

Supporting Information

Pizzato 10.1073/pnas.1001554107

SI Text

VDJ Junction Sequence of the *tcr* of JJ6 and JTag Is Identical. DNA was extracted from JM, JJ6, and CEM cells. A PCR to amplify a TCR gene region encompassing the VDJ junction was performed by using primers 5'-AGTCGGTGACCCAGCTTGGC-3' and 5'-ATCTTGGAAGCACTGCTGTTCGG-3'. A PCR product was only obtained from DNA derived from JTag and JJ6. The DNA sequence of the PCR products was determined and found to be 100% identical in the two cell lines and to the sequence of the Jurkat T cell receptor active α -chain precursor mRNA (accession no. M12423).

Effect of gPr80 and Nef on Cell Surface Expression Levels of CD4 and MHC-I. JTag cells and mouse immortalized fibroblasts MC57 were transfected with vectors encoding HA-gg189 or Nef-HA in combination with a plasmid encoding EGFP. Forty-eight hours after transfection, surface human CD4, and human and mouse

MHC-1 expression levels on cells gated for GFP expression were determined by two-color flow cytometry after staining with mouse anti-CD4, anti-human HLA-ABC, and anti-mouse H-2K^b (BD Pharmingen) followed by APC-conjugated anti-mouse (Jackson ImmunoResearch).

Virus Binding Assay. Virus binding assay for detection of mCAT-1 surface expression levels was performed on JTag cells stably expressing mCAT-1 or control parental JTag cells. Cells were transfected with vectors encoding HA-gg189 or Nef-HA in combination with a plasmid encoding EGFP. Forty-eight hours after transfection, cells were incubated with a MoMLV suspension produced from 293T cells transfected with pNCA. at 4 °C for 60 min. Virus-cell complexes were stained with anti-RLV SU (Quality Biotech) followed by APC-conjugated anti-goat, and cells gated for GFP expression were analyzed.

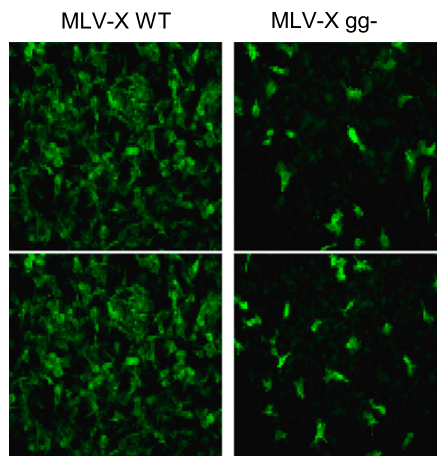


Fig. S1. gPr80 is required for optimal infectivity of MLV. MLV-X restricted to a single cycle of replication produced from JTag cells and normalized for RT activity was inoculated onto HT10180 cells. The figure shows immunofluorescence staining of infected target cells performed 48 h after infection by using an anti-RLV P30 antibody followed by Alexa-488 conjugated anti-goat antibodies.

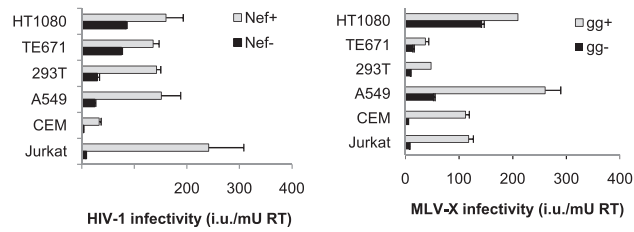


Fig. S2. Absolute infectivity of HIV-1 and MLV-X derived from different producer cell types. Results expressed as percentage of WT in Fig. 5B are here shown as absolute infectivity values.

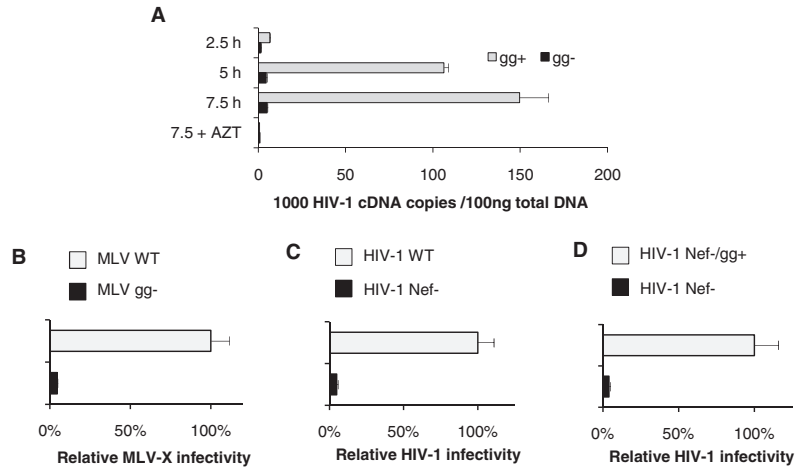


Fig. S3. gPr80 affects progression of HIV-1 reverse transcription. (A) gPr80 affects the steady-state levels of Nef-negative HIV-1 full-length viral cDNA, resembling the effect of Nef (Fig. 5C). (B–D) Relative infectivity of HIV-1 and MLV-X used in Fig. 5C. Viruses produced in JTAG cells and used in Fig. 5C and in A were serially diluted and inoculated onto HT1080 (B) or NP2-CD4/CXCR4 (C and D) respectively. Infected target cells were stained with anti RLV P30 anti HIV-1 p55/p2 and followed by Alexa-488 conjugated antibodies and quantified by scoring clusters of infected cells by fluorescence microscopy.

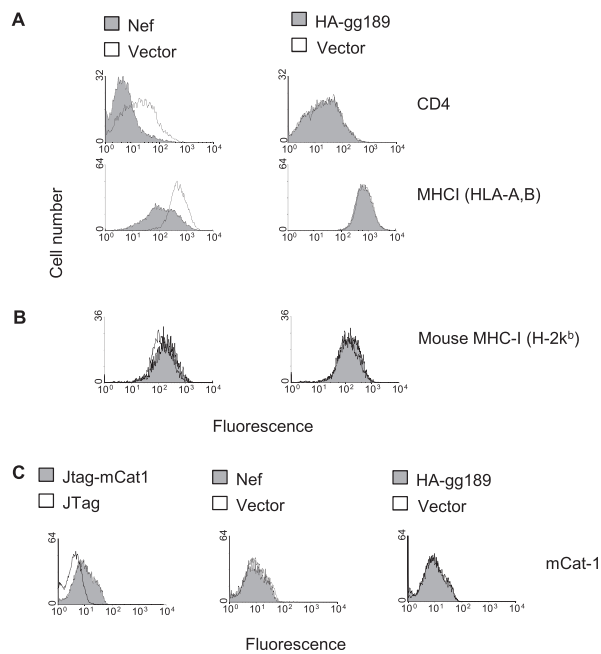


Fig. S6. Unlike Nef, glycoprotein does not down-regulate retrovirus receptors and MHC-I. (A) gPr80 does not affect cell surface expression levels of human CD4 and MHC-I in JTag cells. (B) Neither gPr80 nor Nef affect surface levels of mouse MHC-I (H-2k^b) in MC57 mouse fibroblasts. Of note, the failure of HIV-1 Nef to down-regulate MHC-I in mouse cells has been reported (1) (C) Virus binding assay for detection of cell surface mCat-1 on JTag/mCat-1 cells (*SI Text*). (Left) The presence of mCat-1 on JTag cells promotes virus binding detected by using an anti-SU antibody. (Center and Right) Nef and gPr80 expression in JTag/mCat1 do not affect efficiency of SU binding and, therefore, of mCat-1 surface expression level.

1. Fleis R, Filzen T, Collins KL (2002) Species-specific effects of HIV-1 Nef-mediated MHC-I downmodulation. *Virology* 303:120–129.

Table S1. Properties and origin of the cell lines used in Fig. 1

Cell line	Origin	Comments	Source
C8166	Human T cell leukemia	Parental	Imperial College London
CEM	Acute lymphoblastic leukemia	Parental	University College London
CEM/A3.01	Acute lymphoblastic leukemia	Derivative of CEM, highly permissive to HIV-1 replication	NIH AIDS Research & Reference Reagent Program
CEM/A3.01-F7	Acute lymphoblastic leukemia	Derivative of CEM/A3.01, ectopic expression of CCR5	National Biological Standards Board
CEM-SS	Acute lymphoblastic leukemia	Derivative of CEM, highly susceptible for fusion with HIV	NIH AIDS Research & Reference Reagent Program
DG75/HAD	Burkitt's lymphoma	Parental, EBV-negative	Medical College of Wisconsin
DG75/UW	Burkitt's lymphoma	Parental, EBV-negative	Hadassah Medical Center
H9	Human T cell lymphoma	Derivative of HUT-78, highly permissive for HIV-1 replication	Imperial College London
HBS-2	Acute lymphoblastic leukemia	parental, CD4-negative	NIH AIDS Research & Reference Reagent Program
Jurkat D1.1	Acute T cell leukemia	Derivative of JM, CD4-negative	American Type Culture Collection
Jurkat E6.1	Acute T cell leukemia	Derivative of JM, high producers of IL-2	American Type Culture Collection
Jurkat J6	Acute T cell leukemia	Derivative of JM, high expression of CD3	London Research Institute
Jurkat JM	Acute T cell leukemia	Parental	NIH AIDS Research & Reference Reagent Program
Jurkat TAg	Acute T cell leukemia	Derivative of JM, express SV40 Large T antigen	Dana-Farber Cancer Institute
RAJI	Burkitt's lymphoma	Parental, EBV-positive	Imperial College London
Ramos	Burkitt's lymphoma	Parental, EBV-negative	Imperial College London
SupT1	Lymphoblastic leukemia	Parental	Imperial College London
U937	Histiocytic lymphoma	Parental	Imperial College London