Supporting Information

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Fig. S1. Purification and characterization of MOV10L1-associated proteins from 18- to 20-d-old testes by immunoprecipitation (IP) and mass spectrometry. Protein identity for each band is indicated. In brief, to isolate MOV10L1-associated proteins, eight pairs of 18- to 20-d testes (\approx 300 mg) were homogenized in 3 mL radioimmunoprecipitation assay buffer (RIPA) in the presence of proteinase inhibitor mixture. Protein lysate was centrifuged twice. Supernatants were precleared with protein A agarose beads. Precleared lysate incubated with affinity-purified anti-MOV10L1 antibody, followed by binding to protein A agarose beads. Immunoprecipitated proteins were extensively washed, run on a 4–15% gradient SDS/PAGE gel, and stained with SYPRO Ruby (Bio-Rad).



Fig. 52. Profiles of small RNA reads from MOV10L1, MILI, and TDRD1 libraries. (*A*) Annotation profile of MOV10L1 reads resembles those in MILI and TDRD1 libraries. (*B*) Pie charts showing representation of repeat piRNAs belonging to different transposon classes in indicated libraries. (*C*) Common genomic origins of MOV10L1-, MILI-, and TDRD1-associated piRNAs. All three libraries have similar read-depths (≈ 2 million). The Piwi-interacting RNA (piRNA) density for the top two pachytene clusters was plotted. The chromosome 17 (Chr17) cluster is ≈ 79 kb in length and bidirectional, with piRNAs arising from both strands in a nonoverlapping manner, whereas the Chr2 cluster is ≈ 72 kb and unidirectional, with most piRNAs deriving from the top strand.



Fig. S3. Targeted inactivation of the *Mov10l1* gene. *Top:* Illustration of the predicted RNA helicase domain of mouse MOV10L1 protein. The mouse *Mov10l1* gene consists of 27 exons and spans a >72-kb genomic region. This deletion removes the ATPase and unwindase domains, which are highly conserved among RNA helicases.



Fig. S4. Immunofluorescence analyses of MOV10L1 in testes from adult $Mov10l1^{+/-}$ (A) and $Mov10l1^{-/-}$ (B) mice. Leydig cells give strong autofuorescence. In $Mov10l1^{-/-}$ spermatocytes (B) the internally deleted mutant MOV10L1 protein was barely detectable by immunofluorescence analysis, owing to its sharply reduced abundance. (Scale bar, 25 μ m.)



Fig. S5. MILI and TDRD1 persist in spermatogonia but not in spermatocytes in $Mov10l1^{-/-}$ testes. Frozen testis sections from 2-mo-old (P60) $Mov10l1^{+/-}$ (A, C, and E) and $Mov10l1^{-/-}$ (B, D, and F) were immunostained with anti-MILI, anti-TDRD1, and anti-MVH antibodies. Chromatin was stained with DAPI. $Mov10l1^{-/-}$ seminiferous tubules are demarcated with dash lines. *Insets* (B and D): Enlarged view of indicated spermatogonia. Spg, spermatogonia; Spc, spermatocytes. (Scale bar, 25 μ m.)



Fig. S6. Derepression of LINE1 and IAP retrotransposons in *Mov10l1^{-/-}* perinatal gonocytes. Newborn (P0) testis sections were immunostained with anti-L1 ORF1p and anti-IAP antibodies. Gonocytes (Gc) are indicated by arrows. (Scale bar, 25 μm.)