

## ONLINE APPENDIX - Supplementary Materials

### Experimental Procedures

*Immunofluorescence staining and flowcytometric analysis*— To determine the purity of isolated CD4<sup>+</sup> T lymphocytes, CD4<sup>+</sup> T cells from the spleen, thymus and LN were treated with Fc receptor blocking CD16/32 antibody and subsequently incubated with PE anti-CD3, APC anti-CD4 or PerCP anti-CD8 antibodies (BioLegend, San Diego, CA) for 30 mins at 4°C. Following washing steps, cells were analyzed with an LSRII and a FACS Diva software (BD Biosciences, San Diego, CA).

*Rac1 pull-down activation assay*— GTP-bound Rac1 was measured using a Rac1 activation assay kit (Upstate Technology) according to the manufacturer's instructions. Briefly, DPP-IV (100 mU/ml), GIP (100 nM) or GLP-1 (100 nM) treated CD4<sup>+</sup> T lymphocytes were lysed in ice-cold magnesium lysis buffer and cleared with glutathione-agarose beads. Cell extracts were then incubated with p21-binding domain (PBD, residues 67-150) of human PAK-1 agarose beads, the resulting beads were resuspended and separated on 15 % sodium dodecyl sulfate (SDS)/polyacrylamide gel. GTP-bound Rac1 was detected using an anti-Rac1 antibody.

*Preparation of nuclear extracts*— Nuclear proteins were isolated as described by Schreiber *et al* (1). Briefly, LN CD4<sup>+</sup> T cells were washed with PBS, and disrupted with 200  $\mu$ l ice-cold buffer A (10 mM HEPES pH 7.9, 10 mM KCl, 1.5 mM MgCl<sub>2</sub>, 1 mM EDTA, 1 mM dithiothreitol, 0.1 % Nonidet P40 and protease inhibitors). Following centrifugation, the resulting pellet was re-suspended in 20  $\mu$ l buffer B (20 mM HEPES, pH 7.9, 400 mM NaCl, 1 mM EDTA, 1 mM dithiothreitol, 20 % glycerol and protease inhibitors) and incubated on ice for 10 min. After clarification of the mixture by centrifugation, the supernatant (nuclear extract) was collected and subjected to an NF $\kappa$ B transcription factor activity assay.

*NF $\kappa$ B transcription factor activity assay*— An NF $\kappa$ B transcription factor activity assay kit (Millipore, MA) was used to measure DNA binding activity of NF $\kappa$ B transcription factor in nuclear extracts of LN CD4<sup>+</sup> T cells, according to the manufacturer's protocols. Briefly, nuclear extracts of LN CD4<sup>+</sup> T cells were incubated with biotinylated oligonucleotide containing the consensus sequence for NF $\kappa$ B binding (5'-GGGACTTTCC-3'), and the biotinylated oligonucleotide bound by active NF $\kappa$ B protein was immobilized. The bound NF $\kappa$ B transcription factor subunit p50, was detected with a specific primary antibody,  $\alpha$ -NF $\kappa$ B p50 (active form), and subsequently with an HRP-conjugated secondary antibody. Nuclear NF $\kappa$ B activity is presented as relative activity normalized to protein concentration.

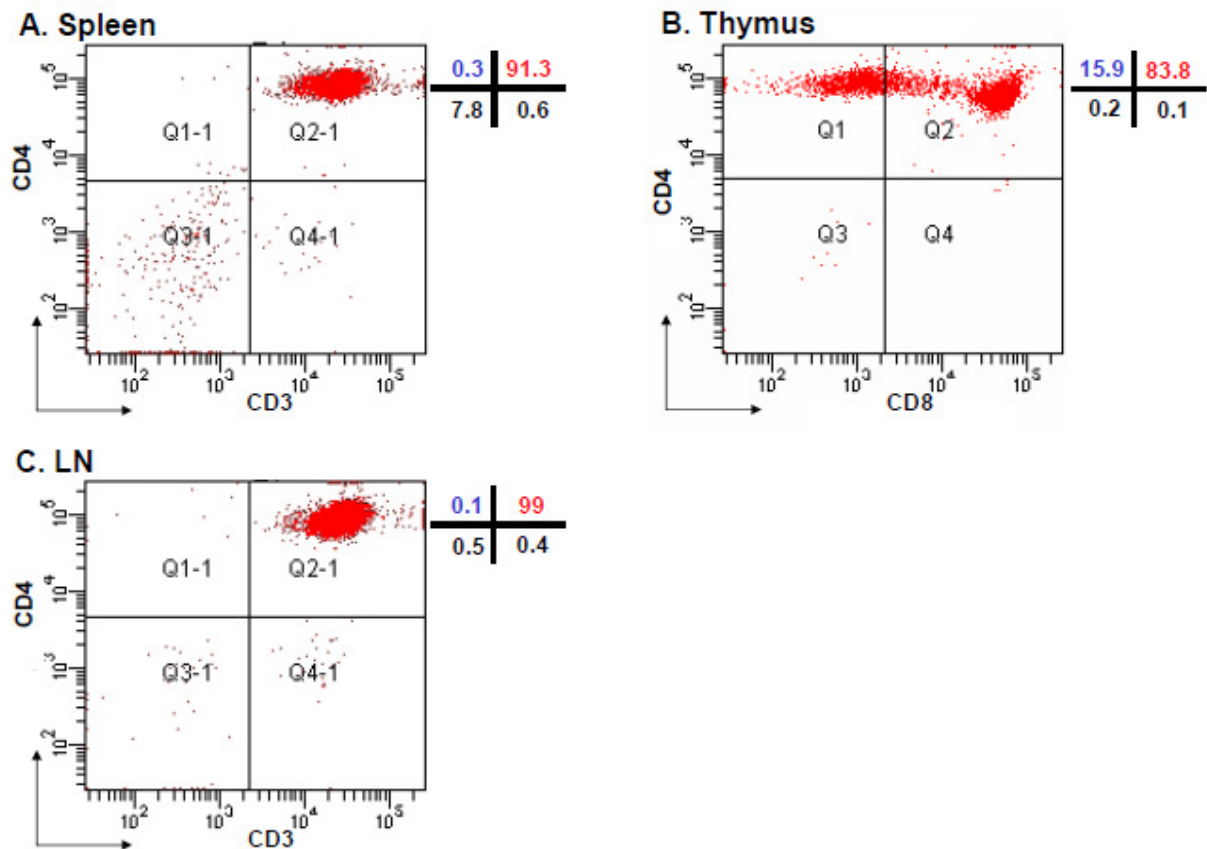
*Western blot analysis*— Total cellular or nuclear extracts were separated on a 15% sodium dodecyl sulfate (SDS)/polyacrylamide gel and transferred onto nitrocellulose membranes (Bio-Rad). Probing of the membranes was performed with antibodies against phospho-Cofilin (Serine3), Cofilin, Phospho-Ezrin (Threonine567)/Radixin (Threonine564)/Moesin (Threonine558), Ezrin/Radixin/Moesin, phospho-VASP (Serine157), phospho-VASP (Serine239), VASP, phospho-I $\kappa$ B $\alpha$  (Ser32), I $\kappa$ B $\alpha$ , phospho-NF $\kappa$ B p65 (Ser536), NF $\kappa$ B p65, phospho-IKK $\alpha$ / $\beta$  (Ser176/180), IKK $\alpha$ , IKK $\beta$ , NF $\kappa$ B p50 and  $\beta$ -actin (Cell Signaling Technology, Beverly, MA), as identified in the figures. Immunoreactive bands were visualized by enhanced chemiluminescence

(Amersham Pharmacia Biotech) using horseradish peroxidase-conjugated IgG secondary antibodies.

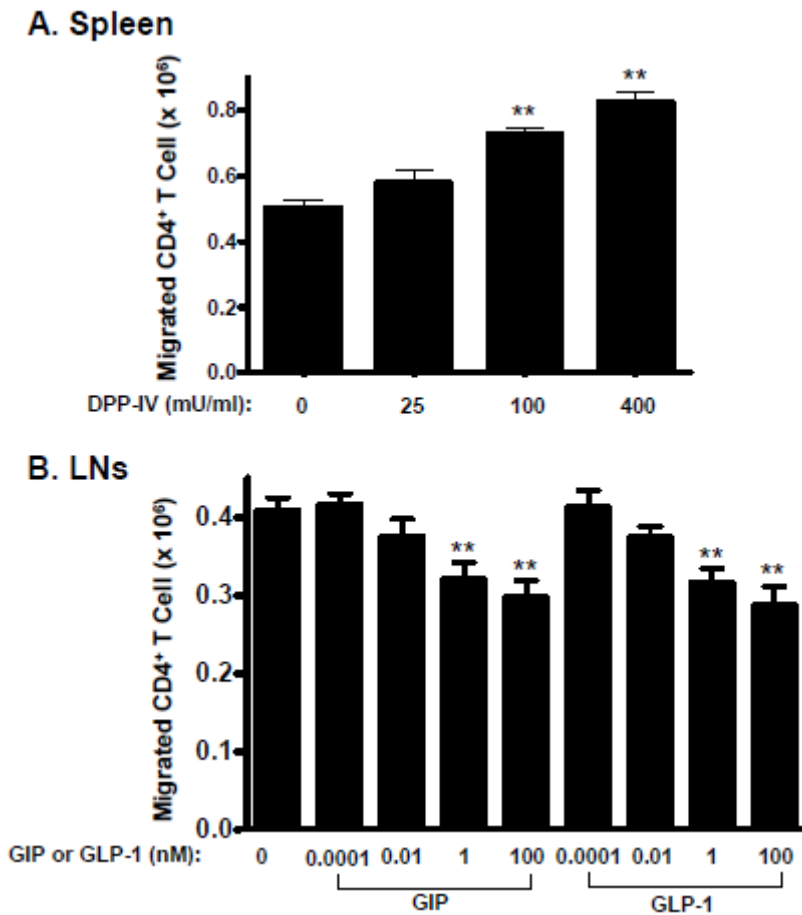
## REFERENCES

1. Schreiber E, Matthias P, Müller MM, Schaffner W: Rapid detection of octamer binding proteins with 'mini-extracts', prepared from a small number of cells. *Nucleic Acids Research* 17: 6419, 1989.

**Supplementary Figure 1. Representative FACS profiles of purified CD4<sup>+</sup> T cells.** For these, and all subsequent, studies presented as Supplementary Figures, CD4<sup>+</sup> T cells were positively isolated from non-diabetic female NOD mice receiving NCD. FACS profile of CD4<sup>+</sup> T cells from Spleen (A) and LN (C) in CD3<sup>+</sup>CD4<sup>+</sup>-gated events, and Thymus (B) in CD4<sup>+</sup> CD8<sup>+</sup>-gated events.

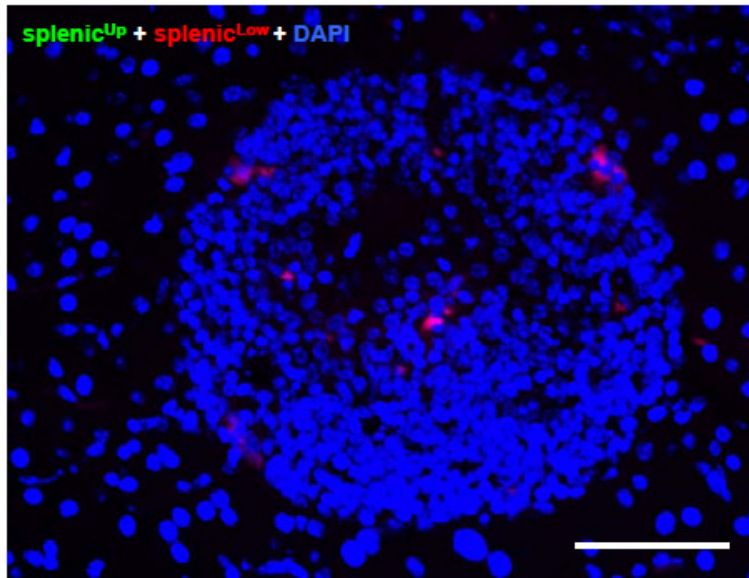


**Supplementary Figure 2. Concentration dependent effects of sDPP-IV and incretins on the migration of CD4<sup>+</sup> T cells.** Splenic (A) or LN (B) CD4<sup>+</sup> T cells isolated from non-diabetic female NOD mice were incubated with indicated concentrations of sDPP-IV, GIP or GLP-1. The migration of CD4<sup>+</sup> T cells was determined using Transwell chamber (Corning) as described in *Experimental Procedures*. All data represent the mean  $\pm$  S.E.M. and significance was tested using ANOVA with a Newman-Keuls post hoc test, where \*\* represents  $p < 0.05$  vs Control.



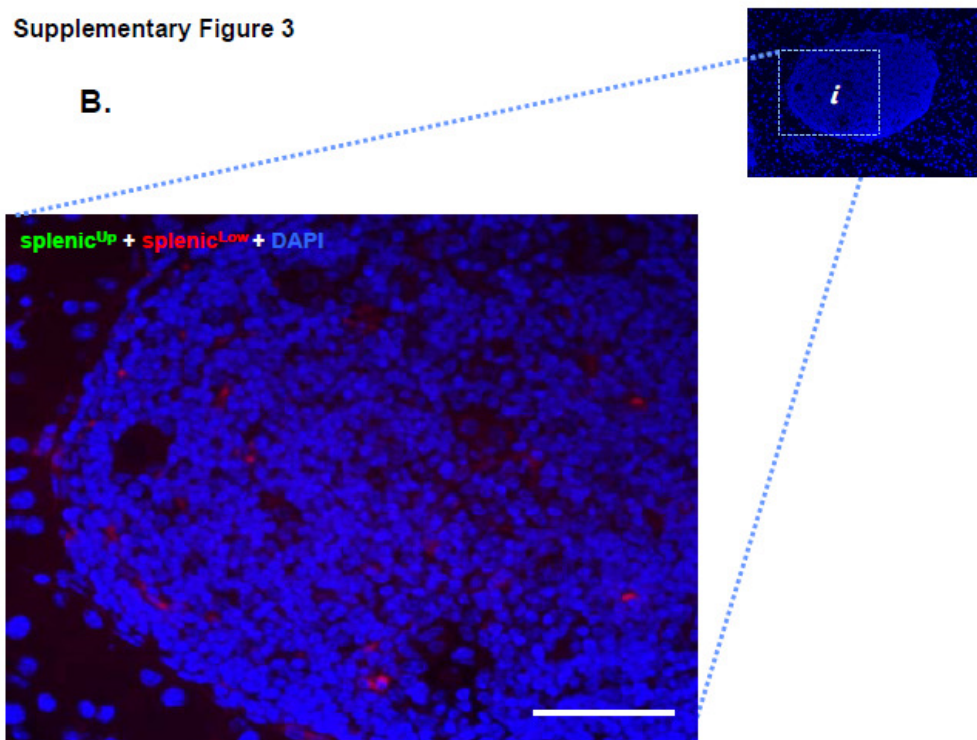
**Supplementary Figure 3. Islet localization of labeled splenic (A-D) and LN (E-H) CD4<sup>+</sup> T lymphocytes following sDPP-IV or incretin stimulation.** splenic<sup>Up</sup> and splenic<sup>Low</sup> (A-D) and LN<sup>Up</sup> and LN<sup>Low</sup> (E-H) = Splenic or LN lymphocytes from upper and lower chambers, respectively. Mixtures of DiI- and CFSE- labeled splenic (A-D) and LN (E-H) CD4<sup>+</sup> T lymphocytes were intravenously injected into diabetic recipient mice and pancreatic homing of labeled lymphocytes determined by confocal fluorescent microscopy. Infiltrated islets are represented with an *i*, and the scale bar indicates 25  $\mu$ m.

**A.**



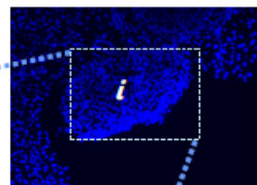
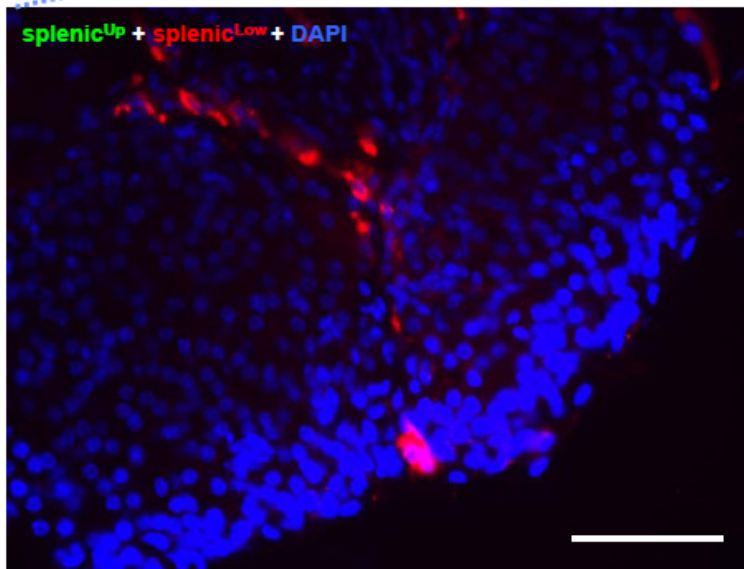
Supplementary Figure 3

**B.**



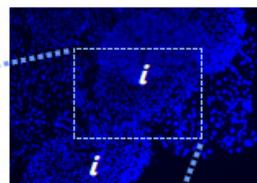
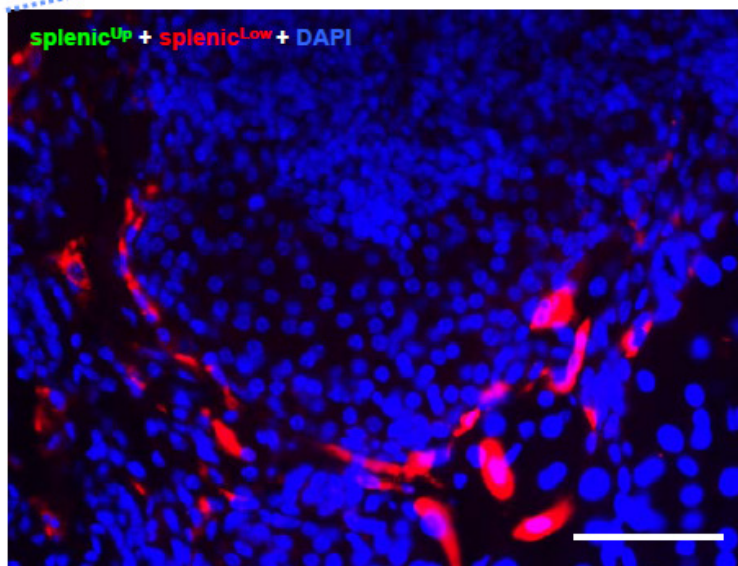
Supplementary Figure 3

C.

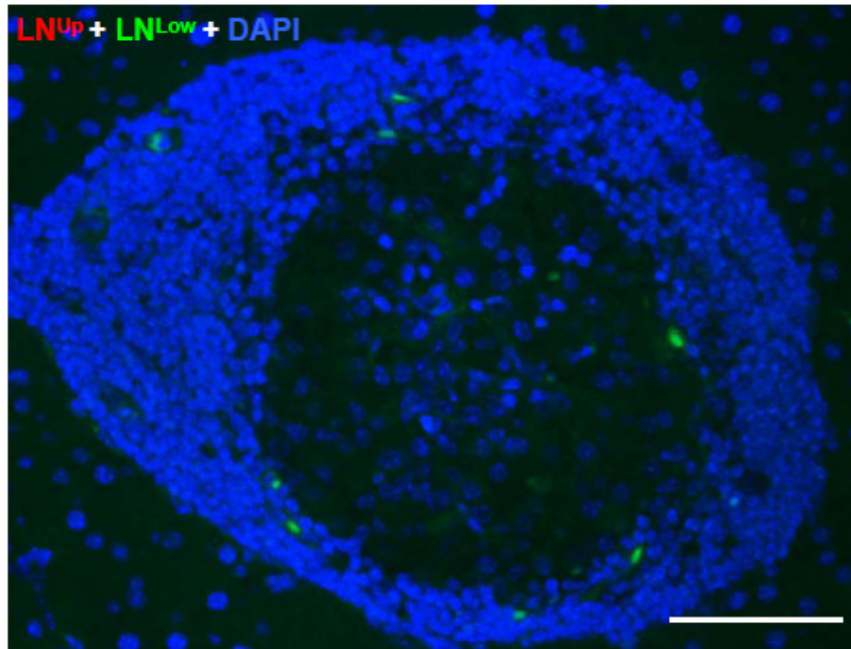


Supplementary Figure 3

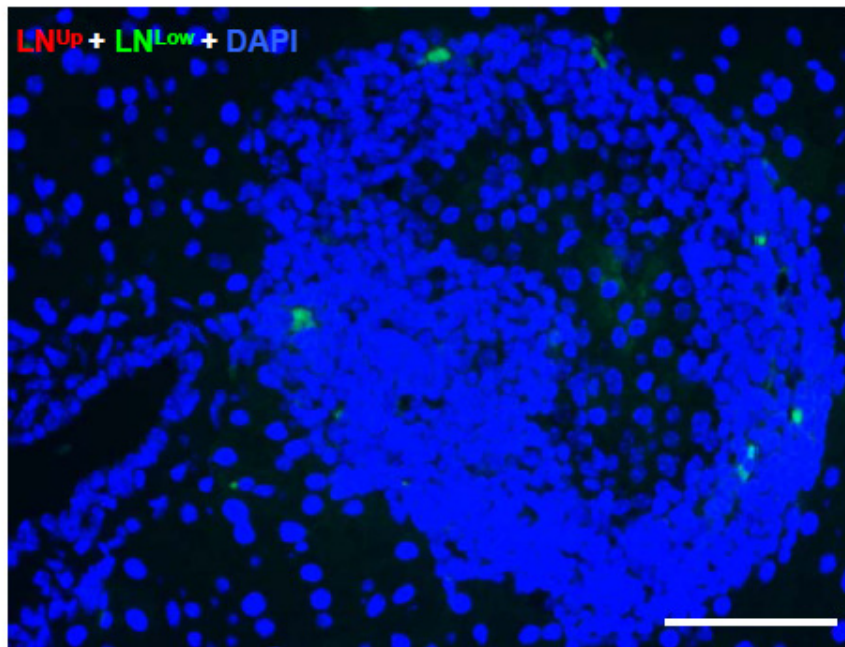
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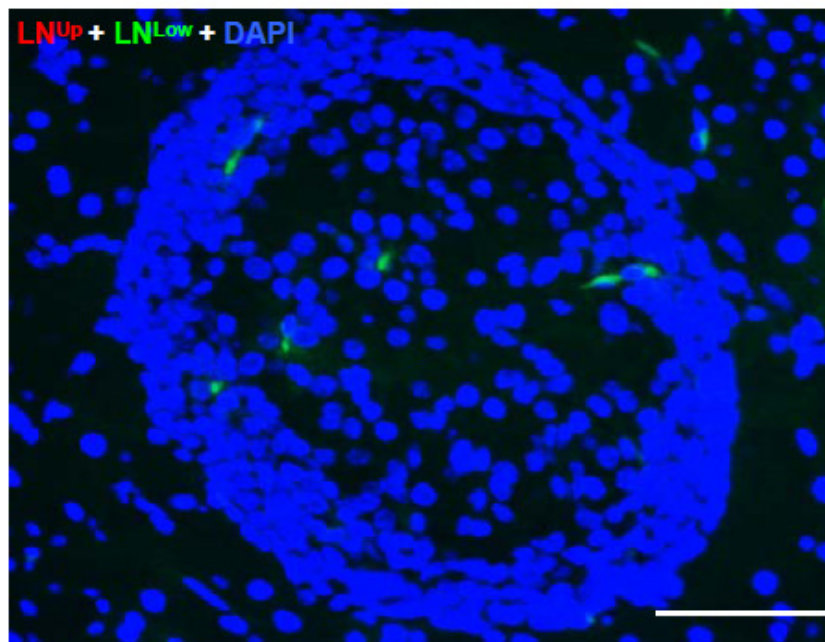
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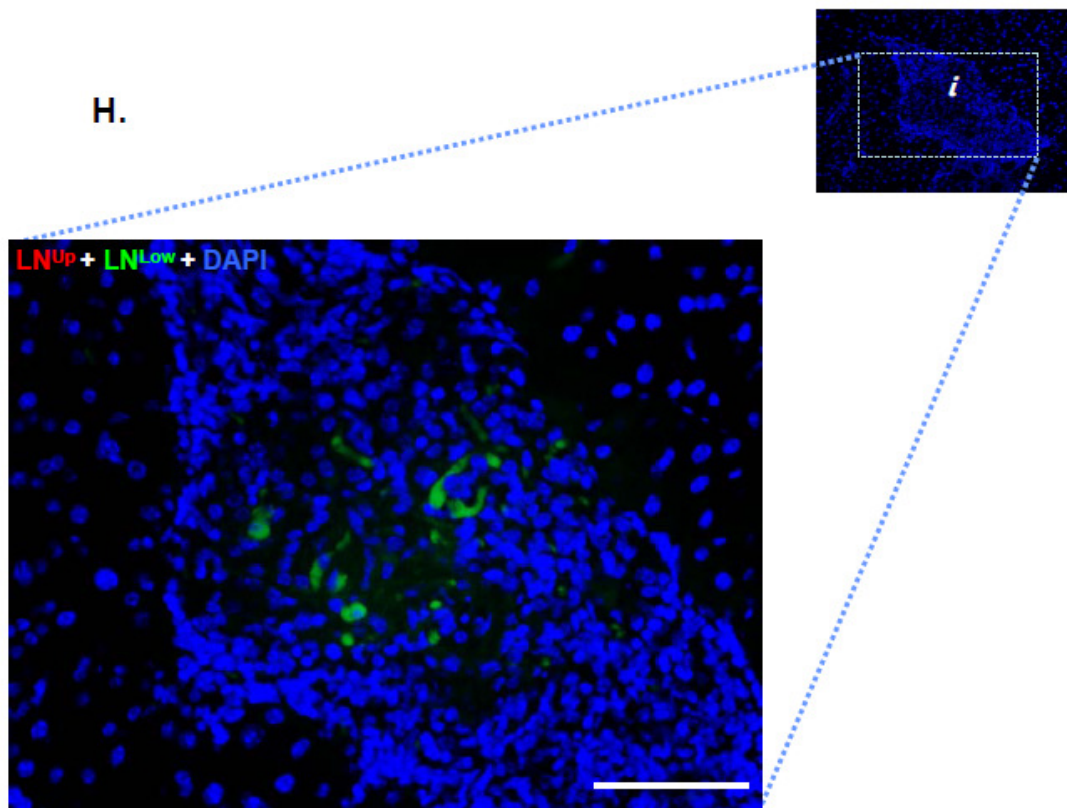
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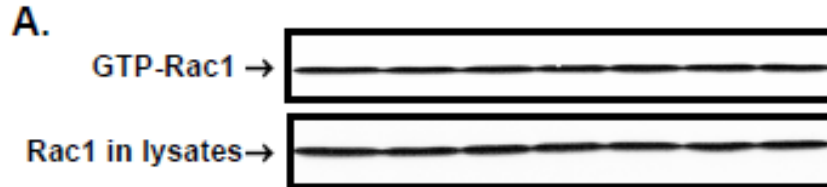
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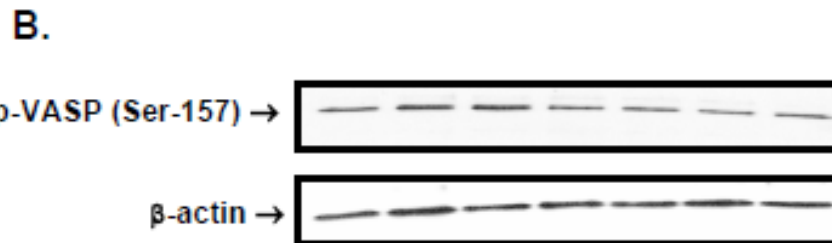
H.



**Supplementary Figure 4. The effect of Sitagliptin on phosphorylation levels of cytoskeletal organizing protein in thymic and LN CD4<sup>+</sup> T cells.** CD4<sup>+</sup> T cells were stimulated for 1 h with sDPP-IV (100 mU/ml), GIP or GLP-1 (100 nM) in the presence or absence of DPP-IV inhibitor (100 μM). Total cellular extracts were isolated from thymic (A-D) or LN (E-H) CD4<sup>+</sup> T cells and Rac1 activation assays (A, E) or Western blot analyses performed with antibodies against phospho-VASP (Ser157) (B, F), Phospho-Ezrin (Thr567)/Radixin (Thr564)/Moesin (Thr558), Ezrin/Radixin/Moesin (C, G), phospho-Cofilin (Ser3), Cofilin (D, H), and β-actin. Western blots are representative of *n* = 3.



<b>GIP (100 nM):</b>	–	–	–	+	+	–	–
<b>GLP-1 (100 nM):</b>	–	–	–	–	–	+	+
<b>DPP-IV (100 mU/ml):</b>	–	+	+	–	–	–	–
<b>Sitagliptin (100 μM):</b>	–	–	+	–	+	–	+



<b>GIP (100 nM):</b>	–	–	–	+	+	–	–
<b>GLP-1 (100 nM):</b>	–	–	–	–	–	+	+
<b>DPP-IV (100 mU/ml):</b>	–	+	+	–	–	–	–
<b>Sitagliptin (100 μM):</b>	–	–	+	–	+	–	+



**C.**

p-Ezrin (Thr-567)/ Radixin (Thr-564)/ Moesin (Thr-558) →

Ezrin/ Radixin/ Moesin →

GIP (100 nM):	-	-	-	+	+	-	-
GLP-1 (100 nM):	-	-	-	-	-	+	+
DPP-IV (100 mU/ml):	-	+	+	-	-	-	-
Sitagliptin (100 μM):	-	-	+	-	+	-	+

**D.**

p-Cofilin (Ser-3) →

Cofilin →

GIP (100 nM):	-	-	-	+	+	-	-
GLP-1 (100 nM):	-	-	-	-	-	+	+
DPP-IV (100 mU/ml):	-	+	+	-	-	-	-
Sitagliptin (100 μM):	-	-	+	-	+	-	+

**E.**

GTP-Rac1 →

Rac1 in lysates →

GIP (100 nM):	-	-	-	+	+	-	-
GLP-1 (100 nM):	-	-	-	-	-	+	+
DPP-IV (100 mU/ml):	-	+	+	-	-	-	-
Sitagliptin (100 μM):	-	-	+	-	+	-	+

**F.**

p-VASP (Ser-157) →

β-actin →

GIP (100 nM):	-	-	-	+	+	-	-
GLP-1 (100 nM):	-	-	-	-	-	+	+
DPP-IV (100 mU/ml):	-	+	+	-	-	-	-
Sitagliptin (100 μM):	-	-	+	-	+	-	+


**G.**

p-Ezrin (Thr-567)/ Radixin  
(Thr-564)/ Moesin (Thr-558) 

Ezrin/ Radixin/  
Moesin 

GIP (100 nM):	-	-	-	+	+	-	-
GLP-1 (100 nM):	-	-	-	-	-	+	+
DPP-IV (100 mU/ml):	-	+	+	-	-	-	-
Sitagliptin (100 μM):	-	-	+	-	+	-	+

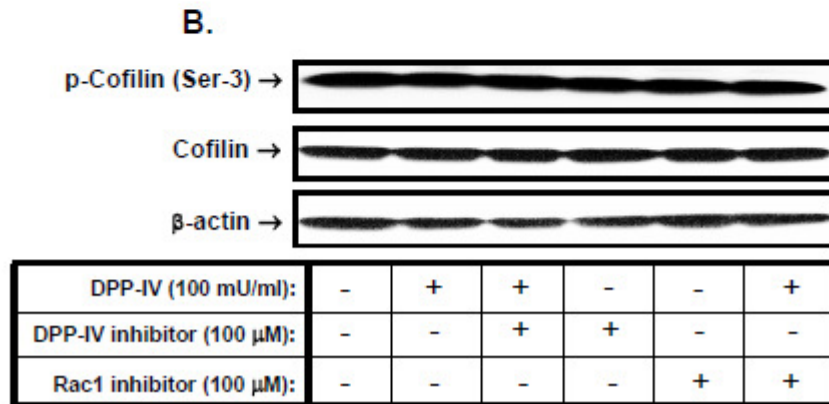
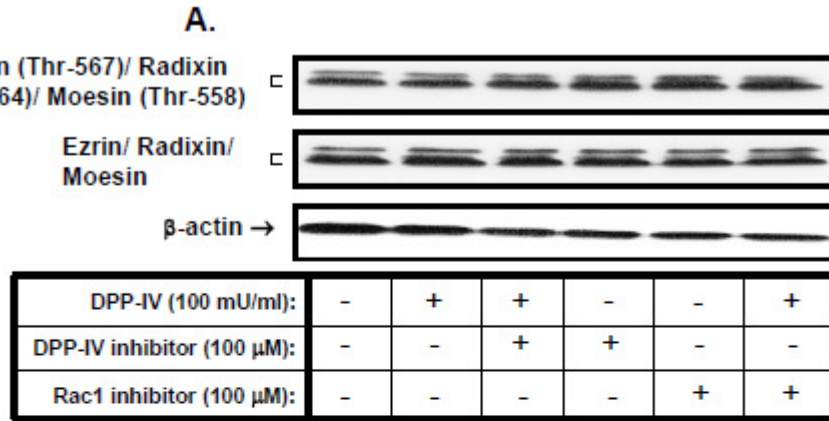
**H.**

p-Cofilin (Ser-3) → 

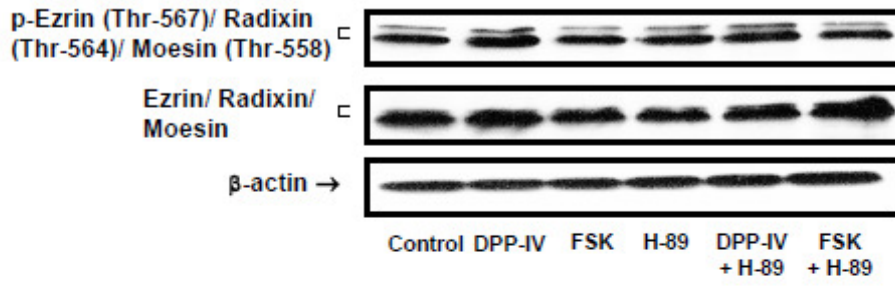
Cofilin → 

GIP (100 nM):	-	-	-	+	+	-	-
GLP-1 (100 nM):	-	-	-	-	-	+	+
DPP-IV (100 mU/ml):	-	+	+	-	-	-	-
Sitagliptin (100 μM):	-	-	+	-	+	-	+

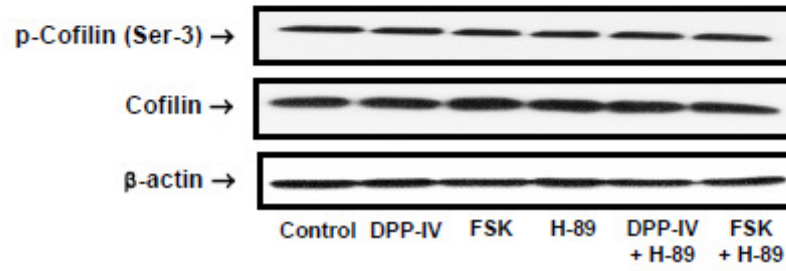
**Supplementary Figure 5. The effect of Rac1 inhibitor and PKA inhibitor on the phosphorylation of cytoskeletal organizing protein Ezrin (Thr567)/Radixin (Thr564)/Moesin (Thr558) and Cofilin (Ser3).** Splenic CD4<sup>+</sup> T lymphocytes were stimulated for 1 h with DPP-IV (100 mU/ml) in the presence or absence of Rac1 inhibitor (100  $\mu$ M) or forskolin (10  $\mu$ M)  $\pm$  H-89 (10  $\mu$ M) and total cellular extracts were isolated A-B. Effects of treatment with Rac1 inhibitor on the level of Ezrin (Thr567)/Radixin (Thr564)/Moesin (Thr558) (A) and Cofilin (Ser3) (B). C-D. Effect of treatment with H-89 on the level of Ezrin (Thr567)/Radixin (Thr564)/Moesin (Thr558) (C) and Cofilin (Ser3) (D). Western blots are representative of  $n = 3$ .



C.



D.



**Supplementary Figure 6. The effect of Sitagliptin on phosphorylation levels of NF- $\kappa$ B signalling modules in splenic and thymic CD4<sup>+</sup> T cells.** Total cellular extracts were isolated from splenic (A-C) or thymic (D-F) CD4<sup>+</sup> T cells and stimulated for 1 h with sDPP-IV (100 mU/ml), GIP or GLP-1 (100 nM) in the presence or absence of DPP-IV inhibitor (100  $\mu$ M). Western blot analyses were performed with antibodies against phospho-I $\kappa$ B (Ser32), I $\kappa$ B (A, D), Phospho-NF $\kappa$ B p65 (Ser536), NF $\kappa$ B p65 (B, E), phospho-IKK $\alpha/\beta$  (Ser176/180), IKK $\alpha$ , IKK $\beta$  (C, F), and  $\beta$ -actin. Western blots are representative of  $n = 3$ .

