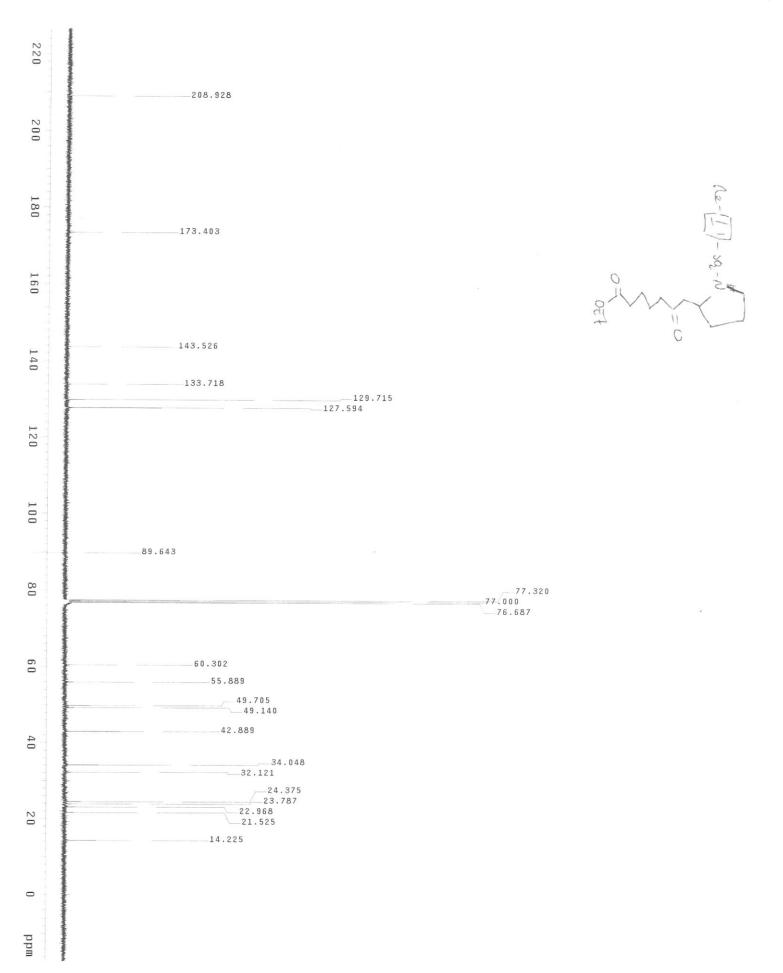


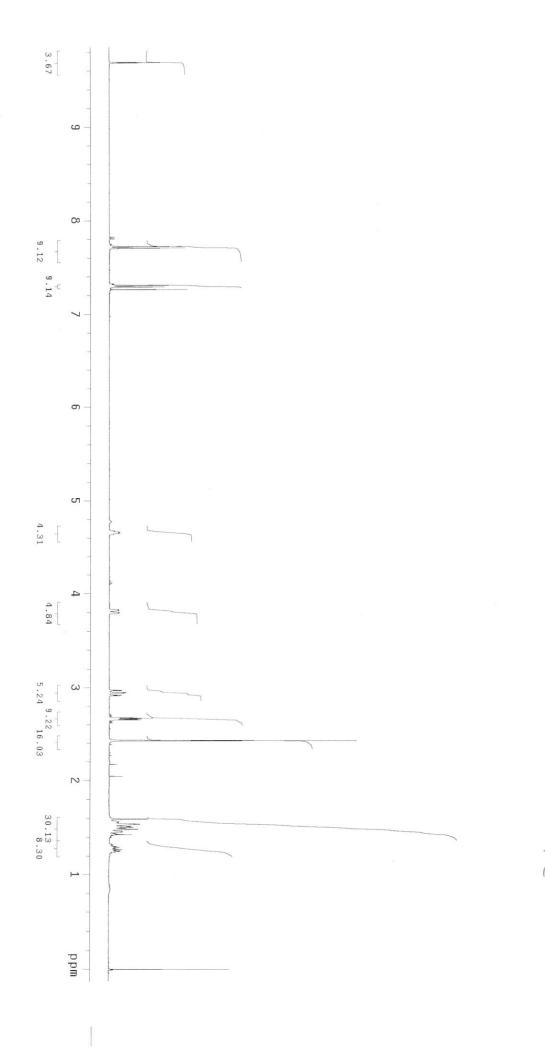


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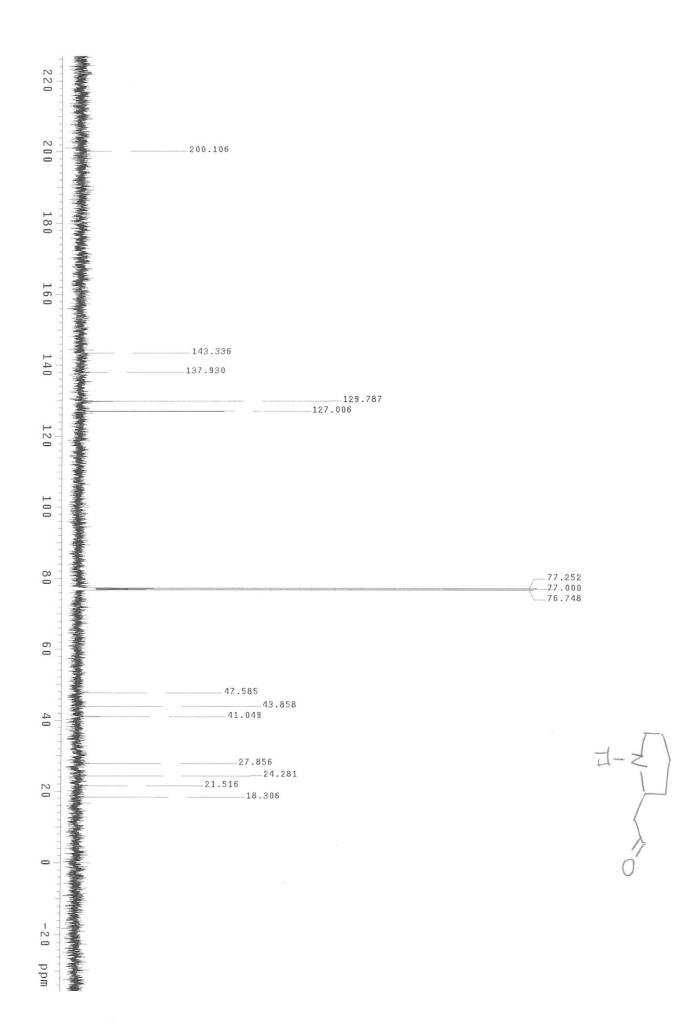
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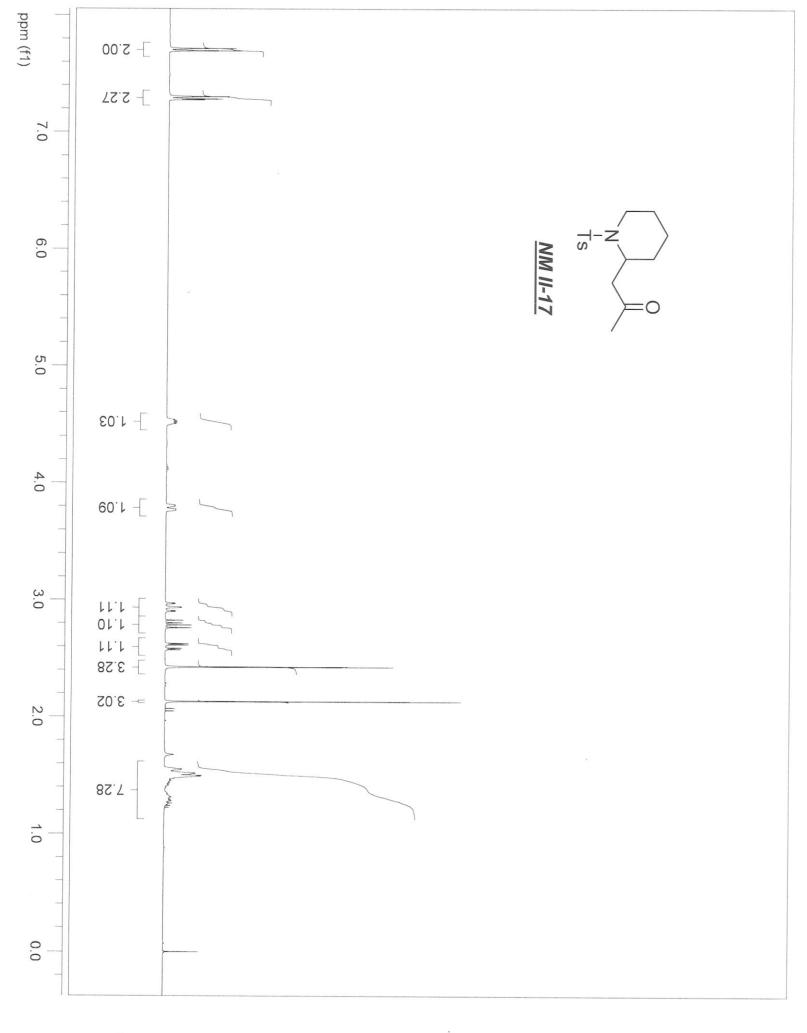


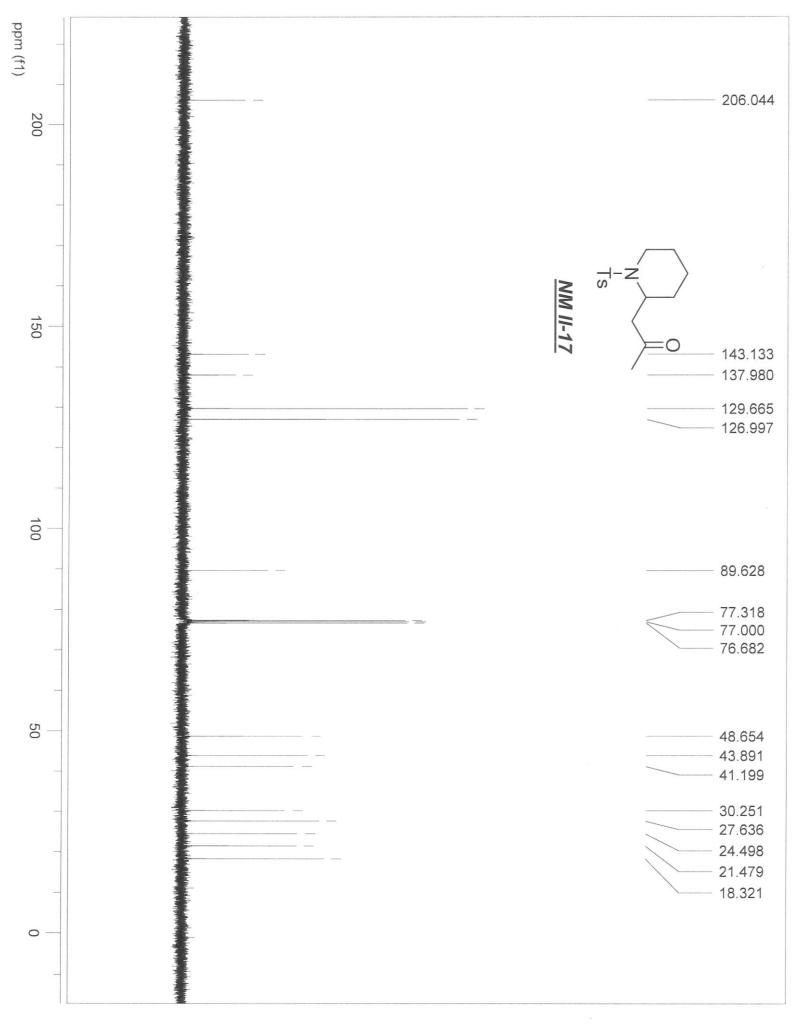


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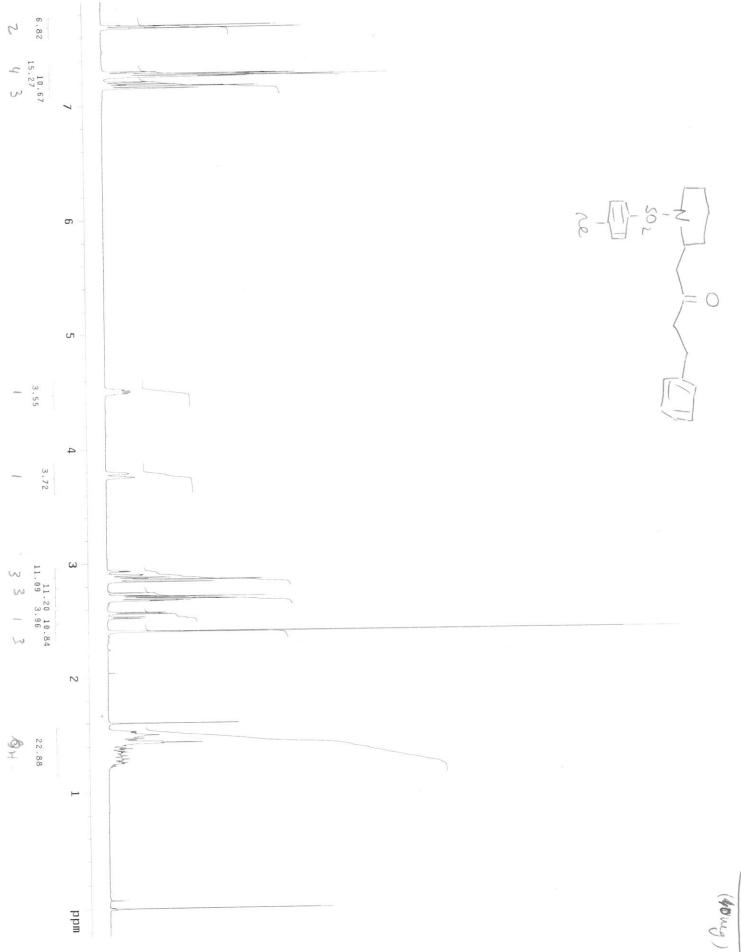
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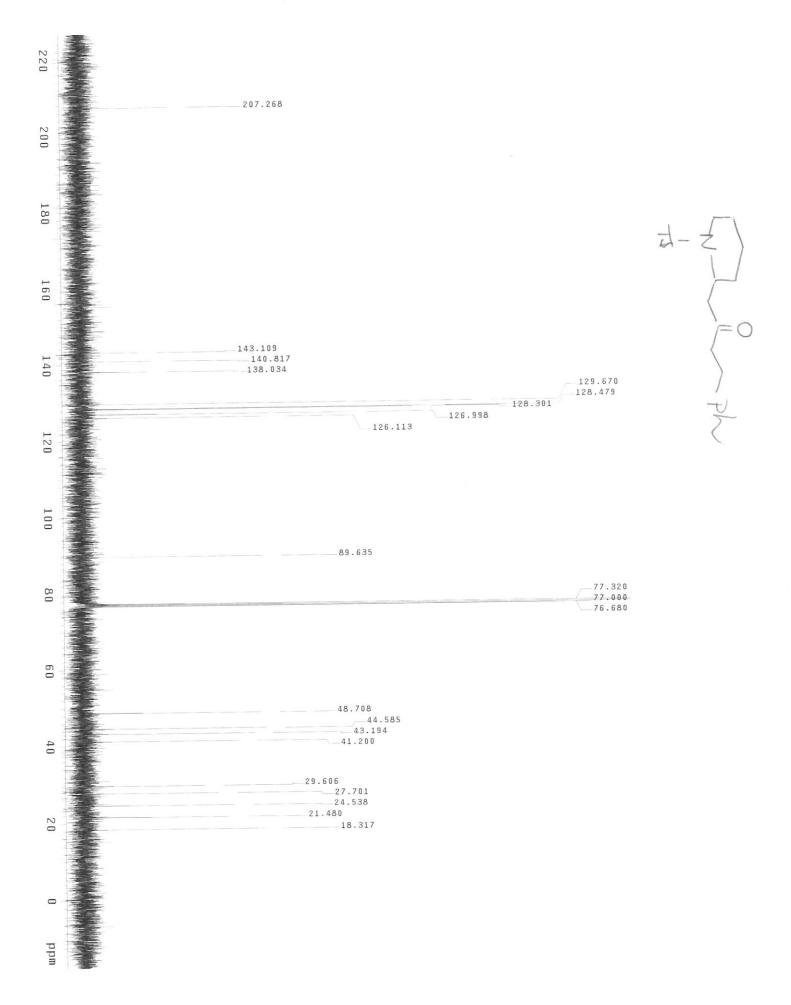


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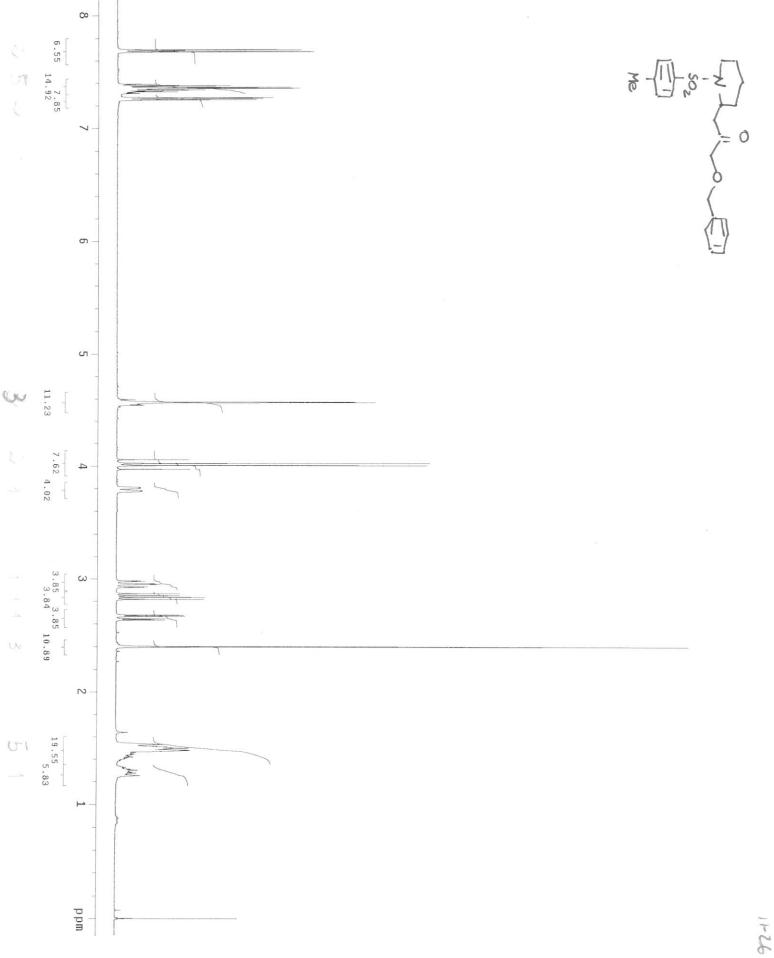


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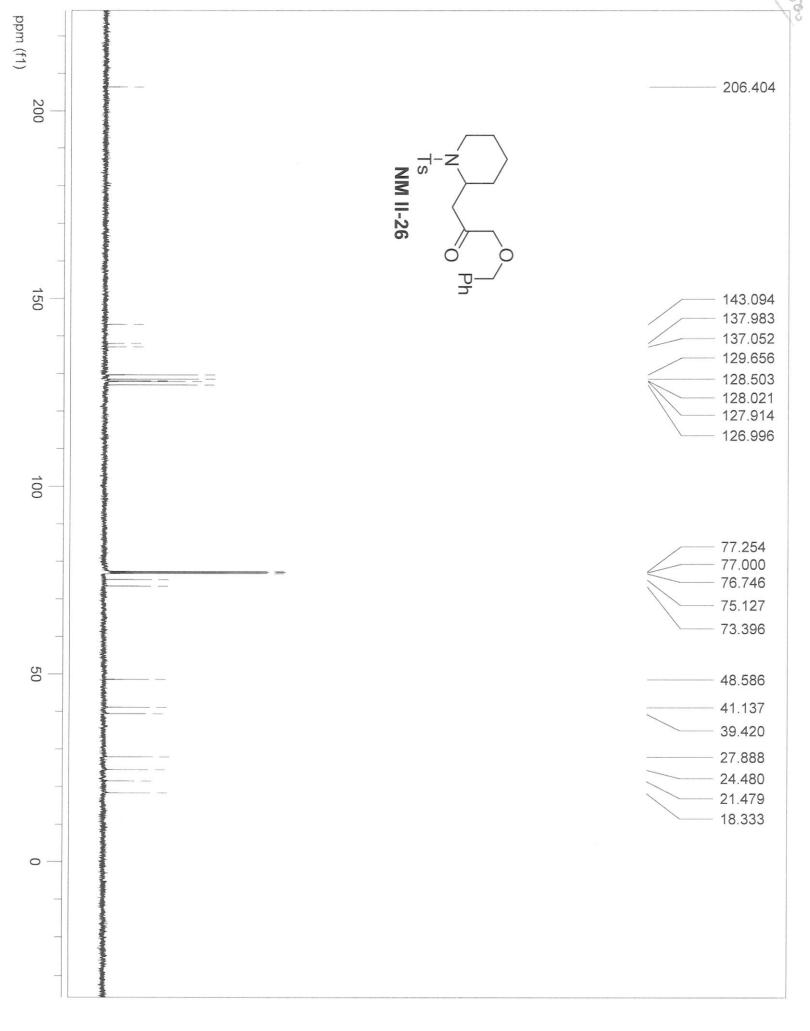


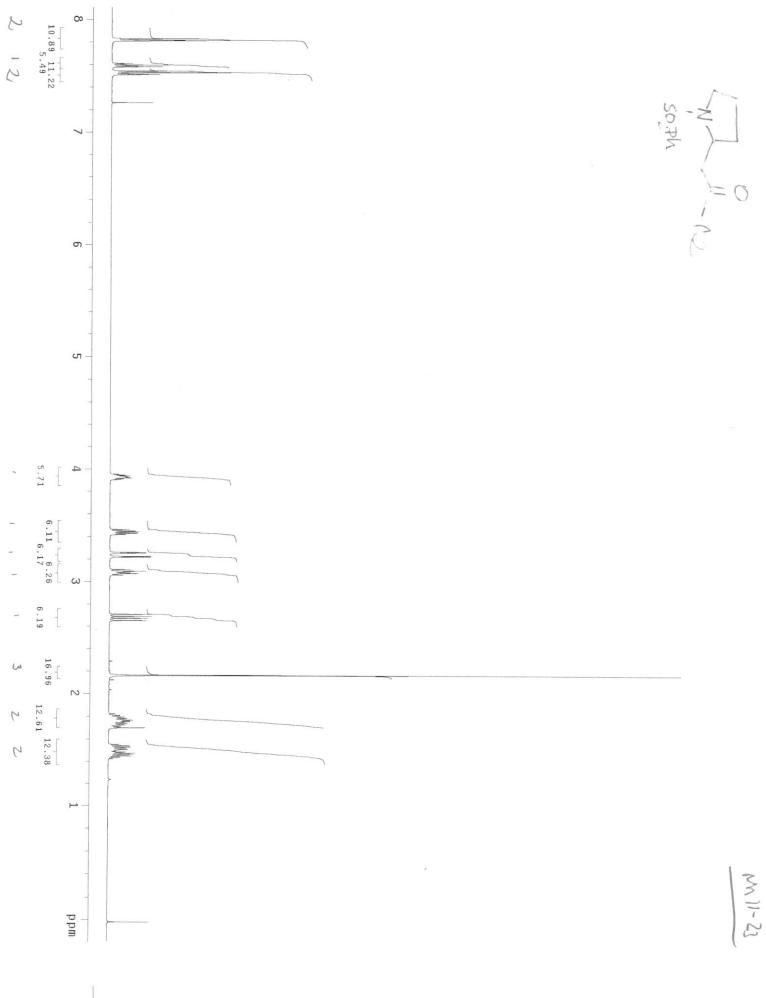
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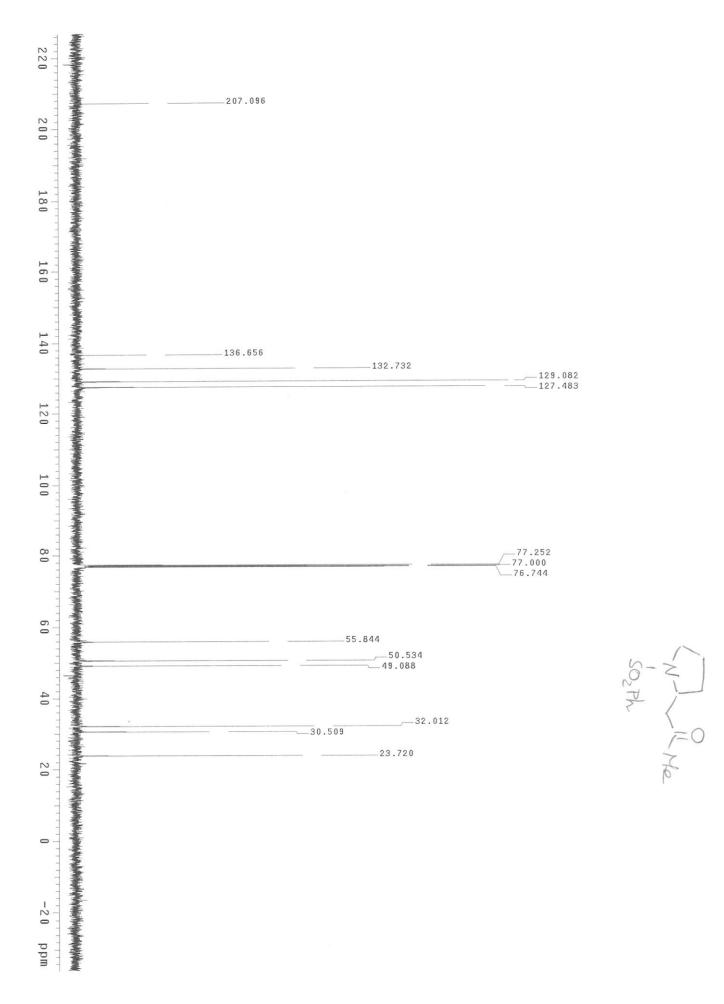


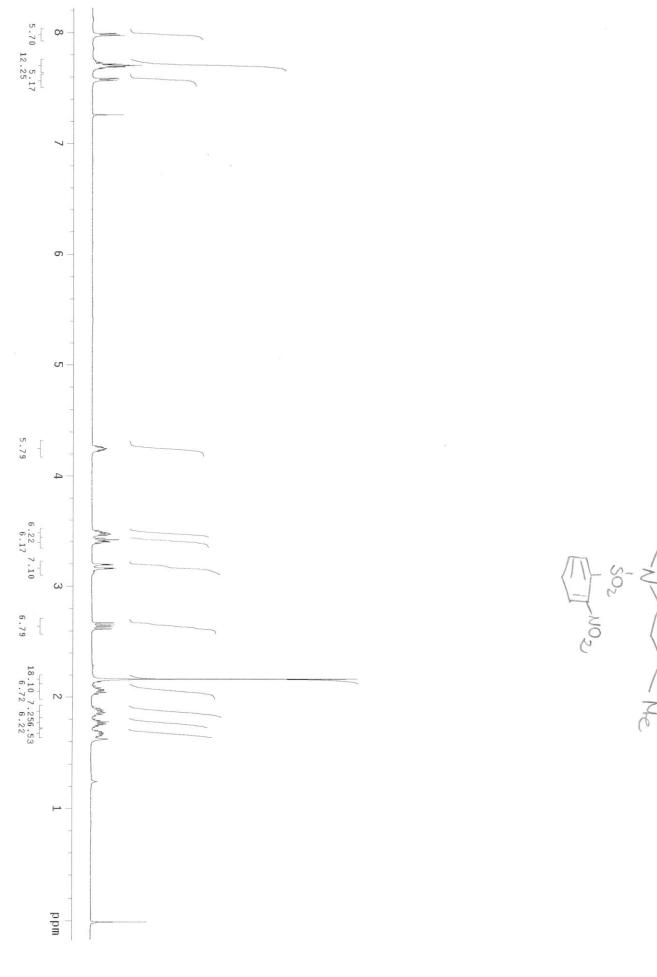
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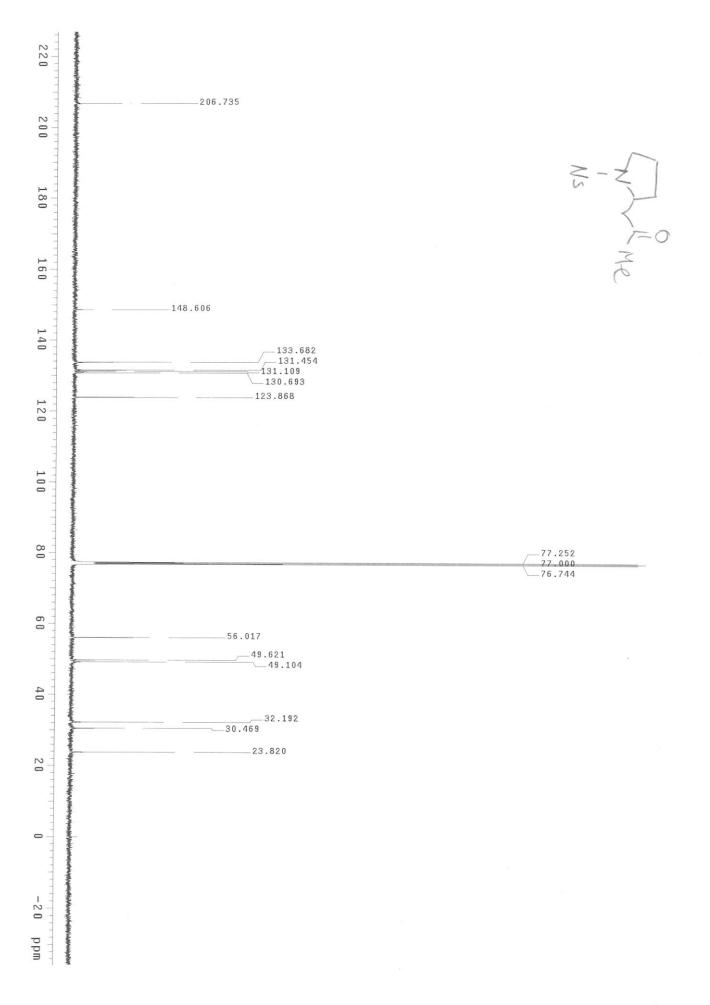




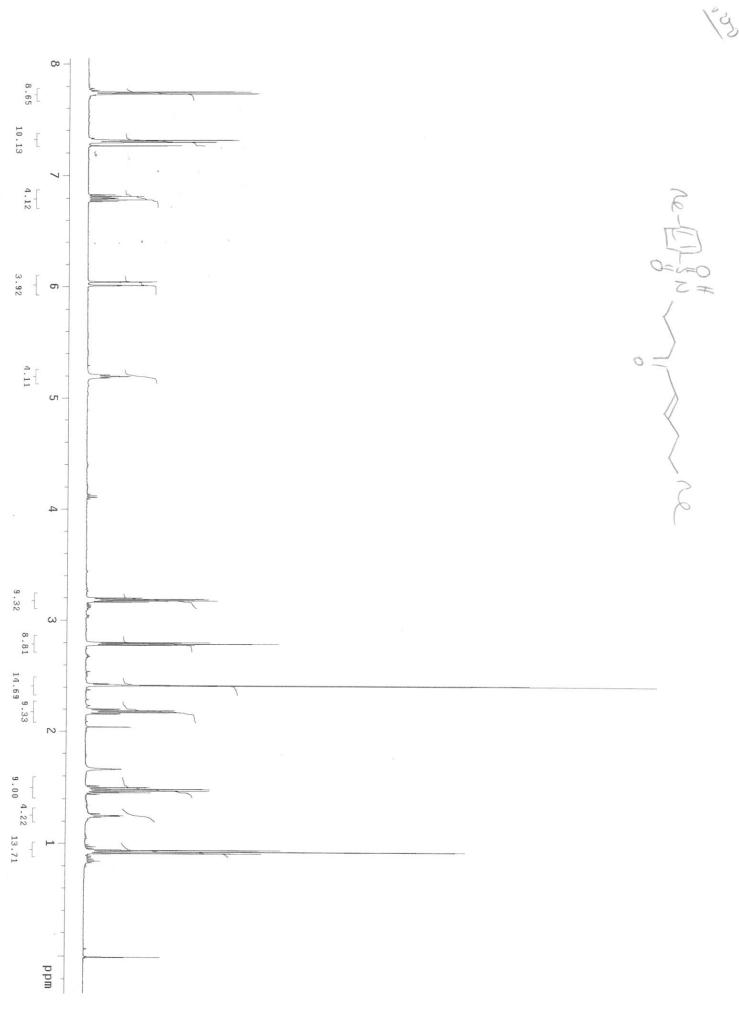
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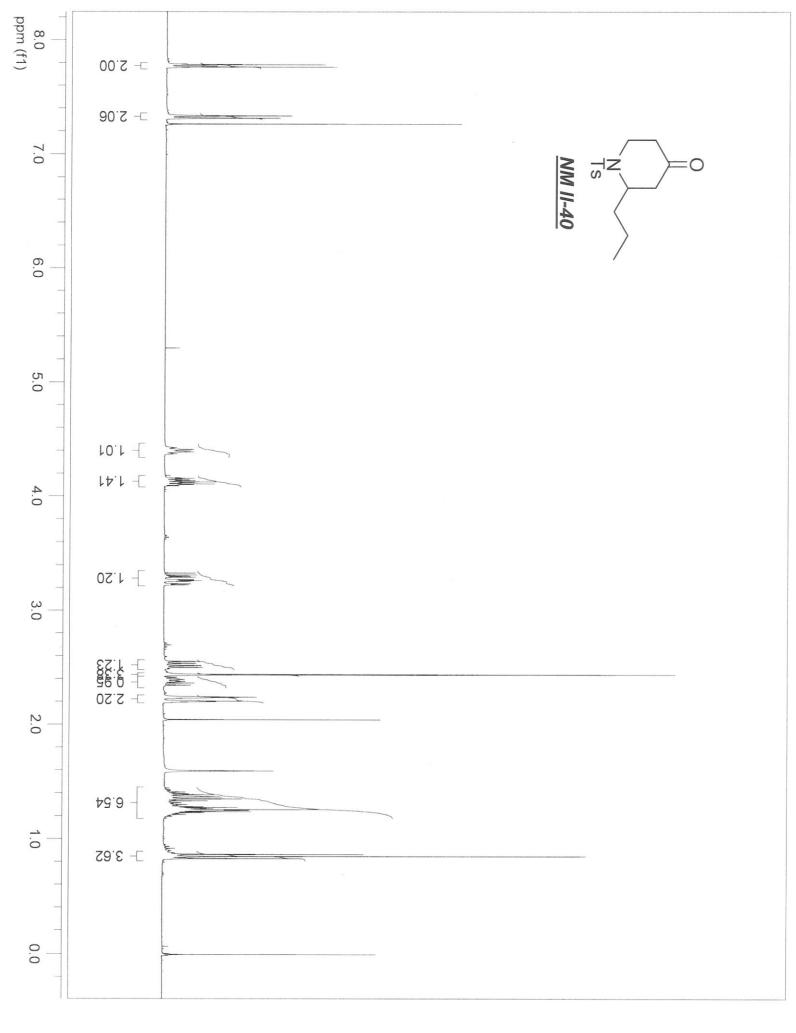
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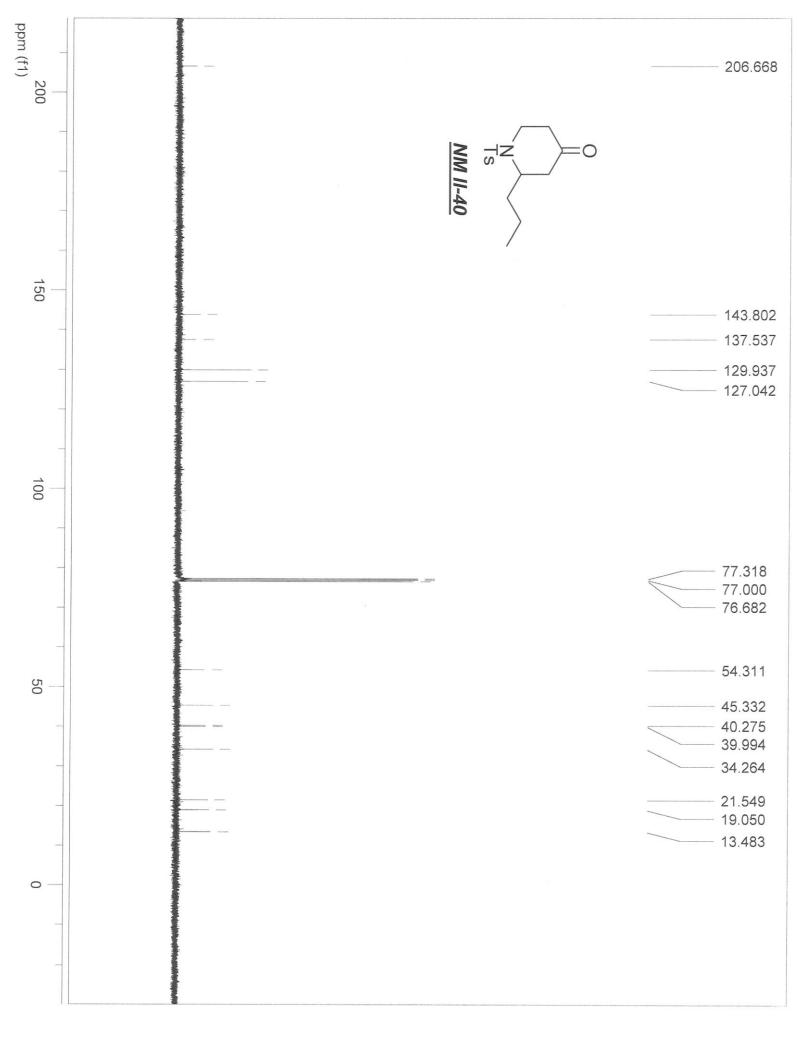
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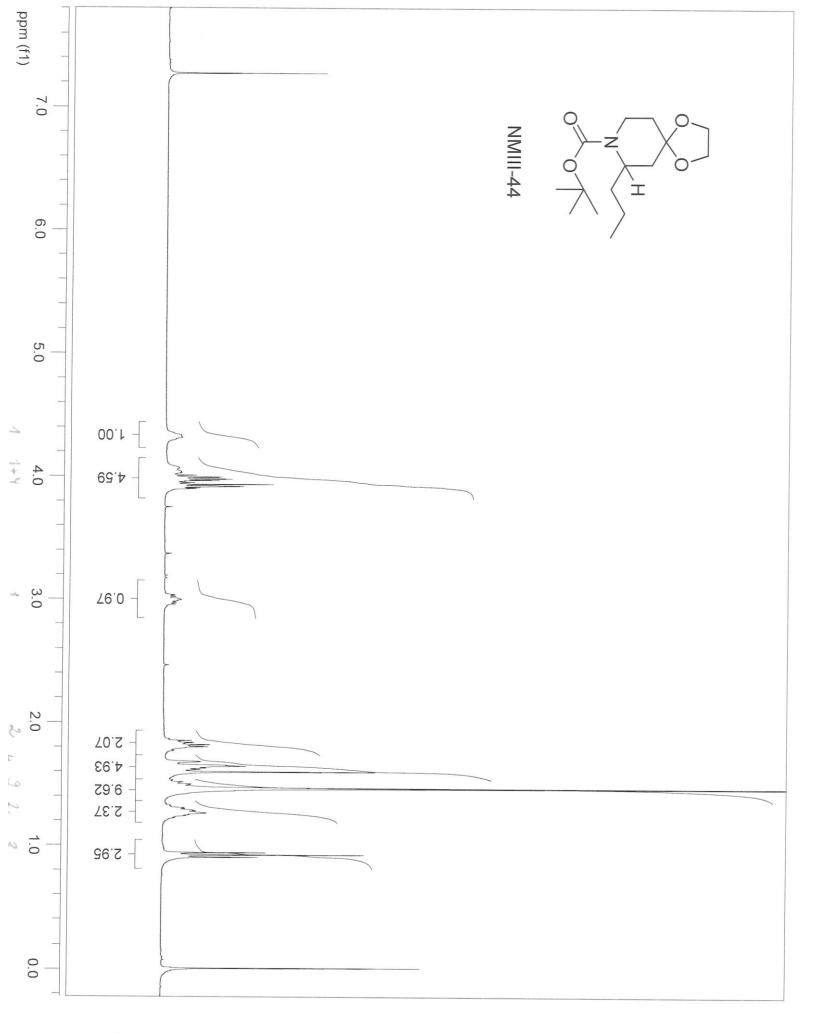


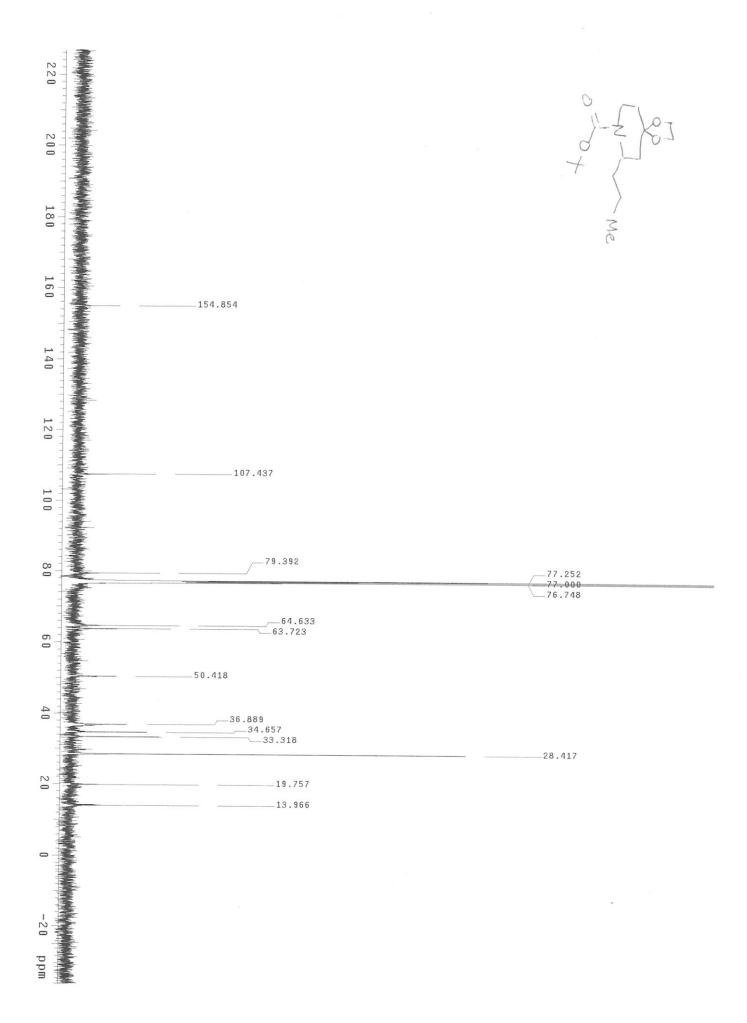
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