

SUPPORTING INFORMATION APPENDIX FOR

**THE END-DEVONIAN EXTINCTION AND A BOTTLENECK IN THE EARLY
EVOLUTION OF MODERN JAWED VERTEBRATES**

LAUREN COLE SALLAN^a and MICHAEL I. COATES^{a,b}

^aDepartment of Organismal Biology and Anatomy, University of Chicago, 1027 E. 57th
Street, Chicago, IL 60637

^bCommittee on Evolutionary Biology, University of Chicago, 1025 E. 57th Street,
Chicago, IL 60637

Corresponding Author: Lauren Cole Sallan, lsallan@uchicago.edu. Phone: (954) 895-
9192, Fax: (773) 702-0037

Table of Contents.

I. Detailed Methods	9
A. Dataset Assembly Procedures.	9
B. Multivariate Ecological Methods.	11
II. Detailed Results	14
A. Total Faunal Dataset.	14
1. Cluster Analysis.	14
2. Canonical Correspondence Analysis (CCA).	14
3. Nonparametric Multidimensional Scaling (NMDS).	15
4. Factor Analysis.	15
5. Analysis of Similarity (ANOSIM).	16
6. Similarity Percentage (SIMPER).	17
B. Extinction Intervals: Marine and Non-marine Environments.	18
1. NMDS.	18
2. Factor Analysis.	18
3. ANOSIM.	19
4. SIMPER.	20
C. Environmental Intervals: Restricted Timespans.	20
1. NMDS.	20
2. Factor Analysis.	21
3. ANOSIM.	22
4. SIMPER.	22

III. Site Descriptions and Faunal Lists	24
A. Givetian Sites	24
1. Orcadian Lake	24
2. Harma	24
3. Gerolstein	24
4. Rockport	25
5. Aztec	25
6. Haikou	25
7. The Cletts	26
8. Mt. Howitt	26
9. Abava	26
10. Cedar Valley	26
11. Gauja	27
B. Frasnian Sites	27
12. Gladbach	27
13. Boghole	27
14. Gogo	28
15. Chahriseh	28
16. Kerman	28
17. North Evans	29
18. Miguasha	29
19. Wietrznia	29
20. Rodebjerg	30

21. Rhinestreet	30
22. Chemung	30
23. Wildungen	31
24. Scaat Craig	31
25. Stolbovo	31
C. Famennian Sites	32
26. Canowindra	32
27. Rosebrae	32
28. Dura Den	32
29. <i>Cheiloceras</i> Beds	32
30. Tervete	33
31. Oryol	33
32. Portishead	33
33. Aina Dal	34
34. Britta Dal	34
35. Grenfell	34
36. Hyner	34
37. Evieux	35
38. Ketleri	35
39. Tafilalt	35
40. Andreyevka-2	36
41. Cleveland Shale	36
42. Chaffee	36

43. Witpoort	37
D. Tournaisian Sites	37
44. Horton Bluff	37
45. Izkchul Village	37
46. Mansfield	38
47. Foulden	38
48. Herbesskaya	38
E. Viséan Sites	38
49. Waiipoort	38
50. Glencartholm	39
51. Burdiehouse	39
52. Ducabrook	39
53. Wardie	40
54. Cheese Bay	40
55. East Kirkton	40
56. Gilmerton	41
57. Ardross Castle	41
58. Abden	41
59. Inchkeith	41
F. Serpukhovian Sites	42
60. Bearsden	42
61. Chokier	42
62. Loanhead	42

63. Niddrie	43
64. Millstone Grit	43
65. Dora	43
66. Bear Gulch	43
IV. Supporting Figures	45
Figure S1. Paleogeography of sampled vertebrate sites.	45
Figure S2. Histogram of site species-level diversity.	46
Figure S3. Cluster dendrogram: Bray-Curtis, raw diversity.	47
Figure S4. Cluster dendrogram: Bray-Curtis, relative diversity.	48
Figure S5. Cluster dendrogram: Kulczynski similarity.	49
Figure S6. CCA of relative diversity.	50
Figure S7. NMDS plots of all sites.	51
Figure S8. NMDS plots of restricted intervals: Bray-Curtis, raw diversity.	52
Figure S9. NMDS plots of restricted intervals: Bray-Curtis, rel. div.	53
Figure S10. NMDS plots of restricted intervals: Kulczynski.	54
Figure S11. Factor analyses of restricted intervals.	55
Figure S12. Ordination plots of end-Famennian and Tournaisian sites.	56
V. Supporting Tables	57
Table S1. Raw diversity data.	57
Table S2. Relative diversity data.	57
Table S3. Genus-level gnathostome occurrences.	57
Table S4. Descriptive statistics for species counts.	58
Table S5. CCA of raw diversity: site scores.	59

Table S6. CCA of raw diversity: taxon scores.	60
Table S7. CCA of relative diversity: site scores.	61
Table S8. CCA of relative diversity: taxon scores.	62
Table S9. Factor analysis of all sites: taxon loadings.	63
Table S10. Factor analysis of all sites: site scores.	64
Table S11. ANOSIM R statistics for raw diversity: extinction intervals.	66
Table S12. ANOSIM p-values for raw diversity: extinction intervals.	67
Table S13. ANOSIM R statistics for raw diversity: other intervals.	68
Table S14. ANOSIM p-values for raw diversity: other intervals.	69
Table S15. ANOSIM R statistics for rel. diversity: extinction intervals.	70
Table S16. ANOSIM p-values for rel. diversity: extinction intervals.	71
Table S17. ANOSIM R statistics for rel. diversity: other intervals.	72
Table S18. ANOSIM p-values for rel. diversity: other intervals.	73
Table S19. SIMPER average dissimilarities.	74
Table S20. SIMPER for all Frasnian and Famennian sites.	74
Table S21. SIMPER for all Famennian and Tournaisian sites.	75
Table S22. Factor analysis of Fra.-Fam. sites: taxon loadings.	76
Table S23. Factor analysis of Fra.-Fam. sites: site scores.	77
Table S24. Factor analysis of Fam.-Miss. sites: taxon loadings.	78
Table S25. Factor analysis of Fam.-Miss. sites: site scores.	79
Table S26. SIMPER for Fra.-Fam. marine sites.	80
Table S27. SIMPER for Fra.-Fam. non-marine sites.	81
Table S28. SIMPER for Fam.-Miss. marine sites.	82

Table S29. SIMPER for Fam.-Miss. non-marine sites.	83
Table S30. Factor analysis of late Fra-early Fam. sites.: taxon loadings.	84
Table S31. Factor analysis of late Fra-early Fam. sites.: site scores.	84
Table S32. Factor analysis of late Fam.-Tour. sites: taxon loadings.	85
Table S33. Factor analysis of late Fam.-Tour. sites.: site scores.	85
Table S34. Factor analysis of end-Fam.-Tour. sites: taxon loadings.	86
Table S35. Factor analysis of end-Fam.-Tour. sites: site scores.	86
Table S36. SIMPER for late Frasnian and early Famennian sites.	87
Table S37. SIMPER for late Famennian and Tournaisian sites.	88
Table S38. SIMPER for end-Famennian and Tournaisian sites.	89
VI. References	90

I. Detailed Methods

A. Dataset Assembly Procedures

Diversity curves presented in this study are based on data from new comprehensive datasets of genus-level and species-level gnathostome vertebrate occurrences from the Givetian to Serpukhovian stages (391-318 million years ago) of the Devonian and Mississippian periods (Tables S1, S2, S3). This compilation was undertaken to address deficiencies in vertebrate sampling in both Sepkoski's marine dataset (1) and the Paleobiology Database – including lack of non-marine taxa (representing much of known vertebrate material) and undersampling of vertebrate museum collections. Agnathan occurrences are presently excluded due to the relative scarcity of macrofossil material in both the late Devonian and post-Devonian deposits. In the case of conodonts, there is an additional lack of relevant assemblage information and large taxonomic uncertainty related to element assignment. There is also strong indication that they tended to practice rapid within-group replacement after extinction intervals – obscuring any signal in analyses binned by stage or at higher taxonomic levels (1, 2, 3). However, it should be noted that there is recent evidence for acute extinction among conodonts during the Hangenberg event (3). Gnathostome occurrence data was taken from variety of sources in the literature including species descriptions and faunal lists. Genus-level diversity curves were generated from data binned by stage for each of the six traditional gnathostome divisions: Placodermi, Acanthodii, Actinopterygii, limbed Tetrapoda (including elpistostegalians), finned Sarcopterygii, and Chondrichthyes (Fig. 1 and Table S3).

Due to the high susceptibility of diversity curve analyses to sources of sampling bias, including (but not limited to) stage length, Lagerstätten, number of localities (4, 5), the paraphyly of some involved groups, and the limited temporal ranges of most vertebrate species and even genera, the occurrence data were subsampled to produce site-specific species lists for use in ecological analyses (Tables S1 and S2). Only macrofossil sites containing at least five named species in three groups in two divisions were eligible for inclusion in this smaller dataset. In addition, an attempt was made to sample each distinct fauna only once, resulting in the consolidation of taxa from multiple temporally concurrent deposits in the same environment into a single sample of the entire ecosystem (e.g. the Orcadian Lake). This culling of the dataset was performed in order to further circumvent the production of false signal due to poor sampling, unrepresentative preservation, and the taxonomic issues common to microfossil localities.

The aforementioned procedures produced a list of 66 eligible sites, including 11 sites from the Givetian, 14 from the Frasnian, 18 from the Famennian, 5 from the Tournaisian, 11 from the Viséan, and 7 from the Serpukhovian. Localities are referred to by their most common name, usually that of a nearby settlement or attraction (e.g. East Kirkton, Bearsden). The name of the formation or other geological trait is used when collections from multiple contemporaneous outcrops contain similar assemblages (e.g. Cleveland Shale), temporally, environmentally, and/or compositionally distinct assemblages are known from a single site (e.g. Rodebjerg, Aina Dal, and Britta Dal in East Greenland), or when no other name is available (e.g. Herbesskaya, Aztec).

Sites are widespread geographically (Fig. S1, Table S1) - although there is an overrepresentation of Euramerican ecosystems for historical reasons (e.g. proximity to workers and financial means) (6). However, this bias is consistent throughout the dataset (7), and visualization procedures, as explained below, should be able to differentiate between turnover signal stemming from geographic sampling and real faunal change. The chosen localities are all primarily aquatic, representing a variety of environments both marine and non-marine (defined here as having any freshwater influence, including marginal marine, brackish and estuarine localities). A bias towards the non-marine was noted for every stage except the Frasnian (Fig. S1 and Table S1). Site diversity ranges from 6 to 82 species, with a mean species count of 19.27 (SD: 13.01) for all sites, and a median of 15.00 (Table S4) – producing a left-skewed distribution with a long right tail representing Lagerstätten (Fig. S2). Analyses were run using both raw species counts and as well as counts converted to relative percentage of the overall faunal diversity. The latter was done in order to check sample size bias without the loss of signal expected to attend the exclusion of lagerstätten. Shannon-Weiner index values – the negative of the sum of the proportion of samples containing a species multiplied by the natural log of that proportion (8) – were calculated to measure the diversity at all sites (Table S4) along with taxonomic richness, which is a merely a count of occupied bins.

As demanded by the ecological analyses performed here, the sub-sampled data was arranged into matrix in which sites served as samples and species counts in taxonomic bins acting as variables (5). These bins represent major gnathostome clades and/or well-differentiated groups extant by the Frasnian. This binning scheme is preferable to the use of single taxonomic levels (such as families, genera, species) due to the high disagreement between classification schemes and usage among gnathostome groups, localities, and even workers. The validity of some well-known vertebrate divisions (e. g. Acanthodii, Placodermi) has recently come under fire due to potential paraphyly (9). While their higher-level standing is problematic, many of their lower-level taxonomic groups might be phylogenetically valid and monophyletic (e.g. Antiarcha and Arthrodira in Placodermi; Gyracanthida and Acanthodida in Acanthodii). Many of the remaining taxonomically problematic groups show enough morphological and/or ecological distinction in the Frasnian to warrant treatment as separate entities capable of responding independently to extinction or turnover. In addition, most of the groups diverged before the first event of interest. Use of these groups represents a conservative approach to measuring extinction – pseudoextinction is avoided but extinction with prompt replacement within divisions goes undetected. Only a high magnitude extinction event involving mostly monophyletic groups will show up in the results.

It should be noted here that it is common in ecological studies to use abundances rather than taxon counts, so as to prevent undue contribution from rare taxa (5). Unfortunately, few vertebrate sites have been subjected to abundance surveys (10), even though there is anecdotal evidence for the composition of many of them. The most likely effect of using diversity is lower sensitivity to real turnover – something that might be preferable when examining extinction events. For methods using presence-absence distance measures, the outcome should be identical.

B. Multivariate Ecological Methods and Outcomes

The faunal composition data was characterized and analyzed using the general statistics program R (version 2.8.1) (11), the ecological package vegan (version 1.15) (12), and the paleobiological statistics program PAST (version 2.98) (13). Ecological analyses used in this study included cluster analysis, canonical correspondence analysis (CCA), detrended correspondence analysis (DCA), factor analysis (FA), non-parametric multidimensional scaling (NMDS), one-tailed analysis of similarity (ANOSIM), and similarity percentage (SIMPER). The first five methods are ordination procedures that produce plots allowing visualization of the data. These procedures are useful for detecting true faunal breaks associated with mass extinction since they do not depend on *a priori* group assignment whether temporal or geographic. Despite commonalities, all the methods characterize different aspects of the data, and so their results can complement and support one another. Cluster analysis is useful for determining the degree and structure of relationships between individual faunas, while correspondence analysis reveals any major environmental and/or temporal gradients underlying the distribution of both sites and taxa, potentially revealing major assemblages. Detrended correspondence analysis corrects the latter for sample size if necessary and linearizes the data. Factor analysis reveals the most important taxa driving separation between different clusters of localities. NMDS is ordination along two axes based on distance metrics, and so can serve as the visual representation of statistical tests predicated on the same (such as ANOSIM). ANOSIM is a statistical permutation test of faunal similarities between *a priori groups* (unlike the previous methods), and SIMPER reveals the relative contribution of the changes to diversity of individual taxa to differentiation between those assigned groups. These methods will be explained in more detail below.

In order to control for sampling, analyses were performed using raw diversity, (with results presented in the text) and relative diversity in order to reveal any effects of sample size (Tables S1 and S2), as well as temporally restricted subsets containing Frasnian, Famennian, and Mississippian sites marked by environment (Table S1). The Tournaisian and Viséan were grouped together in order to correct for a lack of marine sites in the Tournaisian that would reduce statistical power. Since there was no significant difference between Tournaisian and Viséan sites in the overall analyses (Tables S12 and S16), this was unlikely to skew the results. A subset containing late Frasnian and early Famennian sites was analysed to further pinpoint the interval of faunal change (Fig. 2). Additional matrices comprising late Famennian and Tournaisian sites, and end-Famennian (those sites conformably overlain by Hangenberg and/or boundary deposits) and Tournaisian sites were analysed for the same reason (Fig. 2)

Some of the methods (cluster analysis, NMDS, and ANOSIM) require distance or similarity matrices. Distance and similarity metrics used here include: 1. Bray-Curtis distance (14), an abundance-sensitive distance measure representing the sum of absolute differences between sites divided by the sum of actual differences between sites (5); 2. Kulczynski similarity (15), a presence-absence measure modified from Jaccard similarity that groups subsets of faunas with larger biotas containing the same species; 3. Dice similarity (16), a presence-absence measure which puts greater weight on joint

occurrences of taxa, and is calculated from twice the number of matches divided by the sum of twice the number of matches plus the number of dissimilar taxa; 4. Simpson similarity (17), a presence-absence measure which treats subsets of taxa as identical to large samples and is produced by the number of taxon matches divided by the minimum number of presences among two samples; 5. Chord distance (17) an abundance distance measure taken from relative species abundances (normalized to 1) rather than absolute numbers; 6. Morisita similarity (18), an abundance measure recommended by the authors of PAST (17); 7. Raup-Crick index (19), a presence-absence similarity measure which uses randomized pooled data from two sites to produce a distribution of co-occurrences and then compares these to the observed data.

The metrics are all based on different assumptions, and have different sensitivities to faunal differentiation. Using all of them will give information about the structure of the data, potentially avoiding errors. Agreement between such different measures is likely to provide especially compelling evidence of either extinction-related turnover or stasis. Presence-absence metrics do not take into account relative numbers of species, producing results based on faunal composition alone. They are therefore particularly useful for testing whether observed patterns of turnover are due to extinction rather than unrelated diversity changes. Use of these measures also corrects for the effects of oversampling of, or increased taxonomic work on, certain groups at a single locality.

Cluster analysis produces a hierarchical dendrogram of sites based on a distance or similarity matrix, with branching based on a chosen algorithm without *a priori* grouping by either age or environment (5). Dendrograms were produced using the *hclust* function in R based on average linkage clustering (UPGMA) (20), which averages all possible distances between groups. Clustering was performed for both raw and relative diversity datasets using both Bray-Curtis distances (“abundances” – actually species diversity here) and Kulczynski similarities (presence-absence) generated by using the *vegdist* function of the *vegan* package. Data was first square root transformed to correct for the effects of oversampled taxa. The cophenetic correlation, calculated in PAST, shows the relationship between true distances and those represented in the dendrogram. Sudden extinction and turnover is expected to show a large split between pre- and post-event faunas, while stasis and gradual change will result in sites from surrounding temporal intervals clustering together based on some other factor.

Canonical correspondence analysis (CCA) is an indirect ordination procedure that detects environmental (or temporal) gradients and distribution of sites from taxonomic composition without *a priori* group assignment. CCA visualizes the gradients as axes in a two-dimensional ordination plot of both taxa and sites (5, 21), revealing the correspondence between them, as well as any faunal associations. CCA plots were generated from the both the raw (Fig. 3) and relative diversity datasets (Fig. S6) using the *cca* function in the R package *vegan*. Detrended correspondence analysis is a related and popular technique that corrects for correlations between the first and second axes which often produce a horseshoe shaped distortion of the ordinations (5). Since primary gradient produced by CCA was temporal, while the second was environmental, there was little interplay between the two and DCA (generated by the *decorana* function in the R package *vegan*) proved an unnecessary step – turning out similar, but more condensed, plots. Mass extinction and turnover is assumed when a large temporal gap is observed between environmentally similar faunas. Gradual change or stasis is implied when sites

form gradient along both correspondence axes and show overlap by environment or geography.

Factor analysis (FA) is a procedure similar to principal components analysis, in which a variance-covariance matrix is used to produce a visual ordination of samples (5). However, in FA, the axes are rotated to better explain the differentiation between clusters of samples that might otherwise fall diagonally (5). This technique was used by Sepkoski (22) to characterize his evolutionary faunas, and is another method for visualizing the main differences in taxon composition between sites. Factor analysis was performed on PAST using varimax rotation, which scales the loadings to maximize sum of variance. Taxon loadings are produced for four axes. Large-scale differences between pre- and post-extinction faunas are expected to result in their placement primarily along distinct axes representing large amounts of variance. Stasis or gradual change is likely when faunas are ordinated along a single axis, or form a diagonal gradient.

NMDS is a visualization method which ordinated samples into a two-dimensional space while retaining the ranked distances found in an original multidimensional distance matrix data (5). It is often used for non-parametric data such as faunal distributions in place of principal components analysis (5). NMDS plots were produced using PAST, which chooses the best solution of 11 trials based on least “stress” – a value representing the difference between ranks within the distance matrix and in the ordination plot (5). PAST additionally generates a Shepard plot of obtained versus targeted ranks as visual representation of fit. NMDS plots were generated for all raw and relative diversity datasets based on both Bray-Curtis and Kulczynski distances. NMDS results are expected to appear similar to those of CCA depending on the patterns present in the data.

One-tailed ANOSIM is a non-parametric method similar to NMDS in that it depends on ranked distances. However, this is a statistical permutation test, a procedure much like bootstrapping which compares *a priori* group composition (5, 23). ANOSIM obtains these values by comparing within-in group and between-group ranks, under the assumption that the later must be larger than the former if two faunas are truly different. This generates the test statistic R, which can range from 1 (absolute dissimilarity) to 0 and even negative numbers (high similarity). Permutations of samples within groups are then used to determine the significance of differences between the groups themselves by producing pairwise p-values. As a non-parametric test, ANOSIM is a popular method for detecting significant differences in the faunal composition of modern and fossil ecosystems. ANOSIM p-values for all datasets were obtained for all seven distance measures using 1000000 permutations in PAST. Mean R statistics and p-values for raw diversity using all metrics are presented in the text. Means for all metrics, presence-absence, abundance, and individual metrics based on both raw and relative diversity data are additionally presented in the supporting tables S11-S18 for both extinction and selected background interval. Global mass extinction is expected to produce statistically significant ANOSIM p-values and high R statistics in pairwise comparisons of all environments and intervals around such an event using both kinds of metrics and diversity data. Stasis is expected to result in insignificant ANOSIM p-values and low R-statistics. Gradual and even sudden diversity changes without extinction are expected to produce insignificant p-values and low R in ANOSIM tests based on presence-absence metrics.

The final method used here, SIMPER, determines the relative contribution of taxa to dissimilarity between groups based on Bray-Curtis distances, as well as average dissimilarity (5). In other words, SIMPER reveals faunal patterns underlying turnover detected by ANOSIM, showing which groups underwent changes. SIMPER analysis was performed in PAST on both the restricted relative diversity datasets in order to determine the changes in faunal composition driving differentiation independent of environment. The main downside to this method is that the use of percentages might exaggerate the impact of rare taxa, while focusing on a narrow environment might obscure the signal for euryhaline species. However, it is still a useful technique for exploring ANOSIM results. In addition, the average dissimilarity metric can be used to relate SIMPER results from different comparisons.

II. Detailed Results

A. Total Faunal Dataset.

1. Cluster Analysis

The most basal division in all cluster dendrograms, whether based on abundance (Bray-Curtis) or presence-absence (Kulczynski) metrics or on raw or relative diversity, is between all Devonian sites and all Mississippian sites (Figs. S3, S4, S5). This split corresponds to the Hangenberg event at the Devonian-Mississippian boundary. A secondary deep division is observed among the Mississippian sites, falling between a cluster of majority marine sites and a cluster of mostly nonmarine localities. A similar environmental division is observed among the overall Mississippian cluster in dendrograms based on relative diversity and Kulczynski distances (Figs S4 and S5). Otherwise, sites mostly cluster by geography (e.g. Euramerican sites, Fig S1) or finer environmental designation in the (e.g. reef sites such as Gogo, Wildungen, and Wietrzna) (Figs. S3, S4, S5, Table S1). Few clusters of more than two localities, and none of more than four, belong to a single Mississippian or Devonian stage. Therefore, the dendrograms give little indication of any widespread faunal turnover around boundaries or extinction events apart from the Hangenberg.

2. Canonical Correspondence Analysis (CCA)

The correspondence axes generated for both raw and relative diversity are the same. Correspondence axis 1 (raw diversity eigenvalue=0.46, relative diversity eigenvalue=0.47) is purely temporal and does not represent a gradient so much as a split between the Devonian and Mississippian (Fig. 3A and S6A, Tables S5 and S7). Clusters of sites from all stages within the periods show extensive overlap, including those from around the Kellwasser extinction. There is a large gap between clusters containing all Devonian and all Mississippian localities along this axis. This shows deep dissimilarity between faunas before and after the end-Devonian Hangenberg extinction event, as was previously observed in the cluster dendrograms. A similar split is observed between taxa associated with sites from pre- and post-Hangenberg intervals, suggesting that acute faunal turnover as the cause of this pattern (Figs. 3B and S6B, Tables S6 and S8).

Correspondence axis 2 (eigenvalue=0.27) is associated with a salinity gradient evident in sites from both periods, with marine sites having more positive values (Figs. 3A and S5A, Tables S5 and S7). However, post-Hangenberg Tournaisian sites cluster tightly around the origin despite a worldwide geographic spread and differences in reported environment (Fig. S1 and Table S1). The spread of taxa along axis 2 shows that designations based on geological data can be corroborated by vertebrate faunal composition (Figs. 3B and S6B, Tables S6 and S8). On the basis of these results, gnathostome assemblages might be used to determine the depositional environments of fossil sites from this interval in the absence of geological study. In addition, this data could be used to hypothesize the vertebrate faunas of new later Devonian and Mississippian localities with known environments.

3. Nonparametric Multidimensional Scaling (NMDS)

The first NMDS coordinate generated from both raw and relative data is associated with both time and to a lesser extent environment: Devonian and nonmarine sites have lower values than Mississippian and marine sites (Figs. 4A and S7). The temporal signal is much stronger; overlap between Mississippian and Devonian sites along the first coordinate is confined to Serpukhovian terrestrial freshwater sites (Dora, Niddrie) and late Famennian open marine deposits (Cleveland Shale, Tiflalt) in raw data analyses (Figs. 4A, S7A, S7B). This is likely due to underreporting of certain non-tetrapod taxonomic groups, such as actinopterygians, at those two sites, a situation common at Late Viséan and Serpukhovian terrestrial localities (e.g. Niddrie, Ducabrook, and previously East Kirkton). In fact, Dora is clustered with East Kirkton and other non-outliers in the cluster dendrograms (Figs. S3, S4, S5) and Niddrie lies well within a cluster of Tournaisian and other non-marine tetrapod site in the dendrogram based only on presence-absence of groups (Fig S5). This suggests that these sites should contain more actinopterygians and other typical Mississippian taxa. However, if this overlap is real, it either shows convergence (excluding extinct groups) between the faunas of two very different environments after a gap of 40 My, the existence of a marine refugium repopulating inland environments after the Hangenberg, and/or radical habitat shifts in the aftermath of extinction. However, it should be noted that there is no overlap along the first coordinate produced by NMDS ordination based on relative diversity (Figs. S7C and S7D). As in the CCA plots (Figs. 3 and S6), the second coordinate is strictly environmental – marine sites have more positive values.

4. Factor Analysis

Varimax factor analysis of all sites produces identical results from both raw and relative diversity data. Factor 1 (39.68% of variance) is positively associated Devonian non-marine taxa as designated by the CCA plots (Fig. 4B and Table S9). Factor 2 (22.10% of variance) is negatively correlated with Mississippian forms, largely actinopt and chondrichthyans (Table S9). Factor 3 (9.59% of variance) is negatively loaded by Devonian marine taxa, while Factor 4 (5.45% of variance) is positively associated with the same (Table S9). Overall, position along the main axes of variation are correlated with taxa abundant in either pre- or post-Hangenberg sites. This again illustrates the

extreme differentiation of faunal composition of all environments around the Hangenberg event, but not the earlier Kellwasser extinction. The only overlap between pre and post-Hangenberg intervals on the non-environmental Mississippian axis (Factor 2) concerns the same two outlier terrestrial faunas seen on the first NMDS coordinate, and is likely caused the same factors (Fig. 4B and Table S10). All overlap on the other major factors is because Devonian environmental biotas (marine and non-marine) are distinct enough to be highly correlated with separate axes of variation (Tables S9 and S10). This causes contemporary sites representing the opposite environment to fall near the origin, alongside Mississippian localities. High correlation of the first factor with non-marine Devonian sites to exclusion of Mississippian non-marine faunas precludes a role for freshwater refugia in the Hangenberg event.

5. Analysis of Similarity (ANOSIM)

ANOSIM reveals a highly significant difference ($\alpha=0.05$) between Devonian and Mississippian faunal composition, corresponding to the Hangenberg event (Tables S11-S18). In pairwise comparisons of various pre- and post- extinction stages, including the Famennian and Tournaisian, mean R statistics range from 0.74 to 0.81 and mean p-values range from 0* to <0.001* (Tables S11-S18). The R and p-values generated from tests based on different similarity metrics, including those based on abundance and presence-absence, are very consistent - the turnover signal is strong enough to be immune to minor methodological differences (Tables S11-S18). Crucially, this agreement indicates that faunal change at the Hangenberg was primarily due to whole-taxon extinction and basic compositional change rather than relative fluctuations in the diversity of co-existing groups.

In contrast, the ANOSIM results for the Kellwasser extinction are mixed, with a bias towards insignificance (Tables S11-S18). Pairwise p-values for the Frasnian and Famennian based on abundance metrics show a significant difference between faunas before and after the event, despite low R-statistics indicating high faunal similarity (raw diversity abundance mean R=0.12, mean p=0.03*; relative diversity abundance mean R=0.13, mean p=0.03*) (Tables S11, S12, S15, S16). However, no significance is observed in tests based on presence-absence metrics, which have even lower R-statistics (raw diversity presence-absence mean R=0.07, p=0.08; relative diversity presence-absence mean R=0.08, p=0.08) (Tables S11, S12, S15, S16). This suggests that any faunal differentiation over the Frasnian-Famennian is due to relative changes in diversity, rather than turnover related to a first-order mass extinction. In addition, there is no significant difference in abundances in tests comparing the Givetian and Famennian, despite the longer interval and the inclusion of other geological events (raw diversity abundance mean R=0.12, mean p=0.05; relative diversity abundance mean R=0.10, p=0.08) (Tables S13, S14, S17, S18). The Frasnian-Famennian extinction has previously been considered an artefact of sampling, and the mixed signal observed here supports that claim. However, comparison of whole stages says nothing about the temporal spread, environmental extent, or the exact diversity changes underlying the abundance signal. These were examined in analyses of subsets of the data, results of which are presented below.

No significant difference was observed between faunas across other boundaries. This includes those thought to be associated with lesser extinction events, such as the Taghanic at the Frasnian-Givetian boundary (raw diversity mean $R=-0.02$, mean $p=0.6$; relative diversity mean $R=-0.03$, mean $p=0.6$) and the unnamed event of the early Serpukhovian (raw and relative diversity mean $R=0.05$, mean $p=0.3$) (Tables S13, S14, S17, S18). For both events and over Tournaisian-Viséan boundary, results from abundance metrics showed greater faunal differentiation than those based on presence-absence measures – the same signal observed for the Frasnian-Famennian (Tables S11, S12, S15, S16).

ANOSIM results suggest that changes in relative diversity of coexisting taxa are common over long intervals, but extinction and replacement of higher-level monophyletic or morphologically similar groups between two intervals is more rare. Therefore, the results generated for the Frasnian-Famennian are not exceptional compared to background. In contrast, the R -values generated from presence-absence metrics for the Hangenberg event are orders of magnitude larger than those for any other extinction or boundary examined here, while the p -values are orders of magnitude smaller. This shows the Devonian-Mississippian extinction to be a truly anomalous event in the mid-late Paleozoic.

6. Similarity Percentage (SIMPER)

SIMPER average dissimilarity for the Frasnian-Famennian is close to that for Givetian-Frasnian, and so is indistinguishable from background turnover (Table S19). It does appear from SIMPER comparison of the whole stages that arthrodires lose half their diversity over the Kellwasser event, along with ptyctodonts, actinopterygians, climatiids, and other taxa designated as marine or euryhaline in the CCA results (Fig. 3B and S5B), although none go completely extinct (Table S20). This might suggest a marine-restricted turnover event. However, antiarchs more than double, as do the other Devonian non-marine taxa as sorted by the CCA (e.g. tristichopterids, osteolepidids, Table S22). Therefore, environmental sampling is likely behind significant values generated by ANOSIM using abundance metrics, rather than real diversity change, turnover, or extinction (Tables S11, S12, S15, S16). Indeed, there is more than double the number of marine sites known from the Frasnian than from the Famennian (4 vs. 10) (Table S1), even though a greater number of Famennian localities were included in the overall dataset. Analyses performed using single environments in order to examine the impact of sampling, and the results are presented below.

SIMPER reveals the Famennian-Tournaisian average dissimilarity to be anomalously high (83.00), more than double the Tournaisian-Viséan (40.80) (Table S20). Comparisons based on relative diversity data show that 44% of all higher-level groups (all placoderms, many sarcopterygians and acanthodians) were completely lost over the Hangenberg event (Table S21). Increases in the relative diversity of most surviving taxa were as expected from the losses alone. However, actinopterygian species per site increases 20-fold in over the recovery interval, while holocephalans and gyracanthids show similar increases in mean relative abundance. However, these increases contribute less to overall faunal dissimilarity than extinctions, showing the latter to be the main driver of Devonian-Mississippian turnover.

B. Extinction Intervals: Marine and Non-marine Environments.

1. NMDS

Faunas of pre- and post-event stages were subjected to additional ordination analyses with sites marked by environment. Nearly all variation in the NMDS plots for Frasnian and Famennian sites is along the first coordinate (Figs. 4C, S7A, S8A, S9A), which represents a salinity gradient equivalent to CCA axis 2 (Figs. 3 and S6). No real difference is observed between plots generated by Bray-Curtis and Kulczynski metrics, nor from the raw and relative diversity datasets. As with the CCA (Figs. 3 and S6, Tables S5-S8), sites are distributed as expected from *a priori* assignments of environment (Figs. S8A, S9A, S10A). This once again shows that designations based on geological data can be confirmed by vertebrate faunal composition. Unlike the overall NMDS plots (Fig. S7), there appears to be no temporal gradient or gap – Frasnian and Famennian faunas overlap along both coordinates. Clustering of all sites is particularly tight along second axis. Differences between the spread of Frasnian and Famennian sites in the NMDS plots are due to differences in the number of marine and non-marine sites as well as geographic sampling (Figs. S8A, S9A, S10A). In summary, there is little evidence of any break in faunal composition associated with the Kellwasser event in either environment.

The coordinates generated by NMDS ordination of Famennian and Tournaisian-Viséan sites are very similar to the CCA and NMDS axes produced by the total datasets (Figs. 3A, S7, S8A, S9A, S10A) The only real difference is that positive values along axis 1 are associated with the Devonian rather than the Mississippian. Marine sites in the Devonian have more positive values on coordinate 2 than nonmarine sites, again as in CCA and the overall NMDS. Post-Hangenberg sites of all environments are tightly clustered on this coordinate, but marine and nonmarine sites are still well-sorted on the plot. Some other unknown coordinate must describe variation between environments during the recovery interval. These results again show that *a priori* environmental assignment in other analyses to be appropriate. No real difference in results is observed among plots generated by Bray-Curtis and Kulczynski metrics, nor from the raw and relative diversity datasets (Figs. S8A, S9A, S10A). All plots show a large gap between all Famennian sites and all Mississippian sites of the same environment. The closest approach between sites from pre- and post-Hangenberg intervals along coordinate 1 is again between Famennian open marine localities and an inland freshwater fauna from the late Viséan, some 30 million years later (Figs. S8A, S9A, S10A). This precludes the possibility of a single environment (e.g. freshwater) serving as an unaffected refugium during the extinction event – both the marine and nonmarine assemblages were changed.

2. Factor Analysis

Factor analyses plots generated from raw and relative diversity data are identical. Clusters of marine sites from before and after the Kellwasser event are overlapped on all major factors (Fig. S11A and Tables S22 and S23), as are nonmarine localities. Factors are completely environmental like the axes generated in NMDS (Table S22). Factor 1 (54.95% of variance) is positively associated with non-marine taxa from both stages (Figs. S11A and Table S22). Factor 2 (16.82% of variance) is positively associated with

marine taxa (Fig. S11A), while factor 3 (6.06% of variance) and factor 4 (<5.00% of variance) are negatively loaded by the same (Table S22). These results once again show that the *a priori* environmental assignments used here are accurate. The plots of factors 1 and 2 show complete diagonal salinity gradient, with sites ranging from open marine to completely terrestrial (Fig. S11A). There is some geographic signal in the results – Middle Eastern sites from around the boundary are close, as are North American and European/German localities (Table S23). This suggests that both environmental and geographic sampling have a hand in any observed difference between Frasnian and Famennian faunas.

Factor analysis plots of peri-Hangenberg sites exhibit a large gap between all Devonian and all Mississippian sites from both environments (Fig. S11C). Plots generated from raw and relative diversity are completely identical. Factor 1 (42.92% of variance) is associated positive correlated with Mississippian taxa from all environment (Fig. S11C and Table S24). Factor 2 (25.88% of variance) is positively associated with Devonian non-marine taxa (Fig. S11C and Table S24). Factor 3 (9.85% of raw diversity variance) is positively correlated with Devonian marine groups (Table S24), as is Factor 4 (<5.00% of variance). All Mississippian sites fall near the origins of factors loaded by primarily Devonian taxa (Figs. S11C), and do not overlap with pre-Hangenberg localities on any major axis (Table S25).

3. ANOSIM

According to the ANOSIM results, faunas sampled from single environments exhibit great similarity across the Kellwasser event. (Tables S11, S12, S15, S16). Analyses of Frasnian and Famennian marine sites produced higher mean p-values (raw and relative diversity mean $R=0.25$, $p=0.1$) than pairwise comparisons of whole stages (Tables S11, S12, S15, S16). The mean values are similar to those obtained for non-extinction intervals in the overall analyses (Tables S13, S14, S17, S18). However, the results of individual metrics were highly mixed, likely due to the large difference in number of sites between intervals and geographical sampling. Insignificant values were obtained for non-marine faunas (raw diversity mean $R=-0.08$, $p=0.7$; relative diversity mean $R=-0.07$, $p=0.7$) - less saline biotas are largely unchanged over the boundary (Tables S11, S12, S15, S16). These results again show that environmental sampling, not mass extinction or major diversity change, is primarily responsible for the turnover signal observed in comparisons of entire stages.

In contrast to the Kellwasser results, ANOSIM comparisons of marine sites around the Hangenberg event indicate extreme dissimilarity of faunas. Marine R statistics are much higher than those generated for entire intervals, while p -values are higher only because of the relatively low number of Famennian and Tournaisian-Viséan marine sites (raw diversity mean $R=0.90$, mean $p=0.03^*$, relative diversity mean $R=0.91$, mean $p=0.03^*$) (Tables S11, S12, S15, S16). Non-marine values have greater significance (raw and relative diversity mean $R=0.91$, $p=0^*$ in 1000000 permutations) (Tables S11, S12, S15, S16). This suggests that analyzing the overall extinction interval was actually depressing the already strong signal. Results were consistent regardless of which similarity metric was used (Tables S11, S12, S15, S16). The Hangenberg turnover was not restricted to a single environment, but rather appears to have been global in effect.

4. SIMPER

In agreement with the ANOSIM results, average dissimilarities generated by SIMPER for Frasnian-Famennian marine (54.53) and non-marine (49.15) sites were lower than that generated from comparison of the overall stages and not much different from background comparisons (Tables S19, S26, S27). Marine faunal differentiation was driven primarily by increases in elasmobranchs and symmoriformes (Table S26), something which might partly be attributed to the greater representation of open marine shales in the Famennian (e.g. Cleveland Shale, *Cheiloceras* beds, and Tafilalt). These types of deposits exhibit better preservation of chondrichthyan remains. In contrast to the overall SIMPER results and previous assertions, marine arthrodires appear to pass through the boundary unscathed – their per site diversity is unaffected (Table S26). While there are some losses among groups in the purely marine results, these are all represented in marginal marine and estuarine environments of the Famennian – with many of these same taxa apparently rising from the dead (Table S27). This suggests that these might not represent real marine extinctions, but rather slight habitat shifts or changes in marine sampling owing to the paucity of nearshore reef sites documented elsewhere. The SIMPER results for nonmarine sites exhibit no real trends – some decreases in relative diversity are observed, but a greater number of groups are represented in Famennian environments (Table S27).

The average dissimilarities generated for SIMPER comparisons of pre- and post-Hangenberg marine and non-marine sites (80.74 and 81.73 respectively) were similar to the high value produced for the overall Famennian and Tournaisian (83.19) (Tables S19, S28, S29). Extinctions are observed in marine and non-marine faunas, with the composition of post-extinction faunas highly similar in both (Tables S28 and S29). This once again indicates that the Hangenberg extinction struck all environments equally.

B. Extinction Intervals: Restricted Timespans.

1. NMDS

Late Frasnian and early Famennian sites (Fig. 2) were analysed in order to determine whether any faunal change could be associated specifically with the Kellwasser event rather than longer-term phenomena. In NMDS plots of this restricted interval, constructed using both datasets and either Bray-Curtis or Kulczynski distances, coordinate 1 looks similar to that produced by analyses of the whole stages (Figs. 4A, S8B, S9B, S10B). Negative values are associated with marine environments, while positive values are associated with nonmarine faunas. As was observed in the environmental analyses, most of the difference between stage clusters is caused by the relative number of sites from each environment. Pre- and post-extinction clusters are more extensively overlapped in plots based on presence-absence distances (Fig. S10B). Therefore, the overall composition of faunas changed less over the Frasnian-Famennian boundary than the relative diversity of coexisting groups. This agrees with the insignificant values generated using presence-absence metrics in ANOSIM. The second coordinate seems to be geographic and partly environmental. In addition, sites from the

same biogeographic region event are adjacent in the ordination despite being separated by the Kellwasser event (e.g. Stolbovo and Tervete, Scaat Craig and Portishead).

NMDS plots of late Famennian and Tournaisian sites (Figs. 2, S8D, S9D, S10D) exhibit the major axes similar both the overall NMDS plots and the CCA (Figs. 3, 4A, S5). However, the first coordinate is arbitrarily flipped in when using Bray-Curtis distance and raw diversity (Fig. S8D). As with all other ordination plots involving Devonian and Mississippian localities, a large gap is seen between late Famennian and Tournaisian sites on the first axis - although this is not aligned near the origin in the raw diversity results (Figs. S8D and S10D). The second coordinate is environmental, with marine sites holding more positive positions. Distances between marine and non-marine sites across the boundary are about equal, again giving little evidence for freshwater refugia across the Hangenberg event. As in previous analyses, Tournaisian sites form the relatively tight cluster. NMDS plots constructed using only end-Famennian sites conformable with the Hangenberg event show the same exact pattern – no latest Devonian locality is more closely associated with Mississippian faunas than earlier localities (Fig. S12). This restricts the timing of the gap along axis 1, as well all associated extinctions, to the period between the beginning of the Hangenberg event and the deposition of Horton Bluff in the early-mid Tournaisian.

2. Factor Analysis

Factor analysis of late Frasnian and early Famennian sites produced the same factors as the overall stages using both raw and relative diversity data (Fig. S11B and Table S30). Factor 1 (56.84% of variance) was positively associated with non-marine taxa and sites. Factor 2 (19.86% of variance) was positively correlated with marine taxa. Factor 3 (6.04% of variance) and factor 4 (5.44% of variance) were both negatively loaded by marine groups (Tables S30). In the plots themselves, sites from similar environments but different stages overlap (Fig. S11B). Once again, there is no temporal signal or Kellwasser-associate faunal break observed in the data (Fig. S11B and Table S31). Environmental sampling, involving a paucity of marine sites in the early Famennian, plays a significant role in observations of faunal change around the stage boundary.

Factor analysis of late Famennian and Tournaisian sites generated factors slightly different from those of previous analyses of longer intervals, but still associated with the faunas of one period or the other (Figs. S11D and Table S32). Factor 1 (43.50% of variance based on both relative and raw data) is positively loaded by Famennian non-marine taxa, such as antiarchs (Tables S32). Factor 2 (22.78% of variance) is positively correlated with members of the Tournaisian biota, both marine and non-marine (Table S40). Factor 3 (12.45% of raw diversity variance) is negatively correlated with Famennian marine forms (Table S32). Factor 4 (5.11% of variance) is positively correlated with Famennian sarcopterygians and other estuarine taxa (e.g. those near the origin of correspondence axis 2 in CCA analysis (Figs. 3 and S6 and Table S32). There is no overlap between late Famennian and Tournaisian clusters on plots of factors 1, 2, and 3 (Figs. S11D), and no overlap along factor 2, the only factor associated with a single stage (Table S33).

Factor analysis of end-Famennian and Tournaisian sites linked the greatest part of the variance directly with changes observed in previous SIMPER analyses. Plots and scores were identical in analyses of both raw and relative diversity. Factor 1 (45.64% of variance) is more positively associated with all those taxa showing increases or little change in SIMPER comparison of the Famennian and Tournaisian (Tables S21 and S34) and less loaded or even negatively loaded by all groups facing extinction or decreases across the Hangenberg horizons (Tables S21 and S34). Factor 2 (24.04% of variance) is negatively correlated with components of the end-Famennian marine fauna (Table S34). Factor 3 (13.81% of variance) is positively associated with Famennian non-marine taxa, mostly sarcopterygians (Table S34). Factor 4 (5.95% of variance) is positively associated with chondrichthyans, and might represent a Tournaisian marine signal. As with the comparisons of longer intervals around the Hangenberg event, Tournaisian and end-Famennian clusters show little overlap in any plot of the major factors (Fig. S12D). The gap between all Tournaisian and all Famennian sites along factor 1 is very wide, much more so than on other factor plots involving many of the same sites (Fig. S12D and Table S35). Once again, the separation between end-Famennian and Tournaisian faunas, as well as associated higher-level group extinctions, is pinpointed to a short interval coincident with the Hangenberg event and its immediate aftermath.

3. ANOSIM

ANOSIM results for the late Frasnian and early Famennian show the same general pattern as comparisons of the entire stages (Tables S11, S12, S15, S16). The overall faunas are not significantly different (raw diversity mean $R=0.19$, mean $p=0.1$; relative diversity mean $R=0.20$, mean $p=0.1$) (Tables S11, S12, S15, S16). However, there is a significant difference in relative abundance of coexisting groups (raw diversity mean $R=0.31$, mean $p=0.04^*$; relative diversity mean $R=0.33$, $p=0.03^*$) which is not mirrored in values based on presence-absence metrics (raw and relative diversity mean $R=0.10$, $p=0.2$) (Tables S11, S12, S15, S16). Faunal differentiation between the late Frasnian and early Famennian is not driven by any fundamental change in faunal composition, such as extinction of higher-level groups at the Kellwasser event.

ANOSIM results for the late Famennian and Tournaisian are also nearly identical to those produced by analyses involving the entire Famennian (raw diversity mean $R=0.71$, mean $p<0.001^*$; relative diversity mean $R=0.72$, mean $p<0.001^*$) (Tables S11, S12, S15, S16). A consistent signal is once again seen in the values produced by all similarity metrics. Comparison of end-Famennian and Tournaisian sites also produced highly significant values despite the smaller number of samples, revealing the great strength of the Hangenberg extinction signal (raw diversity mean $R=0.64$, mean $p=0.002^*$; relative diversity mean $R=0.65$, mean $p=0.002^*$) (Tables S11, S12, S15, S16).

4. SIMPER

The results of SIMPER comparison for late Frasnian and early Famennian sites display the same environmental signal seen in comparisons of the entire stages (average dissimilarity=64.01) (Tables S19 and S36). Once again, antiarchs and non-marine Devonian sarcopterygians (tristichopterids and osteolepids) exhibit increases that exactly

mirror the large decreases among the more marine arthrodiros (Table S36). Only a single group is lost, Other Placodermi, but these reappear later in the Famennian and contribute little to larger patterns due to their overall rarity. Differences in environmental sampling are once again driving dissimilarity right around the Kellwasser horizon, while any evidence of high-level extinction is missing. This, along with all the other results of this study, casts doubt on the status of the Frasnian-Famennian mass extinction, as it seems to have had little real, demonstrable effect on the vertebrate biota.

SIMPER comparisons of Tournaisian, late Famennian, and end-Famennian sites show that all higher-level taxa are represented in Devonian deposits right through to the Hangenberg horizon (Tables S37 and 38). Therefore, extinction of these groups before the event is extremely unlikely. Between the start of the Hangenberg event and the earliest Tournaisian locality, the vertebrate biota experienced the complete extinction of 44% of gnathostome higher-level groups, involving the total loss of placoderms and several acanthodian and sarcopterygian monophyletic groups (Tables S37 and S38). There were attendant increases in the relative contribution of survivors to the Tournaisian fauna. Arthrodiros remain the most taxonomically diverse segment of the Famennian vertebrate biota right up the boundary. In contrast, actinopterygians, a minor part of Devonian faunas, radiate far beyond what would be expected during the post-extinction interval, becoming the most taxonomically diverse segment of the Mississippian biota. This suggests that actinopterygians opportunistically replaced most of the dead in the Tournaisian recovery fauna (Tables S37 and S38). This acute faunal turnover, involving the extinction of multiple, large, mostly monophyletic groups - observed in all comparisons of Devonian and Mississippian sites in this study - is the expected signal of a true mass extinction. Therefore, on the basis of all analyses, the Hangenberg extinction should be considered an anomalous event, with devastating effects on the worldwide vertebrate biota.

III. Site Descriptions and Faunal Lists

A. Givetian Sites

1. Orcadian Lake.

Stratigraphy/Location. Orcadian basin, Scotland. **Age.** early Givetian.

Environment. Non-marine, Lacustrine/Fluvial.

Taxa. Actinopterygii: *Cheirolepis trailli*. Porolepiformes: *Holoptychius* sp., *Glyptolepis leptopterus*, *Glyptolepis paucidens*. Dipnoi: *Palaeospondylus gunni*, *Dipterus valenciennesi*, Unnamed lungfish. Osteolepididae: *Osteolepis macrolepidotus*, *Gyroptychius milleri*, *Gyroptychius agassizi*. Tristichopteridae: *Tristichopterus alatus*. Arthrodira: *Coccosteus decipiens*, *Homosteus milleri*. Antiarcha: *Pterichthyodes milleri*. Ptyctodontida: *Rhamphodopsis trieplandi*. Climatiida: *Diplacanthus striatus*, *Diplacanthus tenustriatus*, *Diplacanthus (Rhadinacanthus) longispinus*, *Diplacanthus crassisimus*. Acanthodida: *Mesacanthus peachi*, *Cheiracanthus murchisoni*, *Cheiracanthus latus*, *Acanthodes peachi*.

References. 6, 24, 25, 26.

2. Harma.

Stratigraphy/Location. Karski outcrop, Harma substage, Burtnieki regional stage, Estonia. **Age.** early Givetian. **Environment.** Non-marine, deltaic plain.

Taxa. Actinopterygii: *Cheirolepis* sp. Onychodontida: *Onychodus* sp., *Hamodus lutkevitchi*. Porolepiformes: *Glyptolepis karksiensis*, Holoptychiid. Osteolepididae: *Gyroptychius elgae*. Arthrodira: *Heterostius ingens*, *Coccosteus markae*, *Holonema harmae*, *Actinolepis magna*, *Homostius latus*, *Tropinema harmae*. Antiarcha: *Asterolepis dellei*, *Asterolepis* sp. Elasmobranchii: *Karksilepis parva*, Elasmobranchii gen et. sp. nov. Climatiidae: *Minioracanthus* sp., *Rhadinacanthus multisulcatus*, *Diplacanthus carinatus*, *Diplacanthus gravis*, *Diplacanthus* sp., *Ptychodictyon rimosum*, *Ptychodictyon sulcatum*, *Nostolepis gaujensis*. Acanthodida: *Markacanthus alius*, *Cheiracanthus brevicostatus*, *Cheiracanthus longicostatus*, *Cheiracanthus* sp. nov. 1, *Cheiracanthus* sp. nov. 2., *Homacanthus gracilis*, *Homacanthus* sp., Acanthodid gen. et. sp. nov.

References. 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37.

3. Gerolstein.

Stratigraphy/Location. *Stringocephalus* beds, Eifel, Germany. **Age.** Early Givetian. **Environment.** Marine.

Taxa. Dipnoi: *Conchodus elkneri*, *Rhinodipterus* sp., *Archaeonectus pertusus*. Actinistia: *Euporosteus eifelianus*. Arthrodira: *Rhenonema eifeliense*, *Heterostius rhenanus*. Antiarcha: *Gerdalepis dohmi*, *Grossaspis carinata*. Ptyctodonta:

Desmoporella minor, *Palaeomylus predator*, *Ptyctodus sp.*, *Rhychodus rostratus*, *Goniosteus gerolstienensis*. Other Placodermi: *Macropetalichthys pelmensis*, *Epipetalichthys minor*. Ischnacanthida: *Onchus sp.*

References. 27, 28, 29, 30, 38, 39, 40, 41, 42.

4. Rockport.

Stratigraphy/Location. Rockport formation, Traverse group, Michigan. **Age.** mid-Givetian. **Environment.** Marine, inland sea.

Taxa. Onychodontida: *Onychodus sp.* Dipnoi: *Cheirodipterus onawayensis*. Arthrodira: *Protitanichthys rockportensis*, *Dinomylostoma sp.*, *Holonema rugosum*, *Holonema farrowi*, *Dinichthys sp.*, *Mylostoma sp.* Antiarcha: *Bothriolepis sp.*, Ptyctodontida: *Eczematolepis sp.* Elasmobranchii: *Ctenacanthus sp.*, *Acondylacanthus gracillimus*. Ischnacanthida: *Machaeracanthus sp.* Gyracanthida: *Gyracanthus sp.* Acanthodida: *Oracanthus sp.*

References. 27, 28, 43, 44, 45, 46, 47, 48, 49

5. Aztec.

Stratigraphy/Location. Aztec siltstone, Antarctica **Age.** Mid-Givetian. **Environment.** Non-marine, alluvial, lacustrine, fluvial.

Taxa. Actinopterygii: *Donnrosenia schaefferi*. Porolepiformes: *Porolepiformes sp.* Dipnoi: *Howidipterus sp.*, *Ctenodontid sp.*, *Eoetenodus sp.* Osteolepididae: *Gyroptychius antarcticus*, *Koharalepis jarviki*, *Platyethmoidea antarctica*. Megalichthyidae: *Mahalalepis resima*. Rhizodontida: *Notorhizodon mackelveyi*, *Aztecia mahalae*. Actinistia: *Miguashaia grossi*. Arthrodira: *Antarctolepis gunni*, *Groenlandaspis antarcticus*, *Groenlandaspis sp.*, *Boomeraspis goujeti*, *Phlyctaeniid sp. nov.* Antiarcha: *Bothriolepis antarctica*, *Bothriolepis alexi*, *Bothriolepis askinae*, *Bothriolepis barretti*, *Bothriolepis barretti*, *Bothriolepis karawaka*, *Bothriolepis kohni*, *Bothriolepis macphersoni*, *Bothriolepis mawsoni*, *Bothriolepis portalensis*, *Bothriolepis vuwae*, *Venezuelepis antartica*, *Pampulaspis antarctica*. Phyllolepida: *Austrophyllolepis quiltyi*, *Austrophyllolepis youngi*, *Placolepis tingeyi*, *Phyllolepid sp.*, *Antarctaspis mcmurdoensis*. Elasmobranchii: *Mcmurdodus featherensis*, *Antarctilamna prisca*, *Anareodus statei*, *Aztecodus harmeseniae*, *Portalodus bradshawae*. Climatiida: *Milescanthus antarctica*, *Culmacanthus antarctica*, *Nostolepis gaujensis*, *Pechoralepis juozasi*, *Antarctonchus glacialis*. Gyracanthida: *Gyracanthides warreni*. Acanthodida: *Cheiracanthus sp.*

References. 27, 28, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60.

6. Haikou.

Stratigraphy/Location. Haikou formaton, Yunnan, China, **Age.** Mid-Givetian. **Environment.** Marine.

Taxa. Porolepiformes: *Heimenia* sp. Arthrodira: *Jiuchengia longoccipita*, *Yinostius major*, *Kunmingolepis lucaowanensis*, *Quasipetalichthys haikouensis*, *Yangalepis jinningensis*. Antiarcha: *Bothriolepis tungseni*, *Dianolepis liui*, *Xichonolepis qujingensis*, *Hunanolepis tieni*. Other Placodermi: *Exutaspis megista*.

References. 61, 62, 63, 64, 65, 66, 67.

7. The Cletts.

Stratigraphy/Location. Caithness, Middle Old Red Sandstone, Scotland. **Age:** Late Givetian. **Environment.** Non-marine, fluvial/alluvial.

Taxa. Actinopterygii: *Stegotrachelus finlayi*. Dipnoi: *Pentlandia macroptera*, *Dipterus macropterus*. Tristichopteridae: *Tristichopterus* sp. Arthrodira: *Watsonosteus fletti*, *Cocosteus* sp. nov. Antiarcha: *Microbranchius dicki*.

Reference. 6.

8. Mt. Howitt.

Stratigraphy/Location. Mt. Howitt black shale, Victoria, Australia. **Age.** Late Givetian. **Environment.** Non-marine, lacustrine.

Taxa. Actinopterygii: *Howqualepis rostridens*. Dipnoi: *Howidipterus donnae*, *Barwickia downunda*. Osteolepididae: unnamed osteolepidid. Actinistia: *Gavinia syntrips*. Arthrodira: *Groenlandaspis* sp. Antiarcha: *Bothriolepis gipplandiensis*, *Bothriolepis cullodensis*, *Bothriolepis fergusoni*. Phyllolepida: *Austrophyllolepis ritchei* Acanthodida: *Howittacanthus kentoni*, unnamed acanthodid 1, unnamed acanthodid 2, unnamed acanthodid 3.

References. 58, 68, 69, 70, 71, 72, 73.

9. Abava.

Stratigraphy/Location. Abava substage, Burtneki regional stage, Estonia. **Age.** Late Givetian. **Environment.** Non-marine, fluvially dominated basin.

Taxa. Actinopterygii: *Moythomasia* sp. Onychodontida: *Hamodus lutkevitchi*. Porolepiformes: *Laccognathus* sp. Dipnoi: Unnamed dipnoan. Arthrodira: *Livosteus grandis*, *Hybosteus mirabilis*, *Plourdosteus panderi*. Antiarcha: *Byssacanthus* sp., *Microbranchius dicki*. Elasmobranchii: unnamed elasmobranch. Acanthodida: *Cheiracanthus brevicostatus*, *Acanthodes* sp. 1, *Acanthodes* sp. 2, *Acanthodes* sp. 3, *Acanthodes* sp. 4. Tetrapoda: *Panderichthys* sp.

References. 27, 28, 29, 30, 32, 36, 37, 74.

10. Cedar Valley.

Stratigraphy/Location. Cedar Valley Limestone, Milwaukee formation, Iowa. **Age.** late Givetian. **Environment.** Marine, reef.

Taxa. Onychodontida: *Onychodus sigmoides*. Dipnoi: *Straitonia* sp., *Iowadipterus halli*, *Dipterus uddeni*, *Dipterus calvani*. Arthrodira: *Dinichthys pustulosus*, *Dinichthys tuberculatus*. Ptyctodontida: *Palaeomylus greeni*. *Ptyctodus ferox*, *Ptyctodus calceolus*, *Ptyctodus uddeni*, *Rhynchodus excavatus*, *Gamphacanthus politus*, *Ptyctodopsis mezelli*. Elasmobranchii: *Phoebodus macisaacsi*.

References. 27, 43, 75, 76, 77, 78, 79, 81, 82.

11. Gauja.

Stratigraphy/Location. Lode formation, Gauja, Latvia. **Age.** Latest Givetian.

Environment. Non-marine, estaurine, deltaic.

Taxa. Actinopterygii: *Cheirolepis* sp. Onychodontida: *Strunius* sp. nov. Porolepiformes. *Laccognathus grossi*, *Laccognathus panderi*, *Glyptolepis baltica*, *Latvius* sp. Dipnoi: *Grossipterus crassus*, *Griphognathus minutidens*. *Rhinodipterus secans*. Osteolepididae: *Megadonichthys kurikae*. Tristichopteridae: *Eusthenopteron kurshi*. Actinistia: *Miguashaia grossi*. Arthrodira: *Asterolepis ornata*, *Plourdosteus livonicus*, *Plourdosteus panderi*. Antiarcha: *Bothriolepis curonica*, *Grossilepis spinosa*, *Taenolepis speciosa*, *Asterolepis ornata*. Ischnacanthida: *Devononchus concinnus*. Climatiida: *Nostolepis guajensis*, *Nodacosta pauli*. Acanthodida: *Acanthodes* sp., *Lodeacanthus gaujicus*, *Haplacanthus ehrmanensis*. Tetrapoda: *Panderichthys rhombolepis*, *Livoniana multidentata*.

References. 27, 28, 29, 30, 32, 36, 37, 42, 60, 74, 83, 84, 85, 86, 87, 88, 89.

B. Frasnian Sites.

12. Gladbach.

Stratigraphy/Location. Bergish Gladbach, Oberer Plattenkalk, Germany. **Age.** Earliest Frasnian. **Environment.** Marine.

Taxa. Actinopterygii: *Moythomasia nitida*, *Moythomasia striatus*, *Stegotrachelus* sp. Dipnoi: *Dipterus valenciennesi*, *Rhinodipterus ulrichi*, *Griphognathus sculpta*. Osteolepididae: *Gyroptychius* sp., *Thursius* sp., *Latvius niger*. Actinistia: *Holopterygius nudus*, *Diplocercides heiligenstockiensis*. Arthrodira: *Rachioosteus pterygiatus*, *Coccosteus* sp., *Gyroplacosterus* sp., *Plourdosteus* sp. Antiarcha: *Asterolepiformes* sp. Ptyctodontida: *Ptyctodus* sp., *Ctenurella gladbachensis*. Elasmobranchii: *Gladbachus adentatus*. Ischnacanthida: *Atopacanthus* sp. Acanthodida: *Protogonacanthus juergeni*.

References. 27, 28, 42, 74, 90, 91, 92, 93, 94, 95, 96.

13. Boghole.

Stratigraphy/Location. Boghole Beds, Upper Old Red Sandstone, Scotland. **Age.** Early Frasnian. **Environment.** Non-Marine, fluvial.

Taxa. Porolepiformes: *Holoptychius decorata*, *Holoptychius nobillissimus*.
Tristichopteridae: *Eusthenopteron traquairi*. Rhizodontida: *Polyplocodus leptognathus*.
Arthrodira: *Plourdosteus magnus*. Antiarcha: *Asterolepis alta*.

Reference. 6.

14. Gogo.

Stratigraphy/Location. Gogo Crossing, Victoria, Australia. **Age.** Early Frasnian.

Environment. Marine, reef.

Taxa. Actinopterygii: *Gogosardinus coatesi*, “*Mimia*” *toombsi*, *Moythomasia durgaringa*. Onychodontida: *Onychodus jandemarrai*. Dipnoi: *Chirodipterus australis*, “*Griphognathus*” *whitei*, *Pillararhynchus longi*, *Holodipterus gogoensis*, *Holodipterus longi*, *Gogodipterus paddyensis*, *Holodipterus (Holodipteroides) elderae*, *Holodipterus meemanae*, *Adolopas moysmithae*. Tristichopteridae: *Gogonasmus andrewsae*. Arthrodira: *Holonema westolli*, *Harrytoombsia elegans*, *Camuropiscis concinnus*, *Camuropiscis laidlawi*, *Tubonasmus lennardensis*, *Rolfosteus canningensis*, *Kendrickichthys cavernosus*, *Bullerichthys fascidens*, *Bruntonichthys multidentis*, *Incisoscutum ritchiei*, *Torosteus tibericulatus*, *Torosteus pulchellus*, *Compagopiscis croucheri*, *Gogopiscis gracilis*, *Kimberleyichthys cuspidatis*, *Kimberleyichthys whybrowi*, *Simosteus tuberculatus*, *Eastmanosteus calliaspis*, *Fallacosteus turnerae*, *Pinguosteus thulborni*, *Gogosteus sarahae*, *Mcnamaraspis kaprios*. Antiarcha: *Bothriolepis* sp. Ptyctodonta: *Campbellodus decipiens*, *Austroptyctodus gardineri*, *Materpiscis attenboroughi*. Elasmobranchii: *Elasmobranchii* gen. et sp. nov. Climatiidae: *Culmacanthus stewartii*.

References. 69, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125.

15. Chahriseh.

Stratigraphy/Location. Kuh-e-Kaftar, Chahriseh section, central Iran. **Age.** early Frasnian.

Environment. Marine, shallow sea.

Taxa. Actinopterygii: *Moythomasia* sp. Onychodontida: *Strunius rolandi*, *Orvikuina* sp. Dipnoi: *Rhinodipterus* sp., *Dipterus* sp. Arthrodira: *Holonema* sp. Antiarcha: *Bothriolepis* sp. Symmoriformes: *Stethacanthus* sp. Elasmobranchii: *Manberodus fortis*, *Phoebodus latus*, *Protacrodus serra*, *Thrinacodus* sp. Climatiidae: *Nostolepis gaujensis*, *Iranolepis ginteri*.

References. 126, 127, 128, 129.

16. Kerman.

Stratigraphy/Location. Kerman, Tabas Region, eastern Iran. **Age.** Early Frasnian.

Environment. Marine.

Taxa. Actinopterygii. *Moythomasia* sp., “*Rhadinichthys*” *devonicus*. Onychodontida: *Onychodus firouzi*, *Strunius rolandi*. Dipnoi: *Rhinodipterus* sp., *Dipterus* sp., *Chirodipterus* sp., *Iranorhynchus seyedemamii*. Osteolepididae: *Osteolepidid* sp.

Actinistia: *Diplocercides* sp. Arthrodira: *Aspidichthys ingens*, *Holonema radiatum*, *Eastmanosteus* sp., *Plourdosteus mironovi*, *Golshanichthys asiatica*, *Deirosteus* sp. Antiarcha: *Bothriolepis* sp., *Stegolepis* sp., *Byssacanthus dilatatus*, *Asterolepis* sp. Ptyctodontida: *Rhynchodus* sp., *Ptyctodus* sp. Other Placodermi: Unnamed Macropetalichthyid Elasmobranchii: *Cladodus* sp., *Phoebodus* sp., *Protacrodus* sp. Ischnacanthida: *Persacanthus kermanensis*. Climatiida: Unnamed Climatiid.

References. 27, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139.

17. North Evans.

Stratigraphy/Location. Genesee formation, North Evans/Geneseo, New York. **Age.** Early Frasnian. **Environment.** Marine.

Taxa. Actinopterygii: Unnamed actinopterygian. Onychodontida: Unnamed onychodont. Arthrodira: *Dunkleosteus newberryi*, *Dinichthys magnificus*, *Eastmanosteus aduncus*, *Eastmanosteus pustulosus*, “*Heintzichthys*” *denticulatus*, *Heintzichthys insignis*, *Dinomylostoma buffaloensis*, *Copanognathus crassus*, *Machaerognathus woodwardi*, *Acanthaspis* sp., *Aspidichthys notabilis*, *Coccosteus* sp., *Holonema* sp. Ptyctodontida: *Eczematolepis fragilis*, *Palaeomylus hussakofi*, *Palaeomylus lunaeformius*, *Palaeomylus minor*, *Ptyctodus compressus*, *Ptyctodus howlandi*, *Rhynchodus ornatus*, *Rhynchodus telleri*. Other Placodermi: *Oestophorus lilleyi*. Elasmobranchii: *Ctenacanthus* sp., *Coronodus reimanni*. Holocephalii: *Acmoniodus clarkei*, *Synthetodus calvini*. Ischnacanthidae: *Atopacanthus* sp., *Machaeracanthus retustus*. Gyracanthidae: *Gyracanthus sarlei*, *Gyracanthus* sp.

References. 27, 28, 39, 38, 43, 44, 140, 141, 142, 143, 144, 145, 146, 147, 148.

18. Miguasha.

Stratigraphy/Location: Miguasha, Escuminac formation, Quebec. **Age:** Early Frasnian. **Environment:** Non-marine, estuarine.

Taxa. Actinopterygii: *Cheirolepis Canadensis*. Porolepiformes: *Quebecia quebecensis*, *Holoptychius jarviki*. Dipnoi: *Fleurantia denticulate*, *Scaumenacia curta*. Tristichopteridae: *Eusthenopteron fordi*. Actinistia: *Callistiopterus macelveyi*. Arthrodira: *Plourdosteus canadensis*. Antiarcha: *Bothriolepis canadensis*, *Bothriolepis traquairi*. Climatiida: *Diplacanthus ellsii*, *Diplacanthus horridus*. Acanthodida: *Triazeugacanthus affinus*, *Homalacanthus concinnus*. Tetrapoda: *Elpistostege watsoni*.

References. 27, 28, 147, 148, 149, 150, 151, 152, 153.

19. Wietrznia.

Stratigraphy/Location. Zone II, Wietrznia, Kadzielnia, Holy Cross Mountains, Poland. **Age.** Early/mid-Frasnian. **Environment.** Marine.

Taxa. Actinopterygii: “*Rhadinichthys*” *devonicus*. Onychodontida: *Onychodus jaekeli*. Porolepiformes: *Holoptychius giganteus*, *Glyptolepis* sp. Dipnoi: *Holodipterusus santacruzensis*, *Conchodus elkneri*, *Dipterus murchisoni*. Tristichopteridae:

Eusthenopteron sp. Actinistia: *Diplocercides* sp. Arthrodira: *Deveonema obrucevi*, *Gyrolacosterus vialowi*, *Holonema radiatum*, *Oxyosteus* sp., *Plourdosteus grossi*, *Malerosteus gorizdroae*, *Eastmanosteus new* sp., *Brachydeirus scaber*, *Brachydeirus grandis*, *Pachyosteus bulla*, *Erromanosteus* sp., *Rhynchodus* sp. 1, *Rhynchodus* sp. 2, *Holonema* sp., *Gyrolacosteus* sp., *Plourdosteus* sp., *Dinichthys* sp., *Eastmanosteus* sp. 2. Antiarcha: *Bothriolepis panderi*. Ptyctodontida: *Ptyctodus czarnockii*, *Ptyctodus kielcensis*, *Ptyctodus obliquus*, *Rhynchodus marginalis*, *Rhynchodus tetrodon*. Elasmobranchii: *Protacrodus vetustus*, *Phoebodus fastigatus*. Symmoriformes: *Stethacanthus resistens*.

References. 27, 154, 155, 156, 157, 158, 159.

20. Rodebjerg.

Stratigraphy/Location. Rodebjerg Formation/Snehvide Formation, Kap Kolthoff Group, Greenland. **Age.** Early/mid-Frasnian. **Environment.** Non-marine, alluvial, braidplain.

Taxa. Porolepiformes: *Holoptychius* sp. , Dipnoi: *Spodichthys buetleri*. Osteolepididae: *Thursius minor*. Arthrodira: *Clarkosteus* cf. *halmodeus*, Unnamed arthrodira Antiarcha: *Remigolepis tuberculata*, *Asterolepis saevesorderberghi*. Tetrapoda: *Panderichthys* sp.

Reference. 160.

21. Rhinestreet.

Stratigraphy/Location. Rhinestreet shale, Portage formation, New York. **Age.** Late Frasnian. **Environment.** Marine, open marine.

Taxa. Actinopterygii: “*Rhadinichthys*” *devonicus*. Dipnoi: *Dipterus ithacensis*. Arthrodira: *Dunkleosteus magnificus*, *Dinomylostoma beecheri*, “*Heintzichthys*” *ringueberryi*, *Heintzichthys dilochocephalus*, “*Heintzichthys*” *mixeri*, *Callognathus serratus*, *Eastmanosteus pustulosus*. Antiarcha: *Asterolepis maxima*. Ptyctodontida: *Ptyctodus calceolus*. Elasmobranchii: *Cladoselache* sp. Ischnacanthida: *Atopacanthus dentatus*, *Atapeacanthus vetustus*, *Machaeracanthus retustus*. Acanthodida: *Gamphacanthus politus*, *Acanthodes pristis*.

References. 27, 43, 78, 140, 142, 143, 144, 161, 162.

22. Chemung.

Stratigraphy/Location. Chemung facies/Venago Formation, Warren, Pennsylvania. **Age.** Late Frasnian. **Environment.** Marine.

Taxa. Actinopterygii: Unnamed actinopterygian. Onychodontida: *Onychodus hopkinsi*. Porolepiformes: *Holoptychius* sp. Dipnoi: *Ganorhynchus* sp., *Dipterus nelsoni*, *Heliodus lesleyi*. Arthrodira: *Deirosteus abbreviata*, *Holonema rugosus*, *Cocosteus macromus*. Other Placodermi: *Oestophorus lilleyi*. Elasmobranchii: *Ctenacanthus chemungensis*, *Ctenacanthus randalli*. Holocephali: *Helodus gibberulus*. Ischnacanthida: *Onchus rectus*. Acanthodida: *Homacanthus acinaciformis*.

References. 27, 28, 39, 46, 75, 78, 80, 140, 142, 163, 164, 165.

23. Wildungen.

Stratigraphy/Location. Bad Wildungen, Kellwasserkalk, Germany. **Age.** Late Frasnian. **Environment.** Marine.

Taxa. Actinopterygii: *Moythomasia laevigata*. Onychodontida: *Onychodus sp.* Dipnoi: *Chirodipterus wildungensis*. Tristichopteridae: *Devonosteus proteus*. Actinistia: *Nesides heiligenstockiensis*. Arthrodira: *Wildungenichthys grossi*, *Aspidichthys ingens*, *Pholidosteus friedeli*, *Cocosteus bickensis*, *Brachydeirus carinatus*, *Brachydeirus bicarinatus*, *Brachydeirus gracilis*, *Brachydeirus grandis*, *Brachydeirus magnus*, *Brachydeirus minor*, *Belosteus major*, *Synauchenia coalescens*, *Leptosteus bickensis*, *Pholidosteus laevior*, *Tapinosteus heintzi*, *Hadrosteus rapax*, *Erromenosteus (Leiosteus) lucifer*, *Erromenosteus brachyrostris*, *Erromenosteus concavus*, *Erromenosteus (Paraleiosteus) diensti*, *Erromenosteus platycephalus*, *Belosteus elegans*, *Brachyosteus deitrichi*, *Cyrtosteus inflatus*, *Parabelosteus pusillus*, *Parabelosteus acuticeps*, *Parabelosteus tuberculatus*, *Trematosteus fontanellus*, *Braunosteus schmidti*, *Enseosteus jaekeli*, *Enseosteus hermanni*, *Enseosteus pachyostoides*, *Microsteus angusticeps*, *Pachyosteus bulla*, *Rhinosteus traquairi*, *Rhinosteus parvulus*, *Rhinosteus tuberculatus*, *Otonosteus hermanni*, *Walterosteus pachyostoides*. Antiarcha: *Lepadolepis stensioei*. Ptyctodontida: *Rhynchodus tetrodon*, *Rhynchodus eximius*, *Rhynchodus wildungensis*. Other Placodermi: *Jagorina pandora*, *Epipetalichthys wildungensis*. Elasmobranchii: *Cladoselache hassiacus*, *Protacrodus vetustus*, *Cladodus wildungensis*. Acanthodida: *Homacanthus jaekeli*.

References. 1, 2, 27, 28, 30, 49, 138, 148, 166.

24. Scaat Craig.

Stratigraphy/Location. Scaat Craig beds, Upper Old Red Sandstone, Highland, Scotland. **Age:** Late Frasnian. **Environment:** Non-marine, alluvial, fluvial.

Taxa: Porolepiformes: *Holoptychius giganteus*, *Holoptychius nobillissimus*. Dipnoi: *Conchodus ostreiformis*. Rhizodontida: Unnamed rhizodont. Arthrodira: *Cosmacanthus malcolmsoni*. Antiarcha: *Bothriolepis paradoxa*. Tetrapoda: *Elginerpeton pancheni*.

References. 6, 24, 27, 148.

25. Stolbovo.

Stratigraphy/Location. Stolbovo, Syas River, Dubniki beds, Sargayevian region stage, main Devonian field, northwestern Russia. **Age.** Late Frasnian. **Environment.** Non-marine, alluvial.

Taxa. Porolepiformes: *Holoptychius sp.* Dipnoi: *Rhinodipterus stolbovi*, *Conchodus jeroffewi*, *Dipterus sp.* Osteolepididae: *Latvius sp.* Tristichopteridae: *Jarvikina sp.*, *Platycephalichthys bishoffi*. Arthrodira: *Holonema sp.*, *Gyroplacosteus sp.*, *Plourdosteus sp.*, *Eastmanosteus sp.* Antiarcha: *Asterolepis sp.*, *Bothriolepis trauscholdi*.

Ischnacanthida: *Persacanthus* sp. Acanthodida: *Acanthodes* sp. Tetrapoda: *Parapanderichthys stolbovi*.

References. 28, 30, 36, 37, 56, 74, 167.

C. Famennian Sites.

26. Canowindra.

Stratigraphy/Location. Mandangery Sandstone, Canowindra, New South Wales, Australia. **Age.** Early Famennian. **Environment.** Nonmarine, shallow/seasonal basin.

Taxa. Dipnoi: *Chirodipterus potteri*. Osteolepididae: *Canowindra grossi*. Tristichopteridae: *Mandageria fairfaxi*, *Cabonnichthys bursi*. Rhizodontida: *Gooloogongia loomesi*. Arthrodira: *Groenlandaspis* sp. Antiarcha: *Remigolepis walkeri*, *Bothriolepis yeungae*.

References. 27, 56, 168, 169, 170, 171, 172, 173.

27. Rosebrae.

Stratigraphy/Location. Rosebrae beds, Upper Old Red Sandstone, Morayshire, Scotland. **Age.** Early Frasnian. **Environment.** Non-marine, freshwater.

Taxa. Porolepiformes: *Holoptychius nobillissimus*. Dipnoi: *Phaneropleuron* sp., *Rhynchodipterus elginensis*, *Conchodus* sp. Osteolepididae: *Glyptopomus elginensis*. Tristichopteridae: *Eusthenopteron* cf. *traquairi* Antiarcha: *Bothriolepis alvesiensis*, *Bothriolepis cristata*, *Bothriolepis laverocklochensis*.

Reference. 6, 174.

28. Dura Den

Stratigraphy/Location. Dura Den, Upper Old Red Sandstone, Fifeshire, Scotland. **Age.** Early Famennian. **Environment.** Non-marine, freshwater lake.

Taxa. Porolepiformes: *Holoptychius flemingii*. Dipnoi: *Phaneropleuron andersoni*. Tristichopteridae: *Heddeleithys dalgleisiensis*. Antiarcha: *Bothriolepis hydrophilus*, *Bothriolepis cristata*.

References. 6, 24, 174, 175, 176.

29. Cheiloceras Beds

Stratigraphy/Location. *Cheiloceras* beds, Holy Cross Mountains, Poland. **Age.** Early Famennian. **Environment.** Marine.

Taxa. Dipnoi: *Holodipterus kiprijanowi*. Actinistia: *Diplocercides kayseri*, *Diplocercides* sp. Arthrodira: *Dunkleosteus denisoni*, *Dunkleosteus* sp., *Titanichthys kozlowskii*, *Pachyosteus bulla*, *Sentacanthus zelichowskae*. Antiarcha: *Bothriolepis jazwicensis*. Elasmobranchii: *Phoebodus gothicus*, *Phoebodus limpidus*, *Phoebodus australiensis*,

Dittodus sp., *Cladodus sp.*, *Jalodus australiensis*, *Ctenacanthus sp.* Symmoriformes: *Danaea sp.*

References. 1, 2, 27, 39, 41, 155, 156, 165, 177, 178, 179, 180, 181, 182, 183.

30. Tervete.

Stratigraphy/Location. Skujaine river, Tervete formation, Latvia. **Age.** Early Famennian. **Environment.** Non-marine, brackish basin.

Taxa. Porolepiformes: *Holoptychius sp.* Dipnoi: *Dipterus sp.*, *Conchodus sp.* Tristichopteridae: *Platycephalichthys skuensis*. Arthrodira: *Dunkleosteus sp.* Antiarcha: *Bothriolepis ornata*, *Bothriolepis jani*. Phyllolepidida: *Phyllolepis tolli*. Ptyctodontida: *Chelyphorus sp.* Acanthodida: *Homacanthus sweetensis*, *Devononchus tenuispinus*. Tetrapoda: *Obruchevichthys gracilis*.

References. 27, 28, 36, 184, 185.

31. Oryol.

Stratigraphy/Location. Tugenevo beds/Dankoc-Lebedyan beds, Zadoonckian/Plavsk regional stage, central Devonian field, Oryol, Metsensk district, Russia. **Age.** Mid Famennian. **Environment.** Non-marine, deltaic.

Taxa. Actinopterygii: *Moythomasia sp.* Onychodontida: *Strunius rolandi*, *Pycnacanthus fischeri*. Porolepiformes: *Glyptolepis dellei*. Dipnoi: *Holodipterus kiprijanowi*, *Conchodus sp.*, *Jarvikia lebedevi*, *Orlowichthys limnatus*. Osteolepididae: *Glyptopomus bystrowi*, *Cryptolepis grossi*, *Thaumatolepis edelsteini*. Arthrodira: Unnamed pachyosteoromorph. Antiarcha: *Bothriolepis siberica*, *Remigolepis armata*, *Livnolepis zadonica*, *Rossolepis brodensis*. Ptyctodontida: *Chelyphorus verneuli*, *Ptyctodus major*. Elasmobranchii: *Tuberospina nataliae*. Symmoriformes: *Symmorium glabrum*. Ischnacanthida: Unnamed Ischnacanthid. Acanthodida: *Devononchus sp.*, *Acanthodes lopatini*. Tetrapoda: *Jakubsonia livnensis*.

References. 1, 27, 28, 29, 36, 93, 186, 187, 188.

32. Portishead.

Stratigraphy/Location. Woodhill Bay Fish Bed, Upper Old Red Sandstone, Portishead, Avon, England. **Age.** Mid Famennian. **Environment.** Non-marine, alluvial plain.

Taxa. Porolepiformes: *Holoptychius giganteus*, *Holoptychius nobilissimus*. Dipnoi: *Conchodus sp.* Osteolepididae: *Glyptopomus kinnairdi*. Rhizodontida: *Sauripterus anglicus*. Arthrodira: *Groenlandaspis disjunctus*. Antiarcha: *Bothriolepis hydrophila*, *Asterolepis maxima*. Phyllolepidida: *Phyllolepis concentrica*.

References. 6.

33. Aina Dal

Stratigraphy/Location. Aina Dal Formation, Celsius Bjerg Group, East Greenland. **Age.** Late Famennian. **Environment.** Non-marine, fluvial.

Taxa. Porolepiformes: *Holoptychius sp.*, Dipnoi: *Jarvikia arctica*, *Soederberghia groenlandica*. Tristichopteridae: Unnamed tristichopterid Arthrodira: Unnamed arthrodire. Antiarcha: *Remigolepis tuberculata*, *Bothriolepis nielseni*, *Remigolepis kullingi*, *Remigolepis kochi*, *Remigolepis cristata*. Phyllolepidia: *Phyllolepis nielseni*. Acanthodida: Unnamed acanthodid. Tetrapoda: *Acanthostega gunnari*, *Ichthyostega stensioei*, Tetrapoda nov. gen. et. sp.

Reference. 160.

34. Britta Dal

Stratigraphy/Location. Britta Dal Formation, Celsius Bjerg Group, East Greenland. **Age.** Late Famennian. **Environment.** Non-marine, fluvial.

Taxa. Porolepiformes: *Holoptychius sp.* Dipnoi: *Jarvikia arctica*, *Oervigia nordica*, Unnamed dipnoan. Tristichopteridae: *Eusthenodon waengsjoei*. Arthrodira: *Groenlandaspis mirabilis*, Unnamed arthrodire 1, Unnamed arthrodire 2 Antiarcha: *Remigolepis kochi*, *Remigolepis cristata*, *Remigolepis acuta*, *Remigolepis incisa*. Elasmobranchii: Unnamed elasmobranch. Acanthodida: Unnamed acanthodid. Tetrapoda: *Acanthostega gunnari*, *Ichthyostega eigili*, Tetrapoda nov. gen. et. sp.

Reference. 160.

35. Grenfell

Stratigraphy/Location. Hunter Siltstone, Hervey Group, Grenfell, New South Wales, Australia. **Age.** Late Famennian. **Environment.** Non-marine, lacustrine.

Taxa. Dipnoi: *Adelargo schultzei*. Osteolepididae: *Grenfellia meemanae*. Tristichopteridae: *Eusthenodon gavini*, *Yambira thomsoni*. Arthrodira: *Groenlandaspis sp.* Antiarcha: *Bothriolepis grenfellensis*, *Remigolepis redcliffensis*, *Grenfellaspis branagani*. Acanthodida: *Grenfellacanthus zerinae*.

References. 189, 190, 191, 192, 193.

36. Hyner

Stratigraphy/Location. Red Hill, Duncannon Member, Catskill Formation, Hyner, Pennsylvania. **Age.** Late Famennian. **Environment.** Non-marine, fluvial.

Taxa. Actinopterygii: *Limnomis delaneyi*. Porolepiformes: *Holoptychius sp.* Dipnoi: Unnamed dipnoan 1, Unnamed dipnoan 2. Osteolepididae: *Glyptopomus sayerei* Tristichopteridae: *Hyneria lindae*. Megalichthyidae: Megalichthyid sp. nov. Rhizodontidae: Rhizodontidae gen. et sp. nov., *Sauripterus taylori*. Arthrodira: *Groenlandaspis pennsylvanica*, *Turrispis elektor*. Antiarcha: *Bothriolepis minor*,

Bothriolepis nitida. Phyllolepidia: *Phyllolepis rossimontina*, *Phyllolepis concentrina*, *Phyllolepis woodwardi*, *Phyllolepis nielseni*, *Phyllolepis undulata*. Elasmobranchii: *Ageolodus pectinatus*, *Ctenacanthus nodocostatus*. Gyracanthida: *Gyracanthus sherwoodi*. Tetrapoda: *Hynerpeton bassetti*, *Densignathus rowei*.

References. 27, 39, 75, 163, 194, 195, 196, 197, 198.

37. Evieux

Stratigraphy/Location. Evieux Formation, Famenne, Belgium. **Age.** Late/end Famennian. **Environment.** Non-marine, coastal alluvial plain.

Taxa. Actinopterygii: *Osorioichthys marginus*. Porolepiformes: *Holoptychius sp.*, *Glyptolepis sp.* Dipnoi: *Jarvikia sp.*, *Soederberghia sp.* Osteolepididae: *Glyptopomus sp.* Tristichopteridae: *Langliera socqueti*, *Eusthenodon waengsjoei*, *Platycephalichthys sp.* Megalichthyidae: *Megalichthyes sp.* Rhizodontida: *Strepsodus sp.* Arthrodira: *Dunkleosteus belgicus*, *Ardennosteus ubaghsi*, *Deirosteus omaliusi*, *Groenlandaspis thorezi*, Unnamed coccosteid. Antiarcha: *Bothriolepis lohesti*, *Bothriolepis sp.* Phyllolepidia: *Phyllolepis konincki*. Elasmobranchii: *Goodrichthys sp.* Ischnacanthida: *Onchus latus*. Tetrapoda: Unnamed Ichthyostegid.

References. 27, 199, 200, 201, 202, 203, 204, 205, 206.

38. Ketleri

Stratigraphy/Location. Ketleri outcrop, Ketleri Formation, Venta River, Latvia. **Age.** Late/end Famennian. **Environment.** Non-marine, shallow tidal.

Taxa. Onychodontida: *Onychodus sp.* Porolepiformes: *Holoptychius nobilissimus*. Dipnoi: *Orlovichthys limnatus*, *Latvius obrutus*. Osteolepididae: *Glyptopomus bystrowi*, *Cryptolepis grossi*. Tristichopteridae: *Ventalepis ketleriensis*. Arthrodira: *Stenosteus sp.* Antiarcha: *Bothriolepis ciecere*. Acanthodida: *Devononchus ketleriensis*, *Devononchus tenuispinus*. Tetrapoda: *Ventastega curonica*.

References. 27, 28, 32, 36, 184, 185, 195, 207, 208.

39. Tafilalt

Stratigraphy/Location. Tafilalt Basin Limestone, Anti-Atlas Mountains, Southern Morocco. **Age.** Late/end Famennian. **Environment.** Marine, open marine.

Taxa. Actinopterygii: Unnamed actinopterygian. Tristichopteridae: Unnamed eusthenopterid. Actinistia: Unnamed actinistian. Arthrodira: *Dunkleosteus marsaisi*, *Tafilalichthys lavocati*, *Titanichthys termieri*. Elasmobranchii: *Cobelodus sp.*, *Thrinacodus tranquillus*, *Protacrodus*, *Jalodus sp.*, *Siamodus janvieri*. Symmoriformes: *Stethacanthus resistens*, *Denaea sp.*

References. 27, 31, 183, 209, 210, 211, 212.

40. Andreyevka-2

Stratigraphy/Location. Andreyevka-2, Khovanshshina Beds, Zovalzkh Formation, Tula, Siberia. **Age.** Late/end Famennian. **Environment.** Nonmarine, estuarine, epicontinental brackish basin.

Taxa. Actinopterygii: *Moythomasia* sp. Onychodontida: *Strunius* sp. Porolepiformes: *Holoptychius nobilissimus*. Dipnoi: *Andreyevichthys epitomus*. Osteolepididae: *Chrysolepis orlensis*. Tristichopteridae: *Eusthenodon* sp. nov. Megalichthyidae: *Megistolepis* sp. Antiarcha: *Remigolepis armata*, *Bothriolepis* sp. Holocephali: Unnamed eugeneodont. Ischnacanthida: *Onchus latus*. Acanthodida: *Devononchus laevis*, *Devononchus concinnus*, *Cheiracanthus* sp. Tetrapoda: *Tulerpeton curtum*.

References. 27, 28, 36, 213, 214, 215.

41. Cleveland Shale

Stratigraphy/Location. Cleveland Shale Member, Ohio Shale, Cleveland, Ohio. **Age.** Late/end Famennian. **Environment.** Marine, open marine.

Taxa. Actinopterygii: *Tegeolepis clarki*, *Kentuckia hlavini*. Dipnoi: *Proceratodus wagneri*. Arthrodira: “*Hoplonchus*” sp., *Coccosteus cuyahogae*, *Gorgonichthys clarki*, *Heintzichthys gouldii*, *Holdenius holdeni*, *Hussakofia minor*, *Gymnotrachelus hydei*, *Paramylostoma arcualis*, *Selenosteus brevis*, *Stenosteus angustopectus*, *Stenosteus glaber*, *Mylostoma variable*, *Mylostoma eurhinus*, *Mylostoma newberryi*, *Titanichthys agassizi*, *Titanichthys attenuatus*, *Titanichthys clarkii*, *Titanichthys hussakofi*, *Titanichthys rectus*, *Titanichthys* sp., *Bungartius perissus*, *Callognathus regularis*, *Diplognathus mirabilis*, *Glyptaspis verrucosa*, *Trachosteus clarki*, *Dinognathus eurhinus*, *Dunkleosteus terrelli*, *Brontichthys clarki*. Elasmobranchii: *Cladoselache fyleri*, *Cladoselache clarki*, *Cladoselache kepleri*, *Cladoselache pattersoni*, *Ctenacanthus compressus*, *Ctenacanthus clarki*, *Tamiobatus vestustus*, *Phoebodus heslerorum*, *Diademodus hydei*, *Diademodus utahensis*, *Orodus* sp., *Phoebodus politus*. Symmoriformes: *Stethacanthus* sp.

References. 27, 75, 148, 165, 216, 217, 218, 219, 220.

42. Chaffee

Stratigraphy/Location. Fossil Ridge, Parting Formation, Chaffee Group, Gunnison County, Colorado. **Age.** Late/end Famennian. **Environment.** Marine, shallow continental shelf.

Taxa. Onychodontida: Unnamed onychodont. Porolepiformes: *Holoptychius giganteus*. Dipnoi: *Dipterus johnsoni*, *Conchodus parvulus*, *Soederberghia* sp. Osteolepididae: *Gyroptychius* sp., *Litoptychius bryanti*. Arthrodira: *Aspidichthys* sp., *Dinichthys tuberculatus*, Unnamed arthrodire. Antiarcha: *Bothriolepis coloradensis*. Elasmobranchii: *Ctenacanthus* sp. Holocephalii: *Sandalodus minor*.

References. 27, 221, 222, 223, 224, 225.

43. Witpoort

Stratigraphy/Location. Witpoort Formation, Witteberg Group, Cape Supergroup, Grahamstown, South Africa. **Age.** Late/end Famennian. **Environment.** Non-marine, estuarine, brackish lagoon.

Taxa. Actinopterygii: Unnamed actinopterygian. Dipnoi: *Andreyevichthys* sp. Tristichopteridae: *Eusthenodon* sp. Actinistia: *Diplocercides* sp.. Arthrodira: *Groenlandaspis riniensis*, *Africanaspis doryssa*, *Tiaraspis* sp. Antiarcha: *Bothriolepis africana*. Ptyctodontida: Unnamed ptyctodontid. Other Placodermi: Unnamed macropetalichthyid. Elasmobranchii: *Plesioselachus macracanthus*. Climatiida: *Diplacanthus acus*. Gyracanthida: *Gyracanthides warreni*. Tetrapoda: Unnamed elpistostegelian.

References. 226, 227, 228, 229, 230, 231.

D. Tournaisian Sites

44. Horton Bluff

Stratigraphy/Location. Horton Bluff and Albert Formations, Bay of Fundy, Canada. **Age.** Early/mid-Tournaisian. **Environment.** Non-marine, lacustrine with marine influence.

Taxa. Actinopterygii: *Elonichthys elli*, *Elonichthys browni*, *Elonichthys* sp., *Rhadinichthys alberti*, *Rhadinichthys* sp., *Acrolepis hortonensis*, *Palaeoniscus* sp. Dipnoi: Unnamed ctenodont. Rhizodontida: *Letognathus hardingi*. Symmoriformes: *Stethacanthus* sp. Gyracanthida: *Gyracanthides* sp. Acanthodidae: *Acanthodes* sp. Tetrapoda: Unnamed anthracosaur.

References. 232, 233, 234, 235, 236.

45. Izkchul Village

Stratigraphy/Location. Izkchul Village Fish Horizon, Lower Os'kin/Bystraya/Bystr'anskaya Formation, Minussinsk Depression, Siberia. **Age.** Early/mid-Tournaisian. **Environment.** Non-marine, brackish lagoon or lake.

Taxa. Actinopterygii: *Oxypteriscus minimus*, *Ministrella longicaudata*, *Palaeobergia microlepis*, *Bishofia lucida*, *Aetheretmon* sp., *Ganolepis gracilis*, *Gyrolepidotus schmidtii*, *Grassator bidens*. Megalichthyidae: *Rhizodopsis savenkovi*, *Greiserolepis minusensis*. Rhizodontida: *Pyncnotenion sibiriacus*. Elasmobranchii: *Ctenacanthus peregrinus*, *Cladodus* sp. Acanthodida: *Acanthodes lopatini*.

References. 29, 30, 39, 237, 238, 239, 240.

46. Mansfield

Stratigraphy/Location. Broken River fish fauna, Devil's Plain Formation, Mansfield Group, Mansfield Basin, Victoria, Australia. **Age.** Early/mid-Tournaisian. **Environment.** Non-marine, fluvial, floodplain.

Taxa. Actinopterygii: *Novogonatodus kasantsevae*, *Mansfieldiscus sweeti*, *Mansfieldiscus gibbus*. Dipnoi: *Delatitia breviceps*. Rhizodontida: *Barameda decipiens*. Elasmobranchii: *Tristychius sp.*, *Ctenacanthus sp.*, *Ageleodus altus*. Gyracanthida: *Gyracanthus murrayi*. Acanthodida: *Acanthodes australis*, *Eupleurogmus cresswelli*.

References. 71, 241, 242, 243, 244, 245

47. Foulden

Stratigraphy/Location. Foulden fish beds, Cementstone Group, Berwickshire, Scotland. **Age.** Mid/late Tournaisian. **Environment.** Non-marine, brackish alluvial, lacustrine.

Taxa. Actinopterygii: *Styracopterus fulcratus*, *Phanerosteon ovensi*, *Strepheoschema fouldensis*, *Aetheretmon valenticum*, *Cosmoptychius striatus*, *Rhadinichthys sp.* Dipnoi: Unnamed dipnoan. Rhizodontida: *Strepsodus aulaconamensis*, *Strepsodus nov. sp.* Actinistia: *Rhabdoderma sp.* Holocephalii: *Lophosteus sp.* Gyracanthida: *Gyracanthides sp.* Acanthodida: *Acanthodes ovensi*. Tetrapoda: Unnamed anthracosaur.

References. 6, 24, 246, 247, 248.

48. Herbesskaya

Stratigraphy/Location. Senek and Ortokhodan Creeks, Herbesskaya Formation, Shagonar City, Tuva, Western Siberia. **Age.** Late Tournaisian. **Environment.** Marine, shallow sea.

Taxa. Actinopterygii: *Senekichthys hirundo*, *Ganolepis sp.*, *Actinopterygii nov. gen. et sp.*, *Grassator sp.* Megalichthyidae: *Rhizodopsis savinkovi*. Rhizodontida: *Pycnotenion sibiriacus*. Elasmobranchii: *Psammodus linearis*, *Lagarodus angustus*, "*Ctenacanthus*" (*Hybodus*) *panderi*, *Cladodus sp.* Holocephali: *Helodus antiquissimus*, *Helodus contractus*, *Psephodus dentatus*. Acanthodida: *Acanthodes lopatini*.

References. 39, 147, 239, 249.

E. Viséan Sites

49. Waaipoort

Stratigraphy/Location. Waaipoort Formation, Lake Mentz Subgroup, Witteberg Group, Cape Supergroup, South Africa. **Age.** Early Viséan. **Environment.** Non-marine, lacustrine.

Taxa. Actinopterygii: *Adroichthys tuberculatus*, *Mentzichthys walshi*, *Mentzichthys jubbi*, *Mentzichthys maraisi*, *Mentzichthys theroni*, *Aestaurichthys fulcratus*,

Soetendalichthys cromptoni, *Australichthys longidorsalis*, *Willomorichthys striatulus*, *Sundayichthys elegantulus*, *Dwykia analensis*. Elasmobranchii: Unnamed protoacrodontid. Gyracanthida: *Gyracanthides* sp., *Gyracanthus* sp. Acanthodida: *Acanthodes* sp.

References. 230, 250, 251, 252.

50. Glencartholm

Stratigraphy/Location. Glencartholm Fish Bed, Dumfriesshire, Scotland. **Age.** Mid Viséan. **Environment.** Marine, nearshore marine.

Taxa. Actinopterygii: *Protoeurynotus traquairi*, *Paramesolepis tuberculatus*, *Platysomus superbus*, *Cheirodopsis geikiei*, *Rhadinichthys canobiensis*, *Rhadinichthys macconochii*, *Rhadinichthys fusiformis*, *Rhadinichthys formosus*, *Rhadinichthys tuberculatus*, *Rhadinichthys* sp., *Cycloptychius concentricus*, *Mesopoma politum*, *Mesopoma pulchellum*, *Mesopoma crassum*, *Mesopoma macrocephalum*, *Canobius ramseyi*, *Canobius elegantulus*, *Canobius* sp., *Elonichthys serratus*, *Elonichthys pulcherrimus*, *Acrolepis ortholepis*, *Phanerosteon mirabile*, *Holurus parki*, *Styracopterus fulcratus*, *Tarrasius problematicus*. Megalichthyidae: *Megalichthys* sp. Rhizodontida: *Archichthys portlocki*, *Strepsodus saurroides*. Actinistia: *Dumfregia huxleyi*. Elasmobranchii: *Goodrichichthys eskdalensis*, *Onychoselache traquairi*, *Sphenacanthus costellatus*. Holocephalii: *Deltoptychius armigerus*, *Antliodus scoticus*, *Chondrenchelys problematica*. Acanthodida: *Acanthodes nitidus*.

References. 1, 6, 10, 24, 39, 148, 253, 254, 255.

51. Burdiehouse

Stratigraphy/Location. Burdiehouse limestone, Edinburgh, Scotland. **Age.** Mid Viséan. **Environment.** Non-marine, lacustrine, alluvial.

Taxa. Actinopterygii: *Elonichthys robinsoni*, *Elonichthys bucklandi*, *Elonichthys striatulus*, *Rhadinichthys ornatissimus*, *Nematoptychius greenocki*, *Gonatodus punctatus*, *Eurynotus crenatus*, *Elonichthys ovatus*, *Elonichthys striolatus*. Dipnoi: *Uronemus lobatus*, *Ctenodus* sp. Megalichthyidae: *Megalichthys laticeps*. Rhizodontida: *Rhizodus hibberti*, *Screbinodus ornatus*. Elasmobranchii: *Cladodus hibberti*, *Cladodus parvus*, *Diplodus minutus*, *Ptyctacanthus sublaevis*, *Sphenacanthus serratulus*, *Ageolodus pectinatus*. Gyracanthida: *Gyracanthides rectus*. Acanthodida: *Acanthodes sulcatus*. Tetrapoda: *Pholidogaster pisciformis*, *Dolichopareias disjectins*.

References. 6, 24, 41, 148, 256, 257, 258, 259, 260.

52. Ducabrook (**Note:** Locality does not meet the dataset criteria, but is included here due to its high profile and importance to tetrapod workers)

Stratigraphy/Location. Middle Paddock Tetrapod Unit, Ducabrook Formation, Drummond Basin, Queensland, Australia. **Age.** Mid-Viséan. **Environment.** Non-marine, estuarine.

Taxa. Actinopterygii: *Elonichthys* sp., Unnamed actinopterygian. Dipnoi: *Ctenodus* sp. Rhizodontida: *Strepsodus* sp. Elasmobranchii: Unnamed xenacanth, Unnamed hybodont, *Ageolodus* sp., *Psammodus* sp. Gyracanthida: *Gyracanthides hawkinsi*. Tetrapoda: *Ossinodus pueri*.

References. 260, 261, 262, 263, 264, 265, 266.

53. Wardie

Stratigraphy/Location. Wardie Shales, Lower Lothian Group, Lothian, Scotland. **Age.** Late Viséan. **Environment.** Non-marine, brackish lagoon.

Taxa. Actinopterygii: *Cosmoptychius striatus*, *Nematoptychius greenocki*, *Elonichthys robinsoni*, *Elonichthys striatulus*, *Gonatodus punctatus*, *Eurynotus crenatus*, *Wardichthys cyclosoma*, *Rhadinichthys brevis*, *Rhadinichthys carinatus*, *Rhadinichthys ferox*, *Rhadinichthys orenatissimus*, *Chirodus* sp. Dipnoi: Unnamed dipnoan. Megalichthyidae: *Megalichthys* sp. Rhizodontida: *Rhizodus hibberti*. Elasmobranchii: *Onychoselache traquari*, *Sphenacanthus serrulatus*, *Ctenacanthus* sp., *Diplodoselache woodi*. Acanthodida: *Acanthodes sulcatus*. Tetrapoda: *Lethiscus stocki*.

Reference. 6.

54. Cheese Bay

Stratigraphy/Location. Cheese Bay Shrimp Bed, Gullane Formation, Lower Oil Shale Group, Calciferous Sandstone Series, East Lothian, Scotland. **Age.** Late Viséan. **Environment.** Non-marine, brackish pools.

Taxa. Actinopterygii: *Gonatodus punctatus*, *Elonichthys robinsoni*, *Elonichthys intermedius*, *Cosmoptychius striatus*, *Nematoptychius greenocki*, *Rhadinichthys brevis*, *Rhadinichthys elegantulus*, *Rhadinichthys formosus*, *Rhadinichthys ornatissimus*, *Wardichthys cyclosoma*. Rhizodontida: *Rhizodus hibberti*. Actinistia: *Rhabdoderma* sp. Acanthodida: *Acanthodes* sp. Tetrapoda: *Casineria kiddi*.

References. 6, 267.

55. East Kirkton

Stratigraphy/Location. East Kirkton Limestone, West Lothian Oil Shale Formation, Bathgate, Lothian, Scotland. **Age.** Late Viséan. **Environment.** Non-marine, lacustrine.

Taxa. Actinopterygii: *Elonichthys robinsoni*, *Cosmoptychius* sp., *Watsonichthys* sp., *Rhadinichthys carinatus*, *Mesopoma* sp., *Eurynotus* sp., Unnamed actinopterygian. Rhizodontida: Unnamed rhizodont. Elasmobranchii: *Tristychius arcuatus*, *Diplodocanthus woodi*. Gyracanthida: Unnamed gyracanthid. Acanthodida: *Acanthodes* sp. Tetrapoda: *Eucritta melanolimnetes*, *Balanerpeton woodi*, *Ophiderpeton kirktonense*, *Eldeceon rolfei*, *Silvanerpeton miripedes*, *Westlothiana lizziae*.

References. 6, 268, 269.

56. Gilmerton

Stratigraphy/Location. Gilmerton Blackband Ironstone, Gilmerton, Scotland. **Age.** Late Viséan. **Environment.** Non-marine, lacustrine, alluvial.

Taxa. Actinopterygii: *Elonichthys robinsoni*, *Pseudogonatodus macrolepis*, *Eurynotus crenatus*. Dipnoi: *Uronemus splendens*, *Ctenodus interruptus*, *Sagenodus quinquecostatus*. Megalichthyidae: *Megalichthys* sp. Rhizodontida: *Screbinodus ornatus*, *Rhizodus hibberti*. Holocephali: *Harpacanthus fimbriatus*. Acanthodida: *Acanthodes* sp. Tetrapoda: *Crassigyrinus scoticus*, *Eoherpeton watsoni*, *Loxomma allmanni*, *Pholidogaster pisciformis*.

References. 6, 24, 235, 259, 260, 269, 270, 271, 272.

57. Ardross Castle

Stratigraphy/Location. *Crangopsis* Beds, Calciferous Sandstone Series, Ardross Castle, Fife, Scotland. **Age.** Late Viséan. **Environment.** Marine, marine forms stranded in a saline lagoon.

Taxa. Actinopterygii: *Rhadinichthys carinatus*, *Rhadinoniscus wrighti*, *Elonichthys robinsoni*, *Watsonichthys pectinatus*, *Chirodus crassus*. Actinistia: *Rhabdoderma ardrossense*, “*Coelacanthopsis*” *curta*. Elasmobranchii: *Euphyacanthus semistriatus*. Holocephali: *Deltoptychius armigerus*, *Eucentrurus paradoxus*, Unnamed helodont. Acanthodida: *Acanthodes sulcatus*.

Reference. 6.

58. Abden

Stratigraphy/Location. Abden Bone Bed, Calciferous Sandstone Series, Fife, Scotland. **Age.** Late Viséan. **Environment.** Marine, open marine.

Taxa. Actinopterygii: *Chirodus crassus*, *Eurynotus crenatus*, *Nematoptychius greenocki*, *Elonichthys robinsoni*. Rhizodontida: *Archichthys (Strepsodus) portlocki*. Actinistia: *Diplocercides davisii*. Elasmobranchii: *Tristychius arcuatus*. Holocephalii: *Deltoptychius armigerus*. Acanthodida: *Acanthodes* sp.

References. 6, 24, 235, 272.

59. Inchkeith

Stratigraphy/Location. Inchkeith, Upper Oil Shale Group, Firth of Forth, Fife, Scotland. **Age.** Late Viséan. **Environment.** Non-marine, lacustrine.

Taxa. Actinopterygii: *Pseudogonatodus* sp., *Elonichthys robinsoni*, *Nematoptychius* sp., *Eurynotus* sp. Dipnoi: *Sagenodus* sp., *Uronemus* sp. Rhizodontida: *Strepsodus* sp., *Rhizodus hibberti*. Elasmobranchii: *Ageleodus* sp., *Cynopodius crenulatus*, *Pleuracanthus* sp. Gyracanthida: *Gyracanthus* sp. Tetrapoda: *Doragnathus woodi*, *Palaeomolgophis*.

References. 6.**F. Serpukhovian Sites****60. Bearsden**

Stratigraphy/Location. Manse Burn Formation, Bearsden, Scotland. **Age.** Early Serpukhovian. **Environment.** Marine, coastal.

Taxa. Actinopterygii: *Mesopoma carricki*, *Mesopoma smithsoni*, *Mesopoma pancheni*, *Frederichthys musadentatus*, *Melanecta anneiae*, *Woodichthys bearsdeni*, *Chirodus crassus*, Actinopterygii gen. et sp. nov., *Watsonichthys* sp., Unnamed actinopterygian 1 (MIC pers. obs.), Unnamed actinopterygian 2 (MIC pers. obs.). Rhizodontida: Unnamed rhizodont. Actinistia: Unnamed coelacanth. Elasmobranchii: *Tristychius arcuatus*, *Onychoselache traquairi*. Symmoriformes: *Akmonistion zangerli* Holocephali: *Deltoptychius* sp., Unnamed menaspid. Acanthodida: *Acanthodes sulcatus*.

References. 6, 273, 274, 275, 276, 277.

61. Chokier

Stratigraphy/Location. Chokier Formation, Belgium. **Age.** Early Serpukhovian. **Environment.** Marine, open marine.

Taxa. Actinopterygii: *Elonichthys robinsoni*, *Mesonichthys aitkeni*, *Rhadinichthys raineri*. Actinistia: *Rhabdoderma corneti*, *Rhabdoderma stensioei*. Holocephali: *Physonemus konincki*.

References. 205, 206.

62. Loanhead

Stratigraphy/Location. Burghlee Ironstone, Limestone Coal Group, Loanhead, Midlothian, Scotland. **Age.** Mid Serpukhovian. **Environment.** Non-marine, lacustrine.

Taxa. Actinopterygii: *Cryphiolepis striatus*, *Drydenius insignis*, *Elonichthys robinsoni*, *Protohaplolepis scotica*, *Nematoptychius greenocki*, *Pseudogonatodus parvidens*, *Eurynotus crenatus*, *Blairolepis loanheadensis*, *Watsonichthys pectinatus*. Dipnoi: *Ctenodus* sp., *Uronemus splendens*, *Sagenodus quinquecostatus*. Megalichthyidae: *Rhizodopsis* sp. Rhizodontidae: *Rhizodus hibberti*, *Screbinodus ornatus*, *Strepsodus sauroides*, *Strepsodus striatulus*. Elasmobranchii: *Pleuracanthus gracillimus*, *Pleuracanthus horridulus*, *Pleuracanthus elegans*, *Cynopodius crenulatus*, *Dicentrodus bicuspidatus*, *Diplodus parvulus*. Holocephali: *Euctenius elegans*, *Euctenodopsis tenuis*. Gyracanthida: *Aganacanthus striatulus*, *Gyracanthus nobilis*, *Gyracanthus youngi*. Acanthodida: *Acanthodes* sp. Tetrapoda: *Caerorhachis bairdi*, Unnamed aistopod, *Eoherpeton watsoni*, *Acheroniscus caledoniae*, *Adelogyrinus* sp., *Doragnathus woodi*, *Loxomma* sp., *Papposaurus traquair*, *Spathicephalus mirus*.

References. 6, 24, 28, 41, 148, 259, 260, 262, 271, 277, 278, 279, 280, 281, 282.

63. Niddrie

Stratigraphy/Location. Niddrie Colliery Shale overlying South Parrot Coal Seam, Upper Limestone Group, Niddrie, Lothian, Scotland. **Age.** Mid Serpukhovian.

Environment. Non-marine, lacustrine, coal swamp.

Taxa. Dipnoi: *Ctenodus interruptus*. Rhizodontida: *Strepsodus sauroides*, *Rhizodus hibberti*, *Archichthys sulcidens*. Gyracanthida: *Gyracanthus youngi*. Tetrapoda: *Adelogyrinus* sp., *Doracanthus woodi*. Unnamed anthracosaur.

References. 6, 260, 262, 281.

64. Millstone Grit

Stratigraphy/Location. Millstone Grit, Summit, Lancashire, England. **Age.** Late Serpukhovian. **Environment.** Marine, coastal marine.

Taxa. Actinopterygii: *Rhadinichthys planti*, *Rhadinichthys microdon*, *Rhadinichthys monensis*, *Rhadinichthys elegans*, *Elonichthys aitkeni*, *Elonichthys caudalis*, *Elonichthys oblongus*, *Acrolepis hopkinsi*, *Elonichthys obliquus*, *Elonichthys ornatus*, *Platysomus* sp. Dipnoi: *Sagenodus inequalis*. Megalichthyidae: *Megalichthys hibberti*, *Rhizodopsis sauroides*. Rhizodontida: *Archichthys sulcidens*. Actinistia: “*Coelacanthus*” sp. nov. Elasmobranchii: *Cladodus mirabilis*, *Acondylacanthus* sp., *Euctenodopsis tenuis*, *Ctenacanthus major*. Symmoriiformes: *Falcatus falcatus*. Holocephalii: *Psephodus minuta*, *Poecilodus jonesii*, *Janassa linguaeformis*. Gyracanthida: *Marsdenius summiti*, *Oracanthus milleri*. Acanthodida: *Acanthodes wardi*, *Acanthodes striatus*.

References. 283, 284, 285, 286.

65. Dora

Stratigraphy/Location. Dora Bone Bed, Limestone Bone Bed, Cowdenbeath, Fife, Scotland. **Age.** Late Serpukhovian. **Environment.** Non-marine, lacustrine, coal swamp.

Taxa. Actinopterygii: *Psuedogonatodus* sp. Rhizodontida: *Strepsodus* sp. Elasmobranchii: *Dicentrodus bicuspidatus*, *Diplozelache parvulus*, *Xenacanthus elegans*. Gyracanthida: *Gyracanthus young*, *Aganacanthus striatulus*. Tetrapoda: *Eoherpeton watsoni*, *Proterogyrinus* sp., *Crassigyryrinus scoticus*, *Doragnathus woodi*, *Adelogyrinus* sp., *Spathicephalus mirus*, Unnamed anthracosaur.

References. 6, 42, 235, 260, 281, 287, 288.

66. Bear Gulch

Stratigraphy/Location. Bear Gulch Limestone, Heath Formation, Big Snowy Group, Montana. **Age.** Late Serpukhovian. **Environment.** Marine, reef.

Taxa. Actinopterygii: *Paratarrasius hibbardi*, *Kalops monophrys*, *Kalops diophrys*, Unnamed tarrasiiform, *Aesopichthys erinaceus*, *Proceramala montanensis*, *Cyranorhis bergeraci*, *Discoserra pectinodon*, *Wendichthys dicksoni*, *Wendyichthys lautreci*,

Guildayichthys carnegiei, Unnamed actinopterygian “*copiosus*”, Unnamed actinopterygian “*brevis*”, Unnamed actinopterygian “*prolatus*”, *Cornuboniscus sp.*, Unnamed actinopterygian “*gulo*”, *Phanerosteon sp.*, Unnamed actinopterygian “*lundae*”, Unnamed actinopterygian “*worm tail*”, Unnamed actinopterygian “*round tail*”, Unnamed actinopterygian “*spike*”, Unnamed actinopterygian “*BGLP*”, Unnamed actinopterygian “*big antorbital*”, Unnamed actinopterygian “*deep belly*”, Unnamed actinopterygian “*short vertical cheek*”, Unnamed actinopterygian “*nude cheek*”, Unnamed actinopterygian “*horizontal cheek*”, Unnamed actinopterygian “*fat fish*”, Unnamed actinopterygian “*deep scale*”, Unnamed actinopterygian “*long pelvic*”, Unnamed actinopterygian “*barred scale*”, Unnamed actinopterygian “*tube bod*”, Unnamed actinopterygian “*pseudo*”, *Beagiascus pulcherrimus*, *Lineagruan snowyi*, *Lineagruan judithi*, *Platysomus sp.* Rhizodontida: *Strepsodus sp.* Actinistia: *Allenypterus montanus*, *Caridosuctor populosum*, *Hadronector donbairdi*, *Lochmocercus aciculodontus*, *Polyosteorhynchus beargulchensis*, *Actinistia* gen. et sp. nov. Elasmobranchii: *Heteropetalus elegantulus.*, *Squatactinus montanus*, *Orodus daedalus*, *Orodus springeri*, *Orodus sp.*, *Thrinacoselache gracia*, *Listracanthus sp.*, Symmoriiformes: *Damocles serratus*, *Falcatus falcatus*, *Orestiacanthus fergusi*, *Stethacanthus altonensis*, *Stethacanthus productus*. Holocephali: *Bellbonn rogaire*, *Gregorius rexi*, *Srianta dawsoni*, *Srianta iarlis*, *Srianta srianta*, *Debeerius ellefseni*, *Belantsea montana*, *Ctenopetalus serratus*, *Janassa clarki*, *Netsepoye hawesi*, *Petalorhynchus beargulchensis*, *Polyrhizodus digitatus*, *Sikisika ottae*, *Venustodus arguatus*, *Harpacanthus fimbriatus*, *Lisgodus sp.*, *Delphyodontos dacrifomes*, *Harpagofututor volsellorhinus*, *Echinochimaera meltoni*, *Echinochimaera snyderi*, *Traquairius agkistrocephalus*, *Traquairius nudus*, *Traquairius nitidus*, *Rainerichthys zengeri*, *Papillionichthys stahlae*. Acanthodida: *Acanthodes lundi*.

References. 44, 289, 290, 291, 292, 293, 294, 295, Carnegie Museum of Natural History Collections.

IV. Supporting Figures

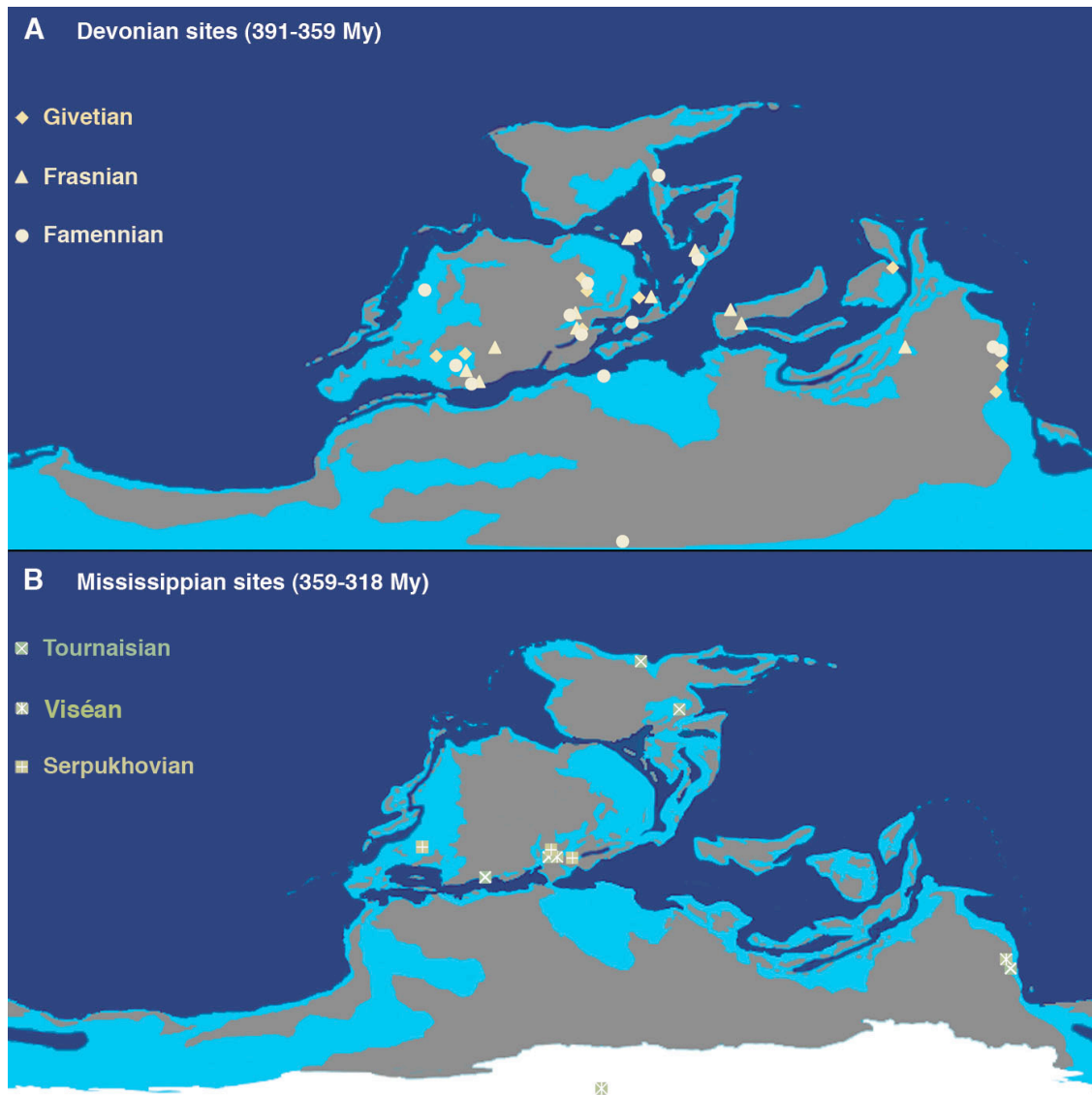


Fig. S1. Paleogeography of sampled vertebrate sites (N=66). (A) Devonian (~370 Ma, N=43) (B) Mississippian (~340 Ma, N=23). (After Blakey, 2007).

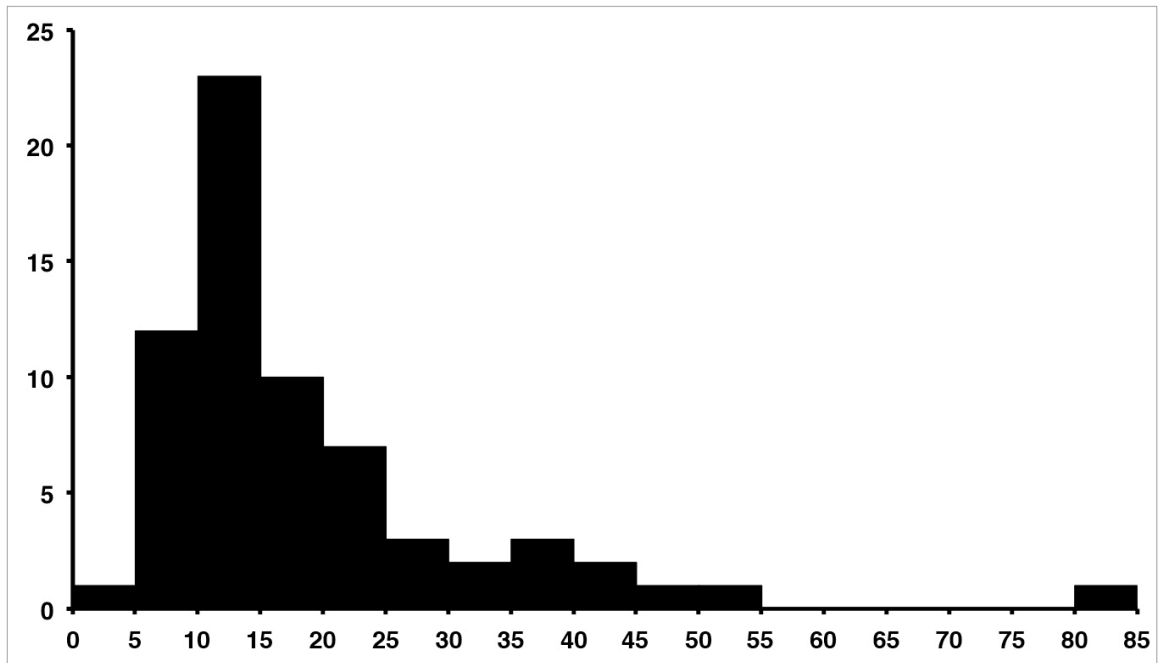


Fig. S2. Histogram of site species-level diversity (N=66, Mean=19.27, SD= 13.10, Median=15). See Table S1 for more details.

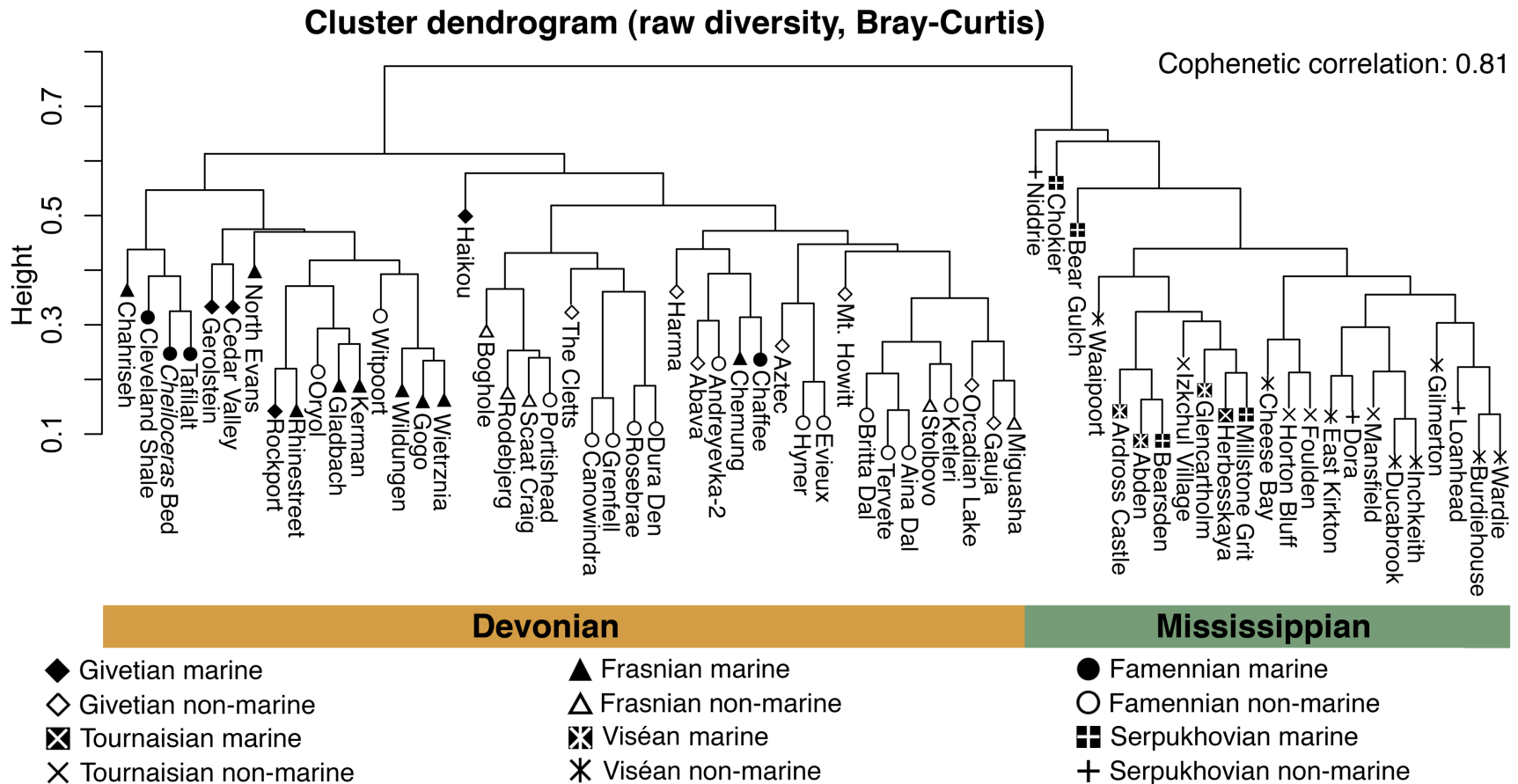


Fig. S3. Cluster dendrogram: Bray-Curtis, raw diversity. Cluster dendrogram of 66 vertebrate sites generated from the raw diversity dataset (Table S1). Sites are coded according to stage and environment. Bars mark clusters associated with geologic period.

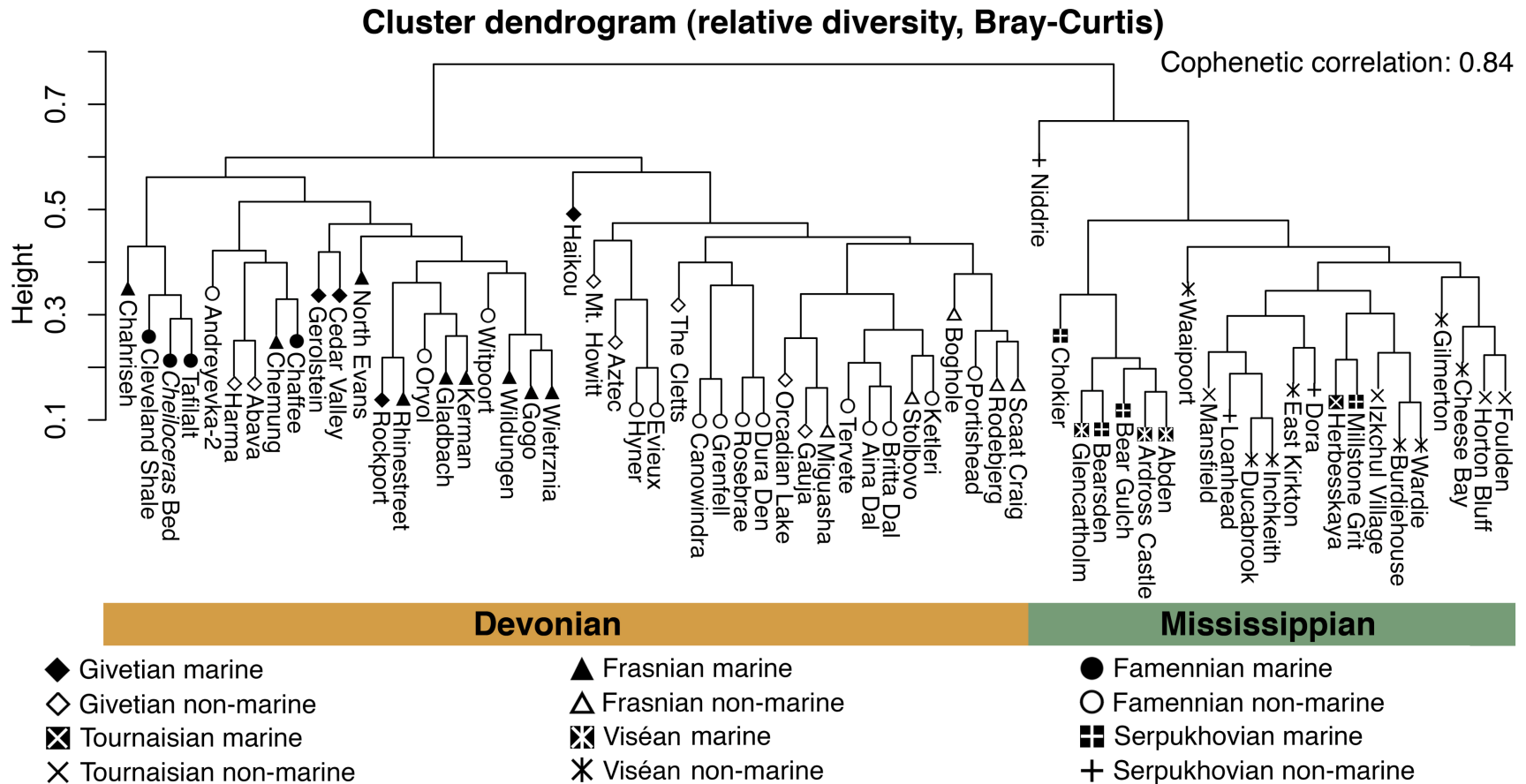


Fig. S4. Cluster dendrogram: Bray-Curtis, relative diversity. Cluster dendrogram of 66 vertebrate sites generated from the relative diversity dataset (Table S2). Sites are coded according to stage and environment. Bars mark clusters associated with geologic period.

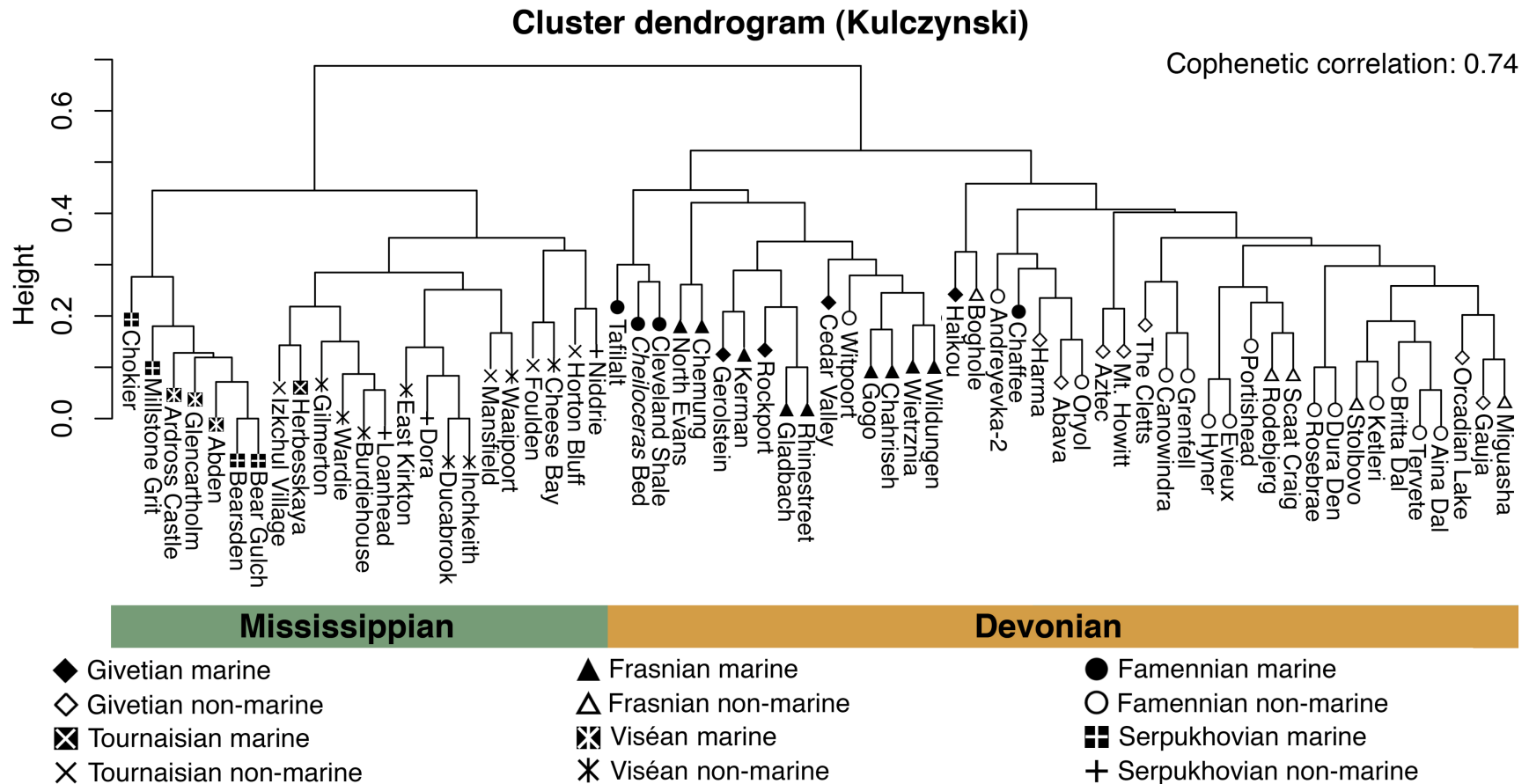


Fig. S5. Cluster dendrogram: Kulczynski similarity. Cluster dendrograms generated from raw and relative diversity were identical since they were constructed from a presence-absence metric. Sites are coded according to stage and environment. Bars mark clusters associated with geologic period.

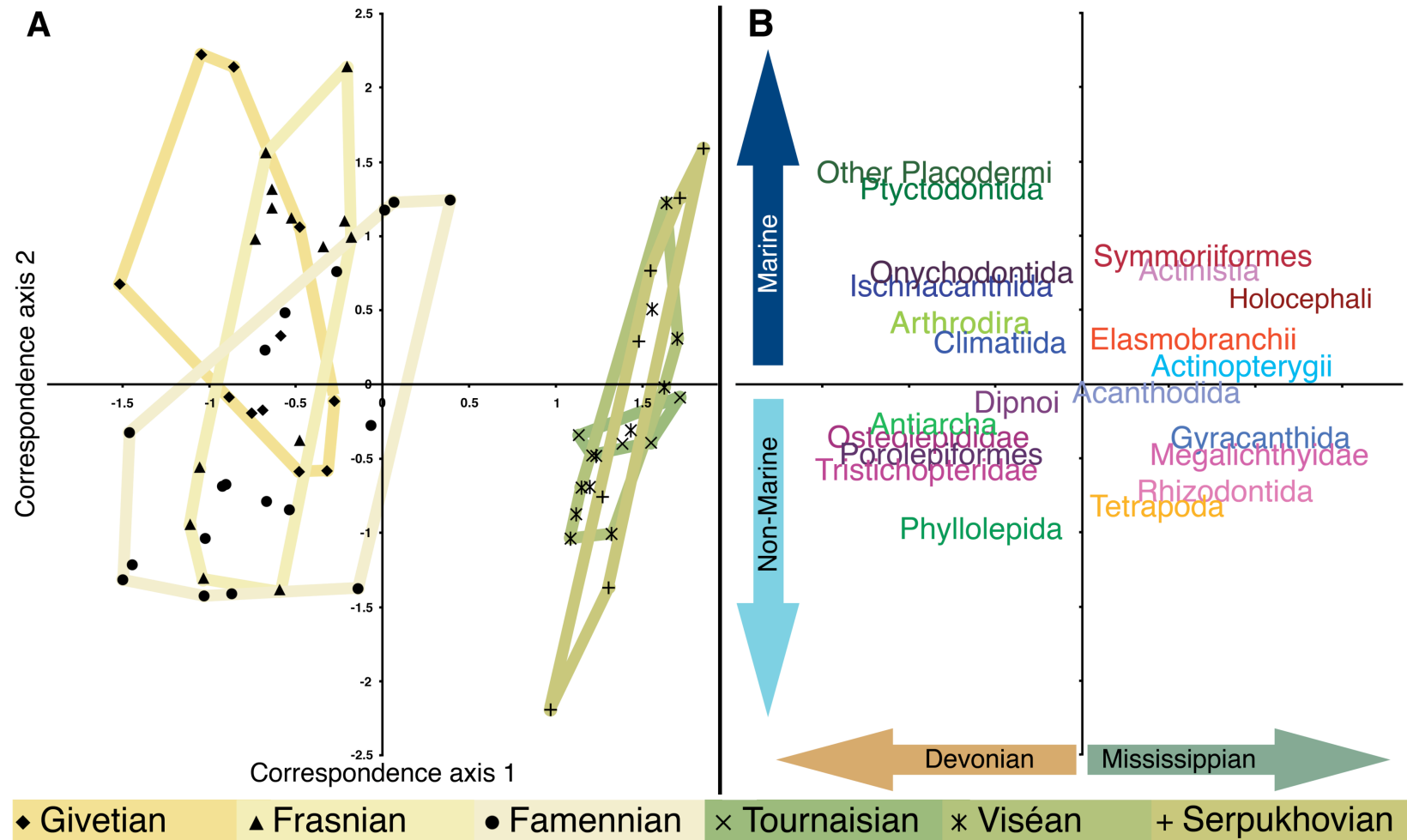


Fig. S6. CCA of relative diversity. (A) Site ordination plot (N=66). (B) taxon ordination and axis gradients (N=22). See Tables S7 and S8 for scores.

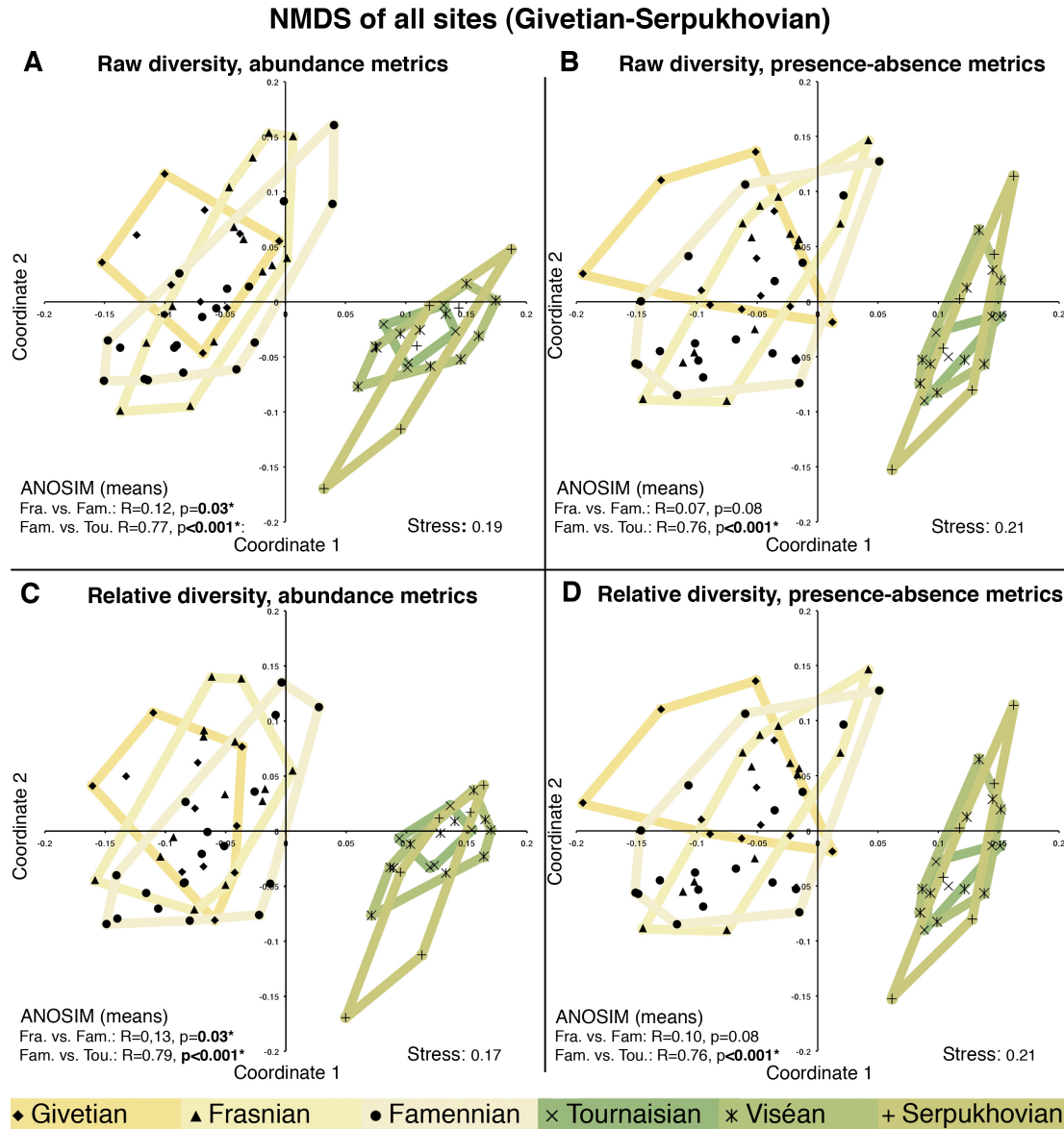


Fig. S7. NMDS plots of all sites. (A) NMDS plot generated from the raw diversity dataset based on Bray-Curtis distances ($N=66$). ANOSIM p-values for stages surrounding extinction events are the mean of those produced by all abundance-based distance measures from the appropriate dataset. Stress is a measure of difference between the rank-order of sites in the distance matrix and the rank-order generated by NMDS. This panel also appeared as text Fig. 4A, but is placed here for comparison to other results. (B) NMDS plot generated from the raw diversity dataset based on Kulczynski similarity ($N=66$). ANOSIM p-values for stages surrounding extinction events are the mean of those produced by all presence-absence-based distance measures from the appropriate dataset. However, the ANOSIM p-values are not, showing some further variability in the Frasnian-Famennian signal. (C) NMDS plot generated from the relative diversity dataset based on Bray-Curtis distances ($N=66$). (D) NMDS plot generated from the relative diversity dataset based on Kulczynski similarity. It is identical to S7B ($N=66$).

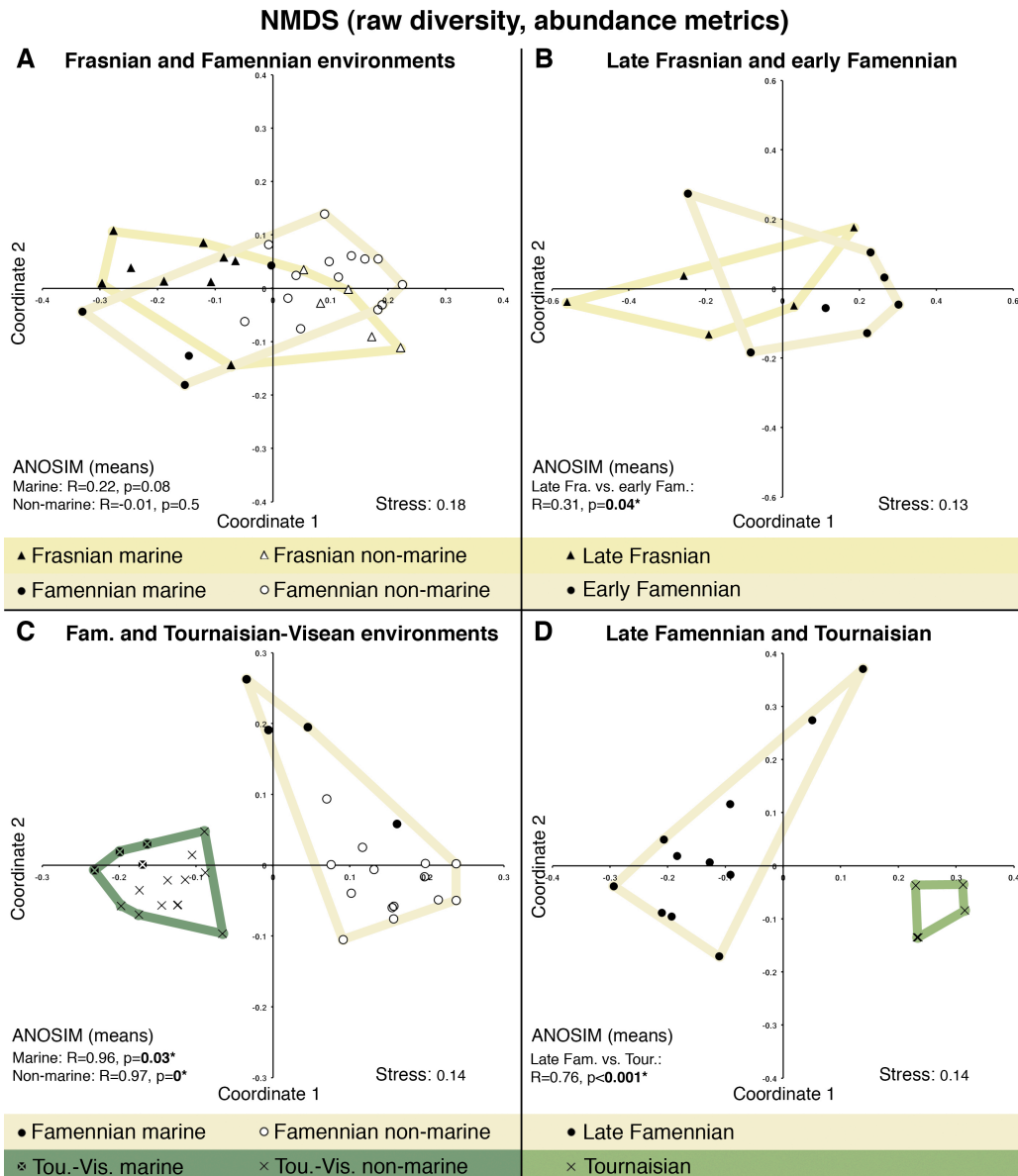


Fig. S8. NMDS of restricted intervals: Bray-Curtis distances, raw diversity. ANOSIM values are the mean of all abundance metric values (Tables S11 and S12). (A) NMDS plot of Frasnian and Famennian sites marked by environment (N=32). This panel also appeared as Fig. 4C, and is included here for comparison (B) NMDS plot of late Frasnian and early Famennian sites (N=13). (C) NMDS plot of Famennian and Tournaisian-Viséan sites marked by environment (N=34). (D) NMDS plot of late Famennian and Tournaisian sites (N=16). This panel also appeared as Fig. 4D and is included here for comparison.

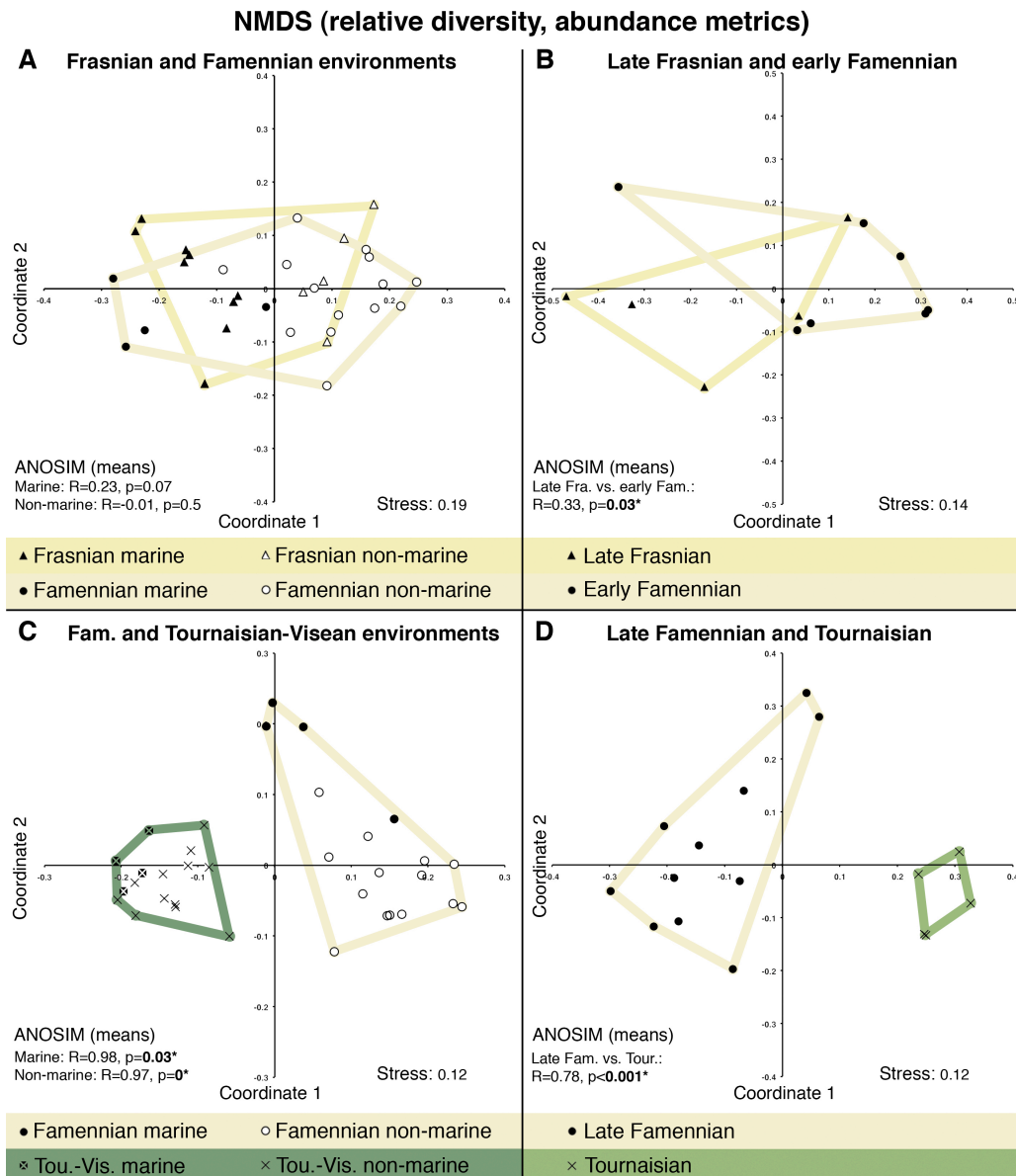


Fig. S9. NMDS of restricted intervals: Bray-Curtis distances, relative diversity. ANOSIM values are the mean of all abundance metric values (Tables S15 and S16). (A) NMDS plot of Frasnian and Famennian sites marked by environment (N=34). (B) NMDS plot of late Frasnian and early Famennian sites (N=13). (C) NMDS plot of Famennian and Tournaisian-Viséan sites marked by environment (N=34). (D) NMDS plot of late Famennian and Tournaisian sites (N=16).

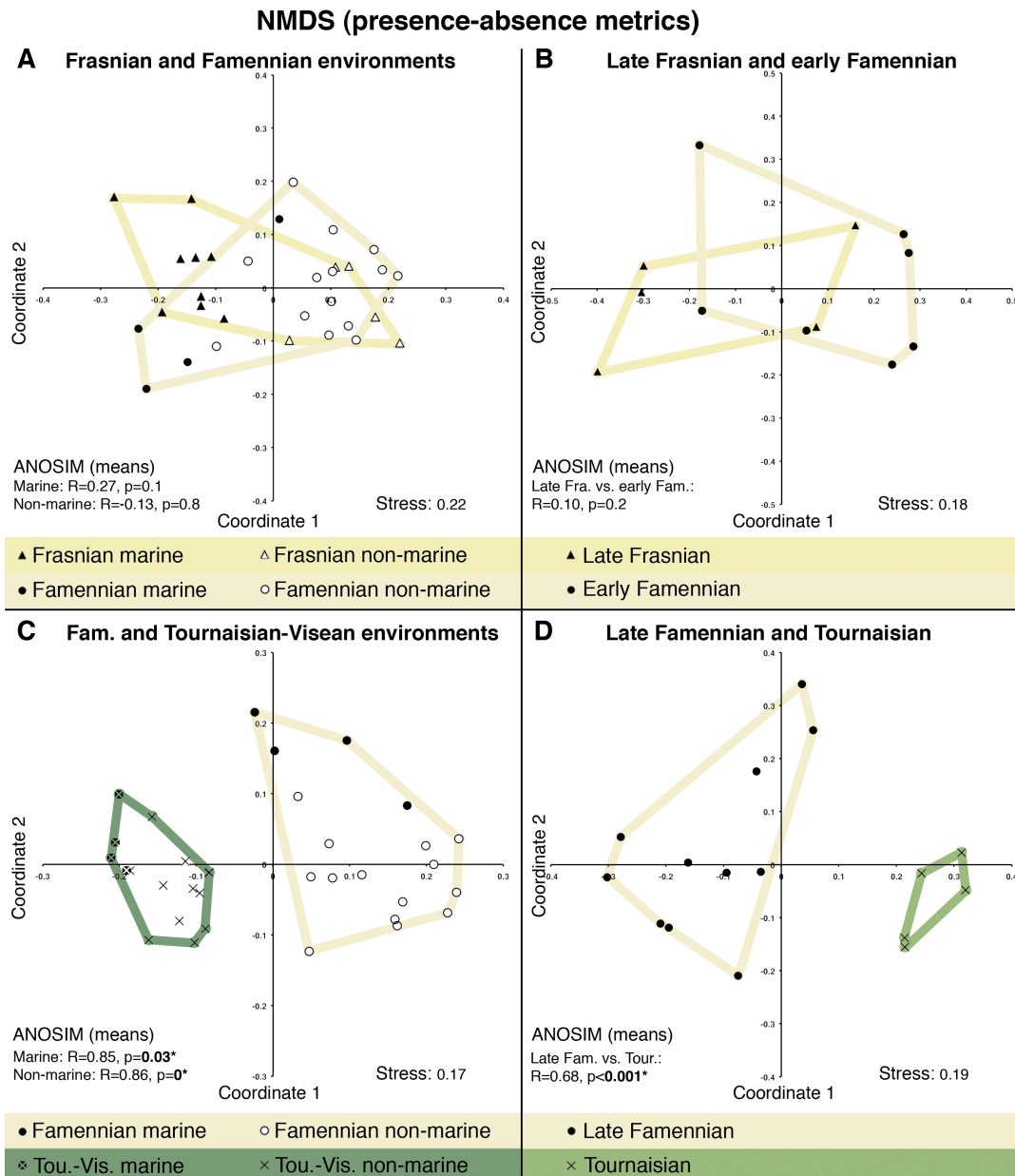


Fig. S10. NMDS of restricted intervals: Kulczynski similarity. ANOSIM values are the mean of all presence-absence metric values from raw diversity. Raw and relative diversity produced identical NMDS plots and similar if not identical ANOSIM values (Tables S11, S12, S15, S16). (A) NMDS plot of Frasnian and Famennian sites marked by environment (N=34). (B) NMDS plot of late Frasnian and early Famennian sites (N=13). (C) NMDS plot of Famennian and Tournaisian-Viséan sites marked by environment (N=36). (D) NMDS plot of late Famennian and Tournaisian sites (N=16).

Factor Analysis Plots

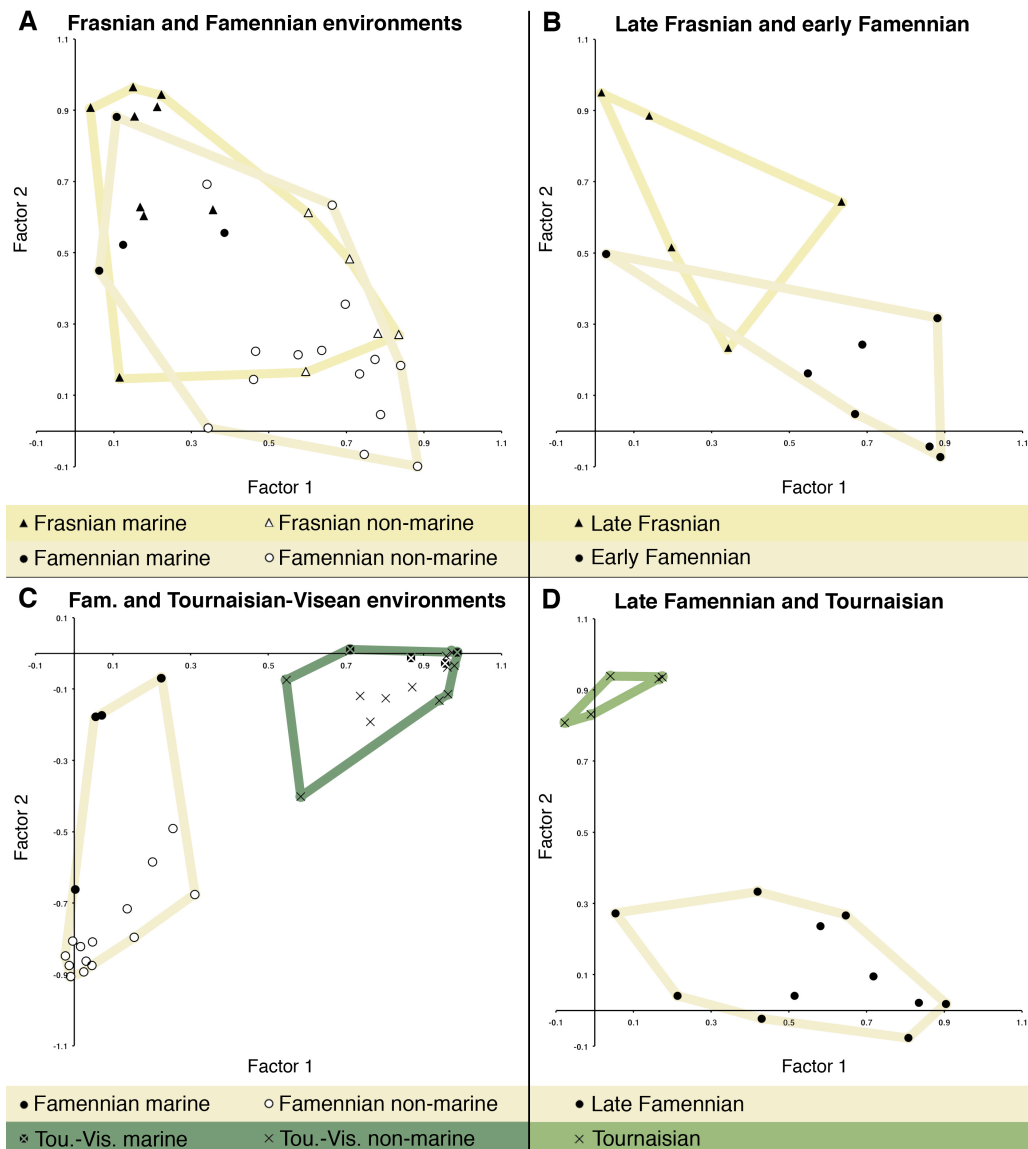


Fig. S11. Factor analysis of restricted intervals (A) Frasnian and Famennian sites marked by environment (N=32) (Tables S24 and S25). Factor 1 (55.17% of variance) is positively loaded by Devonian non-marine taxa as indicated in the CCA plot (Fig. 3B). Factor 2 (16.92% of variance), is positively loaded by Devonian marine taxa. Site placement also displays some paleogeographic signal. (B) Late Frasnian and early Famennian sites (N=13) (Tables S36 and S37). Factors 1 (56.84% of variance) and 2 (19.86% of variance) are highly similar to those generated in analysis of all Frasnian and Famennian sites. (C) Famennian and Tournaisian-Viséan sites marked by environment (N=34) (Tables S28 and S29). Factors are very similar to those produced by factor analysis of all sites (Figs. 4B), except factors 1 (42.92% of variance) and 2 (5.88% of variance) are switched. (D) Late Famennian and Tournaisian sites (N=16) (tables S40 and S41). Factor 1 (43.51% of variance) is positively loaded by Devonian non-marine forms, just as in the overall factor analysis plot (Fig. 4B). Factor 2 (22.77%) is strongly loaded by actinopterygians and other Mississippian taxa.

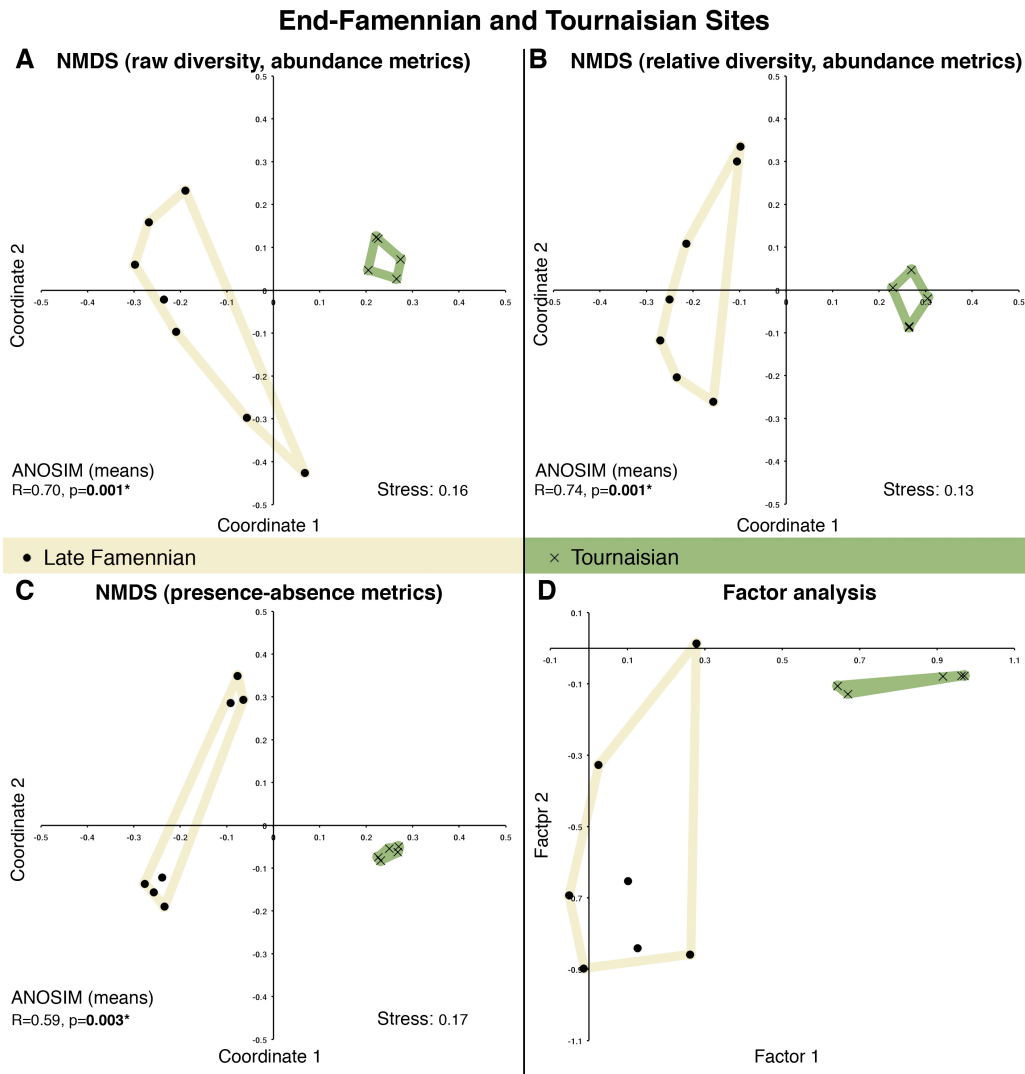


Fig. S12. Ordination plots of end-Famennian and Tournaisian sites ($N=12$). (A) NMDS plot of end-Famennian and Tournaisian relative site diversities based on Bray-Curtis distance. (B) NMDS plot of end-Famennian and Tournaisian raw site diversities based on Bray-Curtis distance (C) NMDS plot of end-Famennian and Tournaisian raw and relative site diversities based on Kulczynski similarity. (D) Factor analysis plot of end-Famennian and Tournaisian sites. Position along factor axis 1 (45.70% of variance) is positively influenced by Tournaisian taxa, such as actinopterygians. Factor 2 (24.02% of variance) is negatively loaded by Devonian freshwater taxa. Plots were the same for both raw and relative diversity data.

V. Supporting Tables

Table S1. Raw diversity data. See separate file in PNAS supporting information.

Table S2. Relative diversity data. See separate file in PNAS supporting information.

Table S3. Genus-level gnathostome occurrences. Raw genus counts for six major divisions of vertebrates binned by stage for the Givetian-Serpukhovian.

Division	Givetian	Frasnian	Famennian	Tournaisian	Viséan	Serpukhovian	Total
Actinopterygii	6	8	11	25	41	46	137
Sarcopterygii	40	48	56	14	16	18	192
Placodermi	83	106	53	0	0	0	242
Acanthodii	29	21	11	8	7	6	82
Chondrichthyes	11	20	36	66	94	77	304
Tetrapoda	2	7	10	4	22	17	62
Total	171	210	177	117	180	164	1019

Table S4. Descriptive statistics for faunal composition. Species counts, Shannon-Weiner diversity, and taxonomic richness for both the overall raw diversity dataset and each stage. Species counts range from 6-82.

Stage	Species Count			Shannon-Wiener Diversity			Taxonomic Richness		
	Mean	SD	Median	Mean	SD	Median	Mean	SD	Median
Giv.	20.64	11.24	16.00	1.85	0.38	1.90	8.45	3.17	9.00
Fra.	22.21	14.29	16.50	1.85	0.32	1.76	8.86	2.07	9.00
Fam.	15.61	8.81	13.50	1.89	0.42	1.91	8.28	2.99	8.00
Tour.	14.00	2.24	14.00	1.61	0.23	1.67	6.60	1.14	7.00
Vis.	17.09	7.69	15.00	1.42	0.31	1.46	6.09	1.14	6.00
Serp.	27.86	26.51	18.00	1.46	0.35	1.35	6.43	2.57	7.00
Overall	<i>19.27</i>	<i>13.01</i>	<i>15.00</i>	<i>1.73</i>	<i>0.40</i>	<i>1.74</i>	<i>7.74</i>	<i>2.61</i>	<i>7.00</i>

Table S5. CCA of raw diversity: site scores. Eigenvalue of correspondence axis 1: 0.46. Eigenvalue of correspondence axis 2: 0.27. See Fig. 3A for plot.

Site	CA1	CA2
Orcadian	-0.95	-0.40
Harma	-0.69	0.00
Gerolstein	-1.16	2.00
Rockport	-0.58	0.94
Aztec	-0.35	-0.82
Haikou	-1.57	0.56
Cletts	-0.78	-0.22
MtHowitt	-0.51	-0.85
Abava	-0.34	-0.25
CedarValley	-0.99	1.95
Gauja	-0.75	-0.43
Gladbach	-0.43	0.75
Boghole	-0.98	-1.35
Gogo	-0.82	0.86
Chahriseh	-0.27	0.81
Kerman	-0.75	1.11
NorthEvans	-0.31	2.06
Miguasha	-0.53	-0.63
Wietrznia	-0.72	1.10
Rodebjerg	-1.11	-1.10
Rhinestreet	-0.61	0.99
Chemung	-0.29	1.01
Wildungen	-0.78	1.48
ScaatCraig	-0.54	-1.44
Stolbovo	-1.07	-0.71
Canowindra	-0.99	-1.12
Rosebrae	-1.43	-1.42
DuraDen	-1.49	-1.48
Cheiloceras	-0.02	1.20
Tervete	-0.93	-0.91
Oryol	-0.62	0.30
Portishead	-0.99	-1.61
AinaDal	-0.86	-1.61
BrittaDal	-0.68	-0.91
Grenfell	-1.48	-0.50
Hyner	-0.12	-1.51
Eviewx	-0.54	-0.97
Ketleri	-0.95	-0.89
Tafilalt	0.32	1.27
Andreyevka-2	-0.09	-0.47
Cleveland	-0.07	1.25
Chaffee	-0.71	0.11
Witpoort	-0.35	0.60
Horton	1.23	-0.43
Izkchul	1.53	-0.39

Mansfield	1.10	-0.32
Foulden	1.39	-0.37
Herbesskaya	1.72	-0.02
Waaipoort	1.55	0.02
Glencartholm	1.69	0.34
Burdiehouse	1.19	-0.68
Ducabrook	1.15	-0.64
Wardie	1.22	-0.49
Cheesebay	1.42	-0.34
EastKirkton	1.33	-1.00
Gilmerton	1.14	-1.05
Ardross	1.58	1.19
Abden	1.54	0.51
Inchkeith	1.13	-0.82
Bearsden	1.53	0.80
Chokier	1.78	1.56
Loanhead	1.29	-0.74
Niddrie	1.08	-2.12
Millstone	1.46	0.32
Dora	1.33	-1.34
BearGulch	1.71	1.32

Table S6. CCA of raw diversity: taxon scores. See Fig. 3B for plot.

Taxon	CA1	CA2
Actinopterygii	0.86	0.11
Onychodontida	-0.60	0.60
Porolepiformes	-0.77	-0.63
Dipnoi	-0.34	-0.16
Osteolepididae	-0.74	-0.48
Tristichopteridae	-0.82	-0.50
Megalichthyidae	0.89	-0.51
Rhizodontida	0.86	-0.65
Actinistia	0.49	0.66
Arthrodira	-0.63	0.42
Antiarcha	-0.76	-0.34
Phyllolepida	-0.51	-1.15
Ptyctodontida	-0.73	1.16
OtherPlacoderms	-0.77	1.31
Elasmobranchii	0.43	0.29
Symmoriiformes	0.58	0.86
Holocephali	1.20	0.60
Ischnacanthida	-0.65	0.54
Climatiida	-0.61	-0.03
Gyracanthida	0.91	-0.33
Acanthodida	0.21	-0.16
Tetrapoda	0.37	-0.87

Table S7. CCA of relative diversity: site scores. Eigenvalue of correspondence axis 1: 0.47. Eigenvalue for correspondence axis 2: 0.27. See Fig. S6A for plot.

Site	CA1	CA2
Orcadian	-0.89	-0.08
Harma	-0.59	0.33
Gerolstein	-1.05	2.23
Rockport	-0.48	1.06
Aztec	-0.32	-0.58
Haikou	-1.52	0.68
Cletts	-0.76	-0.19
MtHowitt	-0.48	-0.58
Abava	-0.28	-0.11
CedarValley	-0.86	2.14
Gauja	-0.69	-0.17
Gladbach	-0.34	0.93
Boghole	-1.03	-1.30
Gogo	-0.73	0.99
Chahriseh	-0.18	0.99
Kerman	-0.64	1.33
NorthEvans	-0.20	2.15
Miguasha	-0.48	-0.37
Wietrznia	-0.64	1.20
Rodebjerg	-1.11	-0.93
Rhinestreet	-0.52	1.13
Chemung	-0.22	1.11
Wildungen	-0.67	1.57
ScaatCraig	-0.59	-1.37
Stolbovo	-1.05	-0.55
Canowindra	-1.03	-1.04
Rosebrae	-1.45	-1.22
DuraDen	-1.50	-1.32
Cheiloceras	0.06	1.23
Tervete	-0.91	-0.68
Oryol	-0.56	0.49
Portishead	-1.03	-1.42
AinaDal	-0.87	-1.41
BrittaDal	-0.67	-0.79
Grenfell	-1.46	-0.32
Hyner	-0.14	-1.37
Eviewx	-0.54	-0.84
Ketleri	-0.92	-0.68
Tafilalt	0.39	1.25
Andreyevka-2	-0.07	-0.27
Cleveland	0.01	1.18
Chaffee	-0.68	0.24
Witpoort	-0.26	0.76
Horton	1.21	-0.48
Izkchul	1.55	-0.39

Mansfield	1.13	-0.34
Foulden	1.39	-0.40
Herbesskaya	1.72	-0.09
Waaipoort	1.63	-0.02
Glencartholm	1.70	0.31
Burdiehouse	1.20	-0.69
Ducabrook	1.15	-0.69
Wardie	1.24	-0.48
Cheesebay	1.44	-0.31
EastKirkton	1.32	-1.00
Gilmerton	1.09	-1.04
Ardross	1.64	1.22
Abden	1.56	0.51
Inchkeith	1.12	-0.87
Bearsden	1.55	0.77
Chokier	1.86	1.59
Loanhead	1.27	-0.76
Niddrie	0.97	-2.20
Millstone	1.48	0.29
Dora	1.31	-1.37
BearGulch	1.72	1.26

Table S8. CCA of relative diversity: taxon scores. See Fig. S6B for plot.

Taxon	CA1	CA2
Actinopterygii	0.91	0.10
Onychodontida	-0.51	0.71
Porolepiformes	-0.81	-0.57
Dipnoi	-0.34	-0.14
Osteolepididae	-0.78	-0.37
Tristichopteridae	-0.85	-0.49
Megalichthyidae	0.91	-0.48
Rhizodontida	0.75	-0.75
Actinistia	0.61	0.76
Arthrodira	-0.61	0.40
Antiarcha	-0.79	-0.27
Phyllolepida	-0.57	-0.99
Ptyctodontida	-0.65	1.30
OtherPlacoderms	-0.71	1.39
Elasmobranchii	0.51	0.29
Symmoriiformes	0.55	0.82
Holocephali	1.18	0.56
Ischnacanthida	-0.62	0.65
Climatiida	-0.52	0.26
Gyracanthida	0.97	-0.39
Acanthodida	0.26	-0.08
Tetrapoda	0.33	-0.84

Table S9. Factor analysis of all sites: taxon loadings. See Fig. 4B for plot of factors 1 and 2.

Taxon	Factor 1	Factor 2	Factor 3	Factor 4
Actinopterygii	0.06	-4.43	-0.04	0.37
Onychodontida	0.20	0.00	-0.29	-0.05
Porolepiformes	1.56	0.12	0.23	0.07
Dipnoi	2.03	-0.09	-0.48	-0.78
Osteolepididae	0.99	-0.02	0.03	0.27
Tristichopteridae	1.05	0.09	0.03	0.12
Megalichthyidae	0.09	-0.21	0.05	-0.14
Rhizodontida	0.35	-0.29	0.28	-1.51
Actinistia	0.08	-0.58	-0.24	0.47
Arthrodira	0.45	0.33	-4.20	0.57
Antiarcha	3.12	0.21	0.25	0.29
Phyllolepida	0.49	0.06	0.14	-0.23
Ptyctodontida	0.01	0.02	-0.92	0.28
OtherPlacoderms	0.00	0.01	-0.26	0.06
Elasmobranchii	-0.73	-0.65	-1.55	-2.71
Symmoriiformes	-0.10	-0.10	-0.20	-0.16
Holocephali	0.01	-0.78	-0.06	0.30
Ischnacanthida	0.19	-0.03	-0.34	0.22
Climatiida	0.39	-0.09	-0.14	0.18
Gyracanthida	-0.13	-0.27	-0.11	-0.99
Acanthodida	1.27	-0.79	0.13	0.61
Tetrapoda	0.81	0.07	0.51	-3.03
Variance				
Explained	39.68%	22.10%	9.59%	5.45%

Table S10. Factor analysis of all sites: site scores. See Fig. 4B for plot of factors 1 and 2.

Site	Factor 1	Factor 2	Factor 3	Factor 4
Orcadian	0.66	-0.18	-0.25	0.10
Harma	0.40	-0.16	-0.44	0.09
Gerolstein	0.42	0.01	-0.46	0.05
Rockport	0.25	-0.01	-0.93	-0.08
Aztec	0.70	-0.07	-0.33	-0.14
Haikou	0.53	0.09	-0.65	0.14
Cletts	0.59	-0.23	-0.59	0.02
MtHowitt	0.78	-0.19	-0.09	0.09
Abava	0.57	-0.24	-0.42	-0.03
CedarValley	0.22	0.00	-0.47	-0.07
Gauja	0.87	-0.13	-0.30	-0.04
Gladbach	0.43	-0.42	-0.64	0.05
Boghole	0.61	0.04	-0.24	-0.03
Gogo	0.29	-0.06	-0.89	0.05
Chahriseh	0.22	-0.26	-0.46	-0.42
Kerman	0.51	-0.20	-0.75	-0.09
NorthEvans	0.07	-0.05	-0.89	0.05
Miguasha	0.82	-0.25	-0.17	-0.03
Wietrznia	0.23	0.00	-0.94	0.06
Rodebjerg	0.77	0.07	-0.47	-0.11
Rhinestreet	0.28	-0.11	-0.86	0.09
Chemung	0.37	-0.26	-0.70	-0.18
Wildungen	0.13	0.03	-0.93	0.09
ScaatCraig	0.70	0.03	-0.23	-0.31
Stolbovo	0.73	0.04	-0.59	-0.04
Canowindra	0.75	0.03	-0.24	-0.04
Rosebrae	0.89	0.03	-0.02	-0.05
DuraDen	0.88	0.04	0.02	0.00
Cheiloceras	0.06	-0.10	-0.78	-0.37
Tervete	0.86	-0.03	-0.22	-0.10
Oryol	0.74	-0.16	-0.28	-0.21
Portishead	0.81	0.04	-0.19	-0.06
AinaDal	0.87	0.03	-0.06	-0.26
BrittaDal	0.79	0.02	-0.40	-0.33
Grenfell	0.78	0.05	-0.22	0.07
Hyder	0.52	-0.15	-0.27	-0.45
Evieux	0.61	-0.09	-0.63	-0.13
Ketleri	0.79	-0.05	-0.21	-0.09
Tafilalt	-0.04	-0.25	-0.70	-0.37
Andreyevka-2	0.73	-0.33	0.02	-0.02
Cleveland	0.04	-0.05	-0.96	-0.12
Chaffee	0.56	-0.02	-0.63	-0.09
Witpoort	0.38	-0.22	-0.74	-0.19
Horton	0.13	-0.93	0.00	-0.09
Izkchul	0.02	-0.95	-0.07	-0.09

Mansfield	0.12	-0.74	-0.21	-0.39
Foulden	0.15	-0.91	0.00	-0.16
Herbesskaya	-0.04	-0.73	-0.19	-0.37
Waaipoort	0.02	-0.96	-0.04	0.00
Glencarholm	0.01	-0.98	-0.04	0.00
Burdiehouse	0.07	-0.85	-0.17	-0.45
Ducabrook	0.01	-0.52	-0.26	-0.71
Wardie	0.04	-0.95	-0.10	-0.19
Cheesebay	0.07	-0.96	0.01	0.00
EastKirkton	0.12	-0.74	0.00	-0.51
Gilmerton	0.39	-0.52	0.04	-0.52
Ardross	0.04	-0.91	-0.08	0.05
Abden	0.06	-0.97	-0.07	-0.06
Inchkeith	0.15	-0.71	-0.15	-0.63
Bearsden	0.02	-0.98	-0.07	-0.03
Chokier	0.02	-0.87	-0.04	0.13
Loanhead	0.15	-0.65	-0.10	-0.72
Niddrie	0.26	-0.05	0.08	-0.73
Millstone	0.04	-0.96	-0.12	-0.15
Dora	0.10	-0.18	-0.03	-0.87
BearGulch	0.00	-0.90	-0.07	0.02

Table S11. ANOSIM R statistics for raw diversity: extinction intervals. Values generated after 1000000 permutations. Values closer to 0 indicate greater similarity between faunas, while values closer to 1 indicate high dissimilarity.

Comparison	<i>Overall Mean</i>	<i>Abundance Mean</i>	<i>Pres.-Abs. Mean</i>	Bray-Curtis	Kulczynski	Dice	Simpson	Chord	Morisita	Raup-Crick
Frasnian-Famennian	0.09	0.12	0.07	0.10	0.07	0.06	0.08	0.13	0.13	0.08
F-F Marine	0.25	0.22	0.27	0.24	0.40	0.53	0.05	0.24	0.18	0.11
F-F Non-Marine	-0.08	-0.01	-0.13	-0.07	-0.11	-0.12	-0.05	0.02	0.01	-0.22
Late Fra.-Early Fam.	0.19	0.31	0.10	0.18	0.09	0.09	0.13	0.39	0.35	0.10
Famennian-Tournaisian	0.76	0.77	0.76	0.73	0.77	0.76	0.74	0.80	0.79	0.75
Fam-Miss. Marine	0.90	0.96	0.85	0.91	0.85	0.84	0.88	0.98	0.98	0.84
Fam-Miss. Non-marine	0.91	0.97	0.86	0.96	0.89	0.90	0.80	0.98	0.98	0.85
Late Fam-Tour	0.71	0.76	0.68	0.71	0.70	0.70	0.65	0.79	0.79	0.65
End-Fam-Tour	0.64	0.70	0.59	0.65	0.63	0.61	0.59	0.73	0.73	0.51

Table S12. ANOSIM p-values for raw diversity: extinction intervals. Values generated from 1000000 permutations. Significant values are marked with an asterisk ($\alpha=0.05$).

Comparison	<i>Overall Mean</i>	<i>Abundance Mean</i>	<i>Pres.-Abs. Mean</i>	Bray-Curtis	Kulczynski	Dice	Simpson	Chord	Morisita	Raup-Crick
Frasnian-Famennian	0.06	0.03*	0.08	0.04*	0.07	0.1	0.07	0.02*	0.02*	0.06
F-F Marine	0.1	0.08	0.1	0.05	0.02*	0.003*	0.4	0.07	0.1	0.2
F-F Non-Marine	0.7	0.5	0.8	0.7	0.8	0.8	0.6	0.4	0.4	0.9
Late Fra.-Early Fam.	0.1	0.04*	0.2	0.08	0.2	0.2	0.1	0.02*	0.02*	0.2
Famennian-Tournaisian	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*
Fam.-Miss. Marine	0.03*	0.03*	0.03*	0.03*	0.03*	0.03*	0.03*	0.03*	0.03*	0.03*
Fam.-Miss. Non-marine	0*	0*	0*	0*	0*	0*	0*	0*	0*	0*
Late Fam-Tour	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*
End-Fam-Tour	0.002*	0.001*	0.003*	0.001*	0.001*	0.003*	0.001*	0.001*	0.001*	0.008*

Table S13. ANOSIM R statistics for raw diversity: other intervals. Values generated from 1000000 permutations. Values closer to 0 indicate greater similarity between faunas, while values closer to 1 indicate high dissimilarity.

Comparison	<i>Overall Mean</i>	<i>Abundance Mean</i>	<i>Pres.-Abs. Mean</i>	Bray-Curtis	Kulczynski	Dice	Simpson	Chord	Morisita	Raup-Crick
Givetian-Frasnian	-0.02	0.06	-0.09	0.02	-0.09	-0.07	-0.10	0.09	0.08	-0.10
Givetian-Famennian	0.11	0.12	0.11	0.14	0.11	0.09	0.14	0.11	0.10	0.10
Givetian-Tournaisian	0.79	0.87	0.72	0.88	0.74	0.71	0.72	0.87	0.87	0.72
Frasnian-Tournaisian	0.75	0.81	0.71	0.67	0.74	0.73	0.71	0.88	0.88	0.67
Frasnian-Viséan	0.79	0.86	0.74	0.76	0.76	0.78	0.70	0.91	0.91	0.72
Frasnian-Serpukhovian	0.75	0.80	0.71	0.71	0.72	0.72	0.69	0.84	0.84	0.72
Famennian-Viséan	0.77	0.81	0.74	0.78	0.76	0.77	0.71	0.83	0.83	0.73
Famennian-Serpukhovian	0.74	0.76	0.73	0.75	0.74	0.74	0.71	0.77	0.77	0.73
Tournaisian-Viséan	-0.10	-0.04	-0.10	-0.18	-0.12	-0.13	-0.11	-0.04	-0.05	-0.06
Tournaisian-Serpukhovian	-0.06	-0.05	-0.06	-0.04	-0.06	-0.10	0.01	-0.05	-0.07	-0.10
Serpukhovian-Viséan	0.05	0.09	0.03	0.12	0.02	0.04	-0.04	0.07	0.07	0.09

Table S14. ANOSIM p-values for raw diversity: other intervals. Value generated after 1000000 permutations. Significant values are marked with an asterisk ($\alpha=0.05$)

Comparison	<i>Overall Mean</i>	<i>Abundance Mean</i>	<i>Pres.-Abs. Mean</i>	Bray-Curtis	Kulczynski	Dice	Simpson	Chord	Morisita	Raup-Crick
Givetian-Frasnian	0.6	0.2	1.0	0.3	1.0	0.9	1.0	0.08	0.1	1.0
Givetian-Famennian	0.06	0.05	0.06	0.03*	0.05	0.08	0.03*	0.06	0.07	0.07
Givetian-Tournaisian	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*
Frasnian-Tournaisian	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*
Frasnian-Viséan	<0.001*	<0.001*	<0.001*	<0.001*	0*	0*	<0.001*	<0.001*	0*	<0.001*
Frasnian-Serpukhovian	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*
Famennian-Viséan	0*	0*	0*	0*	0*	0*	0*	0*	0*	0*
Famennian-Serpukhovian	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*
Tournaisian-Viséan	0.8	0.7	0.8	0.9	0.9	0.9	0.9	0.5	0.6	0.7
Tournaisian-Serpukhovian	0.6	0.6	0.6	0.6	0.6	0.8	0.4	0.6	0.7	0.7
Serpukhovian-Viséan	0.3	0.2	0.4	0.1	0.3	0.3	0.7	0.2	0.2	0.1

Table S15. ANOSIM R statistics for relative diversity: extinction intervals. Values generated from 1000000 permutations. Values closer to 0 indicate greater similarity between faunas.

Comparison	<i>Overall Mean</i>	<i>Abundance Mean</i>	<i>Pres.-Abs. Mean</i>	Bray-Curtis	Kulczynski	Dice	Simpson	Chord	Morisita	Raup-Crick
Frasnian-Famennian	0.10	0.13	0.10	0.11	0.07	0.06	0.08	0.14	0.13	0.08
F-F Marine	0.25	0.23	0.27	0.28	0.40	0.53	0.05	0.24	0.18	0.10
F-F Non-marine	-0.07	-0.01	-0.12	-0.05	-0.11	-0.12	-0.05	0.02	0.01	-0.20
Late Fra.-Early Fam.	0.20	0.33	0.10	0.25	0.09	0.09	0.13	0.39	0.35	0.11
Famennian-Tournaisian	0.77	0.79	0.76	0.76	0.77	0.76	0.74	0.81	0.80	0.75
Fam.-Miss. Marine	0.91	0.98	0.86	0.99	0.85	0.84	0.88	0.98	0.98	0.84
Fam.-Miss. Non-marine	0.91	0.97	0.86	0.96	0.89	0.90	0.80	0.98	0.98	0.84
Late Fam-Tour	0.72	0.78	0.67	0.77	0.70	0.70	0.65	0.79	0.79	0.64
End Fam-Tour	0.65	0.74	0.59	0.75	0.63	0.61	0.59	0.73	0.73	0.51

Table S16. ANOSIM p-values from relative diversity: extinction intervals. Values generated from 1000000 permutations. Significant values are marked with an asterisk ($\alpha=0.05$).

Comparison	<i>Overall Mean</i>	<i>Abundance Mean</i>	<i>Pres.-Abs. Mean</i>	Bray-Curtis	Kulczynski	Dice	Simpson	Chord	Morisita	Raup-Crick
Frasnian-Famennian	0.05	0.03*	0.08	0.03*	0.07	0.1	0.07	0.02*	0.02*	0.06
F-F Marine	0.1	0.07	0.2	0.04*	0.02*	0.003*	0.4	0.07	0.1	0.2
F-F Non-Marine	0.7	0.5	0.8	0.6	0.8	0.8	0.6	0.4	0.4	0.9
Late Fra.-Early Fam.	0.1	0.03*	0.2	0.06	0.2	0.2	0.1	0.02*	0.02*	0.2
Famennian-Tournaisian	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*
F-M Marine	0.03*	0.03*	0.03*	0.03*	0.03*	0.03*	0.03*	0.03*	0.03*	0.03*
F-M Non-marine	0*	0*	0*	0*	0*	0*	0*	0*	0*	0*
Late Fam-Tour	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*
End-Fam-Tour	0.002*	0.001*	0.003*	0.001*	0.001*	0.003*	0.001*	0.001*	0.001*	0.008*

Table S17. ANOSIM R statistics for relative diversity: other intervals. Values generated after 1000000 permutations. Values closer to 0 indicate greater similarity between faunas.

Comparison	<i>Overall Mean</i>	<i>Abundance Mean</i>	<i>Pres.-Abs. Mean</i>	Bray-Curtis	Kulczynski	Dice	Simpson	Chord	Morisita	Raup-Crick
Givetian-Frasnian	-0.03	0.06	-0.09	0.01	-0.09	-0.07	-0.10	0.09	0.08	-0.10
Givetian-Famennian	0.10	0.10	0.11	0.08	0.11	0.09	0.14	0.11	0.10	0.10
Givetian-Tournaisian	0.79	0.88	0.72	0.90	0.74	0.71	0.72	0.87	0.87	0.72
Frasnian-Tournaisian	0.78	0.86	0.71	0.85	0.74	0.73	0.71	0.88	0.88	0.67
Frasnian-Viséan	0.81	0.90	0.74	0.88	0.76	0.77	0.70	0.91	0.91	0.72
Frasnian-Serpukhovian	0.76	0.83	0.71	0.81	0.72	0.72	0.69	0.84	0.84	0.72
Famennian-Viséan	0.77	0.82	0.74	0.80	0.76	0.77	0.71	0.83	0.83	0.73
Famennian-Serpukhovian	0.75	0.77	0.73	0.76	0.74	0.75	0.71	0.77	0.78	0.73
Tournaisian-Viséan	-0.09	-0.07	-0.11	-0.11	-0.12	-0.13	-0.11	-0.04	-0.05	-0.06
Tournaisian-Serpukhovian	-0.06	-0.04	-0.08	-0.02	-0.06	-0.10	0.01	-0.05	-0.07	-0.10
Serpukhovian-Viséan	0.05	0.07	0.03	0.08	0.02	0.04	-0.04	0.07	0.07	0.09

Table S18. ANOSIM p-values from relative diversity: other intervals. Values generated after 1000000 permutations. Significant values are marked with an asterisk ($\alpha=0.05$)

Comparison	Overall Mean	Abundance Mean	Pres.-Abs. Mean	Bray-Curtis	Kulczynski	Dice	Simpson	Chord	Morisita	Raup-Crick
Givetian-Frasnian	0.6	0.2	1.0	0.4	1.0	0.9	1.0	0.08	0.1	1.0
Givetian-Famennian	0.07	0.08	0.06	0.1	0.05	0.08	0.03*	0.06	0.07	0.07
Givetian-Tournaisian	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*
Frasnian-Tournaisian	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*
Frasnian-Viséan	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	0*	0*	0*	<0.001*	<0.001*
Frasnian-Serpukhovian	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*
Famennian-Viséan	0*	0*	0*	0*	0*	0*	0*	0*	0*	0*
Famennian-Serpukhovian	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*	<0.001*
Tournaisian-Viséan	0.7	0.7	0.8	0.8	0.9	0.9	0.9	0.5	0.6	0.7
Tournaisian-Serpukhovian	0.6	0.6	0.6	0.5	0.6	0.8	0.4	0.6	0.7	0.7
Serpukhovian-Viséan	0.3	0.2	0.4	0.2	0.3	0.3	0.7	0.2	0.2	0.1

Table S19. SIMPER average dissimilarities. Results based on relative diversity data.

Boundary	Givetian.- Frasnian	Frasnian- Famennian	Famennian- Tournaisian	Tournaisian- Viséan	Viséan- Serpukhovian	<i>Average.</i>
Average Dissimilarity	58.65	60.80	83.19	40.74	50.52	58.78

Table S20. SIMPER for all Frasnian and Famennian sites. Results based on relative diversity data. Average dissimilarity: 60.80.

Taxon	Contribution	Cumulative %	Mean abund. Frasnian	Mean abund. Famennian
Arthrodira	10.61	17.45	29.40	15.10
Antiarcha	6.43	28.02	8.63	17.00
Elasmobranchii	4.92	36.12	5.93	8.29
Porolepiformes	4.46	43.46	7.59	6.39
Dipnoi	4.14	50.27	11.4	12.90
Tristichopteridae	3.87	56.63	3.06	7.80
Osteolepididae	3.09	61.71	2.61	5.91
Tetrapoda	2.78	66.28	2.84	4.78
Ptyctodontida	2.72	70.75	5.29	1.32
Acanthodida	2.71	75.21	3.19	3.97
Actinopterygii	2.06	78.60	4.48	2.20
Ischnacanthida	1.87	81.68	3.23	1.50
Rhizodontida	1.84	84.70	2.21	2.05
Onychodontida	1.74	87.55	2.73	1.75
Phyllolepida	1.46	89.94	0.00	2.91
Actinistia	1.34	92.14	1.74	1.48
Climatiida	1.32	94.31	2.40	0.40
Symmoriiformes	1.00	95.96	0.71	1.54
Holocephali	0.77	97.23	0.92	0.82
Other Placoderms	0.74	98.44	1.22	0.40
Gyracanthida	0.50	99.27	0.45	0.64
Megalichthyidae	0.45	100.00	0.00	0.89

Table S21. SIMPER for all Famennian and Tournaisian sites. Results based on relative diversity data. Average dissimilarity: 83.19.

Taxon	Contribution	Cumulative %	Mean abund. Famennian	Mean abund. Tournaisian
Actinopterygii	19.08	22.94	2.20	40.40
Antiarcha	8.48	33.13	17.00	0.00
Arthrodira	7.55	42.21	15.10	0.00
Elasmobranchii	7.07	50.71	8.29	13.00
Dipnoi	4.68	56.34	12.90	4.69
Gyracanthida	4.59	61.85	0.64	9.55
Acanthodida	4.14	66.82	3.97	9.11
Tristichopteridae	3.90	71.51	7.80	0.00
Rhizodontida	3.74	76.00	2.05	8.63
Porolepiformes	3.19	79.84	6.39	0.00
Osteolepididae	2.96	83.39	5.91	0.00
Holocephali	2.53	86.44	0.82	4.86
Tetrapoda	2.53	89.47	4.78	2.87
Megalichthyidae	2.12	92.02	0.89	4.03
Phyllolepada	1.46	93.77	2.91	0.00
Symmoriiformes	1.32	95.35	1.54	1.54
Actinistia	1.18	96.78	1.48	1.33
Onychodontida	0.87	97.83	1.75	0.00
Ischnacanthida	0.75	98.73	1.50	0.00
Ptyctodontida	0.66	99.52	1.32	0.00
Other Placoderms	0.20	99.76	0.40	0.00
Climatiida	0.20	100.00	0.40	0.00

Table S22. Factor analysis of Frasnian-Famennian sites: taxon loadings. See Fig. S11A for plot of factors 1 and 2.

Taxon	Factor 1	Factor 2	Factor 3	Factor 4
Actinopterygii	-0.39	0.36	-0.62	-0.88
Onychodontida	-0.46	-0.07	-0.68	-1.17
Porolepiformes	2.21	-0.06	0.39	0.44
Dipnoi	0.89	0.21	-0.71	-2.61
Osteolepididae	0.21	0.05	0.13	-1.54
Tristichopteridae	1.74	-0.07	-0.05	0.49
Megalichthyidae	0.04	-0.02	-0.01	-0.17
Rhizodontida	1.21	-0.05	-0.05	1.05
Actinistia	-0.12	0.14	-0.59	-0.15
Arthrodira	0.65	4.49	-0.15	0.40
Antiarcha	3.06	-0.65	-0.38	-0.97
Phyllolepida	0.70	-0.18	-0.33	0.35
Ptyctodontida	-0.41	0.91	0.34	-0.65
OtherPlacoderms	-0.11	0.20	-0.11	-0.12
Elasmobranchii	-0.25	0.00	-4.19	0.29
Symmoriiformes	-0.04	-0.17	-1.04	0.11
Holocephali	-0.23	0.14	0.05	-0.51
Ischnacanthida	-0.23	0.45	0.25	-0.76
Climatiida	-0.05	-0.14	-0.68	-0.31
Gyracanthida	0.04	0.13	-0.08	0.08
Acanthodida	-0.46	0.16	0.69	-2.44
Tetrapoda	0.76	-0.02	0.20	-0.66
Variance				
Explained	54.95%	16.82%	6.06%	4.56%

Table S23. Factor analysis of Frasnian-Famennian relative diversity: site scores. See Fig. S11A for plot of factors 1 and 2

Site	Factor 1	Factor 2	Factor 3	Factor 4
Tafilalt	0.06	0.45	-0.82	0.08
Cleveland	0.11	0.88	-0.40	0.07
Cheiloceras	0.12	0.52	-0.79	0.01
Chaffee	0.38	0.56	-0.29	-0.48
Miguasha	0.59	0.17	-0.11	-0.54
Stolbovo	0.60	0.61	-0.07	-0.42
ScaatCraig	0.78	0.28	-0.02	-0.14
Rodebjerg	0.71	0.48	-0.06	-0.34
Boghole	0.83	0.27	0.01	0.14
NorthEvans	0.04	0.91	-0.12	-0.05
Gogo	0.21	0.91	-0.15	-0.18
Chemung	0.18	0.60	-0.44	-0.46
Wietrznia	0.22	0.94	-0.16	-0.06
Rhinestreet	0.15	0.88	-0.12	-0.24
Kerman	0.36	0.62	-0.46	-0.43
Chahriseh	0.11	0.15	-0.87	-0.32
Gladbach	0.17	0.63	-0.24	-0.55
Wildungen	0.15	0.97	-0.11	0.03
Witpoort	0.34	0.69	-0.35	-0.20
Andreyevka	0.34	0.01	0.01	-0.78
Evieux	0.66	0.63	-0.20	-0.15
Portishead	0.84	0.18	-0.06	-0.20
Grenfell	0.73	0.16	-0.09	-0.33
BrittaDal	0.70	0.36	-0.21	-0.43
AinaDal	0.79	0.05	-0.07	-0.42
Hyner	0.57	0.21	-0.36	-0.14
Ketleri	0.46	0.22	-0.02	-0.74
Oryol	0.46	0.15	-0.37	-0.71
Tervete	0.64	0.23	-0.02	-0.59
Canowindra	0.77	0.20	-0.10	-0.23
DuraDen	0.88	-0.10	-0.09	-0.29
Rosebrae	0.75	-0.06	-0.13	-0.53

Table S24. Factor Analysis of Famennian-Mississippian sites: taxon loadings. See Fig. S11C for plot of factors 1 and 2.

Taxon	Factor 1	Factor 2	Factor 3	Factor 4
Actinopterygii	4.50	0.04	0.16	0.82
Onychodontida	-0.05	-0.33	0.09	-0.39
Porolepiformes	-0.07	-1.18	0.19	0.01
Dipnoi	0.07	-2.24	0.02	-1.35
Osteolepididae	-0.06	-1.07	0.01	0.14
Tristichopteridae	0.05	-1.28	-0.33	1.40
Megalichthyidae	0.22	-0.14	0.04	-0.25
Rhizodontida	0.63	-0.32	-0.07	-0.63
Actinistia	0.33	0.05	-0.51	0.52
Arthrodira	-0.27	-1.01	-3.21	1.64
Antiarcha	-0.10	-3.07	0.38	0.77
Phyllolepida	-0.08	-0.49	-0.09	-0.33
Ptyctodontida	-0.03	-0.23	-0.08	-0.10
OtherPlacoderms	0.01	-0.03	-0.15	0.15
Elasmobranchii	0.53	0.41	-3.19	-2.27
Symmoriiformes	0.00	0.04	-0.46	-0.03
Holocephali	0.50	-0.09	0.03	-0.01
Ischnacanthida	0.03	-0.29	0.04	0.18
Climatiida	0.01	-0.03	-0.15	0.15
Gyracanthida	0.42	0.05	-0.31	-0.62
Acanthodida	0.53	-0.83	0.66	-1.30
Tetrapoda	0.32	-0.94	0.40	-2.44
Variance				
Explained	42.92%	25.88%	9.85%	4.05%

Table S25. Factor Analysis of Famennian-Mississippian sites: site scores. See Fig. S11C for plot of factors 1 and 2.

Site	Factor 1	Factor 2	Factor 3	Factor 4
Glencarholm	0.99	0.00	-0.04	0.10
Ardross	0.87	-0.01	-0.09	0.05
Herbesskaya	0.71	0.01	-0.36	-0.29
Abden	0.95	-0.03	-0.11	-0.02
Waaipoort	0.97	0.00	-0.03	0.08
Foulden	0.94	-0.13	0.03	-0.06
Ducabrook	0.55	-0.07	-0.54	-0.54
Gilmerton	0.58	-0.40	0.09	-0.47
Cheesebay	0.98	-0.03	0.04	0.09
EastKirkton	0.80	-0.13	-0.06	-0.35
Inchkeith	0.76	-0.19	-0.29	-0.45
Burdiehouse	0.87	-0.09	-0.31	-0.32
Mansfield	0.74	-0.12	-0.35	-0.41
Izkchul	0.96	-0.01	-0.11	-0.01
Horton	0.96	-0.11	0.04	-0.02
Wardie	0.96	-0.04	-0.16	-0.09
Tafilalt	0.22	-0.07	-0.90	-0.12
Chaffee	0.00	-0.66	-0.50	-0.03
Cleveland	0.05	-0.18	-0.89	0.13
Cheiloceras	0.07	-0.17	-0.93	-0.17
Andreyevka-2	0.31	-0.68	0.17	-0.26
Portishead	0.00	-0.81	-0.13	0.06
Ketleri	0.05	-0.81	-0.06	-0.20
Evieux	0.14	-0.72	-0.54	0.19
Oryol	0.15	-0.80	-0.19	-0.24
DuraDen	-0.01	-0.87	0.05	0.13
Hyner	0.20	-0.58	-0.35	-0.25
Grenfell	-0.02	-0.85	-0.14	0.29
Tervete	0.05	-0.87	-0.05	-0.18
Witpoort	0.25	-0.49	-0.64	0.09
BrittaDal	0.03	-0.86	-0.30	-0.17
AinaDal	0.02	-0.89	0.01	-0.15
Rosebrae	-0.01	-0.91	0.05	-0.01
Canowindra	0.02	-0.82	-0.20	0.25

Table S26. SIMPER for Frasnian-Famennian marine sites. Results based on relative diversity data. Average dissimilarity: 54.53

Taxon	Contribution	Cumulative %	Mean abund. Frasnian marine	Mean abund. Famennian marine
Elasmobranchii	10.57	19.45	9.22	28.60
Arthrodira	10.51	38.78	36.00	34.80
Dipnoi	5.00	47.99	11.20	7.81
Ptyctodontida	4.12	55.57	8.24	0.00
Symmoriiformes	2.85	60.81	1.10	5.88
Actinistia	2.48	65.38	1.97	4.86
Osteolepididae	2.42	69.83	1.98	3.85
Actinopterygii	2.30	74.05	6.22	3.06
Onychodontida	2.21	78.11	4.25	1.92
Ischnacanthida	2.16	82.09	4.32	0.00
Antiarcha	2.15	86.04	4.34	3.39
Acanthodida	1.39	88.60	2.78	0.00
Holocephali	1.32	91.03	1.44	1.92
Porolepiformes	1.30	93.42	1.36	1.92
Tristichopteridae	1.16	95.55	0.78	1.92
Climatiida	1.13	97.62	2.25	0.00
Other Placoderms	0.95	99.36	1.90	0.00
Gyracanthida	0.35	100.00	0.69	0.00
Megalichthyidae	0.00	100.00	0.00	0.00
Phyllolepida	0.00	100.00	0.00	0.00
Tetrapoda	0.00	100.00	0.00	0.00
Rhizodontida	0.00	100.00	0.00	0.00

Table S27. SIMPER for Frasnian-Famennian non-marine sites. Results based on relative diversity data. Average dissimilarity: 49.15.

Taxon	Contribution	Cumulative %	Mean abund. Frasnian Non-marine	Mean abund. Famennian Non-marine
Porolepiformes	6.65	13.52	18.80	7.66
Arthrodira	5.35	24.40	17.50	9.48
Antiarcha	5.02	34.61	16.40	20.80
Tristichopteridae	4.21	43.19	7.17	9.48
Dipnoi	3.56	50.44	11.80	14.40
Rhizodontida	3.36	57.27	6.19	2.63
Tetrapoda	3.35	64.10	7.94	6.14
Acanthodida	3.20	70.61	3.92	5.11
Osteolepididae	3.13	76.98	3.75	6.50
Phyllolepida	1.87	80.79	0.00	3.74
Climatiida	1.49	83.81	2.67	0.51
Ischnacanthida	1.29	86.43	1.25	1.93
Actinopterygii	1.27	89.00	1.33	1.95
Elasmobranchii	1.24	91.52	0.00	2.47
Ptyctodontida	0.85	93.25	0.00	1.70
Onychodontida	0.85	94.98	0.00	1.70
Actinistia	0.83	96.66	1.33	0.51
Megalichthyidae	0.57	97.82	0.00	1.15
Gyracanthida	0.41	98.66	0.00	0.82
Other Placoderms	0.26	99.18	0.00	0.51
Holocephali	0.26	99.70	0.00	0.51
Symmoriiformes	0.15	100.00	0.00	0.30

Table S28. SIMPER for Famennian-Mississippian marine sites. Results based on relative diversity data. Average dissimilarity: 80.74.

Taxon	Contribution	Cumulative %	Mean abund. Famennian marine	Mean abund. Mississippian marine
Actinopterygii	20.86	25.83	3.06	44.80
Arthrodira	17.40	47.39	34.80	0.00
Elasmobranchii	9.20	58.78	28.60	12.80
Holocephali	6.80	67.20	1.92	15.50
Dipnoi	3.90	72.04	7.81	0.00
Actinistia	3.51	76.39	4.86	7.64
Acanthodida	3.51	80.74	0.00	7.02
Symmoriiformes	2.94	84.38	5.88	0.00
Rhizodontida	2.82	87.87	0.00	5.64
Gyracanthida	2.21	90.60	0.00	4.41
Osteolepididae	1.92	92.99	3.85	0.00
Antiarcha	1.70	95.09	3.39	0.00
Megalichthyidae	1.08	96.43	0.00	2.17
Porolepiformes	0.96	97.62	1.92	0.00
Tristichopteridae	0.96	98.81	1.92	0.00
Onychodontida	0.96	100.00	1.92	0.00
OtherPlacoderms	0.00	100.00	0.00	0.00
Ptyctodontida	0.00	100.00	0.00	0.00
Phyllolepida	0.00	100.00	0.00	0.00
Tetrapoda	0.00	100.00	0.00	0.00
Climatiida	0.00	100.00	0.00	0.00
Ischnacanthida	0.00	100.00	0.00	0.00

Table S29. SIMPER for Famennian-Mississippian non-marine sites. Results based on relative diversity data. Average dissimilarity: 81.73.

Taxon	Contribution	Cumulative %	Mean abund. Famennian Non-marine	Mean abund. Mississippian Non-marine
Actinopterygii	20.90	25.58	1.95	43.80
Antiarcha	10.42	38.32	20.80	0.00
Elasmobranchii	6.48	46.25	2.47	13.7
Arthrodira	4.74	52.05	9.48	0.00
Tristichopteridae	4.74	57.85	9.48	0.00
Dipnoi	4.66	63.55	14.40	6.74
Tetrapoda	4.46	69.00	6.14	9.91
Porolepiformes	3.83	73.69	7.66	0.00
Rhizodontida	3.58	78.07	2.63	8.39
Acanthodida	3.41	82.24	5.11	6.22
Osteolepididae	3.25	86.22	6.50	0.00
Gyracanthida	2.82	89.67	0.82	5.86
Phyllolepida	1.87	91.96	3.74	0.00
Megalichthyidae	1.47	93.76	1.15	2.49
Ischnacanthida	0.96	94.94	1.93	0.00
Ptyctodontida	0.85	95.98	1.70	0.00
Onychodontida	0.85	97.02	1.70	0.00
Actinistia	0.75	97.94	0.51	1.15
Holocephali	0.73	98.83	0.51	1.11
Symmoriiformes	0.44	99.38	0.30	0.64
Other Placoderms	0.26	99.69	0.51	0.00
Climatiida	0.26	100.00	0.51	0.00

Table S30. Factor analysis of late Frasnian-early Famennian sites: taxon loadings. See Fig. S11B for plot of factors 1 and 2.

Taxon	Factor 1	Factor 2	Factor 3	Factor 4
Actinopterygii	-0.04	0.09	-0.68	0.06
Onychodontida	0.05	-0.29	-1.05	0.09
Porolepiformes	-0.13	-0.44	-0.05	-3.79
Dipnoi	2.03	-0.27	-1.93	-0.71
Osteolepididae	1.17	-0.20	-0.38	0.25
Tristichopteridae	2.41	0.75	1.23	1.32
Megalichthyidae	0.00	0.00	0.00	0.00
Rhizodontida	0.03	0.22	0.73	-1.44
Actinistia	-0.12	0.02	-0.58	0.12
Arthrodira	-0.11	4.49	-0.39	-0.63
Antiarcha	3.18	-0.28	-0.02	-0.78
Phyllolepida	0.03	-0.03	0.13	-0.76
Ptyctodontida	0.30	0.09	-0.50	0.15
OtherPlacoderms	-0.13	0.08	-0.38	-0.06
Elasmobranchii	-0.47	-0.10	-3.54	0.50
Symmoriiformes	0.03	-0.17	-0.63	0.14
Holocephali	-0.12	-0.03	-0.44	-0.06
Ischnacanthida	0.09	0.67	-0.48	0.23
Climatiida	0.00	0.00	0.00	0.00
Gyracanthida	0.00	0.00	0.00	0.00
Acanthodida	0.41	0.54	-0.50	0.10
Tetrapoda	-0.06	0.03	-0.03	-1.15
<i>Variance Explained</i>	56.84%	19.86%	6.04%	5.44%

Table S31. Factor analysis of late Frasnian-early Famennian sites: site scores. See Fig. S11B for plot of factors 1 and 2.

Site	Factor 1	Factor 2	Factor 3	Factor 4
Rhinestreet	0.14	0.89	-0.31	-0.11
Chemung	0.20	0.52	-0.70	-0.26
Wildungen	0.02	0.95	-0.17	-0.12
ScaatCraig	0.34	0.23	-0.12	-0.87
Stolbovo	0.63	0.64	-0.22	-0.27
Canowindra	0.88	0.32	0.03	-0.09
Rosebrae	0.89	-0.07	-0.24	-0.31
DuraDen	0.86	-0.04	-0.06	-0.38
Cheiloceras	0.03	0.50	-0.72	-0.02
Tervete	0.69	0.24	-0.23	-0.38
Oryol	0.67	0.05	-0.60	-0.24
Portishead	0.55	0.16	-0.12	-0.73

Table S32. Factor analysis of late Famennian-Tournaisian sites: taxon loadings. See Fig. S11D for plot of factors 1 and 2.

Taxon	Factor 1	Factor 2	Factor 3	Factor 4
Actinopterygii	0.38	4.13	0.24	-0.63
Onychodontida	-0.28	-0.09	-0.05	1.44
Porolepiformes	0.49	-0.10	-0.06	1.04
Dipnoi	1.34	0.14	-0.32	1.68
Osteolepididae	0.09	-0.24	-0.21	2.02
Tristichopteridae	1.50	-0.14	-0.30	-0.35
Megalichthyidae	0.12	0.44	0.04	0.28
Rhizodontida	0.38	0.94	0.05	-0.38
Actinistia	0.14	0.19	-0.34	-0.32
Arthrodira	1.39	-0.55	-3.47	-0.25
Antiarcha	3.14	-0.18	0.76	0.43
Phyllolepida	1.11	0.14	0.07	-0.83
Ptyctodontida	0.18	0.02	-0.16	-0.25
OtherPlacoderms	0.18	0.02	-0.16	-0.25
Elasmobranchii	-0.85	1.07	-2.84	0.55
Symmoriiformes	-0.15	0.15	-0.50	-0.07
Holocephali	-0.40	0.47	-0.17	0.99
Ischnacanthida	0.36	-0.01	0.16	0.33
Climatiida	0.18	0.02	-0.16	-0.25
Gyracanthida	0.09	1.04	-0.20	-0.20
Acanthodida	-0.30	0.89	0.47	2.92
Tetrapoda	1.74	0.32	0.47	0.03
Variance Explained	43.59%	22.78%	12.45%	5.11%

Table S33. Factor analysis of late Famennian-Tournaisian sites: site scores. See Fig. S11D for plot of factors 1 and 2.

Site	Factor 1	Factor 2	Factor 3	Factor 4
AinaDal	0.90	0.02	0.04	0.27
BrittaDal	0.83	0.02	-0.30	0.32
Grenfell	0.81	-0.08	-0.11	0.23
Hynier	0.65	0.27	-0.31	0.06
Evieux	0.72	0.10	-0.57	0.16
Ketleri	0.51	0.04	-0.14	0.79
Tafilalt	0.06	0.27	-0.86	0.01
Andreyevka-2	0.42	0.33	0.13	0.77
Cleveland	0.21	0.04	-0.92	0.01
Chaffee	0.43	-0.02	-0.58	0.52
Witpoort	0.58	0.24	-0.64	-0.01
Horton	0.16	0.93	0.05	-0.01
Izkchul	0.04	0.94	-0.08	-0.02
Mansfield	-0.01	0.83	-0.31	0.28
Foulden	0.17	0.94	0.04	0.01
Herbesskaya	-0.08	0.81	-0.33	0.14

Table S34. Factor analysis of end-Famennian- Tournaisian sites: taxon loadings. See Fig. S12D for plot of factors 1 and 2.

Taxon	Factor 1	Factor 2	Factor 3	Factor 4
Actinopterygii	4.40	-0.28	-0.24	-0.05
Onychodontida	-0.20	0.09	1.19	0.11
Porolepiformes	-0.12	-0.30	1.23	-0.25
Dipnoi	0.24	-0.91	1.98	-0.50
Osteolepididae	-0.32	-0.22	1.90	-0.10
Tristichopteridae	0.04	-1.01	0.37	-0.45
Megalichthyidae	0.32	0.08	0.45	0.22
Rhizodontida	0.94	-0.08	-0.02	-0.02
Actinistia	0.24	-0.47	-0.23	-0.06
Arthrodira	-0.34	-4.10	-0.13	-0.28
Antiarcha	-0.04	-0.46	1.61	-0.37
Phyllolepida	0.04	-0.20	0.02	-0.18
Ptyctodontida	0.12	-0.36	-0.12	-0.28
OtherPlacoderms	0.12	-0.36	-0.12	-0.28
Elasmobranchii	-0.13	-1.01	-0.27	4.18
Symmoriiformes	0.00	-0.25	-0.22	0.61
Holocephali	0.17	0.27	0.72	0.91
Ischnacanthida	0.00	0.01	0.52	-0.03
Climatiida	0.12	-0.36	-0.12	-0.28
Gyracanthida	0.90	-0.09	-0.07	0.56
Acanthodida	0.40	0.96	2.62	1.19
Tetrapoda	0.50	-0.39	0.82	-0.71
<i>Variance</i>				
<i>Explained</i>	45.68%	24.04%	13.81%	5.95%

Table S35. Factor analysis of end-Famennian-Tournaisian sites: site scores. Fig. S12D for plot of factors 1 and 2

Site	Factor 1	Factor 2	Factor 3	Factor 4
Evieux	0.13	-0.84	0.38	-0.05
Ketleri	0.02	-0.33	0.91	-0.04
Tafilalt	0.10	-0.65	-0.08	0.69
Andreyevka-2	0.28	0.01	0.89	0.11
Cleveland	-0.01	-0.90	-0.04	0.30
Chaffee	-0.05	-0.69	0.57	0.08
Witpoort	0.26	-0.86	0.15	0.04
Horton	0.97	-0.08	0.10	0.02
Izkchul	0.91	-0.08	0.03	0.24
Mansfield	0.67	-0.13	0.24	0.63
Foulden	0.96	-0.08	0.13	0.05
Herbesskaya	0.64	-0.11	0.09	0.65

Table S36. SIMPER for late Frasnian and early Famennian sites. Results based on relative diversity data. Average dissimilarity: 64.01.

Taxon	Contribution	Cumulative %	Mean abund. late Frasnian	Mean abund. early Famennian
Arthrodira	13.42	20.96	34.50	9.36
Antiarcha	8.39	34.07	6.90	22.80
Porolepiformes	5.26	42.28	8.30	9.40
Elasmobranchii	4.74	49.69	4.95	7.07
Tristichopteridae	4.56	56.80	2.87	9.21
Dipnoi	4.55	63.91	12.20	16.6
Osteolepididae	3.28	69.04	1.25	6.75
Ischnacanthida	3.00	73.72	6.11	0.60
Acanthodida	2.97	78.37	5.31	2.98
Rhizodontida	2.44	82.18	2.86	3.37
Tetrapoda	2.29	85.76	4.11	1.79
Ptyctodontida	1.68	88.39	2.29	2.38
Actinopterygii	1.45	90.65	2.88	0.60
Phyllolepida	1.39	92.82	0.00	2.78
Onychodontida	1.20	94.70	1.70	1.19
Other Placoderms	1.04	96.32	2.07	0.00
Actinistia	0.97	97.84	0.37	1.68
Symmoriiformes	0.72	98.96	0.00	1.44
Holocephali	0.67	100.00	1.33	0.00
Megalichthyidae	0.00	100.00	0.00	0.00
Gyracanthida	0.00	100.00	0.00	0.00
Climatiida	0.00	100.00	0.00	0.00

Table S37. SIMPER for late Famennian and Tournaisian sites. Results based on relative diversity data. Average dissimilarity: 80.79.

Taxon	Contribution	Cumulative %	Mean abund. late Famennian	Mean abund. Tournaisian
Actinopterygii	18.57	22.98	3.22	40.40
Arthrodira	9.38	34.60	18.80	0.00
Elasmobranchii	6.82	43.04	9.06	13.00
Antiarcha	6.61	51.22	13.20	0.00
Gyracanthida	4.46	56.75	1.04	9.55
Acanthodida	4.10	61.82	4.60	9.11
Rhizodontida	3.81	66.54	1.20	8.63
Dipnoi	3.76	71.20	10.60	4.69
Tristichopteridae	3.45	75.47	6.90	0.00
Tetrapoda	3.07	79.26	6.68	2.87
Osteolepididae	2.69	82.59	5.38	0.00
Holocephali	2.59	85.80	1.35	4.86
Porolepiformes	2.23	88.57	4.47	0.00
Megalichthyidae	2.19	91.27	1.46	4.03
Phyllolepada	1.50	93.13	3.00	0.00
Symmoriiformes	1.39	94.85	1.60	1.54
Actinistia	1.10	96.21	1.35	1.33
Onychodontida	1.05	97.51	2.11	0.00
Ischnacanthida	1.04	98.79	2.07	0.00
Other Placoderms	0.32	99.20	0.65	0.00
Ptyctodontida	0.32	99.60	0.65	0.00
Climatiida	0.32	100.00	0.650	0.00

Table S38. SIMPER for end-Famennian and Tournaisian sites. Results based on relative diversity data. Average dissimilarity: 79.19.

Taxon	Contribution	Cumulative %	Mean abund. end-Famennian	Mean abund. Tournaisian
Actinopterygii	17.96	22.68	4.44	40.40
Arthrodira	11.59	37.32	23.20	0.00
Elasmobranchii	7.21	46.42	12.20	13.00
Acanthodida	4.72	52.38	5.44	9.11
Gyracanthida	4.47	58.02	1.02	9.55
Rhizodontida	3.99	63.06	0.65	8.63
Dipnoi	3.61	67.62	9.34	4.69
Antiarcha	3.32	71.82	6.65	0.00
Osteolepididae	3.12	75.77	6.25	0.00
Holocephali	2.69	79.16	2.12	4.86
Tristichopteridae	2.63	82.48	5.26	0.00
Porolepiformes	2.30	85.39	4.61	0.00
Megalichthyidae	2.22	88.19	1.67	4.03
Tetrapoda	1.92	90.61	3.88	2.87
Symmoriiformes	1.75	92.81	2.52	1.54
Onychodontida	1.65	94.90	3.31	0.00
Actinistia	1.35	96.60	2.12	1.33
Ischnacanthida	0.84	97.66	1.67	0.00
OtherPlacoderms	0.51	98.30	1.02	0.00
Ptyctodontida	0.51	98.95	1.02	0.00
Climatiida	0.51	99.59	1.02	0.00
Phyllolepida	0.32	100.00	0.65	0.00

VI. References

1. Sepkoski JJ (2002) A compendium of fossil marine animal genera. *Bull Am Paleont* 363:1-560.
2. Benton MJ (1993) *The Fossil Record II* (Chapman and Hall, London).
3. Kaiser SI, Steuber T, Becker RT (2008). Environmental change during the Late Famennian and Early Tournaisian (Late Devonian-Early Carboniferous): implications from stable isotopes and conodont biofacies in southern Europe. *Geol J* 43:241-260.
4. Peters SE, Foote M (2002) Determinants of extinction in the fossil record. *Nature* 416:420-424.
5. Hammer Ø, Harper DAT (2006) *Paleontological Data Analysis* (Blackwell, Malden).
6. Dineley DL, Metcalfe SJ (1999) *Fossil Fishes of Great Britain* (JNCC, Petersborough).
7. Purvis A (2008) Phylogenetic approaches to the study of extinction. *Ann Rev Ecol Evol Syst* 39:301-319.
8. Shannon CE, Weaver W (1949) *The Mathematical Theory of Communication* (University of Illinois, Urbana).
9. Brazeau M (2009) The braincase and jaws of a Devonian 'acanthodian' and modern gnathostome origins. *Nature* 457:305-308.
10. Schram FR (1983) Lower Carboniferous biota of Glencartholm, Eskdale, Dumfriesshire. *Scot J Geol* 19:1-15.
11. R Development Core Team. (2008) R: a language and environment for statistical computing. <http://www.R-project.org>.
12. Oksanen J *et al.* (2008) Vegan: Community Ecology Package. R package version 1.15-1. <http://cran.r-project.org/>, <http://vegan.r-forge.r-project.org/>.
13. Hammer Ø, Harper DAT, Ryan PD (2001) PAST: Paleontological software package for education and data analysis. *Palaeont Electr* 4:9pp.
14. Bray JR, Curtis JT (1957) An ordination of the upland forest communities of southern Wisconsin. *Ecol Monog* 27:325-349.
15. Cheetham AH, Hazel JE (1969) Binary (presence-absence) similarity coefficients. *J Paleont* 43:1130-1136.
16. Dice LR (1945) Measures of the amount of ecological association between species. *Ecology* 26:297-302.

17. Hammer Ø (2002) Palaeontological community and diversity analysis – brief notes. <http://folk.uio.no/ohammer/past/community.pdf>
18. Morisita M (1959) Measuring of interspecific association and similarity between communities. *Mem Fac Kyushu Univ E* 3:65-80.
19. Raup D, Crick RE (1979) Measurement of faunal similarity in paleontology. *J Palaeont* 53:1212-1217.
20. Sokal RR, Michener CD (1958) A statistical method for evaluating systematic relationships. *Univ Kansas Sci Bull* 38:1409-1438.
21. Greenacre MJ (1984) *Theory and Applications of Correspondence Analysis* (Academic, London).
22. Sepkoski JJ (1981) A factor analytic description of the Phanerozoic marine fossil record. *Paleobiology* 7:36-53.
23. Clarke KR (1993) Non-parametric multivariate analysis of changes in community structure. *Austral J Ecol* 18:117-143.
24. Woodward AS, Sherborn CD (1890) *A Catalogue of British Fossils. Vertebrata* (Dalau, London).
25. Watson DMS (1938) On *Rhamphodopsis*, a ptyctodont from the Middle Old Red Sandstone of Scotland. *Trans R Soc Edin Earth Sci* 59:397-410.
26. Traquair RH (1890a) On the fossil fishes found at Achanarras Quarry, Caithness. *Ann Mag Nat Hist* 6:249-252.
27. Denison RH (1978) *Handbook of Paleoichthyology, Vol. 2: Placodermi* (Fischer, Stuttgart).
28. Denison RH (1979) *Handbook of Paleoichthyology, Vol. 5. Acanthodii* (Gustav Fischer Verlag, Stuttgart).
29. Obruchev DV (1967) in *Fundamentals of Paleontology, Vol XI Agnatha, Pisces*, ed, YA Orlov (Israel Program for Scientific Translations, Jerusalem), pp. 168-259.
30. Vorobyeva EI, Obruchev DV (1967) in *Fundamentals of Paleontology, Vol XI Agnatha, Pisces*, ed, YA Orlov (Israel Program for Scientific Translations, Jerusalem), pp. 420-509.
31. Marss T, Kleesment A, Niit M (2008). *Karksilepis parva* gen. et sp. nov. (Chondrichthyes) from the Burtnieki Regional Stage, Middle Devonian of Estonia. *Eston J Earth Sci* 57:219-230.
32. Kleesment A, Mark-Kurik E (1997) in *Geology and Mineral Resources of Estonia*, eds. A Ruakas, A Teedumae (Estonian Academy, Tallinn), pp. 107-121.

33. Valiukevicius J (1988) New species of acanthodians from the Middle Devonian of the Baltic region and Belorussia. *Palaeont J* 22:75-81.
34. Valiukevicius J (1995) Acanthodians from marine and non-marine early and middle Devonian deposits. *Geobios* 19:393-397.
35. Mark-Kurik E (1973) *Actinolepis* (Arthrodira) from the Middle Devonian of Estonia. *Paleontographica* 143:89-108.
36. Lebedev O *et al.* (2009) Bite marks as evidence of predation in early vertebrates. *Acta Zool* 90:344-356.
37. Luksevics E (2001) Bothriolepid antiarchs (Vertebrata, Placodermi) from the Devonian of the Northwestern part of the East European Platform. *Geodiversitas* 23:489-609.
38. Hussakof L (1906) Studies on the Arthrodira. *Mem Am Mus Nat Hist* 9:106-150.
39. Maisey JG (1984) Studies on the Paleozoic selachian genus *Ctenacanthus* Agassiz. No. 3. Nominal species referred to *Ctenacanthus*. *Amer Mus Nov* 2774:1-20.
49. Stensio EA (1925) On the head of macropetalichthyids with certain remarks on the head of other arthrodiras. *Feldiana* 4:89-198.
41. Andrews SM, Gardiner BG, Miles RS, Patterson C (1967) in *The Fossil Record* (Geol Soc London, Northern Ireland), pp. 637-683.
42. Forey P, Ahlberg PE, Luksevics E, Zupins I (2000) A new coelacanth from the middle Devonian of Latvia. *J Vert Paleo* 20:243-252.
43. Eastman CR (1898) Dentition of Devonian ptyctodontidae. *Am Nat* 32:545-560.
44. Frickhinger KA (1991) *Fossilien Atlas Fische* (Mergus, Germany).
45. Case EC (1931) Arthrodiran remains from the Devonian of Michigan. *Contrib Mus Paleont Univ Michigan* 3:163-182.
46. Wells JW (1942) Arthrodiran fish plates from the Enfield formation (Upper Devonian) of New York. *J Paleont* 16:651-656.
47. Mitchell SW (1971) A new occurrence of the Devonian arthrodire *Holonema*. *Ohio J Sci* 71:120-124.
48. Dorr JA, Eschman DF (1970) *Geology of Michigan* (Univ. Michigan, Ann Arbor).
49. Schultze HP (1982) A dipterid dipnoan from the Middle Devonian of Michigan, USA. *J Vert Paleont* 2:155-162.

50. Ritchie A (1975) *Groenlandaspis* in Antarctica, Australia and Europe. *Nature* 254:569-573.
51. Young GC (1988) Antiarchs (placoderm fishes) from the Devonian Aztec Siltstone, southern Victoria Land, Antarctica. *Palaeontographica A* 202:1-125.
52. Young GC (1989) The Aztec fish fauna (Devonian) of Southern Victoria Land: evolutionary and biogeographic significance. *Geol Soc London* 47:43-62.
53. Young GC, Long JA, Ritchie A (1992) Crossopterygian fishes from the Devonian of Antarctica: systematics, relationships, and biogeographic significance. *Rec Austral Mus* 14:1-77.
54. Turner S, Young GC (1992) Thelodont scales from the Middle-Late Devonian Aztec Siltstone, southern Victoria Land, Antarctica. *Antarct Sci* 4:89-105.
55. Long JA, Young GC (1995) Sharks from the Middle-Late Devonian Aztec Siltstone, southern Victoria Land, Australia. *Rec West Austral Mus* 17:287-308.
56. Johansen Z, Ahlberg PA (2001) Devonian rhizodontids and tristichopterids (Sarcopterygii: Tetrapodomorpha) from East Gondwana. *Trans R Soc Edinburgh Earth Sci* 92:43-74.
57. Young GC, Burrow CJ (2004) Diplacanthid acanthodians from the Aztec Siltstone (late Middle Devonian) of southern Victoria Land, Antarctica. *Fossils & Strata* 50:23-43.
58. Young GC, Long JA (2005) Phyllolepid placoderm fish remains from the Devonian Aztec Siltstone, southern Victoria Land, Australia. *Antarct Sci* 17:387-408.
59. Long JA, Choo B, Young GC (2008) A new basal actinopterygian from the Middle Devonian Aztec Siltstone of Antarctica. *Antarct Sci* 20:393-412.
60. Burrow CJ, Long JA, Trinajstić K (2009) Disarticulated acanthodian and chondrichthyan remains from the upper Middle Devonian Aztec Siltstone, southern Victoria Land, Antarctica. *Antarct Sci* 21:71-88.
61. Chang K (1965) New antiarchs from the Middle Devonian of Yunnan. *Vert Palasiat* 7:342-351.
62. P'an K, Wang S-T (1978) in *Symposium on the Devonian System of south China* (Geol Publ House, Beijing), pp. 298-333.
63. Wang J, Wang N (1983) A new genus of Coccosteidae. *Vert Palasiat* 21:1-8.
64. Wang J, Wang N (1984) New material of the arthrodira from the Wuding Region, Yunnan. *Vert Palasiat* 22:1-7.

65. Liu Y, Wang J (1981) On three new arthrodires from the Middle Devonian of Yunnan. *Vert Palasiat* 19:295-304.
66. Liu S (1983) Biogeography of Silurian and Devonian vertebrates in China. *Vert Palasiat* 21:97-102.
67. Pan J, Dineley DL (1988) A review of early (Devonian and Silurian) vertebrate biogeography and biostratigraphy of China. *Proc R Soc London B* 235:29-61.
68. Marsden MAH (1976) Upper Devonian Carboniferous. *Spec Publ Geol Soc Austral* 5:77-124.
69. Long JA (1983) New bothriolepid fishes from the Late Devonian of Victoria, Australia. *Palaeontology* 26:295-320.
70. Long JA (1984) New phyllolepid fish from Victoria, and the relationships of the group. *Proc Linn Soc NSW* 107:263-308.
71. Long JA (1988) New palaeoniscoid fishes from the Late Devonian and Early Carboniferous of Victoria. *Mem Ass Australas Paleontols* 7:1-64.
72. Long JA (1992) Cranial anatomy of two new Late Devonian lungfishes (Pisces, Dipnoi) from Mt. Howitt, Victoria. *Records Austral Mus* 44: 299-318.
73. Long, JA (1999) A new genus of fossil coelacanth (Osteichthyes: Coelacanthiformes) from the middle Devonian of southeastern Australia. *Records West Austral Mus* 57:169-174.
74. Moloshnikov SV (2008) Devonian antiarchs (Pisces, Antiarchi) from central and southern European Russia. *Paleo J* 7:691-773.
75. Newberry JS (1889) *The Paleozoic Fishes of North America* (USGS, Washington).
76. Claypole EW (1883) Note on a large fish-plate from the Upper Chemung Beds of North Pennsylvania. *Proc Am Philos Soc* 20:664-666.
77. Claypole EW (1890) Paleontological notes from Indianapolis. *Am Geol* 6:255-260.
78. Eastman CR (1897) On the relations of certain plates in the Dinichthyids. *Bull Mus Comp Zool* 31:19-44.
79. Miller SA (1892) *North American Geology and Paleontology* (Western Methodist Book Concern, Cincinnati)
80. Eastman CR (1908) *Devonian fishes of Iowa* (Iowa Geol Surv, Des Moines).
81. Denison RH (1985) A new ptyctodontid placoderm, *Ptyctodopsis*, from the Middle Devonian of Iowa. *J Paleo* 59:511-522.
82. Schultze H-P (1992) A new long-headed dipnoan (Osteichthyes) from the Middle Devonian of Iowa, USA. *J Vert Paleo* 12:42-58.

83. Upeniece I (1995) A new species of *Strunius* (Sarcopterygii, Onychodontida) from Latvia, Lode Quarry (Upper Devonian). *Geobios* 19:281-284.
84. Upeniece I (2001) The Unique fossil assemblage of the Lode Quarry (Upper Devonian, Latvia). *Mitt. Mus. Nat.kd. Berl., Geowiss. Reihe*
85. Vorobyeva EI (1995) The shoulder girdle of *Panderichthyes rhombolepis* (Gross) (Crossopterygii), Upper Devonian, Latvia. *Geobios* 19:285-288.
86. Vorobyeva EI (2006) A new species of *Laccognathus* (Porolepiform Crossopterygii) from the Devonian of Latvia. *Paleont J* 40: 312-332.
87. Ahlberg PE, Luksevics E, Mark-Kurik E (2000) A near tetrapod from the Baltic Middle Devonian. *Palaeontology* 43:533-548.
88. Zupins I (2008) A new tristichopterid (Pisces, Sarcopterygii) from the Devonian of Latvia. *Proc Latv Acad Sci* 62:40-46.
89. Ebbighausen V, Becker RT, Bockwinkel RT, Aboussalam ZS (2008) in *Devonian Events and Correlations*, eds. RT Becker and WT Kirchgasser (Geological Society, Bath), pp. 157-172.
90. Schultze, H-P (1969) *Griphognathus* Gross, ein langschnauziger Dipnoer aus dem Oberdevon von Bergisch-Gladbach (Rheinisches Schiefergebirge) und von Lettland. *Geol et Palaeo* 3:21-79.
91. Orvig T (1960) New finds of acanthodians, arthrodires, crossopterygians, ganoids and dipnoans in the Upper Middle Devonian Calcareous Flags (Oberer Plattenkalk) of the Bergisch Gladbach-Paffrath Trough. Part 1. *Palaont Zeit* 34:295-335
92. Orvig T (1961) New finds of acanthodians, arthrodires, crossopterygians, ganoids and dipnoans in the Upper Middle Devonian Calcareous Flags (Oberer Plattenkalk) of the Bergisch Gladbach-Paffrath Trough. Part 2. *Palaont Zeit* 35:10-27.
93. Jessen H (1968) *Moythomasia nitida* Gross und *M. cf. striata* Gross, devonische Palaeonisciden aus dem Oberen Plattenkalk der Bergisch-Gladbach --- Paffrather Mulde (Rheinisches Schiefergebirge). *Palaeontographica A* 128:87-114.
94. Miles RS (1966) The acanthodian fishes of the Devonian Plattenkalk of the Paffrath Trough in the Rhineland. *Ark Zool* 18:147-194
95. Miles RS (1971) The Holonematidae (placoderm fishes): a review based on new specimens of *Holonema* from the Upper Devonian of Western Australia. *Phil Trans R Soc London B*. 263:101-234.
96. Friedman M, Coates MI (2005) A newly recognized fossil coelacanth highlights the early morphological diversification of the clade. *Proc R Soc B* 273:245-250.

97. Long JA, Trinajstic K, Young GC, Senden T (2008) Live birth in the Devonian period. *Nature* 453:650-652.
98. Choo B, Long JA, Trinajstic K (2009) A new genus and species of basal actinopterygian fish from the Upper Devonian Gogo Formation of Western Australia. *Acta Zool* 90:194-210.
99. Andrews SM *et al.* (2006) The structure of the sarcopterygian *Onychodus jandemarrai*, n. sp. from Gogo, Western Australia: with a functional interpretation of the skeleton. *Trans R Soc Edinburgh* 96:197-307.
100. Campbell KSW, Barwick RE (1990) Palaeozoic dipnoan phylogeny: functional complexes and evolution without parsimony. *Palaeontology* 16:143-169.
101. Campbell KSW, Barwick. RE (1998) A new tooth-plated dipnoan from the Upper Devonian Gogo formation and its relationships. *Mem Queensland Mus* 42: 403-437.
102. Dennis KD, Miles RS (1979a) A second eubrachythoracid arthrodire from Gogo, Western Australia. *Zool J Linn Soc* 67:1-29.
103. Dennis KD, Miles, RS. (1979b) Eubrachythoracid arthrodires with tubular rostral plates from Gogo, Western Australia. *Zool J Linn Soc* 67:297-328.
104. Dennis KD, Miles RS. (1980) New durophagous arthrodires from Gogo, Western Australia. *Zool J Linn Soc* 69:43-85.
105. Dennis KD, Miles RS (1981) A pachyosteomorph arthrodire from Gogo, Western Australia. *Zool J Linn Soc* 73:213-258.
106. Dennis KD, Miles, RS (1982) A eubrachythoracid arthrodire with a snub-nose from Gogo, Western Australia. *Zool J Linn Soc* 75:153-166.
107. Dennis-Bryan K (1987) A new species of eastmanosteid arthrodire (Pisces: Placodermi) from Gogo, Western Australia. *Zool J Linn Soc* 90:1-64.
108. Dennis-Bryan K, Miles RS (1983) Further eubrachythoracid arthrodires from Gogo, Western Australia. *Zool J Linn Soc* 67:1-29.
109. Andrews SM, Miles RS, Walker AD (1977) *Problems in Vertebrate Evolution* (Academic, London).
110. Gardiner BG, Miles RS (1990) A new genus of eubrachythoracid arthrodire from Gogo, Western Australia. *Zool J Linn Soc* 99:159-204.
111. Gardiner BG, Miles RS (1994) Eubrachythoracid arthrodires from Gogo, Western Australia. *Zool J Linn Soc* 112:443-477.
112. Long JA (1985a) The structure and relationships of a new osteolepiform fish from the Late Devonian of Victoria, Australia. *Alcheringa* 9:1-22.

113. Long JA (1985b) A new osteolepidid fish from the upper Devonian Gogo formation, Western Australia. *Rec West Austral Mus* 12:361-377.
114. Long JA (1987) A description of the lungfish *Eoectenodus microsoma* (Hills 1929) with a reassessment of the genus *Dipterus* (Sedgwick and Murchison 1928) in Australia. *Rec West Austral Mus* 13:297-314.
115. Long JA (1988) A new camuropiscid arthrodire (Pisces: Placodermi) from Gogo, Western Australia. *Zool J Linn Soc* 94:233-258.
116. Long JA (1990) Two new arthrodires (placoderm fishes) from the upper Devonian Gogo formation, Western Australia. *Mem Queensland Mus* 28:51-63.
117. Long JA.(1992) *Gogodipterus paddyensis* gen. nov. a new chirodipterid lungfish from the late Devonian Gogo formation, Western Australia. *Beagle NT Mus* 9:10.
118. Long JA (1994) A second incisoscutid arthrodire from Gogo, Australia. *Alcheringa* 18:59-69.
119. Long JA (1995) A new plourdosteid arthrodire from the Upper Devonian Gogo formation of Western Australia. *Palaeontology* 38:39-62.
120. Long JA (1997) Ptyctodont fishes from the Late Devonian Gogo Formation, Western Australia, with revision of the European genus *Ctenurella* Orvig 1960. *Geodiversitas* 19:515-555.
121. Long JA (2007) *Swimming in Stone: The Amazing Gogo Fossils of the Kimberley* (Fremantle, Fremantle).
122. Miles RS (1977) Dipnoan (lungfish) skulls from the Upper Devonian of Western Australia. *Zool J Linn Soc* 61:31-62.
123. Miles RS, Dennis K (1979) A primitive eubrachythoracid arthrodire from Gogo, Western Australia. *Zool J Linn Soc* 66:31-62
124. Miles RS, Young GC (1977) in *Problems in Vertebrate Evolution*, eds. SM Andrews, RS Miles, AD Walker (Academic, London), pp. 123-198.
125. Trinajstić K (1999) The Late Devonian palaeoniscoid *Moythomasia durgaringa* Gardiner and Bartram 1977. *Alcheringa* 23:9-19.
126. Hairapetian J, Gholamalian H (1998) First report on the Late Devonian fish remains and microvertebrate fragments in the Chahriseh area, north east of Esfahan, Iran. International Geological Correlation Program 421, Esfahan, Iran. 5-20 December, 1998. Abstract 15.
127. Hairapetian V, Ginter M, Yazdi M (2008) Early Frasnian sharks from central Iran. *Acta Geol Polon* 58:173-179.

128. Hairapetian V, Ginter M (2009) Famennian chondrichthyan remains from the Chahriseh section, central Iran. *Acta Geol Polon* 59:173-200
129. Hairapetian V, Valiuvicius J, Burrow, CJ (2006) Early Frasnian acanthodians from central Iran. *Acta Palaeo Polon* 51:499-520.
130. Walliser OH (1965) Preliminary notes on Devonian, Lower and Upper Carboniferous goniatites in Iran. *Geol Surv Iran Rep* 6:7-24.
131. Schultze H-P (1973) Large Upper Devonian arthrodires from Iran. *Fieldiana* 23:53-78.
132. Janvier P (1974) A preliminary report on Late Devonian fishes from Central and Eastern Iran. *Geol Surv Iran Rep* 31:5-47.
133. Janvier P (1977) Les Poissons devoniens de l'Iran central et de L'Afghanistan. *Mem h ser Soc geol France* 8:277-289.
134. Janvier P, Martin M (1978) Les vertebrates devoniens de L'Iran Central I: Dipnuestes. *Geobios* 11:819-833.
135. Janvier P, Martin M (1979) Les Vertebrates devoniens de L'Iran central II- Coelacanthiformes, Struniiformes, Osteolepiformes. *Geobios* 12:497-511.
136. Janvier P (1979) Les vertebrates devoniens de L'Iran Central III: antiarches. *Geobios* 12:605-608.
137. Lelievre H, Janvier P, Goujet D (1981) Les Vertebrates devoniens de L'Iran central IV – arthrodires et ptyctodontes. *Geobios* 14:677-709.
138. Maisch MW (1998) *Wildungenichthys grossi* n. gen., n. sp. – a new selenosteid arthrodire (Placodermi, Arthrodira) from the Kellwasserkalk (late Frasnian, Upper Devonian) of Bad Wildungen (Hessen, W-Germany). *Palaont Zeit* 72:373-382.
139. Wendt J, et al. (2005) Devonian/Lower Carboniferous stratigraphy, facies patterns and paleogeography of Iran Part II. Northern and central Iran. *Acta Geol Pol* 55:31-97.
140. Newberry JS (1875) Description of Fossil Fishes. *Geol Surv Ohio Rep* 2:1-64.
141. Clarke JM (1885) On the higher Devonian faunas of Ontario County, New York. *US Geol Surv Bull* 16:1-80.
142. Eastman CR (1907) Devonian fishes of the New York formations. *New York St Mus Mem* 10:1-235.
143. Hussakof L, Bryant WL (1918) Catalogue of fossil fishes in the museum of the Buffalo Society of Natural Sciences. *Buffalo Soc Nat Sci Bull* 12:1-346.
144. Bryant WL (1923) New Upper Devonian fishes from western New York. *Buffalo Soc Nat Hist Bull* 17:18-42

145. Youngquist W, Hibbard RR, Reiman IG (1948) Additions to the Devonian Conodont faunas of Western New York. *J Paleont* 22:48-59
146. Burrow CJ (2004) A redescription of *Atopacanthus dentatus* Hussakof and Bryant, 1918 (Acanthodii, Ischnacanthidae). *J Vert Paleo* 24:257-267.
147. Stahl BJ (1999) *Handbook of Paleichthyology, Vol. 4, Chondrichthyes III, Holocephalii*. (Verlag Dr. Friedrich Pfiel, Munchen).
148. Zangerl R (1981) *Handbook of Paleichthyology, Vol. 3A. Chondrichthyes I, Paleozoic Elasmobranchii*. (Verlag Dr. Friedrich Pfiel, Munchen).
149. Vezina D (1996) in *Devonian Fishes and Plants of Miguasha, Quebec, Canada*, eds. H-P Schultze, R Cloutier (Verlag Dr. Friedrich Pfiel, Munchen), pp. 141-148.
150. Schultze H-P (1996) in *Devonian Fishes and Plants of Miguasha, Quebec, Canada*, eds. H-P Schultze, R Cloutier (Verlag Dr. Friedrich Pfiel, Munchen), pp. 120-122.
151. Schultze H-P, Cloutier R (1996) in *Devonian Fishes and Plants of Miguasha, Quebec, Canada*, eds. H-P Schultze, R Cloutier (Verlag Dr. Friedrich Pfiel, Munchen), pp. 348-368.
152. Prichonnet G, Di Vergilio, M, Chidiac Y (1996) in *Devonian Fishes and Plants of Miguasha, Quebec, Canada*, eds. H-P Schultze, R Cloutier (Verlag Dr. Friedrich Pfiel, Munchen), pp. 23-36.
153. Parent N, Cloutier R (1996) in *Devonian Fishes and Plants of Miguasha, Quebec, Canada*, eds. H-P Schultze, R Cloutier (Verlag Dr. Friedrich Pfiel, Munchen), pp. 54-78.
154. Gorizdro-Kulczycka Z (1934) Ptyctodontidae gornodewonskie z Gor Swietokrzyskich. *Prace Polsk Inst Geol* 3:1-38.
155. Gorizdro-Kulczycka Z (1950) Les Dipnuestes devonienes du Massif de S-te Croix. *Acta Geol Pol* 1:82-105
156. Kulczycki J (1957) Upper Devonian fishes from the Holy Cross Mountains. *Acta Paleo Polon* 1:103-111.
157. Ginter M (2002) Chondrichthyan fauna of the Frasnian-Famennian boundary beds in Poland. *Acta Paleo Polon* 47:329-338.
158. Ivanov A, Ginter M (1997) Comments on the Late Devonian placoderms from the Holy Cross Mountains (Poland). *Acta Paleo Polon* 42:413-426.
159. Ginter M, Ivanov A (1992) Devonian phoebodont shark teeth. *Acta Paleo Polon* 37:55-75.
160. Blom H, Clack JA, Ahlberg PE, Friedman M (2007) Devonian vertebrates from East Greenland: a review of faunal composition and distribution. *Geodiversitas* 29:119-141.
161. McGhee GR, Sutton RG (1983) Evolution of late Frasnian Marine environments in

New York and the central Appalachians. *Alcheringa* 7:9-21.

162. Rontondo KA, Over, DJ. 2000. Biostratigraphic age of the Belpre Ash (Frasnian), Chattanooga and Rhinestreet shales in the Appalachian Basin. Geological Society of America, Northeastern Section, 35th annual meeting, Abstracts with Programs 32:A-70.
163. Daeschler EB (2000) An early actinopterygian fish from the Catskill formation (Late Devonian, Famennian) in Pennsylvania, USA. *Proc Acad Nat Sci Philadelphia* 150: 181-192.
164. Maisey JG (1981) Studies on the Paleozoic selachian genus *Ctenacanthus* Agassiz no. 1. Historical review and revised diagnosis of *Ctenacanthus*, with a list of referred taxa. *Am Mus Nov* 2718:1-22.
165. Williams ME (1985) The “Cladodont level” sharks of the Pennsylvanian Black Shales of Central North America. *Palaeontographica A* 190:83-158.
166. Gross W (1953) Devonische palaeonisciden-reste in Mittel- und Osteuropa. *Palaont Zeit* 27:85-112.
167. Krupina N (1995) New species of *Rhinodipterus* (Dipnoi) from the Upper Devonian of Northwestern Russia. *Geobios* 19:269-274.
168. Thomson KS (1973) Observations on a new rhipidistian fish from the Upper Devonian of Australia. *Palaeontographica A* 143:209-220.
169. Johansen Z, Ahlberg PE (1997) A new tristichopterid (Osteolepiformes: Sarcopterygii) from the Mandagery Sandstone (Late Devonian, Famennian) near Canowindra, NSW, Australia. *Trans R Soc Edinburgh* 88:39-68.
170. Ahlberg PE, Johansen Z (1997) Second tristichopterid (Sarcopterygii, Osteolepiformes) from the Upper Devonian of Canowindra, New South Wales, Australia and the phylogeny of the Tristichopteridae. *J Vert Paleont* 17:653-673.
171. Johansen Z (1997) New *Remigolepis* from Canowindra, NSW, Australia. *Geol Mag* 134:813-846.
172. Johansen Z, Ahlberg PE (1998) A complete primitive rhizodont from Australia. *Nature* 394:569-573.
173. Ahlberg PE, Johansen Z, Daeschler EB (2001) The late Devonian lungfish *Soederberghia* (Sarcopterygii, Dipnoi) from Australia and North America and its biogeographical implications. *J Vert Paleont* 21:1-12.
174. Snitting D (2009) *Heddleichthys* – a new tristichopterid genus from the Dura Den formation, Midland Valley, Scotland (Famennian, Late Devonian). *Acta Zool* 90:273-284.

175. Anderson J (1859) *Dura Den; a monograph of the Yellow Sandstone and its remarkable fossil remains* (Thomas Constable, Edinburgh).
176. Traquair RH (1892) A new fossil fish from Dura Den. *Ann Scot Nat Hist* 1:233-255.
177. Ginter M, Ivanov A (1990) Late Famennian shark teeth from the Holy Cross Mountains, Central Poland. *Acta Geol Pol* 40:69-81.
178. Ginter M, Ivanov A (1992) Devonian phoebodont shark teeth. *Acta Paleo Pol* 37:55-75.
179. Liszkowski J, Racki G (1993) Ichthyoliths and deepening event in the Devonian platform of the Holy Cross Mountains. *Acta Paleo Pol* 42:413-426.
180. Ivanov A, Ginter M (1997) Comments on the Late Devonian placoderms from the Holy Cross Mountains (Poland). *Acta Paleo Pol* 42:401-426.
181. Szrek P (2004) The first articulated antiarch (Vertebrata, Placodermi) from the Upper Devonian of the Holy Cross Mountains (central Poland). *Acta Geol Pol* 54:401-406.
182. Szrek P (2007) Coelacanths (Actinistia, Sarcopterygii) from the Famennian (Upper Devonian) of the Holy Cross Mountains, Poland. *Acta Geol Pol* 57:403-413.
183. Derycke C, Spalletta C, Perri MC, Corradini C. (2008) Famennian chondrichthyan microremains from Morocco and Sardinia. *J Palaeont* 82:984-995.
184. Luksevics E (1992) Palaeoichthyocenoses of the Famennian brackish seas of the Baltic area. *Academia* 1:273-280.
185. Blicek A, et al. (2007) in *Devonian Events and Correlations*, eds RT Becker and WT Kirchgasser (Geol Soc London, Northern Ireland), pp. 219-235.
186. Lebedev O (1995) Middle Famennian (Upper Devonian) chondrichthyans and sarcopterygians from Oryol region, central Russia. *Geobios* 19:361-368.
187. Krupina N (1999) A new dipnoan from the Upper Devonian locality Rybnitsa in the Oryol region. *Palaeont J* 33:627-629.
188. Lebedev O (2004) A new tetrapod *Jakubsonia livnensis* from the Early Famennian (Devonian) of Russia and palaeoecological remarks on the Late Devonian tetrapod habitats. *Acta Univ Latv* 679:79-98.
189. Young GC (1999) Preliminary report on the biostratigraphy of new placoderm discoveries in the Hervey Group (Upper Devonian) of central New South Wales. *Records West Austral Mus* 57:139-150.
190. Long JA, Burrow CJ, Ritchie A (2004) A new Late Devonian acanthodian fish from the Hunter Formation near Grenfell, New South Wales. *Alcheringa* 28:147-156.

191. Johansen Z, Ritchie A (2000) Rhipidistians (Sarcopterygii) from the Hunter Siltstone (Late Famennian) near Grenfell, NSW, Australia. *Mitt Mus Natkd Berl Geowiss Reihe* 3:111-136.
192. Ritchie A, Wang ST, Young GC, Zhang GR (1992) The Sinolepidae, a family of antiarchs (placoderm fishes) from the Devonian of south China and eastern Australia. *Rec Austral Mus* 44:319-370.
193. Johansen Z (1997) New antiarchs (Placodermi) from the Hunter Siltstone (Famennian) near Grenfell, NSW. *Alcheringa* 21:191-217.
194. Daeschler EB, Clack JA, Shubin NH (2009) Devonian tetrapod remains from Red Hill, Pennsylvania, USA: how much diversity? *Acta Zool* 90:306-317.
195. Friedman M, Daeschler EB (2007) Late Devonian (Famennian) lungfishes from the Catskill Formation of Pennsylvania, USA. *Palaeontology* 49:1167-1183.
196. Daeschler EB, Frumes AC, Mullison CF (2003) Groenlandaspid placoderm fishes from the Late Devonian of North America. *Rec Austral Mus* 55:45-60
197. Lane JA, Cuffey RJ (2005) *Phyllopepis rossimontina* sp. nov. (Placodermi) from the uppermost Devonian at Red Hill, north-central Pennsylvania. *Revist Brasil Paleont* 8:117-126.
198. Daeschler EB, Shubin NH, Thomson KS, Amaral WW (1994) A Devonian tetrapod from North America. *Science* 265:639-642.
199. Daeschler EB (2000) Early tetrapod jaws from the Late Devonian of Pennsylvania, USA. *J Paleont* 74:301-308.
200. Taverne L (1997) *Osorioichthyes marginis*, "Paleonisciforme" du Famennian de Belgique et la phylogenie des Actinopterygiens devoniens (Pisces). *Bull Inst R Sci Nat Belg* 67:57-78.
201. Clement G (2002) Large Tristichopteridae(Sarcopterygii, Tetrapodamorpha) from the Late Famennian Evieux Formation of Belgium. *Palaeontology* 45:577-593.
202. Clement G, et al. (2004) Palaeogeography: Devonian tetrapod from western Europe. *Nature* 427:412-413.
203. Janvier P, Clement G (2005) A new groenlandaspidid arthrodire (Vertebrata: Placodermi) from the Famennian of Belgium. *Geol Belg* 8:51-67.
204. Blicq A, Lelievre H (1995) Palaeozoic vertebrates of northern France and Belgium. Part I: Heterostraci, Osteostraci, Thelodonti, Placodermi (Devonian). *Geobios* 19:311-317.
205. Derycke C, Cloutier R, Candilier A-M (1995) Palaeozoic vertebrates of northern France and Belgium: Part II – Chondrichthyes, Acanthodii, Actinopterygii (Uppermost Silurian to Carboniferous) *Geobios* 19:343-350.

206. Cloutier R, Candilier A-M (1995) Paleozoic vertebrates of northern France and Belgium: Part III – Sarcopterygii (Devonian to Carboniferous). *Geobios* 19:335-341.
207. Luksevics E (1991) Bothriolepids of the Ketleri Beds of the Upper Devonian of Latvia (Pisces, Placodermi) *Daba un muzejs* 3:38-50.
208. Ahlberg PE, Luksevics E, Lebedev O (1994) The first tetrapod finds from the Devonian (Upper Famennian) of Latvia. *Phil Trans R Soc B* 343:303-328.
209. Lehman JP (1976) Nouveaux poissons fossiles du Devonien du Maroc. *Ann Paleont* 62:1-34.
210. Lehman JP (1977) Nouveaux arthrodières du Tafilalet et de ses environs. *Ann Paleont* 63:105-132.
211. Lelievre JP, Janvier P (1986) L'Eusthenoptéride (Osteichthyes, Sarcopterygii) du Famennien (Devonien supérieur) du Tafilalet (Maroc): nouvelle description. *Bull Mus Nat Hist Paris* 8:351-365.
212. Lelievre JP, Janvier P (1988) Un actinoptérien (Sarcopterygii, Vertebrata) dans le Devonien supérieur du Maroc. *Comptes-rendus Acad Sci Paris, Paleont* 307:1425-1430.
213. Lebedev O (1992) The latest Devonian, Khovanian vertebrate community of Andreyevka-2 locality, Tula region, Russia. *Academia* 1:265-272.
214. Lebedev O, Clack JA (1993) Devonian tetrapod remains from Andreyevka, Tula, Russia. *Palaeontology* 36:721-734.
215. Alekseev AA *et al.* (1994) On the stratigraphic positions of the Famennian and Tournaisian fossil vertebrate beds in Andreyevka, Tula region, central Russia. *Proc Geol Ass* 105:41-52.
216. Carr RK, Jackson GL (2010) in *Guide to the Geology and Paleontology of the Cleveland Member of the Ohio Shale*, ed. JT Hannibal (Ohio Geol Surv, Ohio), Chapter 5.
217. Carr RK (1995) Opportunity knocked and no one was home: aspinothoracid arthrodières (placodermi) from the Ohio Shale Formation (Upper Devonian, North America). *Geobios* 19:81-83.
218. Dunkle D (1964) Preliminary description of a paleoniscoid fish of the Upper Devonian. *Sci Pub Cleveland Mus Nat Hist* 3:1-16.
219. Ginter M (2008) Devonian filter-feeding sharks. *Acta Geol Pol* 58:147-153.
220. Pilgrim BL, Franz-Odenaal TA (2009) Comparative study of the ocular skeleton of fossil and living chondrichthyans. *J Anat* 214:248-258.
- 221 Bryant WL, Johnson JH (1936) Devonian fish from Colorado. *J Palaeont* 10:656-659.

222. Denison RH (1951) Late Devonian fresh-water fishes from the western United States. *Fieldiana* 11:225-261.
223. Pampe WR (1969) Late Devonian fish remains from central Colorado. *J Paleont* 43:1111-1113.
224. Ginter M (2001) Chondrichthyan biofacies in the Late Famennian of Utah and Nevada. *J Vert Paleont* 21:714-729.
225. Schultze H-P, Chorn J (1998) Sarcopterygian and other fishes from the marine Upper Devonian of Colorado, USA. *Mitt Mus Natkd Berl Geowiss Reihe* 1:53-72.
226. Anderson ME, Hiller N, Gess RW (1994) The first *Bothriolepis*-associated Devonian fish fauna from Africa. *So Afr J Sci* 90:397-403.
227. Gess RW, Hiller N (1995) A preliminary catalogue of the fossil algal, plant, arthropod and fish remains from a Late Devonian black shale near Grahamstown, South Africa. *Ann Cape Prov Mus Nat Hist* 19:225-304.
228. Long JA, Anderson ME, Gess RW, Hiller N (1997) Placoderm fishes from the Late Devonian of South Africa. *J Vert Paleont* 17:253-268.
229. Anderson ME, Long JA, Gess RW, Hiller N (1999). A new fossil shark (Pisces: Chondrichthyes) from the Late Devonian of South Africa. *Records West Austral Mus* 57:151-156.
230. Murray AM (2000) The Palaeozoic, Mesozoic, and Early Cenozoic fishes of Africa. *Fish Fisher* 1:111-145.
231. Gess RW (2001) A new species of *Diplacanthus* from the Late Devonian (Famennian) of South Africa. *Ann Paleont* 87:49-60.
232. Gardiner BG (1966) Catalogue of Canadian fossil fishes. *Life Sci, R Ontario Mus Univ Toronto* 68:1-154.
233. Miller RF, McGovern JH (1996) in *Current Research 1996: New Brunswick Department of Natural Resources and Energy; Minerals, Policy and Planning Division, Mineral Resource Report MRR 97-4*, ed. BMS Carroll (New Brunswick Dept of Natural Resources, New Brunswick), pp. 191-200.
234. Brazeau MD (2005) A new genus of rhizodontid (Sarcopterygii, Tetrapodomorpha) from the Lower Carboniferous Horton Bluff Formation of Nova Scotia, and the evolution of the lower jaws in this group. *Can J Earth Sci* 42:1481-1499.
235. Jeffery JE (2006) The Carboniferous fish genera *Strepsodus* and *Archichthyes* (Sarcopterygii: Rhizodontida): clarifying 150 years of confusion. *Palaeontology* 49:113-132.
236. Rygel MC *et al.* (2006) Tournaisian forested wetlands of the Horton Group of Atlantic Canada. *GSA Spec Pap* 399:103-126.

237. Berg LS, Kazantseva AA, Obruchev DV (1967) in *Fundamentals of Paleontology, Vol XI Agnatha, Pisces*, ed, YA Orlov (Israel Program for Scientific Translations, Jerusalem), pp. 528-572.
238. Kazantseva AA (1968) in *Ocherki po filogenii i sistematike beschelyustnykh i ryb*, ed. VV Menner (Nauka, Moscow), pp. 98-115.
239. Prokofiev AM (2004) A remarkable new paleoniscoid fish (Osteichthyes:Actinopterygii) from the lower Carboniferous of Tuva Republic (Russia). *Paleont Res* 8:109-114.
240. Beznosov P (2009) A redescription of the Early Carboniferous acanthodian *Acanthodes lopatini*. *Acta Zool* 90:183-193.
241. Woodward AS (1906) On a Carboniferous fish fauna from the Mansfield District, Victoria. *Mem Nat Mus Melbourne* 1:1-32.
242. Warren A, Currie BP, Burrow CJ, Turner S (2000) A redescription and reinterpretation of *Gyracanthides murrayi* Woodward 1906 (Acanthodii, Gyracanthidae) from the Lower Carboniferous of Mansfield Basin, Victoria, Australia. *J Vert Paleont* 20:225-242.
243. Holland TM, Long JA, Warren A, Garvey JM (2006) Second specimen of the lower actinopterygian *Novogonatodus* from the Early Carboniferous of Mansfield, Victoria. *Proc R Soc Victoria* 118:1-10.
244. Garvey JM, Turner S (2006) Vertebrate microremains from the presumed earliest Carboniferous of Mansfield Basin, Victoria. *Alcheringa* 30:43-62.
245. Garvey JM, Hasiotis ST (2008) Microfossil assemblage from the Lower Carboniferous Snowy Plains Formation, Mansfield Basin, Australia. *Palaeogeog Palaeoclim Palaeoecol* 258:257-276.
246. White EI (1925) The fish fauna of the Cementstones of Foulden, Berwickshire. *Trans R Soc Edinburgh Earth Sci* 55:255-287.
247. Moy-Thomas JA (1938) Carboniferous palaeoniscoids from Northumberland and Berwickshire. *Geol Mag* 75:308-318.
248. Gardiner BJ (1985) Actinopterygian fish from the Dinantian of Foulden, Berwickshire, Scotland. *Trans R Soc Edinburgh Earth Sci* 76:61-66.
249. Lebedev O (1996) Fish assemblages in the Tournaisian-Viséan environments of the East European Platform. *Geol Soc Spec Publ* 107:387-415.
250. Gardiner BG (1969) New palaeoniscoid fish from the Witteberg series of South Africa. *Zool J Linn Soc* 48:423-452.
251. Jubb RA, Gardiner BG (1975) A preliminary catalogue of identifiable fossil fish material from Southern Africa. *Ann S Afr Mus* 6:381-440.

252. Evans FJ (1999) Palaeobiology of the Early Carboniferous lacustrine biota of the Waaipoort Formation (Witteberg Group), South Africa.
253. Moy-Thomas JA (1936) On the structure and affinities of the fossil elasmobranch fishes from the Lower Carboniferous of Glencartholm, Eskdale. *Proc Zool Soc London B* 1936:762-788.
254. Moy-Thomas JA, Bradley-Dyne M (1938) Actinopterygin fishes from the Carboniferous of Glencartholm, Eskdale, Dumfriesshire. *Trans R Soc Edinburgh Earth Sci* 59:437-480.
255. Coates MI, Gess RW (2007) A new reconstruction of *Onychoselache traquairi*, comments on early chondrichthyan pectoral girdles and hybodontiform phylogeny. *Palaeontology* 50:1421-1446.
256. Traquair RH (1877a) On new and little-known fossil fishes from the Edinburgh district, No. I. *Proc R Soc Edinburgh* 9:262-272.
257. Traquair RH (1877b) On new and little-known fossil fishes from the Edinburgh district, No. II. *Proc R Soc Edinburgh* 9:275-282
258. Traquair RH (1877b) On new and little-known fossil fishes from the Edinburgh district, No. III. *Proc R Soc Edinburgh* 9:427-444.
259. Traquair RH (1890) On the fossil Dipnoi and Ganoidea of Fife and the Lothians. *Proc R Soc Edinburgh* 18:385-400.
260. Turner S, Burrow CJ, Warren A (2005) *Gyracanthides hawkinsi* sp. nov. (Acanthodii, Gyracanthidae) from the Lower Carboniferous of Queensland, Australia, with a review of gyracanthid taxa. *Palaeontology* 48:963-1006.
261. Turner S (1982) Devonian and Carboniferous vertebrates from Queensland. *Fossil Collect Bull* 9:20-21.
262. Turner S, Kemp A, Warren A (1999) First Early Carboniferous lungfish (Dipnoi, Ctenodontidae) from central Queensland. *Alcheringa* 23:177-183.
263. Johansen Z, Turner S, Warren A (2000) First East Gondwanan record of *Strepsodus* (Sarcopterygii: Rhizodontida) from the Lower Carboniferous Ducabrook Formation, central Queensland, Australia. *Geodiversitas* 22:161-169.
264. Warren AA, Ptasznick R (2002) The earliest fractured tetrapod bone. *Alcheringa* 26:459-463.
265. Warren A, Turner S (2004) First stem tetrapod from the Lower Carboniferous of Gondwana. *Palaeontology* 47: 151-184.
266. Parker KE, Webb JA (2008) Estuarine deposition of a mid-Viséan tetrapod unit, Ducabrook Formation, central Queensland: implication for tetrapod dispersal. *Austral J Earth Sci* 55:509-530.

267. Paton RL, Smithson TR, Clack JA (1999) An amniote-like skeleton from the Early Carboniferous of Scotland. *Nature* 398:508-513.
268. Coates MI (1994) Actinopterygian and acanthodian fishes from the Viséan of East Kirkton, West Lothian, Scotland. *Trans R Soc Edinburgh Earth Sci* 84:317-327.
269. Clack JA (2001) *Eucritta melanolimnetes* from the Early Carboniferous of Scotland, a stem tetrapod showing a mosaic of characters. *Trans R Soc Edinburgh Earth Sci* 92:75-95.
270. Traquair RH (1877-1914) *The Ganoid Fishes of the British Carboniferous Formations* (Palaeontographical Society).
271. Gardiner BG (1967) Further notes on palaeoniscoid fishes with a classification of the Chondrostei. *Bull Brit Mus Nat Hist* 14:143-206.
272. Traquair RH (1897) A list of fish remains occurring the bone bed at Abden, near Kinghorn, Fife. *Proc Geol Ass* 15:143-145.
- 273.. Wood SP (1982) New basal Namurian (Upper Carboniferous) fishes and crustaceans found near Glasgow. *Nature* 297:574-577.
274. Coates MI (1993) New actinopterygian fish from the Namurian Manse Burn Formation of Bearsden, Scotland. *Palaeontology* 36:123-146.
275. Coates MI (1998) Actinopterygians from the Namurian of Bearsden, Scotland, with comments on early actinopterygian neurocrania. *Zool J Linn Soc* 122:27-59.
276. Coates MI, Sequeira SK (2001) A new stethacanthid chondrichthyan from the Lower Carboniferous of Bearsden, Scotland. *J Vert Paleont* 21:438-459.
277. Traquair RH (1881) Notice of new fish remains from the Black-band Ironstone of Borough Lee near Edinburgh. *Geol Mag* 8:34-37.
278. Traquair RH (1882) Notice of new fish remains from the Blackband Ironstone of Borough Lee Edinburgh. No. III. *Geol Mag* 12:540-546.
279. Traquair RH (1903) On the distribution of fossil fish remains in the Carboniferous remains on the Edinburgh district. *Trans R Soc Edinburgh* 14:687-707.
280. Lowney KA (1983) The earliest known (Namurian A, E1) haplolepid (Osteichthyes: Actinopterygii). *Trans R Soc Edinburgh Earth Sci* 74:69-78.
281. Smithson TR (1985) The morphology and relationships of the Carboniferous amphibian *Eoherpeton watsoni* Panchen. *Zool J Linn Soc* 85:317-410.
282. Ruta M, Milner AR, Coates MI (2001) The tetrapod *Caerorhachis bairdi* Holmes and Carroll from the Lower Carboniferous of Scotland. *Trans R Soc Edinburgh Earth Sci* 92:229-261.

283. McCoy F (1848) On some new fossil fish of the Carboniferous period. *Ann Mag Nat Hist* 2:1-10,115-133.
284. Wellburn ED (1901) On the fish fauna of the Yorkshire Coal Measures. *Proc Yorkshire Geol Polytech Soc* 14:159-177
285. Wellburn ED (1903) On some new species of fossil fish from the Millstone Grit rocks, with an amended list of genera. *Proc Yorkshire Geol Polytech Soc* 15:70-78.
286. Johnson GA, Hodge L, Fairbairn RA (1962) The base of the Namurian and of the Millstone Grit in north-eastern England. *Proc Yorkshire Geol Soc* 33:341-362.
287. Clack JA (2002) *Gaining Ground: The Origin and Evolution of Tetrapods* (Indiana Univ, Bloomington).
288. Hampe O, Johnson GD, Turner S (2006) *Dicentroodus* (Chondrichthyes: Xenacanthida) from the Early Carboniferous (Viséan: upper St Louis Formation) of Iowa, USA. *Geol Mag* 143:545-549.
289. Lowney KA (1980) Certain Bear Gulch (Namurian A, Montana) Actinopterygii (Osteichthyes) and a reevaluation of the evolution of the Paleozoic actinopterygians. PhD thesis (New York Univ).
290. Lund R, Poplin C (1999) Fish diversity of the Bear Gulch Limestone, Namurian, Lower Carboniferous of Montana, USA. *Geobios* 32:285-295.
291. Lund R, Poplin C, McCarthy K (1995) Preliminary analysis of the interrelationships of some Palaeozoic actinopterygii. *Geobios* 19:215-220.
292. Lund R, Grogan ED (2005) Fossil fishes of the Bear Gulch Limestones. http://www.sju.edu/research/bear_gulch/
293. Grogan ED, Lund R (2008) A basal elasmobranch, *Thrinacoselache gracia* n. gen and sp., (Thrinacodontidae, new family) from the Bear Gulch limestone, Serpukhovian of Montana, USA. *J Vert Paleont* 28:970-988.
294. Grogan ED, Lund R (2009) Two new inipterygians (Chondrichthyes) from the Mississippian (Serpukhovian) Bear Gulch Limestone of Montana with evidence of a new form of chondrichthyan neurocranium. *Acta Zool* 90:134-151.
295. Mickle KE, Lund R, Grogan ED (2009) Three new palaeoniscoid fishes from the Bear Gulch Limestone (Serpukhovian, Mississippian) of Montana (USA) and the relationships of lower actinopterygians. *Geodiversitas* 31:623-668.