

## Supplementary Methods: Modelling synaptic depression

The justification of the model was described previously (Wang and Manis 2008; Yang and Xu-Friedman 2008), and was based primarily on the model of Dittman *et al.* (2000), which is itself similar to models going back to Liley and North (1953). Briefly, we calculated the relative size of the  $i^{\text{th}}$  EPSC in a train of stimuli by:

$$EPSC_i = F D_i S_i \quad (1)$$

where  $F$  is the probability of release,  $D_i$  is the proportion of release sites that are ready to release on the  $i^{\text{th}}$  pulse,  $S_i$  is the proportion of receptors that are available (i.e. not desensitized). We did not model changes in  $F$ , as facilitation does not appear to be present under our conditions.

The number of release-ready sites,  $D$ , is reduced by release, such that immediately after a release event,  $D$  decreases by  $FD_i$ , and then recovers to resting levels at a rate depending on a calcium-dependent process:

$$\frac{dD(t)}{dt} = (1 - D(t)) \left( \frac{k_{\max} - k_0}{1 + K_D / CaD(t)} + k_0 \right), \quad (2)$$

where  $k_0$  is the resting recovery rate, and  $CaD(t)$  is the calcium-bound state of a sensor that drives a rapid recovery process of rate  $k_{\max}$ . The interaction between the rapid recovery process and the calcium-bound sensor is modelled as a simple affinity given by  $K_D$ .  $CaD$  increments after each EPSC, and decays to resting levels by a simple exponential, given by:

$$CaD_{i+1} = CaD_i e^{-\Delta t / \tau_D} \quad (3)$$

These equations yield an analytical expression for  $D_{i+1}$  based on the preceding pulse:

$$D_{i+1} = 1 - (1 - (1 - F)D_i) e^{-k_0 \Delta t} \left( \frac{K_D / CaD_i + 1}{K_D / CaD_i + e^{-\Delta t / \tau_D}} \right)^{-(k_{\max} - k_0) \tau_D} \quad (4)$$

where  $\Delta t$  is the interval between pulses.

Extracellular glutamate in the synaptic cleft drives desensitization according to a simple binding reaction:

$$S = K_S / (K_S + [\text{glutamate}]), \quad (5)$$

where  $K_S$  is the binding affinity of the receptor for extracellular glutamate. After the  $i^{\text{th}}$  release event, the glutamate concentration increments by the amount just released ( $FD_i$ ), and then decays exponentially back to resting levels, so that the glutamate concentration at the  $(i + 1)^{\text{th}}$  pulse is given by:

$$[\text{glutamate}]_{i+1} = ([\text{glutamate}]_i + FD_i)e^{-\Delta t / \tau_S}, \quad (6)$$

where  $\Delta t$  is the interval between pulses, and  $\tau_S$  is the rate of glutamate clearance.

The 7 free parameters of the model were fit to recorded PPR and trains data for a given cell using a least-squares approach. Parameters for the high-, middle-, and low-depressing endbulbs are given in Supplementary Table 3.

Supplementary Table 1: Correlations between parameters measured off PPR recovery or train depression curves.

	$\tau_f$	$\tau_s$	$A_f$	$A_s$	$SS_{100}$	$SS_{200}$	$SS_{333}$	$\tau_{100}$	$\tau_{200}$	$\tau_{333}$
$\tau_f$	1									
$\tau_s$	-0.016	1								
$A_f$	-0.52	-0.045	1							
$A_s$	0.40	-0.48	-0.51	1						
$SS_{100}$	-0.54	-0.14	0.16	-0.36	1					
$SS_{200}$	-0.36	0.054	-0.076	-0.17	0.70	1				
$SS_{333}$	0.12	0.18	-0.52	0.32	0.12	0.50	1			
$\tau_{100}$	-0.65	0.36	0.17	-0.47	0.61	0.67	0.22	1		
$\tau_{200}$	-0.64	0.28	0.15	-0.46	0.70	0.72	0.20	0.95	1	
$\tau_{333}$	-0.53	0.31	0.035	-0.47	0.71	0.69	0.15	0.91	0.95	1

Supplementary Table 2: Principal components analysis. Rows represent the eigenvectors for each principle component  $X_i$ .

$X_i$	$\tau_f$	$\tau_s$	$A_f$	$A_s$	SS <sub>100</sub>	SS <sub>200</sub>	SS <sub>333</sub>	$\tau_{100}$	$\tau_{200}$	$\tau_{333}$	Eigenvalue	Cum. var.
1	-0.32	0.13	0.11	-0.26	0.36	0.34	0.08	0.43	0.44	0.42	4.9	0.49
2	0.26	0.01	-0.58	0.38	0.04	0.31	0.59	0.05	0.06	0.08	2.1	0.7
3	-0.23	-0.8	0.13	0.35	0.36	0.17	-0.09	-0.09	-0.01	-0.06	1.4	0.84
4	-0.53	0.14	0.39	0.31	-0.37	0	0.47	0.14	0	-0.27	0.57	0.9
5	-0.27	-0.03	-0.38	0.42	-0.28	-0.41	-0.38	0.27	0.23	0.3	0.47	0.94
6	0.53	-0.14	0.42	0.21	-0.46	0.35	-0.17	0.23	0.21	0.16	0.25	0.97
7	0.34	-0.14	0.29	0.04	0.28	-0.68	0.4	0.18	0.16	0.15	0.16	0.99
8	-0.07	-0.54	-0.25	-0.59	-0.45	-0.07	0.23	0.19	0.07	-0.05	0.13	1
9	0.09	0.02	-0.08	0.03	0.18	0.03	-0.13	0.77	-0.47	-0.34	0.046	1
10	0.1	0.05	-0.12	0	0.11	-0.02	-0.13	0.03	0.68	-0.7	0.027	1

Supplementary Table 3. Parameters used for the models in Fig. 4. The model is described in detail in Yang & Xu-Friedman (2008), and the Supplementary Methods.

Parameter	Meaning	low	middle	high
$F$	probability of release	0.35	0.65	0.9
$k_0$	baseline recovery rate from depletion	1/s	0.5	0.6
$k_{\max}$	maximal recovery rate from depletion	20/s	27	15
$\tau_D$	decay time constant for calcium-dependent recovery	10 ms	10	30
$K_D$	affinity of fast recovery process for calcium sensor	0.1	0.01	0.2
$\tau_S$	decay time constant of glutamate clearance	5 ms	5	5
$K_S$	affinity of receptor desensitization for glutamate	1	1	1

Supplementary Table 4. Steady-state conductance amplitudes predicted by the three depression models at three stimulation frequencies. In addition, the rightmost column indicates the postsynaptic threshold measured in current-clamp experiments after a train of 30 pulses normalized to threshold at the beginning ( $N = 4$  experiments).

Frequency (Hz)	Depression model			Threshold
	high	middle	low	
333	0.045	0.105	0.138	$1.47 \pm 0.12$
200	0.074	0.170	0.213	$1.93 \pm 0.23$
100	0.14	0.310	0.355	$2.38 \pm 0.34$

Supplementary Table 5. Comparison between sibling and non-sibling inputs onto the same target bushy cell using the Kolmogorov-Smirnov (K-S) test for individual measures. Data are from 10 pairs of siblings compared against the overall distribution of distances (i.e. 190 comparisons). *P* values below 0.05 are highlighted in bold. The main text and Fig. 6 report the plots and *P*-values for measures grouped together.

Parameter	<i>P</i> -value
EPSC <sub>1</sub>	0.0657752
PPR ( $\Delta t$ in ms)	
3	0.0820644
5	0.212364
10	0.140841
20	0.249175
50	0.179377
100	0.334117
200	0.304269
500	0.334117
1000	0.101354
4000	0.349546
100 Hz trains (pulse #)	
<b>2</b>	<b>0.0242287</b>
<b>3</b>	<b>0.0521546</b>
4	0.200975
<b>5</b>	<b>0.0242287</b>
<b>6</b>	<b>0.0112116</b>
<b>7</b>	<b>0.00592289</b>
<b>8</b>	<b>0.0221065</b>
<b>9</b>	<b>0.00529738</b>
<b>10</b>	<b>0.020144</b>
<b>11</b>	<b>0.00820421</b>
<b>12</b>	<b>0.0037559</b>
<b>13</b>	<b>0.0183318</b>
14	0.0881462
<b>15</b>	<b>0.0289917</b>
<b>16</b>	<b>0.00205284</b>
<b>17</b>	<b>0.0112116</b>
<b>18</b>	<b>0.0183318</b>
<b>19</b>	<b>0.0265205</b>
<b>20</b>	<b>0.00529738</b>
200 Hz trains (pulse #)	
<b>2</b>	<b>0.0221065</b>
<b>3</b>	<b>0.00592289</b>
<b>4</b>	<b>0.0443896</b>

<b>5</b>	<b>0.0289917</b>
<b>6</b>	<b>0.0316522</b>
<b>7</b>	<b>0.0443896</b>
8	0.140841
9	0.0564202
10	0.275905
11	0.159329
12	0.108498
13	0.249175
14	0.304269
15	0.464839
<b>16</b>	<b>0.0242287</b>
<b>17</b>	<b>0.0481476</b>
18	0.0564202
19	0.628767
20	0.108498
333 Hz trains (pulse #)	
<b>2</b>	<b>0.020144</b>
<b>3</b>	<b>0.0183318</b>
4	0.212364
5	0.319018
6	0.189984
7	0.289892
8	0.289892
9	0.159329
10	0.108498
11	0.289892
12	0.51742
13	0.554358
14	0.304269
15	0.482079
16	0.249175
17	0.249175
18	0.200975
19	0.349546
20	0.36528