

A

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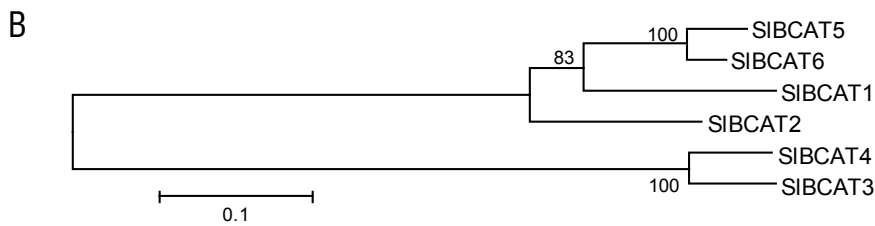


Figure S1. Evolutionary relationships of mature SIBCAT proteins. Mature peptides were predicted by omitting highly divergent N-terminal residues thought to be transit peptides.

A, Alignment of amino acid sequences using ClustalW (Larkin et al., 2007). B, The evolutionary history was inferred using the Neighbor-Joining method (Saitou and Nei, 1987). The optimal tree with the sum of branch length = 1.21366717 is shown. The percentage of replicate trees in which the proteins clustered together in the bootstrap test (500 replicates) are shown next to the branches (Felsenstein, 1985). The tree is drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. There were a total of 332 amino acids in the final dataset. Phylogenetic analyses were conducted in MEGA4 (Tamura et al., 2007).

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SIBCAT5 1  LDWDLGRFGLGIDYMYRGRSGEGRGRCGLLRVGNLSPSAGVLYNGQGLFEGKRA
SIBCAT6 1  LDWDLGRFGLGIDYMYRGRSGEGRGRCGLLRVGNLSPSAGVLYNGQGLFEGKRA
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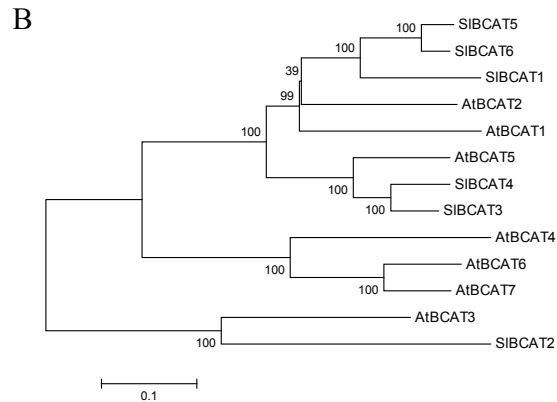


Figure S2. Evolutionary relationships of mature SIBCAT and AtBCAT proteins. Mature peptides were predicted by omitting highly divergent N-terminal residues thought to be transit peptides. A, Alignment of amino acid sequences using ClustalW (Larkin et al., 2007). B, The evolutionary history was inferred using the Neighbor-Joining method (Saitou and Nei, 1987). The optimal tree with the sum of branch length = 2.55921922 is shown. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (500 replicates) are shown next to the branches (Felsenstein, 1985). The tree is drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. There were a total of 331 positions in the final dataset. Phylogenetic analyses were conducted in MEGA4 (Tamura et al., 2007).

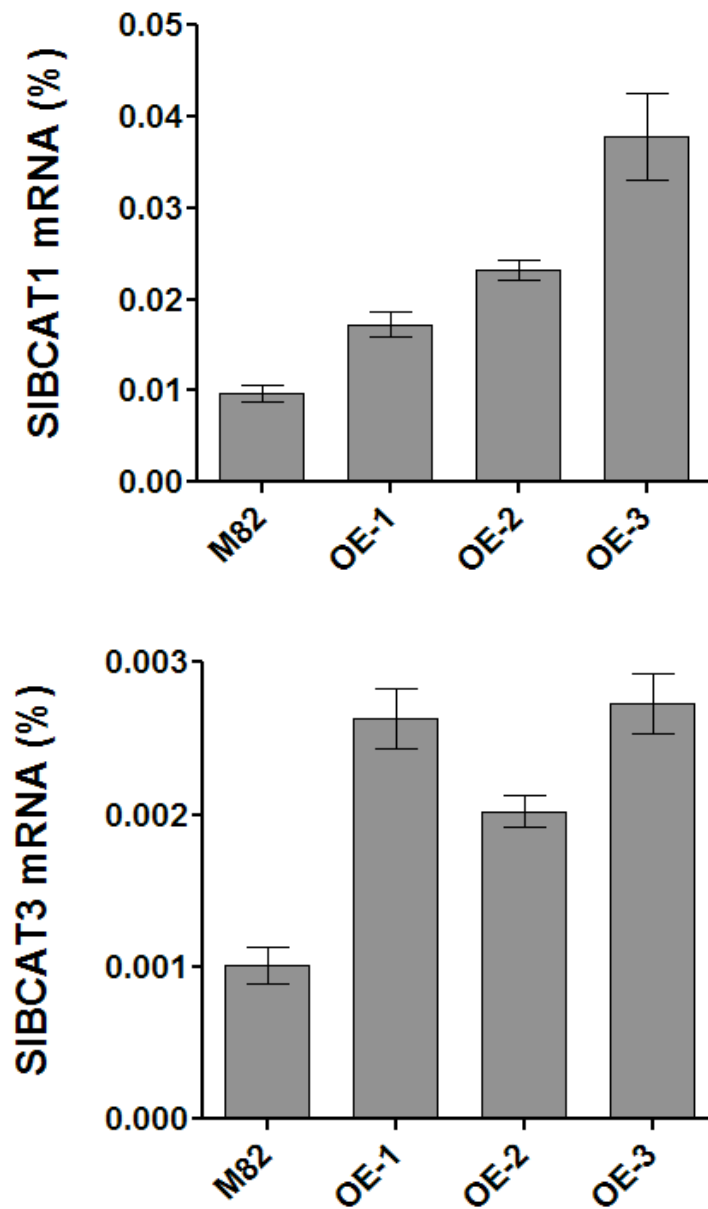


Figure S3. Transcript analysis in fruits of transgenic plants overexpressing *SIBCATs*. Analysis was performed on three biological and three technical replicates of ripe fruit for each sample. Values represent percentage of total mRNA per sample \pm SD, calculated from a standard curve for each cDNA. Data were analyzed by one-way ANOVA and values of each line are significantly different compared to control as determined by Tukey's test ($p < 0.05$).

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M82 1 ATGATCATCAAAGGGCTTCATCCTTTCAAAGGGCTCTATTTACTTCTTTTGAGAAGAAG
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M82 1141 TGA
Lp 1141 TGA

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Figure S4. Comparison of nucleotide sequences of BCAT1 from *S. lycopersicum* cv. M82 and *S. pennellii* (Lp). Alignment of the sequences was carried out using the ClustalW and output processed using the Boxshade program. White boxes indicate nucleotide changes between variants. The black boxes indicate identical base pairs, gray indicates conservative substitutions.

Table S1. Levels of free amino acids in red ripe fruit of M82 and *SIBCAT* over-expression lines.

Line	Amino acid quantity (ng gFW ⁻¹)						
	Val	Leu	Ile	Ala	Pro	Met	Lys
M82	0.79 ± 0.06	1.12 ± 0.14	2.19 ± 0.26	6.69 ± 0.48	1.91 ± 0.28	0.65 ± 0.10	3.01 ± 0.22
SIBCAT1-OE-1	0.61 ± 0.06	0.95 ± 0.09	2.22 ± 0.18	3.25 ± 0.29	3.73 ± 0.35	0.60 ± 0.05	2.58 ± 0.19
SIBCAT1-OE-2	0.70 ± 0.10	1.06 ± 0.09	2.21 ± 0.18	6.92 ± 1.27	3.45 ± 0.54	0.47 ± 0.05	2.04 ± 0.21
SIBCAT1-OE-3	0.72 ± 0.17	1.13 ± 0.15	2.64 ± 0.40	6.20 ± 1.65	3.32 ± 0.41	0.62 ± 0.04	2.62 ± 0.45
SIBCAT3-OE-1	0.73 ± 0.12	1.19 ± 0.20	3.38 ± 0.60	3.44 ± 0.50	2.88 ± 0.06	0.74 ± 0.16	1.87 ± 0.26
SIBCAT3-OE-2	0.64 ± 0.07	1.08 ± 0.12	3.52 ± 0.42	5.02 ± 0.68	2.61 ± 0.29	0.62 ± 0.06	2.37 ± 0.26
SIBCAT3-OE-3	1.47 ± 0.32	2.61 ± 0.45	7.96 ± 1.81	7.92 ± 1.00	5.57 ± 1.05	1.00 ± 0.03	4.83 ± 0.72

Values are expressed as means ± SD.

Table S2. Primer sequences used in present study.

Primer name	Primer sequence (5'→ 3')
<i>ilvE</i> -FlankF	GATGCAACATCAGGTCAATGT
<i>ilvE</i> -FlankR	CGCAATGGTGTGAACTCTT
<i>tyrB</i> -FlankF	CTTATTACGCGCCTGACTTC
<i>tyrB</i> -FlankR	CACAGGCAATAAGGCAAAGC
<i>SIBCAT3</i> -pET28bF	GCTAGCGAGAGCGCCGCCGTATTT
<i>SIBCAT3</i> -pET28bR	GTCGACTTTGAGCTCAACAATCCAACCC
<i>SIBCAT1</i> -pET28bF	GCTAGCTCTGCACAACCTTCAACTTATAG
<i>SIBCAT1</i> -pET28bR	GTCGACCTTAATCTCAATAATCCAACCCCT
<i>SIBCAT4</i> -pET28bF	GCTAGCTTTCAGAAGCAGTCACATTTTGC
<i>SIBCAT4</i> -pET28bR	GTCGACTCATTTTGTAGCTCAACAGTCCAATT
<i>SIBCAT2</i> -pET28bF	GCTAGCTACTACACAGCTCAGGTTG
<i>SIBCAT2</i> -pET28bR	GTCGACTCATTCAATGTCAACAATCCAATC
<i>SIBCAT5</i> -pET28bF	GCTAGCGCTTCTTCTCAATCTGTTCTCT
<i>SIBCAT5</i> -pET28bR	GTCGACTTCGATCTCCACGATCCAATT
<i>SIBCAT6</i> -pET28bF	GCTAGCTGTTATACAGCTCAGGCGG
<i>SIBCAT6</i> -pET28bR	GTCGACTCATTCAATCTCCACGATCCAAT
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<i>SIBCAT3</i> -pDONRR	AGAAAGCTGGGTCTTTGAGCTCAACAATCCAACC
<i>SIBCAT1</i> -pDONRF	AAAAAGCAGGCTCCATGATCATCCAAAGGGCTTCA
<i>SIBCAT1</i> -pDONRR	AGAAAGCTGGGTCTTAAATCTCAATAATCCAACCC
<i>SIBCAT4</i> -pDONRF	AAAAAGCAGGCTCCATGGAGAGCGGCGGCG
<i>SIBCAT4</i> -pDONRR	AGAAAGCTGGGTCTTTTGTAGCTCAACAGTCCAATTCA
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Attb2-pDONR	GGGGACCACTTTGTACAAGAAAGCTGGGT
<i>SIBCAT1</i> -pENTR/SD/D-TOPOF	ATGATCATCCAAAGGGCTTCA
<i>SIBCAT1</i> -pENTR/SD/D-TOPOR	AATCTCAATAATCCAACCCCTCTT
<i>SIBCAT3</i> -qRT-PCRF	GTCACCATAACCACCTTCTGG
<i>SIBCAT3</i> -qRT-PCRR	GGACTCAACTCAATGTTACCG
<i>SIBCAT1</i> -qRT-PCRF	AGGGCTCTATTTACTTCTTTTGTAG
<i>SIBCAT1</i> -qRT-PCRR	CATACACATTCTTTTGTAGACCAATT
<i>SIBCAT4</i> -qRT-PCRF	TACTTGCCACCACCTTCCC
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<i>SIBCAT2</i> -qRT-PCRF	AATTGTTTGAATTTTCATCTCTGCG
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<i>SIBCAT5</i> -qRT-PCRF	CTCACCTTCTCTACACCAC
<i>SIBCAT5</i> -qRT-PCRR	AATAGCCGTTTGTCTAACAGCC
<i>SIBCAT6</i> -qRT-PCRF	GGTGTATACAGCTCAGGCG
<i>SIBCAT6</i> -qRT-PCRR	CAGGACGAAATAAAAAATACTCTCC
<i>UBI3F</i>	GGTTAAGCTCGCTGTGTTGCA
<i>UBI3R</i>	CGAAGCCTCTGAACCTTTCCA

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