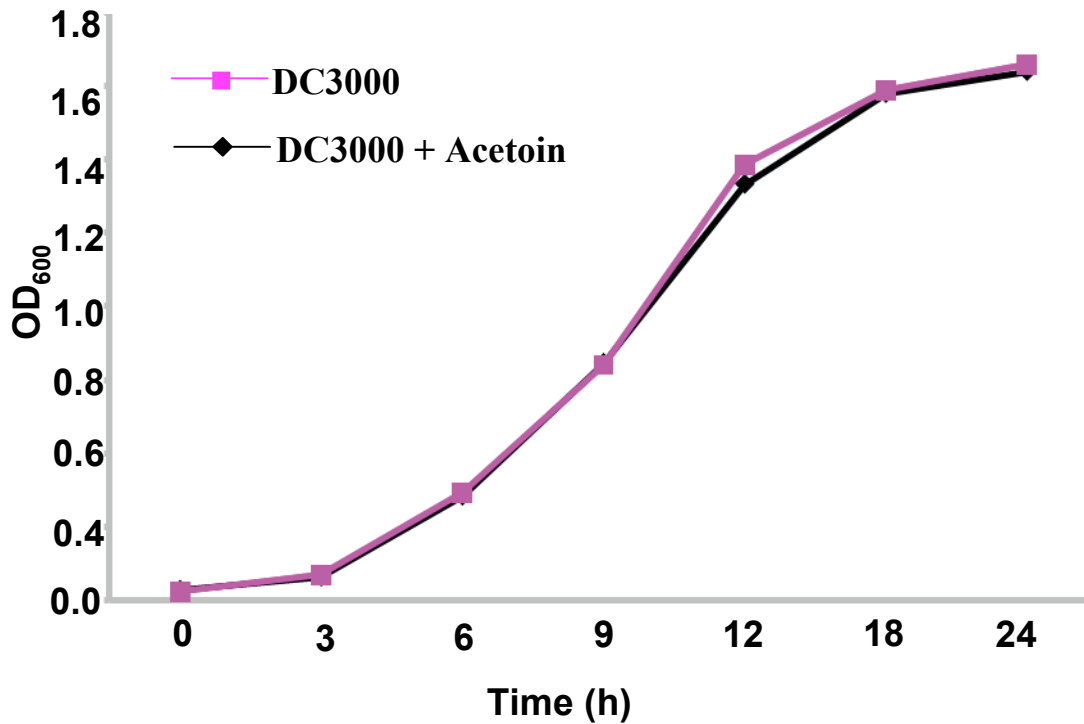
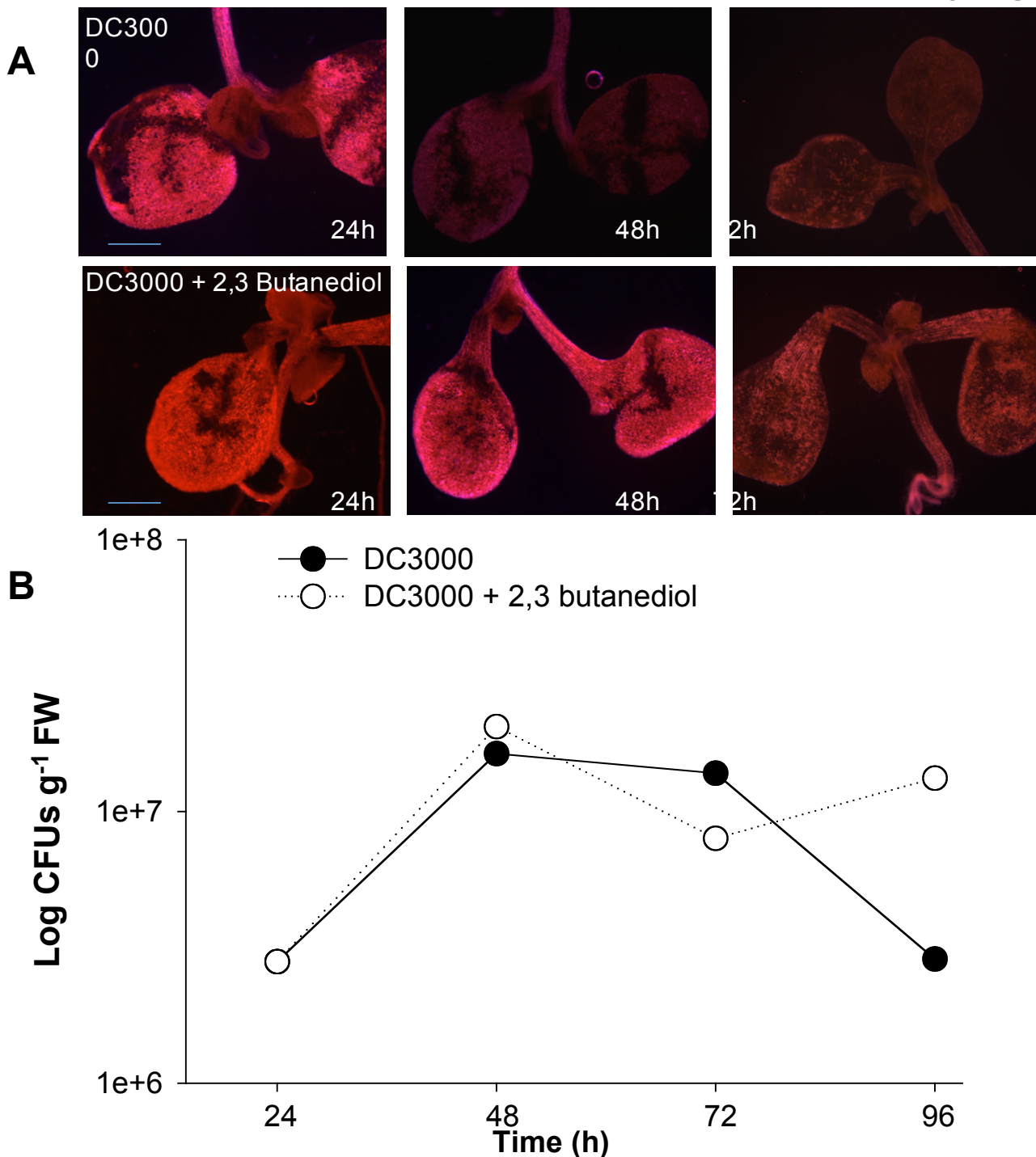


## Supplementary Figure 1



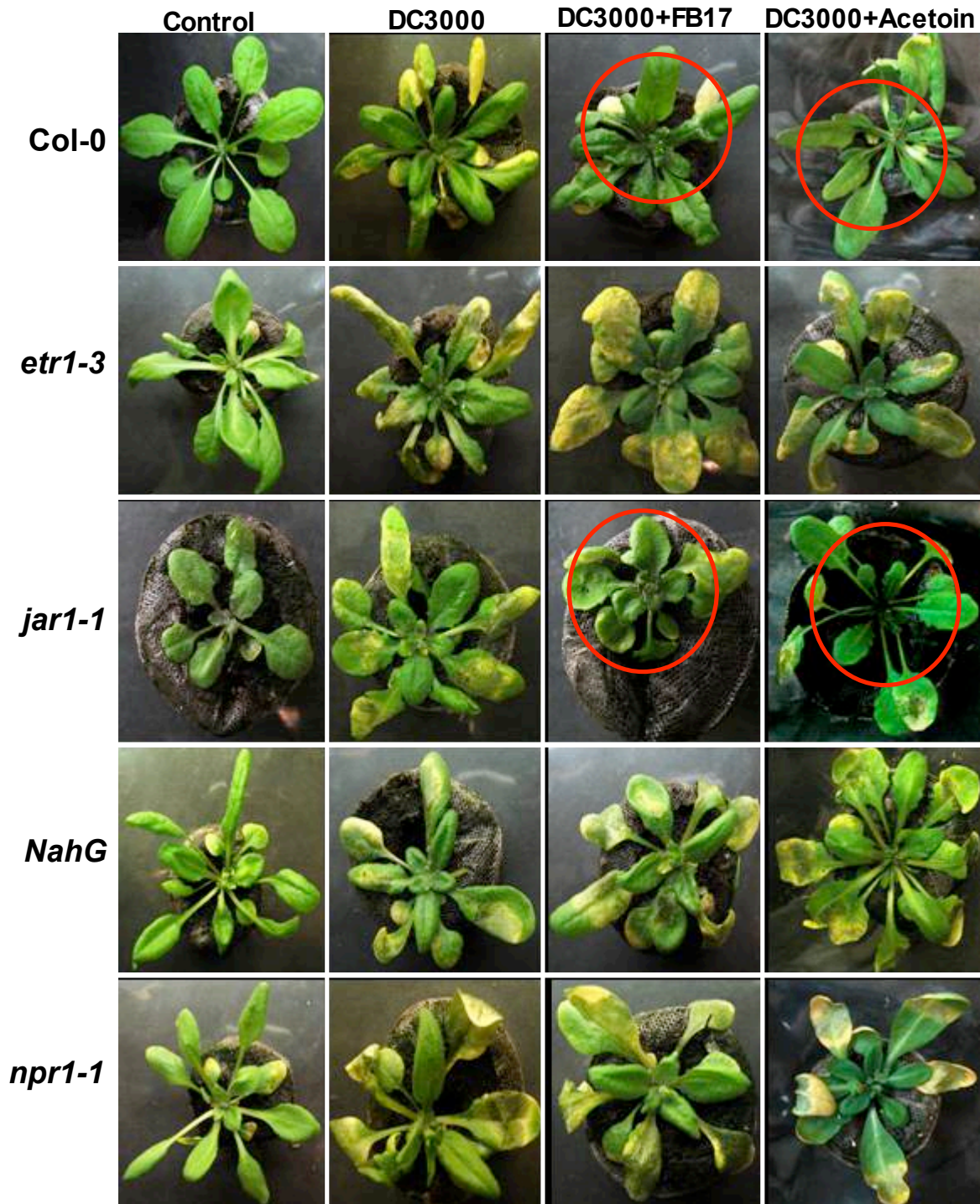
**Supplementary Figure 1.** Broth micro-dilution assay with acetoin (1ml of 10mM stock ~ 88 $\mu$ g) against *P. syringae* DC3000. An overnight culture of DC3000 (0.02 OD<sub>600</sub>) supplemented with acetoin was initiated; samples collected at regular time periods were checked for growth at OD<sub>600</sub> for 24hrs.

## Supplementary Figure 2



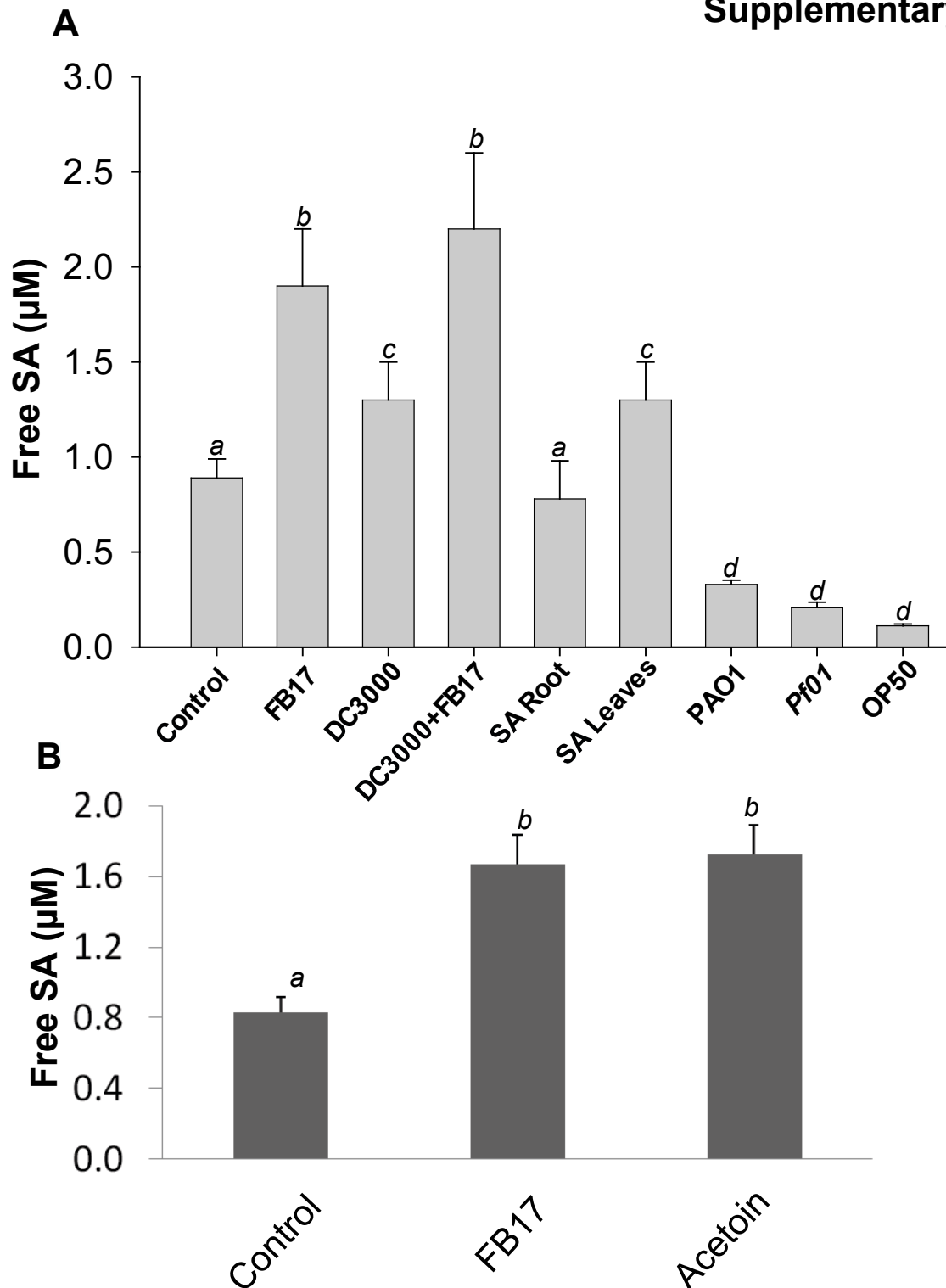
**Supplementary Figure 2.** A) *P. syringae* DC3000 infection symptoms in 2,3 butanediol treated and untreated Col-0 plants. 2,3 Butanediol was supplemented in the seedlings as a volatile, where in a 10 mM stock was prepared by weighing appropriately and dissolving in a known volume of distilled water. 1 ml from this stock solution was used for in vitro DC3000 assays. A bright field microscopy of whole-mount seedlings, illustrating the bleaching phenotype used previously (Schreiber et al. 2008) was used to monitor the disease progression and DC3000 infection. Progression of chlorophyll degradation in infected cotyledons. (red, chlorophyll fluorescence). Note the nearly complete loss of chlorophyll in 72h treatment in both DC3000 and DC3000 + 2,3 butanediol treatments. The images are a representative example of n=20 and the data is an average of two separate experiments each with twenty replicates. Scale bars= 1cm. Effect of 2,3 butanediol on pathogen proliferation (B) in the seedlings of *A. thaliana* Col-0 plants inoculated with DC3000. The numbers on x - axis represent time (in hours) post inoculation with DC3000.

### Supplementary Figure 3



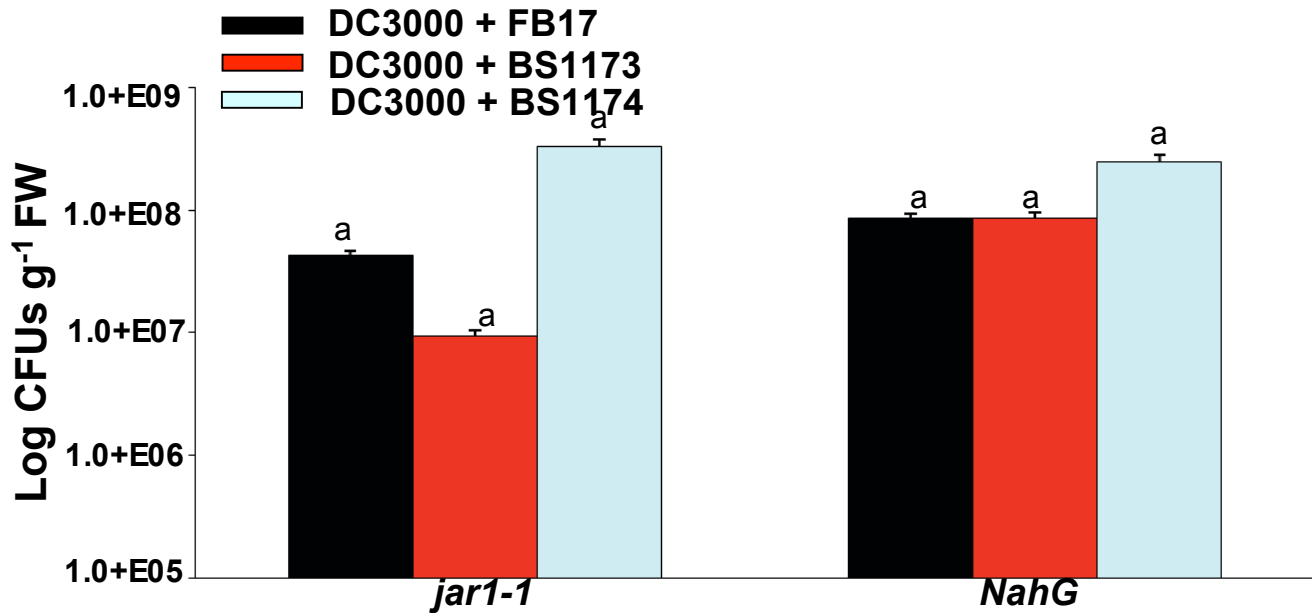
**Supplementary Figure 3.** DC3000 infection symptoms in wild type and disease compromised mutants of *A. thaliana* root inoculated with FB17 and treated with acetoin. Plants were either untreated or root inoculated with FB17 before leaf infection with DC3000. The images are a representative sample of n=6 and the data is an average of two separate experiments with six replicates. The circles in the inset shows induced resistance with FB17 and acetoin treated wild type and *jar1-1* plants.

## Supplementary Figure 4



**Supplementary Figure 4.** Free SA levels in the leaves of Col-0 plants treated with FB17, DC3000 and FB17+DC3000 (A). Free SA levels were also estimated in plants exposed to other rhizobacteria such as *Pfo-1*, PAO1 and OP50. The data shows significantly higher SA in the leaves of FB17 treated plants. The data is an average of six replicates of two experiments conducted separately (Mean  $\pm$  S.D; n=6). Different letters on the bars indicate a statistically significant difference between treatments ( $F_{(8,55)}=175.2$ ,  $P<0.05$ ). (B) Free SA levels in the leaves of Col-0 plants treated with FB17 and acetoin. The data shows significantly higher SA in the leaves of FB17 and acetoin treated plants compared to untreated control ( $F_{(2,19)}=131.2$ ,  $P<0.05$ , ANOVA). The data is an average of six replicates of two experiments conducted separately and the images are a representative of six replicates. Different letters on the bars indicate a statistically significant difference.

## Supplementary Figure 5



**Supplementary Figure 5.** Effect of *B. subtilis* acetoin biosynthetic mutants (BS1173 and BS1174) on leaf colony titer response of DC3000. Twenty one days old wild type and disease compromised mutants were root inoculated with FB17 and the leaves were infected with DC3000. The infected plant leaves were used post 96 hr of treatment to enumerate total leaf CFU counts for DC3000. The data is an average of six replicates of two experiments conducted (Mean  $\pm$  S.D; n=6). Different letters on the bars indicate a statistically significant difference ( $F_{(2, 29)}=231.2, P\leq 0.05$ , ANOVA) .