

## SUPPLEMENTARY MATERIAL

**Fig.S1 Specificity of the binding between Bcl-2 and SOD1.** **(A)** Spinal cords from SOD1-G93A mice (onset and end stage) were homogenized and immunoprecipitated with either the anti-loop Bcl-2, the anti-SOD1 antibody or equal amounts of their corresponding IgGs. Co-precipitates were analyzed by WB with anti-SOD1 and anti-Bcl-2 antibodies respectively. The mutSOD1/Bcl-2 complex is immunoprecipitated only by the precipitating antibodies and not by the corresponding IgGs, clearly indicating the specificity of SOD1/Bcl-2 binding. TL= total lysates **(B)** HEK293T cells which do not express detectable levels of Bcl-2 were transfected with either Bcl-2 or empty plasmid (mock) and immunoprecipitation with anti-SOD1 or IgG was performed, followed by WB for Bcl-2 or SOD1 in order to determine the specificity of the binding between SOD1 and Bcl-2. As shown, the anti-SOD1 antibody co-precipitates Bcl-2 only in transfected, but not in naïve, mock transfected cells. Control IgGs failed to precipitate Bcl-2 even in cells transfected with Bcl-2. In mock transfected cells, control IgGs failed to aspecifically precipitate other proteins in the 25-30 KDa range. TL=total lysates. **(C)** HEK293T cells are a suitable system to study mutSOD1-toxicity in the absence of Bcl-2. Cells were transfected with Bcl-2, fixed and stained with anti Bcl-2 (blue) and the mitochondrial marker mitotracker (300 nM, red). There is a negative staining with the anti-Bcl-2 antibody in non-transfected cells confirming that HEK293T cells do not express detectable levels of endogenous Bcl-2 (scale bar 20  $\mu\text{m}$ ). WB shows expression levels of Bcl-2 prior and after transfection.

**Figure S2. Schematic representation of Bcl-2 and the antibodies used in IP/WB experiments.** **(A)** Linear representation of Bcl-2 with its BH (1-4) domains and the epitopes of the antibodies used in our experiments. **(B)** FACS analysis of

conformational changes in Bcl-2 induced by mutSOD1. Exposure of the BH3 domain in SH-SY5Y was also assessed by FACS analysis using the conformation-specific  $\alpha$ Bcl-2/BH3 antibody. The shift in fluorescence in presence of mutSOD1s indicates an increased exposure of the BH3 domain.

**Figure S3. Lack of SOD1-G93A toxicity in H4 cells expressing nuclear but not**

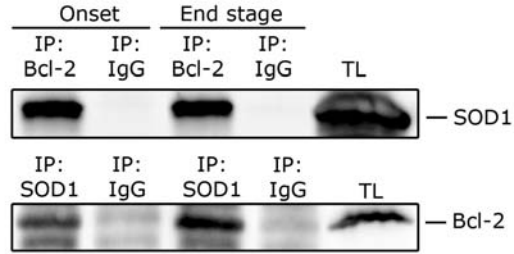
**mitochondrial Bcl-2. (A)** H4 cells (ATCC cat. # HTB-148) were stained with DAPI (as nuclear marker) and for Bcl-2 and Cytochrome C (as mitochondrial marker). Bcl-2 co-localizes with DAPI (nuclear localization) but not with Cytochrome C (mitochondria localization). The bottom panel represents expression of Bcl-2 in the total lysate (TL), nuclei (Nuc), mitochondria (Mito) and cytoplasm (Cyto) (scale bar 20  $\mu$ m). **(B)** In the absence of mitochondrial Bcl-2, SOD1-G93A does not induce Cytochrome C release. H4 cells were co-transfected with SOD1-G93A-EGFP, immunostained for Bcl-2 (blue) and Cytochrome C (red) to assess mitochondrial integrity. In the absence of mitochondrial Bcl-2, mutSOD1-expressing (green) H4 cells show punctate Cytochrome C staining indicating mitochondrial integrity (scale bar 20  $\mu$ m).

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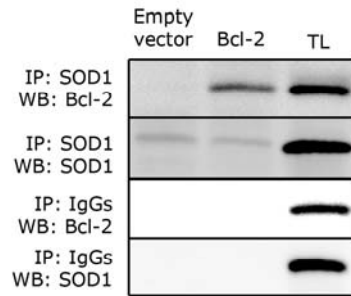
*FACS analysis:* SH-SY5Y cells were grown in DMEM media, 5% FBS with antibiotics. 2 days before transfection, cells were plated at a concentration of 100,000/well in 12-well plates and transfected with 500 ng of SOD1-EGFP constructs using Lipofectamine 2000 following manufacturer's instructions. After 24 hrs, cells were detached, washed twice in PBS, incubated for 1 hr at RT with PBS 5% goat serum and then fixed and permeabilized in Cytofix/Cytoperm (BD, Franklin Lakes, NJ, USA). Following permeabilization, cells were incubated with rabbit  $\alpha$ -Bcl2-BH3 domain

(Abgent, San Diego, CA, USA) 1: 250 in Cytoperm/Cytowash 5% goat serum for 1 hr and subsequently in anti-rabbit Alexa Fluor 546 (Carlsbad, CA, USA) 1:400 in Cytoperm/Cytowash 5% goat serum for 1 hr. Cells were then analyzed using FACScalibur (BD, USA).

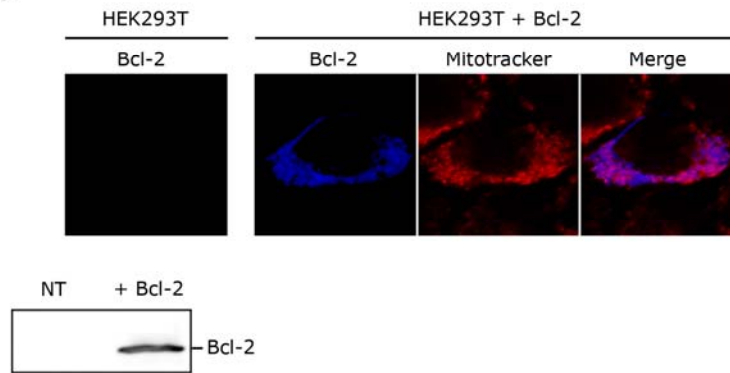
**A**



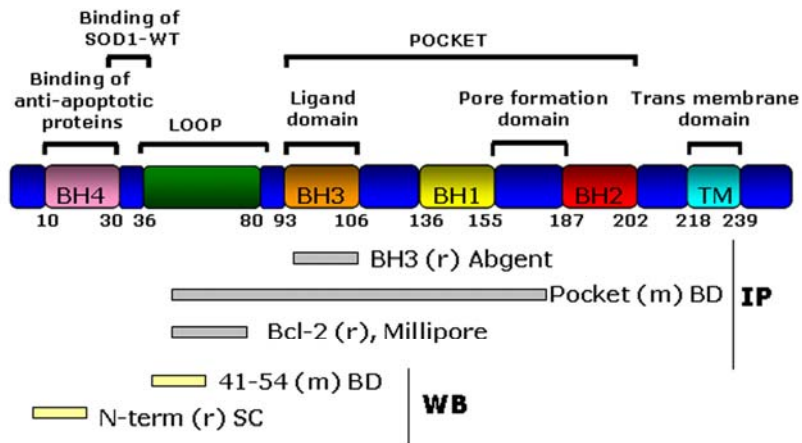
**B**



**C**



**A**



**B**

