

### *Basic theory of MANOVA, LDA and LOO methods*

Because of the statistical methods are classical methods that have been widely used in data analyses, their description here is brief. For MANOVA, it is an extension of the univariate analysis of variance. This method is used in the cases where there are two or more dependent variables.

LDA is a classical pattern recognition method and can provide a classification model based on the combination of variables to predict a category or group to which a material belongs (Fisher 1936). The basic theory of LDA is to classify the dependent by dividing a  $n$ -dimensional parameters space into two regions. That space is separated by a hyperplane defined by a linear discriminant function as follows:

$$Y = a_0 + a_1X_1 + a_2X_2 + \dots + a_nX_n$$

Where  $Y$  is a discriminant score, that is the dependant variable;  $X_1, \dots, X_n$  represents the specific descriptor (in this case, metabolite levels); and  $a_0 \dots a_n$  are the weights of the corresponding parameters. The two regions formed by the hyperplane correspond to the two classes. Once the model was constructed, it is important to validate the predictability of that model. The simplest and a commonly used method of crossvalidation is the LOO method. In this method, the learning algorithm is trained multiple times, using all but one of the training set data points. At last, the mean error over all data points was calculated.

Table 1S: Mass and isotope balance equations for the metabolic calculation:

Mass balance equations:

$$dGlc_{brain}/dt = V_{max\ in} Glc_{blood}/(K_M + Glc_{blood}) - V_{max\ out} Glc_{brain}/(K_{Mout} + Glc_{brain}) - CMR_{glc}$$

$$d(Glu_N)/dt = d(Glu_A)/dt = d(KG_N)/dt = d(KG_A)/dt = d(Asp_N)/dt = d(Asp_A)/dt$$

$$= d(OAA_N)/dt = d(OAA_A)/dt = 0$$

Isotopic balance equations:

$$d(Glc_{brain,C_1^*})/dt = V_{max\ in} (Glc_{blood,C_1^*})/(K_{m\ in} + Glc_{blood}) - V_{max\ out} (Glc_{brain,C_1^*})/(K_{m\ out} + Glc_{brain})$$

$$- CMR_{glc} (Glc_{brain,C_1^*} / Glc_{brain})$$

$$d(L_3^*)/dt = CMR_{Glc} (Glc_{brain,C_1^*} / Glc_{brain}) + V_{dilLac} (0) - (V_{pdh_N} + V_{pdh_A} + V_{pc} + V_{dilLac} + V_{pdh_{GABA}})(L_3^* / L)$$

$$d(Glu_{N_4^*})/dt = V_{cycle} (Gln_4^* / Gln) + V_X (KG_{N_4^*} / KG_N) - (V_{cycle} + V_X)(Glu_{N_4^*} / Glu_N)$$

$$d(Glu_{A_4^*})/dt = V_{cycle} (Glu_{N_4^*} / Glu_N) + (V_X + V_{ana})(KG_{A_4^*} / KG_A) - (V_{Gln} + V_X)(Glu_{A_4^*} / Glu_A)$$

$$d(Gln_4^*)/dt = V_{Gln} (Glu_{A_4^*} / Glu_A) + V_{dilGln} (0) - (V_{cycle} + V_{efflux} + V_{cycle_{GABA_{Gln}}})(Gln_4^* / Gln)$$

$$d(KG_{A_4^*})/dt = V_{pdh_A} (L_3^* / L) + V_X (Glu_{A_4^*} / Glu_A) - (V_{TCA_A\ Net} + V_X)(KG_{A_4^*} / KG_A)$$

$$d(KG_{A_3^*})/dt = V_X (Glu_{A_3^*} / Glu_A) + V_{TCA_A} (OAA_{A_2^*} / OAA_A) - (V_{TCA_A\ Net} + V_X)(KG_{A_3^*} / KG_A)$$

$$d(Glu_{A_3^*})/dt = V_{cycle} (Glu_{N_3^*} / Glu_N) + V_X (KG_{A_3^*} / KG_A) - (V_{Gln} + V_X)(Glu_{A_3^*} / Glu_A)$$

$$d(Glu_{N_3^*})/dt = V_{cycle} (Gln_3^* / Gln) + V_X (KG_{N_3^*} / KG_N) - (V_X + V_{cycle})(Glu_{N_3^*} / Glu_N)$$

$$d(Gln_{A_3^*})/dt = V_{Gln} (Glu_{A_3^*} / Glu_A) + V_{dilGln} (0) - (V_{cycle} + V_{efflux} + V_{cycle_{GABA_{Gln}}})(Gln_3^* / Gln)$$

$$d(KG_{N_4^*})/dt = V_{TCA_N} (L_3^* / L) + V_X (Glu_{N_4^*} / Glu_N) + V_{dilN} (0) - (V_{TCA_N} + V_X)(KG_{N_4^*} / KG_N)$$

$$d(KG_{N_3^*})/dt = V_X (Glu_{N_3^*} / Glu_N) + V_{TCA_N} (OAA_{N_2^*} / OAA_N) - (V_{TCA_N} + V_X)(KG_{N_3^*} / KG_N)$$

$$d(Asp_{N_3^*})/dt = V_{X_{N_{OAAA_{Asp}}}} [(OAA_{N_3^*} / OAA_N) - V_{X_{N_{OAAA_{Asp}}}} (Asp_{N_3^*} / Asp_N)]$$

$$d(Asp_{A_3^*})/dt = V_{X_{A_{OAAA_{Asp}}}} [(OAA_{A_3^*} / OAA_A) - V_{X_{A_{OAAA_{Asp}}}} (Asp_{A_3^*} / Asp_A)]$$

$$d(OAA_{N_2^*})/dt = V_{X_{N_{AspOAA}}} (Asp_{N_2^*} / Asp_N) + 0.5V_{TCA_N} [(KG_{N_4^*} / KG_N) + (KG_{N_3^*} / KG_N)]$$

$$- (V_{X_{N_{AspOAA}}} + V_{TCA_N})(OAA_{N_2^*} / OAA_N)$$

$$d(OAA_{A_2^*})/dt = 0.5V_{TCA_A\ Net} (KG_{A_4^*} / KG_A + KG_{A_3^*} / KG_A) + V_{X_{A_{AspOAA}}} (Asp_{A_2^*} / Asp_A) + 0.5V_{PC} (L_3^* / L)$$

$$+ V_{cycle_{GABA_{Gln}}} (GABA_2 / GABA) - (V_{X_{A_{AspOAA}}} + V_{TCA_A})(OAA_{A_2^*} / OAA_A)$$

$$d(GABA_2^*)/dt = V_{gad} (Glu_{GA_4^*} / Glu_{GA}) - [V_{shunt} + V_{cycle_{GABA_{Gln}}}] (GABA_2^* / GABA)$$

$$d(KG_{GA_4}^* / dt) = V_{pdh_{GABA}} (L_3^* / L) + V_{X_{GGKG}} (Glu_{GA_4}^* / Glu_{GA}) + V_{dil_{GG}} (0)$$

$$- [V_{TCA_{GABA}Net} + V_{X_{KGGG}}](KG_{GA_4}^* / KG_{GA})$$

$$d(Glu_{GA_4}^* / dt) = V_{cycle_{GABAGln}} (Gln_4^* / Gln) + V_{X_{KGGG}} (KG_{GA_4} / KG_{GA}) - [V_{gad} + V_{X_{KGGG}}](Glu_{GA_4} / Glu_{GA})$$

$$d(KG_{GA_3}^* / dt) = V_{X_{KGGG}} (Glu_{GA_3}^* / Glu_{GA}) + V_{TCA_{GABA}} (OAA_{GA_3}^* / OAA_{GA})$$

$$- [V_{TCA_{GABA}Net} + V_{X_{KGGG}}](KG_{GA_3} / KG_{GA})$$

$$d(GABA_3^*) / dt = V_{gad} (Glu_{GA_3}^* / Glu_{GA}) - [V_{shunt} + V_{cycle_{GABAGln}}](GABA_3^* / GABA)$$

$$d(Glu_{GA_3}^*) / dt = V_{cycle_{GABAGln}} (Gln_3^* / Gln) + V_{X_{KGGG}} (KG_{GA_3}^* / KG_{GA}) - [V_{gad} + V_{X_{KGGG}}](Glu_{GA_3}^* / Glu_{GA})$$

$$d(Asp_{GA_3}^*) / dt = V_{X_{GA}OAAAsp} (OAA_{GA_2}^* / OAA_{GA}) - V_{X_{GA}AspOAA} (Asp_{GA_3}^* / Asp_{GA})$$

$$d(OAA_{N_3}^*) / dt = V_{X_{AspOAA}} (Asp_{N_2}^* / Asp_N) + 0.5V_{TCA_N} (KG_{N_3}^* / KG_N) + 0.5V_{TCA_N} (KG_{N_4}^* / KG_N)$$

$$- [V_{X_{OAAAspN}} + V_{TCA_N}](OAA_{N_3}^* / OAA_N)$$

$$d(OAA_{A_3}^*) / dt = V_{cycle_{GABAGln}} (GABA_3 / GABA) + V_{X_AAspOAA} (Asp_{A_2}^* / Asp_A) + 0.5V_{pc} (L_3^* / L) + 0.5V_{TCA_{Net}}$$

$$(KG_{A_4}^* / KG_A) + 0.5V_{TCA_{A}Net} (KG_{A_3}^* / KG_A) - [V_{X_{OAAAsp}} + V_{TCA_A}](OAA_{A_3} / OAA_A)$$

$$d(Asp_{A_2}^*) / dt = V_{X_AOAAAsp} (OAA_{A_2}^* / OAA_A) - V_{X_AAspOAA} (Asp_{A_2}^* / Asp_A)$$

$$d(Asp_{N_2}^*) / dt = V_{X_NOAAAsp} (OAA_{N_2}^* / OAA_N) - V_{X_NAspOAA} (Asp_{N_2}^* / Asp_N)$$

$$d(Asp_{GA_2}^*) / dt = V_{X_{GA}OAAAsp} (OAA_{GA_2}^* / OAA_{GA}) - V_{X_{GA}AspOAA} (Asp_{GA_2}^* / Asp_{GA})$$

$$d(OAA_{GA_2}^*) / dt = 0.5V_{TCA_{GABA}Net} (KG_{GA_3}^* / KG_{GA}) + 0.5V_{TCA_{GABA}Net} (KG_{GA_4}^* / KG_{GA}) + 0.5V_{shunt}$$

$$(GABA_2^* / GABA) + 0.5V_{shunt} (GABA_3^* / GABA) + V_{X_{GA}AspOAA} (Asp_{GA_2}^* / Asp_{GA})$$

$$- [V_{TCA_{GABA}} + V_{X_{GA}OAAAsp}](OAA_{GA_2}^* / OAA_{GA})$$

$$d(OAA_{GA_3}^*) / dt = 0.5V_{TCA_{GABA}Net} (KG_{GA_4}^* / KG_{GA}) + 0.5V_{TCA_{GABA}Net} (KG_{GA_3}^* / KG_{GA}) + 0.5V_{shunt}$$

$$(GABA_2^* / GABA) + 0.5V_{shunt} (GABA_3^* / GABA) + V_{X_{GA}AspOAA} (Asp_{GA_3}^* / Asp_{GA})$$

$$- [V_{TCA_{GABA}} + V_{X_{GA}AspOAA}](OAA_{GA_3}^* / OAA_{GA})$$

*Combination Pools:*

$$Glu_{4\_total} = Glu_{A4} + Glu_{N4} + Glu_{GA4}$$

$$Glu_{3\_total} = Glu_{A3} + Glu_{N3} + Glu_{GA3}$$

The following are one group of rates and concentrations from the parietal cortex in a saline-treated group using the upper equations and combinational pools.

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

Rates:

$CMR_{gl} = (V_{pdhA} + V_{pdhN} + V_{pdhGABA} + V_{pc})/2 = 0.54 \mu\text{mol}/\text{min}/\text{g}$ ; rate of glucose consumption

$K_{m\_in} = 13.9 \text{ mM}$ ; Michaelis-Menten half-saturation constant for blood-brain glucose Transport (Mason et al. 1992b)

$K_{m\_out} = K_{m\_in} * V_d = 10.70 \mu\text{mol}/\text{g}$ ; Michaelis-Menten half-saturation constant for brain blood glucose transport

$R_{\_AspGlu} = 0.29 \mu\text{mol}/\text{min}/\text{g}$ ; Fraction of Asp that is in glutamatergic neurons

$V_{cycle} = 0.49 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of Glu-Gln cycling

$V_{cycleGABAGln} = 0.46 * V_{tcaGABA} = 0.053 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of GABA-Gln cycling, fraction of 0.46 from (Patel et al. 2005)

$V_d = 0.77 \text{ ml}/\text{g}$ ; Brain water space (Buschiazzo et al. 1970)

$V_{dilGG} = 0.046 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of dilution in GABAergic neurons

$V_{dilGln} = 0.35 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of diluting Gln exchange with blood

$V_{dilN} = 0 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of dilution in glutamatergic neurons

$V_{efflux} = V_{pc} + V_{dilGln} = 0.42 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of loss of carbon from the astrocytic TCA cycle via efflux of Gln from the brain, partly balanced by entry of glutamine from the blood, also at the rate  $V_{dilGln}$ .

$V_{gad} = V_{shunt} + V_{cycleGABAGln} = 0.14 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of GABA synthesis

$V_{gln} = V_{cycle} + V_{cycleGABAGln} + V_{pc} = 0.60 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of Gln synthesis

$V_{max\_in} = 5.8 * CMR_{gl} = 3.14 \mu\text{mol}/\text{min}/\text{g}$ ;  $V_{max}$  for glucose flow from blood to brain (Mason et al. 1992b).

$V_{max\_out} = V_{max\_in} = 3.14 \mu\text{mol}/\text{min}/\text{g}$ ;  $V_{max}$  for glucose flow from brain to blood

$V_{pc} = V_{tcaA} - V_{tcaANet} - V_{cycleGABAGln} = 0.065 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of anaplerosis

$V_{pdhA} = 0.176 * V_{tcaN} = 0.14 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of astrocytic pyruvate dehydrogenase (Patel et al. 2005)

$V_{pdhGABA} = 0.070 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of pyruvate dehydrogenase in the GABAergic neuron

$V_{shunt} = V_{tcaGABA} - V_{tcaGABANet} = 0.088 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of GABA degradation in the GABAergic neuron

$V_{tcaGABA} = V_{pdhGABA} + V_{dilGG} = 0.12 \mu\text{mol}/\text{min}/\text{g}$ ; Rate of GABAergic TCA cycle from

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

*citric acid through KG*

$V_{\text{tcaGABANet}} = 0.029 \mu\text{mol/min/g}$ ; Rate of GABAergic TCA cycle flow from KG to succinate

$V_{\text{tcaN}} = 0.81 \mu\text{mol/min/g}$ ; Rate of glutamatergic neuronal TCA cycle rate

$V_{\text{xAspOAA}} = V_{\text{xOAAAsp}} = 0.5 \mu\text{mol/min/g}$ ; Rate of flow from Glu to KG in GABAergic neurons; value indeterminate

$V_{\text{XGGKG}} = 0.5 \mu\text{mol/min/g}$ ; Rate of flow from Glu to KG in astrocytes; value indeterminate

$V_{\text{XKGGG}} = V_{\text{XGGKG}} + V_{\text{gad}} - V_{\text{cycleGABAGlu}} = 0.59 \mu\text{mol/min/g}$ ; Rate of flow from KG to Glu in astrocytes

$V_{\text{xOAAAsp}} = V_{\text{XGGKG}} = 0.5 \mu\text{mol/min/g}$ ; Rate of flow from OAA to Asp in GABAergic neurons

$X_1 = V_{\text{max\_in}} * \text{Glucose}_{\text{blood}} / (\text{Km\_in} + \text{Glucose}_{\text{blood}}) = 0.89 \mu\text{mol/min/g}$ ; Variable used to calculate steady-state brain glucose concentrations

$X_2 = X_1 - \text{CMR}_{\text{gl}} = 0.35 \mu\text{mol/min/g}$ ; Variable used to calculate steady-state brain glucose concentrations

$V_{\text{dilA}} = 0.0058 \mu\text{mol/min/g}$ ; Rate of dilution in astrocytes

$V_{\text{tcaA}} = V_{\text{ac}} + V_{\text{pdhA}} + V_{\text{dilA}} = 0.15 \mu\text{mol/min/g}$ ; Rate of TCA cycle in astrocytes

$V_{\text{tcaANet}} = 0.029 \mu\text{mol/min/g}$ ; Rate of TCA from citric acid to KG in astrocytes

$V_{\text{XAAspOAA}} = V_{\text{XAOAAAsp}} = 1 \mu\text{mol/min/g}$ ; Rate of flow from Asp to OAA in astrocytes

$V_{\text{X}} = V_{\text{XAGluKG}} = 1 \mu\text{mol/min/g}$ ; Rate of flow from OAA to Asp in astrocytes

$V_{\text{XAKGGlu}} = V_{\text{XAGluKG}} + V_{\text{glu}} - V_{\text{cycle}} = 1.12 \mu\text{mol/min/g}$ ; Rate of flow from KG to Glu in astrocytes

$V_{\text{XAOAAAsp}} = V_{\text{XAGluKG}} = 1 \mu\text{mol/min/g}$ ; Rate of flow from OAA to Asp in astrocytes

$V_{\text{pdhN}} = V_{\text{tcaN}} - V_{\text{dilN}} = 0.81 \mu\text{mol/min/g}$ ; Rate of flow through glutamatergic neuronal pyruvate dehydrogenase

$V_{\text{XNASpOAA}} = V_{\text{XNOAAAsp}} = 3.64 \mu\text{mol/min/g}$ ; Rate of flow from Asp to OAA in glutamatergic neurons

$V_{\text{XNGluKG}} = 3.64 \mu\text{mol/min/g}$ ; Rate of flow from Glu to KG in glutamatergic neurons

$V_{\text{XNKGGlu}} = V_{\text{XNGluKG}} = 3.64 \mu\text{mol/min/g}$ ; Rate of flow from KG to Glu in glutamatergic neurons

$V_{\text{XNOAAAsp}} = V_{\text{XNGluKG}} = 3.64 \mu\text{mol/min/g}$ ; Rate of flow from OAA to Asp in

*glutamatergic neurons*

*Pool Concentrations:*

$Asp_{GA} = Asp_{Total} - Asp_A - Asp_N = 1.43 \mu\text{mol/g}$ ; *GABAergic neuronal Asp*

$Asp_{Total} = 3.16 \mu\text{mol/g}$ ; *Total tissue Asp (measured)*

$Glucose_{brain} = Km_{out} * X_2 / (V_{max_{out}} - X_2) = 1.33 \mu\text{mol/g}$ ; *Brain glucose (Mason et al. 1992a)*

$GABA = 1.29 \mu\text{mol/g}$ ; *GABA concentration (measured)*

$Gln = 6.52 \mu\text{mol/g}$ ; *Gln concentration (measured)*

$Glu_{GA} = 0.02 * Glu_{Total} = 0.21 \mu\text{mol/g}$ ; *Glu concentration in GABAergic neurons (small but indeterminate concentration)*

$Glu_{Total} = 10.48 \mu\text{mol/g}$ ; *Total tissue Glu (measured)*

$KG_{GA} = 0.01 \mu\text{mol/g}$ ; *KG in GABAergic neurons, estimated from (Hawkins & Mans 1983)*

$L = 1.5 \mu\text{mol/g}$ ; *Tissue lactate concentration (Hawkins & Mans 1983)*

$OAA_{GA} = 0.05 \mu\text{mol/g}$ ; *OAA concentration in GABAergic neurons, estimated from (Hawkins & Mans 1983)*

$OAA_{Total} = 0.2 \mu\text{mol/g}$ ; *Total tissue OAA (Hawkins & Mans 1983)*

$Asp_A = R_{AspGlu} * Glu_A = 0.30 \mu\text{mol/g}$ ; *Asp in astrocytes*

$Glu_A = 0.1 * Glu_{Total} = 1.05 \mu\text{mol/g}$ ; *Glu in astrocytes (Patel et al. 2010, Lebon et al. 2002)*

$KG_A = 0.09 \mu\text{mol/g}$ ; *KG in astrocytes, estimated from (Hawkins & Mans 1983)*

$OAA_A = OAA_{Total} - OAA_N - OAA_{GA} = 0.05 \mu\text{mol/g}$ ; *OAA in astrocytes, estimated from (Hawkins & Mans 1983)*

$Asp_N = 0.5 * (Asp_{Total} - Asp_A) = 1.43 \mu\text{mol/g}$ ; *Asp in glutamatergic neurons*

$Glu_N = Glu_{Total} - Glu_A - Glu_{GA} = 9.22 \mu\text{mol/g}$ ; *Glu in glutamatergic neurons (Patel et al. 2010, Lebon et al. 2002)*

$KG_N = 0.1 \mu\text{mol/g}$ ; *KG in glutamatergic neurons, estimated from (Hawkins & Mans 1983)*

$OAA_N = 0.1 \mu\text{mol/g}$ ; *OAA in glutamatergic neurons, estimated from (Hawkins & Mans 1983)*

*Note: Glc, glucose; L, lactate; Glu, glutamate; Gln, glutamine; Asp, aspartate; OAA, oxaloacetate;*

*KG,  $\alpha$ -ketoglutarate; Subscript 'N', 'A' and 'GA' stand for glutamatergic neuron, astrocytes and*

1  
2  
3 GABAergic neurons, respectively. Numeric subscripts represent the position of  $^{13}\text{C}$  in the  
4 corresponding molecule. Parameters listed in bold-faced type were determined by iterative fitting.  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

For Peer Review

Table 2S. Mass of tissue samples (mg) from each region across all the rats.

No.	OB	CE	MEA	THA	HYP	HP	FC	PC	OC	ST	TC	MID
Mean	65.6	217.5	182.8	92.4	65.5	110.1	153.7	152.2	108.4	151.1	195.8	98.2
SD	9.7	21.7	21.4	12.7	15.5	10.7	23.1	22.3	14.0	30.9	23.5	12.5

For Peer Review

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60



1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

Table 3S. The descriptors selected by the linear discriminant analysis and their corresponding statistical parameters for the whole group. Their selection was based on the ability to differentiate rats that received nicotine from those that received saline.

Region	Metabolite	Tolerance	F to remove	Wilks' Lambda	CDF
ST	GABA	0.7235	9.7047	0.4500	0.7563
ST	NAA	0.6113	20.4191	0.5890	1.0432
THA	GABA	0.5524	10.6467	0.4622	-0.8944
HYP	Glu	0.4518	11.3550	0.4714	1.0114
TC	GABA	0.6021	7.8397	0.4258	-0.7659

Note: CDF: Standardized Canonical Discriminant Function Coefficients.

Table 4S: Effect of nicotine on cerebral cortical metabolic fluxes ( $\mu\text{mol}\cdot\text{g}\cdot\text{min}^{-1}$ ), including  $V_X$  (exchange rate between KG and glutamate in astrocytes),  $V_{\text{dilGln}}$  (the diluting inflow rate of glutamine),  $V_{\text{tcaN}}$  (TCA cycle flux in neuron),  $V_{\text{dil}}$  (total diluting flow into KG),

Brain	$V_X$		$V_{\text{tcaN}}$		$V_{\text{dilGln}}$		$V_{\text{dil}}$	
	Saline	Nicotine	Saline	Nicotine	Saline	Nicotine	Saline	Nicotine
CE	0.59±0.01	0.59±0.01	0.64±0.03	0.63±0.02	0.33±0.04	0.32±0.03	0.08±0.05	0.03±0.04
FC	0.57±0.01	0.57±0.00	0.77±0.05	0.66±0.02	0.30±0.04	0.27±0.02	0.02±0.04	0.01±0.02
TC	0.58±0.01	0.57±0.01	0.68±0.03	0.59±0.03	0.38±0.03	0.33±0.03	0.01±0.03	0.01±0.03
HP	0.58±0.01	0.57±0.01	0.64±0.03	0.52±0.02	0.37±0.04	0.30±0.03	0.00±0.04	0.02±0.03
HYP	0.58±0.01	0.57±0.01	0.53±0.02	0.47±0.02	0.36±0.03	0.36±0.03	0.01±0.03	0.01±0.03
MEA	0.59±0.01	0.57±0.00	0.53±0.02	0.47±0.02	0.39±0.03	0.34±0.03	0.00±0.03	0.02±0.03
MID	0.61±0.01	0.60±0.01	0.60±0.03	0.54±0.02	0.35±0.03	0.31±0.03	0.02±0.03	0.00±0.02
OB	0.56±0.01	0.57±0.00	0.58±0.04	0.52±0.03	0.33±0.04	0.34±0.04	0.02±0.05	0.02±0.04
OC	0.59±0.01	0.58±0.01	0.67±0.05	0.62±0.03	0.36±0.03	0.30±0.02	0.07±0.02	0.01±0.02
PC	0.59±0.01	0.57±0.01	0.81±0.03	0.67±0.03	0.35±0.03	0.26±0.02	0.01±0.03	0.00±0.02
ST	0.58±0.01	0.55±0.01	0.67±0.02	0.55±0.02	0.30±0.03	0.28±0.02	0.00±0.03	0.01±0.02
THA	0.58±0.01	0.58±0.00	0.64±0.03	0.65±0.03	0.33±0.03	0.33±0.03	0.01±0.02	0.00±0.02

Table 5S: The p-values of pair-wise comparisons of rates of metabolism after nicotine injection in different regions, with indications of which rates survived adjustments for multiple comparisons.

	CE	FC	FDC	HP	HYP	MEA	MID	OB	OC	PC	ST	THA
CMR <sub>gl(ox)</sub>	0.449	0.007	0.045	0.010	0.031	0.038	0.022	0.008	<b>0.000*</b>	<b>0.000*</b>	0.004	0.433
V <sub>cycle</sub>	0.196	0.212	0.238	0.303	0.522	0.742	0.153	0.313	<b>0.001*</b>	<b>0.000*</b>	0.267	0.537
V <sub>gad</sub>	0.578	0.109	0.427	0.314	0.096	0.035	0.354	0.883	0.037	0.039	<b>0.025*</b>	0.717

Note: \*: The rates that remained significant after adjustment for multiple comparisons.

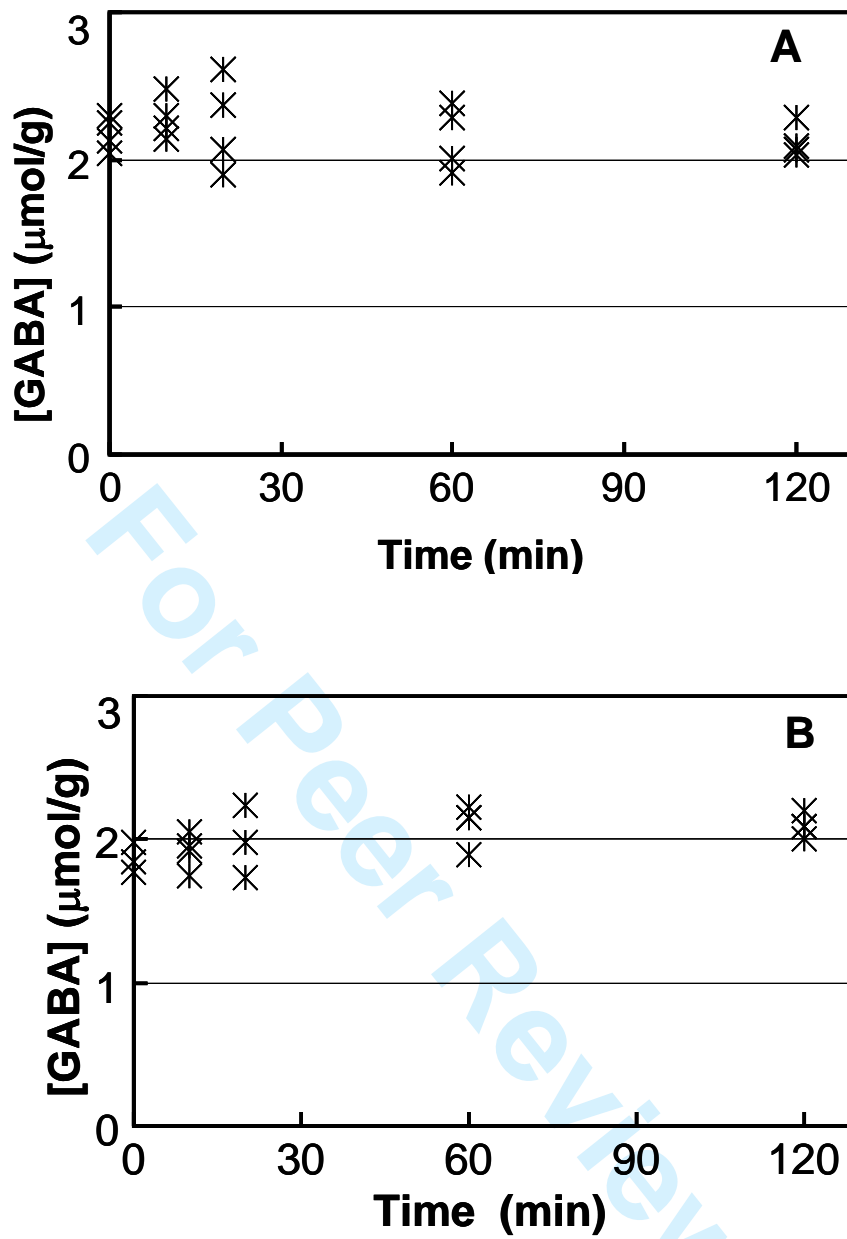


Fig. 1S. Time course of the concentration of GABA in striatum (A) Saline, (B) Nicotine.

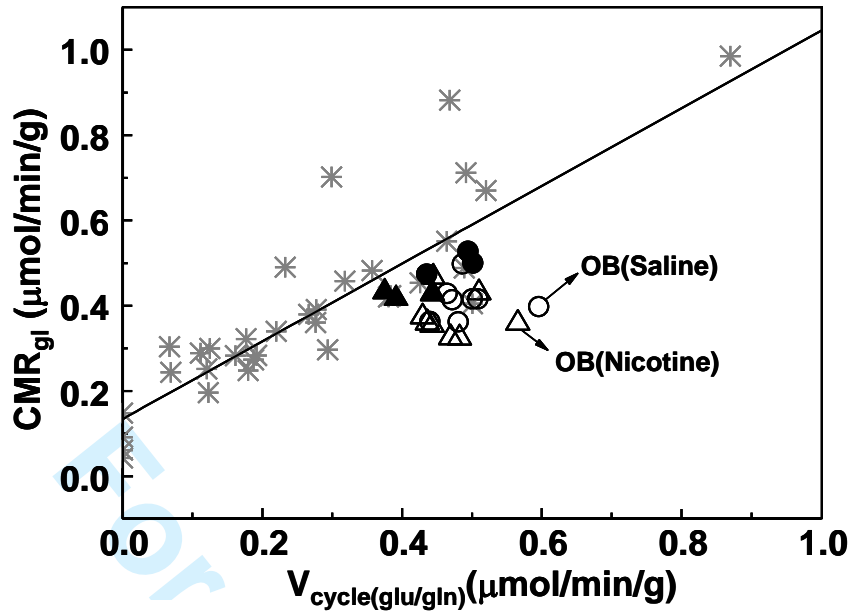


Fig. 2S. Correlation between the rate of glucose oxidation ( $\text{CMR}_{\text{gl(ox)}}$ ) and glutamate neurotransmitter cycling ( $V_{\text{cycle}}$ ). The plot combined the data from the present study (control group = ●; nicotine = ▲) and the data from former studies (\*) (Patel et al. 2004, Sibson et al. 1998). The data from other regions were also added (saline = ○; nicotine = △). The line represents the regression analysis of both the wake and anaesthetized rat cortex, given by the equation  $y = 0.912x + 0.134$ , with a correlation coefficient of  $r = 0.8672$ .

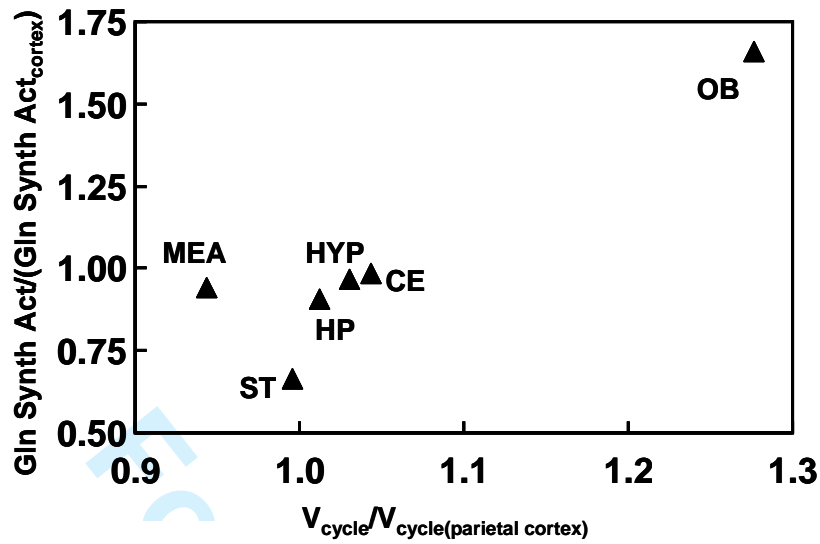


Fig. 3S: Relationship between glutamine synthetase activity in each region, taken as a ratio relative to the cortex, and the ratio of  $V_{\text{cycle}}$  in each region as a ratio relative to the value we measured for  $V_{\text{cycle}}$  in the parietal cortex. The regions for which glutamine synthetase data were available were the OB, ST, CE, HP, HYP, and MEA.

**Bibliography**

- 1  
2  
3  
4  
5  
6 Buschiazzo, P. M., Terrell, E. B. and Regen, D. M. (1970) Sugar transport across the  
7 blood-brain barrier. *Am J Physiol*, **219**, 1505-1513.
- 8 Fisher, R. A. (1936) The use of multiple measurements in taxonomic problems. *Ann.*  
9 *Eugen.*, **7**, 179-188.
- 10 Hawkins, R. A. and Mans, A. M. (1983) Intermediary metabolism of carbohydrates and  
11 other fuels. I. In: *Biochem J*, (A. Lajtha ed.), Vol. 122, pp. 259-294. Plenum Press,  
12 New York.
- 13  
14 Lebon, V., Petersen, K. F., Cline, G. W., Shen, J., Mason, G. F., Dufour, S., Behar, K. L.,  
15 Shulman, G. I. and Rothman, D. L. (2002) Astroglial contribution to brain energy  
16 metabolism in humans revealed by <sup>13</sup>C nuclear magnetic resonance spectroscopy:  
17 Elucidation of the dominant pathway for neurotransmitter glutamate repletion and  
18 measurement of astrocytic oxidative metabolism. *J. Neurosci.*, **22**, 1523-1531.
- 19  
20 Mason, G. F., Behar, K. L., Rothman, D. L. and Shulman, R. G. (1992a) NMR  
21 Determination of Intracerebral Glucose-Concentration and Transport Kinetics in  
22 Rat-Brain. *J. Cereb. Blood Flow Metab.*, **12**, 448-455.
- 23  
24 Mason, G. F., Rothman, D. L., Behar, K. L. and Shulman, R. G. (1992b) NMR  
25 determination of the TCA cycle rate and alpha-ketoglutarate/glutamate exchange  
26 rate in rat brain. *J Cereb Blood Flow Metab*, **12**, 434-447.
- 27  
28 Patel, A. B., de Graaf, R. A., Mason, G. F., Kanamatsu, T., Rothman, D. L., Shulman, R.  
29 G. and Behar, K. L. (2004) Glutamatergic Neurotransmission and Neuronal  
30 Glucose Oxidation Are Coupled During Intense Neuronal Activation. *J. Cereb.*  
31 *Blood Flow Metab.*, **24**, 972-985.
- 32  
33 Patel, A. B., de Graaf, R. A., Mason, G. F., Rothman, D. L., Shulman, R. G. and Behar, K.  
34 L. (2005) The contribution of GABA to glutamate/glutamine cycling and energy  
35 metabolism in the rat cortex in vivo. *Proc Natl Acad Sci USA*, **102**, 5588-5593.
- 36  
37 Patel, A. B., de Graaf, R. A., Rothman, D. L., Behar, K. L. and Mason, G. F. (2010)  
38 Evaluation of Cerebral Acetate Transport and Metabolic Rates in the Rat Brain In  
39 Vivo using <sup>1</sup>H-[<sup>13</sup>C]NMR. *J. Cereb. Blood Flow Metab.*, **in press**.
- 40  
41 Sibson, N. R., Dhankhar, A., Mason, G. F., Rothman, D. L., Behar, K. L. and Shulman, R.  
42 G. (1998) Stoichiometric coupling of brain glucose metabolism and glutamatergic  
43 neuronal activity. *P. Nati. Cad. Sci. USA*, **95**, 316-321.
- 44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60