## **Supporting Information**

## Yang et al. 10.1073/pnas.0914506107



**Fig. S1.** Supplemental method for real-time quantitative RT-PCR (qRT-PCR) analysis. Microarray data were validated using real-time qRT-PCR as described previously (1) except that the BioRad MyiQ2 Two-Color Real-Time PCR Detection System (BioRad Laboratories) and Roche FastStart SYBR Green Master reagent (Roche Applied Science) were used for this experiment. Twelve genes representing different functional categories and a range of gene expression values based on microarray results over different time point were analyzed using qRT-PCR. Primer pairs were designed as described previously (1) and the oligonucleotide sequences are listed in Table S4.

1 Yang S, et al. (2009) Transcriptomic and metabolomic profiling of Zymomonas mobilis during aerobic and anaerobic fermentation. BMC genomics 10:34.



**Fig. S2.** Volcano plot summary of microarray analysis showing all significantly differentially expressed genes between AcR and the ZM4 wild-type strains for all conditions (sodium acetate and sodium chloride in exponential and stationary phase cells). The X axis shows the difference values (log 2) between AcR and ZM4 expression profiles. The Y axis shows statistical significance values (– log 10 (*p*-values)) for expression differences. The red dashed line shows the statistical significance cut-off used in this study.



**Fig. S3.** ZM4 *nhaA* had no effect on inhibitor furfural, HMF, and vanillin tolerance. *Z. mobilis* strains were grown in RM (pH5.0) overnight, and then 5  $\mu$ L of culture seed was transferred into 250- $\mu$ L RM media in the Bioscreen plate (Growth Curves USA). The growth differences of different strains were monitored by using a Bioscreen C machine (Growth Curves USA) under anaerobic conditions in RM with 0.5 g/L furfural, pH 5.0 (A); RM with 0.5 g/L HMF, pH 5.0 (B); RM with 0.5 g/L vanillin, pH 5.0 (C); RM with 4 g/L furfural, pH 5.0 (D); RM with 4 g/L HMF, pH 5.0 (E); RM with 2 g/L vanillin, pH 5.0 (F). Strains included in this study were: ZM4: *Zymomonas mobilis* ZM4 wild-type; AcR: previously described ZM4 acetate tolerant mutant; ZM4 (p42-0119): ZM4 containing a gateway plasmid p42-0119 for *nhaA* (*ZMO0119*) expression; ZM4IM0117: ZM4 *hcp* (*ZMO0117*) insertional mutant; ZM4DM0117: deletion mutant affecting *ZMO0117* and part of upstream *nhaA* (Fig. 2 and Table S4). This experiment has been repeated at least two times with similar results. Duplicates were used for each condition.



**Fig. S4.** Overexpression of ZM4 *nhaA* in *E. coli* had a negative effect on sodium acetate tolerance. Two *E. coli* strains (WM3064 and BL21-DE3, the latter is used for protein overexpression) were grown in LB overnight, and then 5 µL of culture seed was transferred into 250-µL LB broth in the Bioscreen plate (Growth Curves USA). The growth differences of different strains were monitored by Bioscreen C (Growth Curves USA) in LB broth, pH 7.0 (*A*); LB with 3 g/L NaAc, pH 7.0 (*B*); LB with 6 g/L NaAc, pH 7.0 (*C*), LB with 12 g/L NaAc, pH 7.0 (*D*). Strains included in this study were: *E. coli* BL21-DE3 (p42-0119): *E. coli* BL21-DE3 containing a gateway plasmid p42-0119 for *nhaA* (ZMO0119) expression; *E. coli* WM3064 (p42-0119): *E. coli* BL21-DE3 containing a gateway plasmid p42-0119 for *nhaA* (ZMO0119) expression. Duplicates were used for each condition.





## **Other Supporting Information Files**

Table S1 (DOC)Table S2 (DOC)Table S3 (DOC)Table S4 (DOC)Table S5 (DOC)