

SUPPLEMENTARY DATA

SUPPLEMENTARY EXPERIMENTAL PROCEDURES

Mutagenesis of EGFP-MAP1LC3 β — A plasmid encoding human MAP1LC3 β fused to the COOH-terminus of green fluorescent protein was obtained from Addgene (1) and used to introduce silent mutations within the MAP1LC3 β sequence using the oligonucleotide primers 5'-CATCCAACCAAAATCCCGGTAATAATAGAGGCGATACAAGGGTGAGA-3' (sense) and 5'-TCTCACCTTGTATCGCTCTATTATTACCGGGATTTTGGTTGGATG-3' (antisense), and the Quikchange™ mutagenesis kit (Stratagene). The resulting plasmid was sequenced to verify the specific mutations and the absence of PCR errors.

Rescue of TLR3-mediated Type I Interferon Signaling by Mutant EGFP-MAP1LC3 β — To restore type I interferon signaling activity in WT TLR3-HA expressing 293 cells following LC3 β siRNA-A-mediated knockdown of MAP1LC3 β expression, cells were transfected with mutant EGFP-MAP1LC3 β encoding plasmid as follows. WT cells treated with wild type LC3 β siRNA-A duplex and growing on a 60-mm culture dish were harvested and one fourth of the cells was pelleted and suspended in a transfection mixture consisting of 0.5 μ g of mutant EGFP-MAP1LC3 β plasmid, 2.5 μ g of pISRE-LUC reporter, 100 pmoles of wild type LC3 β siRNA-A duplex and 10 μ l of Lipofectamine 2000™ in 350 μ l of Opti®-MEM plus 2 ml of DMEM without antibiotics. One hundred microliter samples of the suspended cells were then plated in 8 wells of a 96-well plate and grown overnight at 37°C. Cells were stimulated with 25 μ g/ml of poly I:C and the luciferase activity was measured with 100 μ l of Bright-Glo™ reagent. Luciferase activity was normalized to the expression of TLR3-HA and expressed relative to the activity detected in the unstimulated WT-TLR3 expressing cells transfected with GFP-MAP1LC3 β encoding plasmid alone. Expression of GFP-MAP1LC3 β fusion proteins was monitored by Western blotting with a monoclonal antibody to GFP (Santa Cruz Biotechnology) or polyclonal anti-MAP1LC3 β antibodies (Novus Biologicals).

Mutagenesis of Beclin 1— The plasmid pcDNA3-Beclin 1 encoding the human Beclin 1 protein was purchased from Addgene (2) and used as a template to introduce silent point mutations corresponding to those present in the Beclin 1 siRNA-C duplex (*see* Table 1) sequence using the oligonucleotides 5'CTTAGAGCAAATGAATGAGGACCGATAGTGAACAATTACAGATGGAGCTAAAGGAG-3' (sense) and 5'-CTCCTTTAGCTCCATCTGTAATTGTTCACTATCGTCCTCATTGCTCTAAG-3' (antisense), and the Quikchange™ mutagenesis kit. The resulting construct was fully sequenced to verify the presence of the desired mutations and the absence of PCR errors.

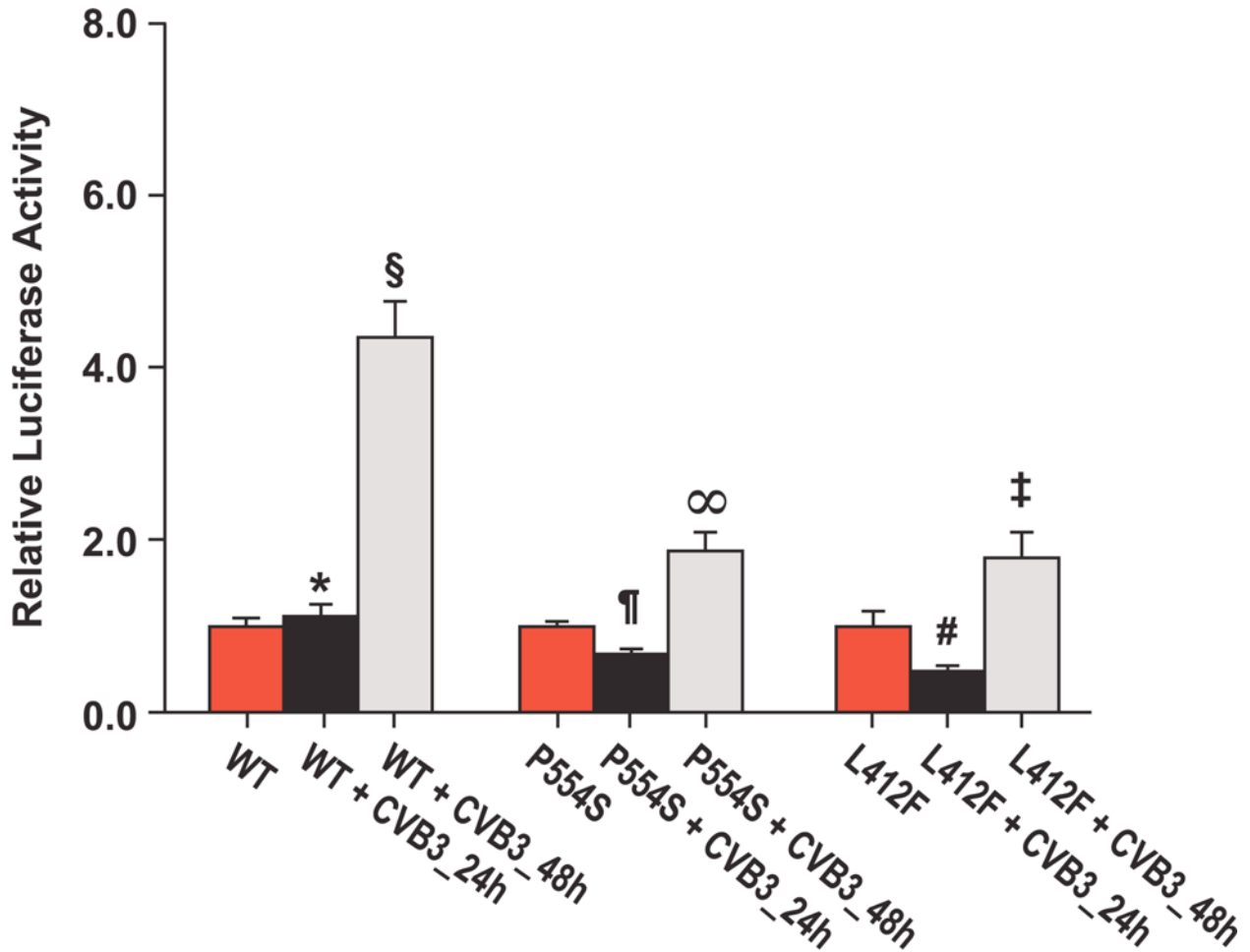
Rescue of TLR3-mediated Type I Interferon Signaling by Wild Type Beclin 1— To restore type I interferon signaling in WT TLR3-HA cells subjected to two rounds of Beclin 1 RNAi, cells were co-transfected a third time with siRNA-C duplex and either WT Beclin 1 encoding plasmid, or the mutant pcDNA3-Beclin 1 construct as follows. The harvested cells were suspended in 14 ml of media, dispensed into 4-ml aliquots and centrifuged. The cell pellets were then resuspended in transfection mixture consisting of either 50 pmoles of siRNA-C plus 2.5 μ g of pISRE-LUC reporter, or 50 pmoles of siRNA-C, 2.5 μ g of reporter plus 0.5 μ g of either WT or mutant pcDNA3-Beclin 1 plasmid in 330- μ l of Opti™-MEM containing 13.2 μ l of Lipofectamine 2000™. 1.9-ml of DMEM without antibiotics was added to each cell suspension, and 100- μ l aliquots were then plated in 16 wells of a 96-well plate and grown overnight at 37°C. Cells in eight of the wells were stimulated for 24 h in 100 μ l of DMEM plus antibiotics containing 25 μ g/ml poly I:C and the luciferase activity was measured as described above. Luciferase activity was normalized to TLR3-HA expression and expressed relative to the activity detected in unstimulated WT TLR3 cells transfected with reporter alone. Expression of Beclin 1 was monitored by Western blotting

with a monoclonal antibody (Santa Cruz Biotechnology) in the whole cell extracts from ~0.6-ml samples of the cell suspensions described above grown in parallel in a 24-well plate.

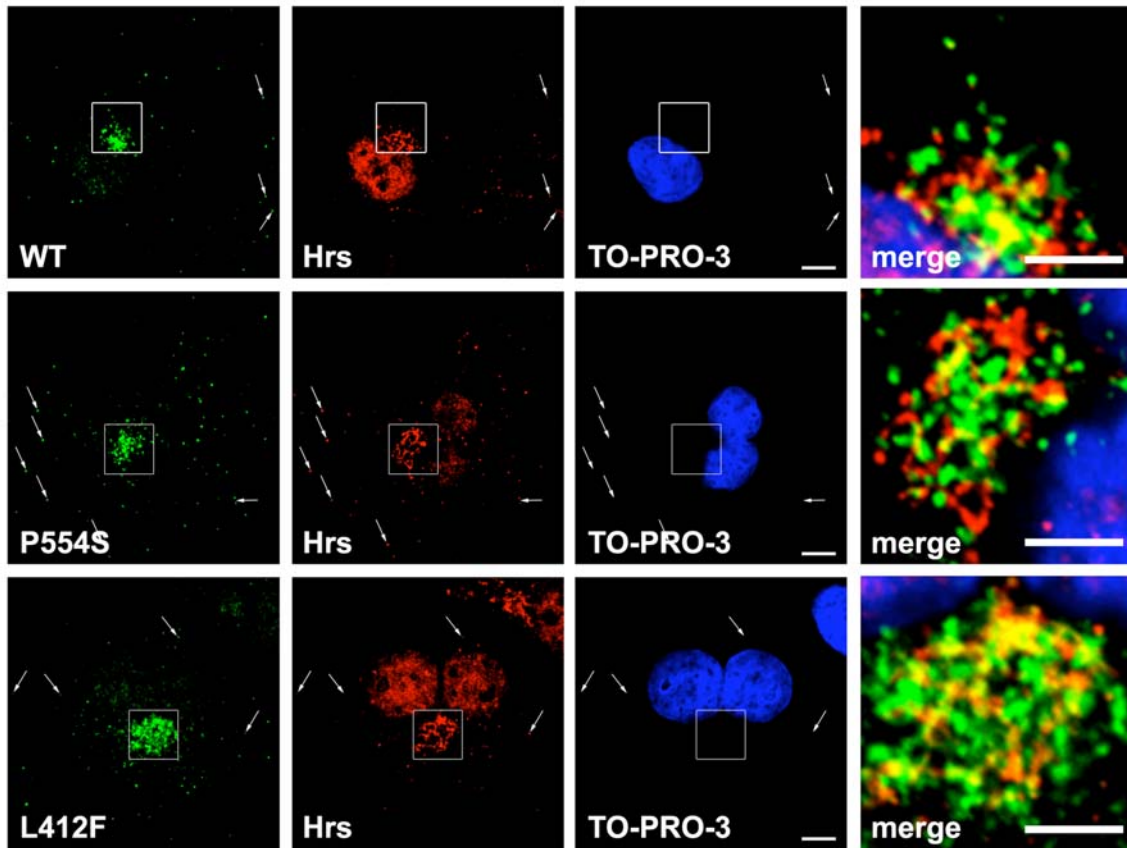
SUPPLEMENTARY REFERENCES

1. Jackson, W. T., Giddings, T. H., Jr., Taylor, M. P., Mulinyawe, S., Rabinovitch, M., Kopito, R. R., and Kirkegaard, K. (2005) *PLoS Biol.* **3**, e156
2. Shihibata, M., Lu, T., Furuya, T., Degterev, A., Mizushima, N., Yoshimori, T., MacDonald, M., Yankner, B., and Yuan, J. (2006) *J. Biol. Chem.* **281**, 14474-14485

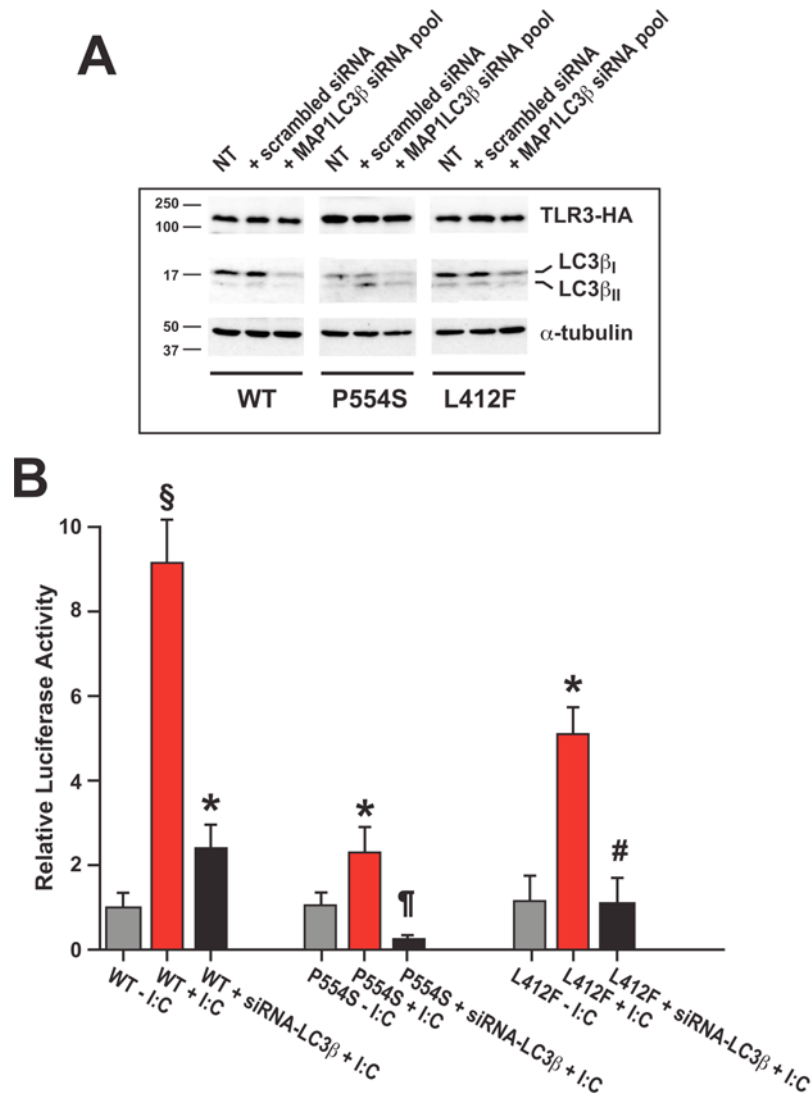
SUPPLEMENTARY FIGURES



Supplementary Figure 1: Analysis of the effect of TLR3 variants on type I interferon signaling following enterovirus infection (MOI = 0.01). Stable cell lines expressing WT, Ser554 (P554S) or Phe412 (L412F) TLR3-HA were transfected with pISRE-LUC and then stimulated infected with Coxsackievirus B3. Luciferase activity was measured at 0 (red bars), 24 (black bars) and 48 (grey bars) hours post-infection, normalized to exogenous TLR3 expression and expressed relative to uninfected cells. The data shown are the *means* \pm *SEM* of eight independent measurements. * P=NS vs. unstimulated WT. § P<0.002 vs. uninfected WT and all other groups. ¶ P=NS vs. uninfected cells in all groups. ∞ P<0.002 versus uninfected Ser554 or infected Ser554 for 24 hours. # P=NS vs. uninfected or 24-hour infected cells in all groups. ‡ P<0.002 versus uninfected cells in all groups or Phe 412 cells infected for 24 hours; P=NS vs. 48-hour infected Ser554 cells or WT cells infected for 24 hours.



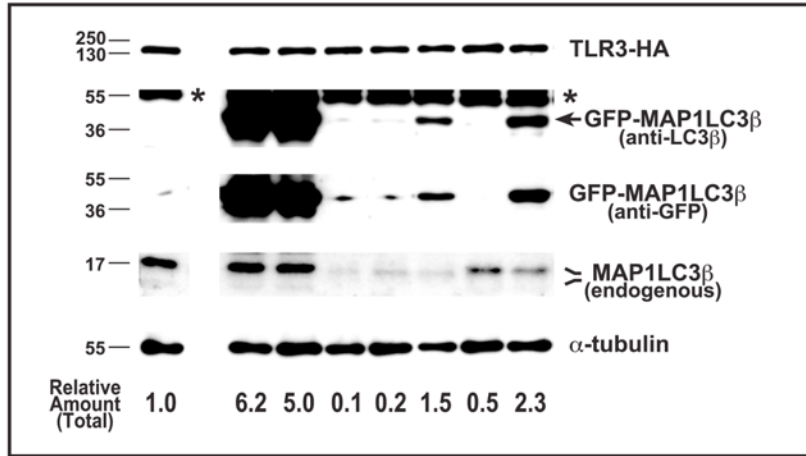
Supplementary Figure 2: TLR3 variants localize to late endosomes/multivesicular bodies (MVB). Stable COS7 cells expressing WT, Ser554 (P554S) or Phe412 (L412F) TLR3-HA were permeabilized, fixed with 3% paraformaldehyde, and stained with an antibody against HA (to detect exogenous TLR3), an affinity-purified antibody to hepatocyte growth-factor regulated tyrosine kinase substrate or Hrs (to detect MVB: a gift from Harald Stenmark), and with TO-PRO-3™ iodide (to detect nuclei). The *right panels* show the merged and magnified images of the enclosed areas directly to the left and show the partial overlap of TLR3 and Hrs staining. The *white arrows* indicate endosomes/MVB to which exogenous TLR3-HA variants and Hrs co-localize. *Bars*, full-size images: 10 μm ; enlarged images: 5 μm .



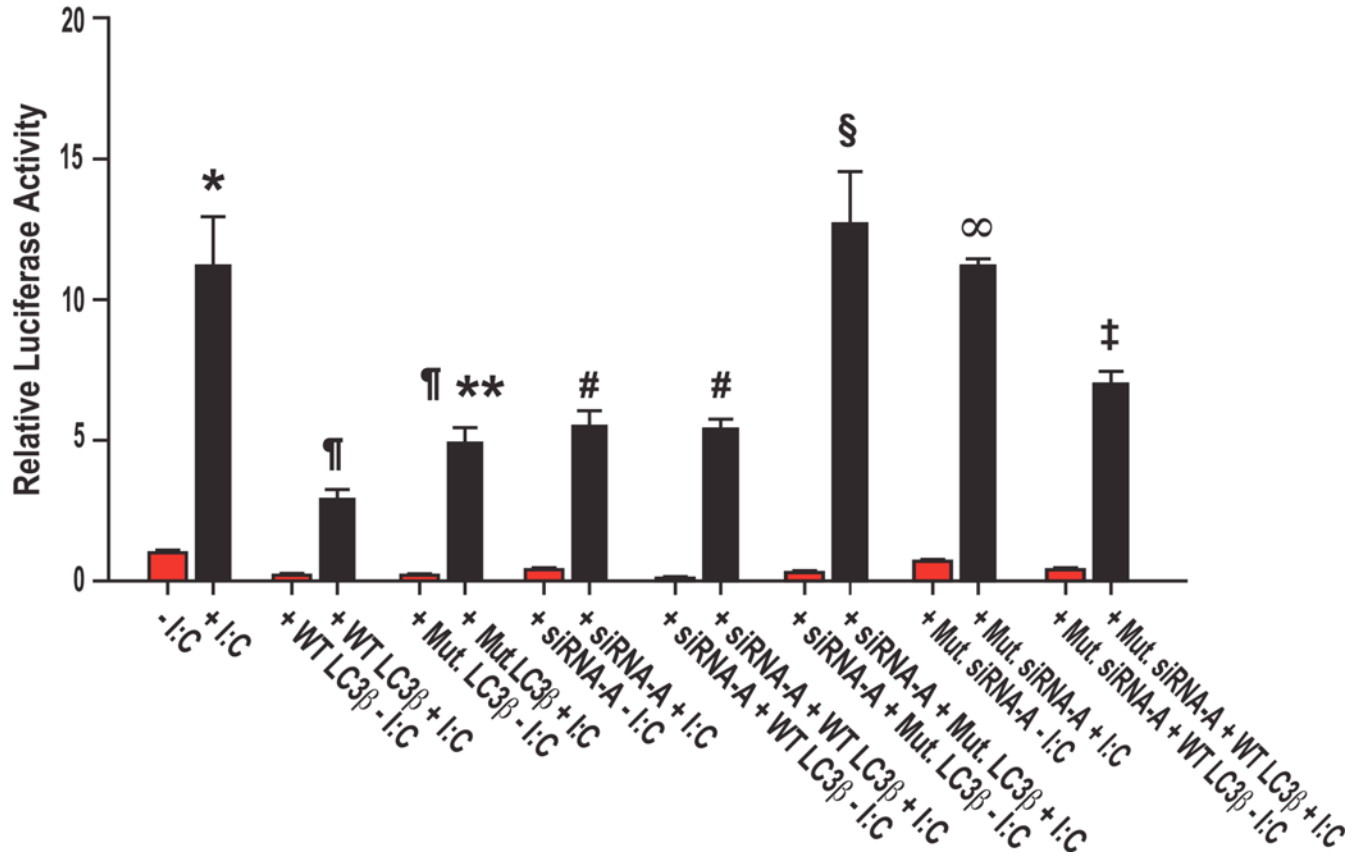
Supplementary Figure 3: Gene silencing of MAP1LC3 β by siRNA inhibits type I interferon signaling by TLR3 variants. *Panel A.* Levels of TLR3-HA, α -tubulin and MAP1LC3 β (LC3 β), with or without MAP1LC3 β knockdown. Stable 293 cells expressing WT, Ser554 (P554S) or Phe412 (L412F) TLR3-HA were transfected with a scrambled siRNA control or a pool of three MAP1LC3 β siRNAs, and equal amounts of protein from the whole cell extracts were then analyzed by SDS-PAGE and immunoblotting with anti-HA, anti- α -tubulin, or a polyclonal antibody to MAP1LC3 β . *Panel B.* WT, Ser554 (P554S) or Phe412 (L412F) TLR3-HA expressing cells were transfected with pISRE-LUC and either a scrambled siRNA duplex, or a MAP1LC3 β (siRNA-LC3 β) siRNA pool, and then stimulated with poly I:C (+ I:C: red and black bars) or vehicle (grey bars). Relative luciferase activity in stimulated WT cells transfected with scrambled siRNA-A duplex was 9.1 ± 1.0 , whereas the luciferase activity in the Ser554 and Phe412 TLR3 expressing cells was 2.3 ± 0.6 and 5.1 ± 0.6 , respectively. By contrast, the luciferase activity in WT cells transfected with the MAP1LC3 β siRNA pool was reduced 74% (2.4 ± 0.5), while the activity in the Ser554 and Phe412 TLR3 expressing cells decreased by 91% and 78%, respectively: 0.2 ± 0.04 in Ser554, and 1.1 ± 0.5 in Phe412. * $P < 0.01$ vs. stimulated WT. ¶ $P < 0.03$ versus stimulated Ser554 TLR3 cells. # $P < 0.01$ versus stimulated Phe412 TLR3 expressing cells. § $P < 0.001$ vs. unstimulated wild type TLR3 cells.

A

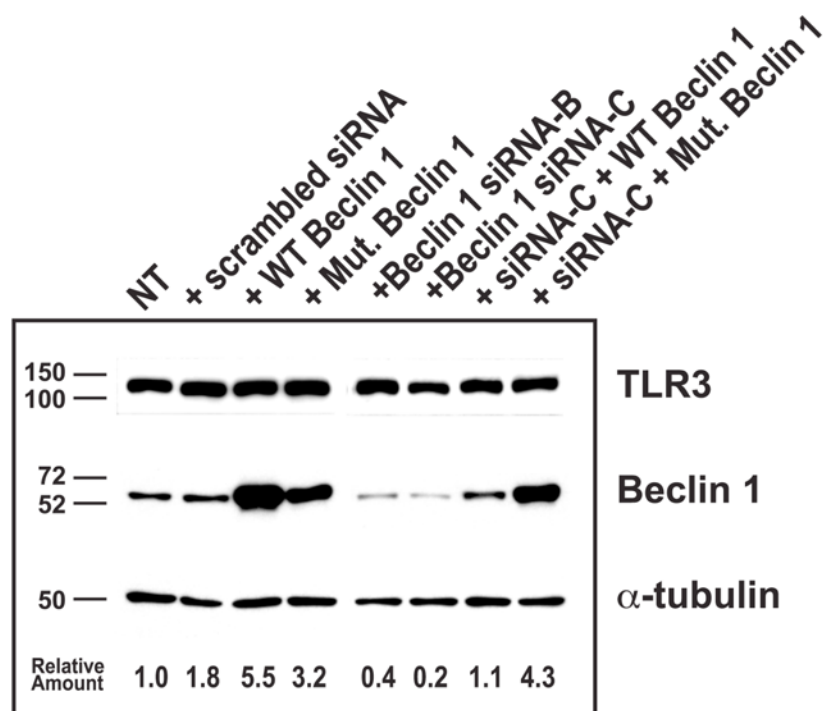
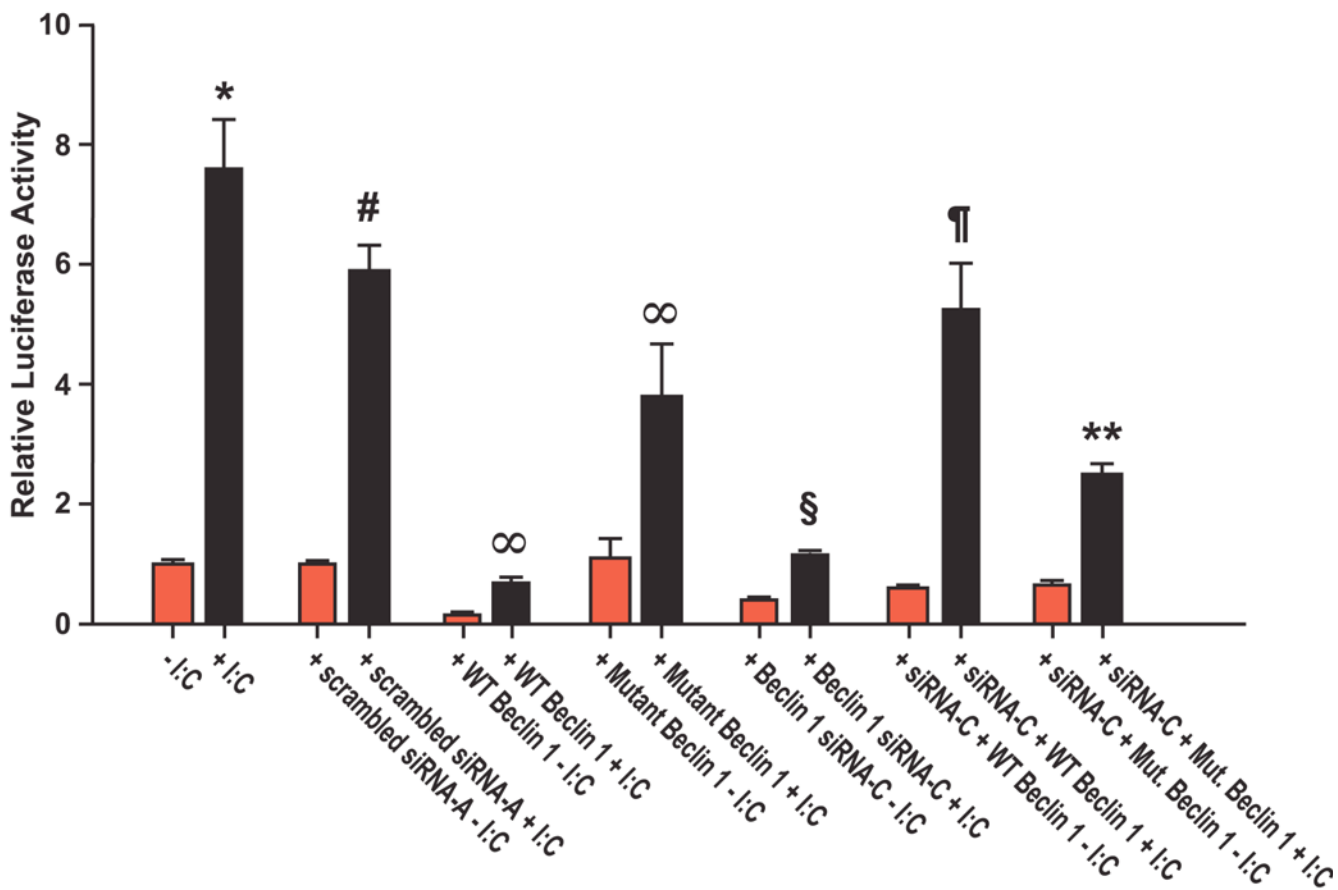
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| - | + | - | - | + | - | - | + | WT GFP-MAP1LC3 β |
| - | - | + | - | - | + | - | - | Mut GFP-MAP1LC3 β |
| - | - | - | + | + | + | - | - | WT LC3 β siRNA-A |
| - | - | - | - | - | - | + | + | Mut LC3 β siRNA-A |



B



Supplementary Figure 4: *Panel A.* Rescue of MAP1LC3 β expression following transfection of plasmid encoding mutant GFP-MAP1LC3 β . Stable 293 cells expressing WT TLR3-HA were subjected to RNAi using wild type LC3 β siRNA-A followed by co-transfection of the siRNA duplex with a plasmid encoding either wild type or mutant GFP-MAP1LC3 β fusion protein. Levels of TLR3-HA, α -tubulin and MAP1LC3 β were determined by Western blotting of the whole cell extracts with anti-HA, anti- α -tubulin, and either a monoclonal antibody to GFP (to detect the GFP-MAP1LC3 β fusion protein) or polyclonal anti-MAP1LC3 β (to detect both endogenous and recombinant MAP1LC3 β) followed by densitometry using NIH image 1.63f. The total (endogenous + recombinant) amount of MAP1LC3 β in the extract is expressed relative to the level of MAP1LC3 β in non-transfected cells. Transfection of either WT or mutant MAP1LC3 β encoding plasmid alone increased the overall level of MAP1LC3 β in the cell extracts 5- and 6-fold, respectively. Note that endogenous MAP1LC3 β protein migrates on the gel near the 17-kDa marker and its level was considerably lower in cells subjected to RNAi with wild type LC3 β siRNA-A, whereas the mutant GFP-MAP1LC3 β fusion protein migrates between the 55- and 36-kDa markers (predicted $M_r = 43$ K) and restored the overall level of MAP1LC3 β in LC3 β siRNA-A treated cells. For comparison, we also co-transfected WT cells with a mutated (*Mut.*) version of LC3 β siRNA-A and plasmid encoding wild type GFP-MAP1LC3 β , in which case the overall amount of MAP1LC3 β in the extract increased more than 2-fold relative to the control WT cells. The *asterisks* denote a protein that reacts non-specifically with the polyclonal antibody to MAP1LC3 β . *Panel B.* Rescue of TLR3-mediated type I interferon signaling by mutant GFP-MAP1LC3 β expression. WT TLR3-HA expressing cells were subjected to RNAi by transfection with wild type LC3 β siRNA-A, and grown for 72 hours prior to co-transfection of the siRNA duplex with the pISRE-LUC reporter and plasmid encoding either wild type or mutant GFP-MAP1LC3 β . Twenty-four hours after transfection the cells were stimulated with poly I:C (+ I:C: *black bars*) or vehicle (- I:C: *red bars*). The relative luciferase activity in WT TLR3 expressing cells transfected with reporter alone was 11.2 ± 1.7 (* $P < 0.001$ vs. unstimulated WT), whereas transfection of either the WT or the mutant GFP-MAP1LC3 β encoding plasmid alone significantly decreased interferon signaling following poly I:C stimulation to 5.5 ± 0.5 and 2.9 ± 0.3 , respectively ($\#\ P < 0.001$ vs. stimulated WT; $\#\#\ P < 0.01$ vs. stimulated WT + wild type GFP-MAP1LC3 β). This indicates that high levels of exogenously expressed MAP1LC3 β inhibit TLR3 signaling. The luciferase activity in WT cells transfected with reporter plus wild type LC3 β siRNA-A duplex was 4.9 ± 0.5 ($\# P < 0.001$ vs. stimulated WT; $P = 0.005$ vs. unstimulated WT), which upon expression of recombinant GFP-MAP1LC3 β failed to return to the activity level present in control cells (5.4 ± 0.4 : $P = \text{NS}$ vs. stimulated WT + LC3 β siRNA-A). In contrast, transfection of mutant GFP-MAP1LC3 β encoding plasmid restored both the overall MAP1LC3 β protein (*Panel A*) and signaling to the levels in WT cells transfected only with the pISRE-LUC reporter (12.8 ± 1.7 : $\S P < 0.001$ vs. stimulated WT plus wild type LC3 β -siRNA-A; $P = \text{NS}$ vs. stimulated WT). By comparison, the luciferase activity in WT cells transfected with Mut. LC3 β siRNA-A was 11.2 ± 0.2 ($\infty P = \text{NS}$ vs. stimulated WT) and was reduced following transfection of both mutant duplex and wild type GFP-MAP1LC3 β encoding plasmid ($\ddagger P < 0.001$ vs. stimulated WT or stimulated WT cells plus mutant LC3 β siRNA-A).

A**B**

Supplementary Figure 5: *Panel A.* Rescue of Beclin 1 expression following transfection of plasmid encoding wild type or mutant Beclin 1. Stable 293 cells expressing WT TLR3-HA were subjected to two rounds of RNAi using Beclin 1 siRNA-C followed by co-transfection of the siRNA-C duplex with pcDNA3-Beclin 1 encoding either WT Beclin 1, or a version of the protein bearing three silent mutations within the siRNA-C sequence (*Mut. Beclin 1*). Levels of TLR3-HA, α -tubulin and Beclin 1 were determined by Western blotting of the whole cell extracts using monoclonal antibodies to the HA epitope, α -tubulin and Beclin 1. The amount of Beclin 1 in the extracts was determined by densitometry using the NIH image 1.63 software package and is expressed relative to the amount of Beclin 1 in non-transfected (NT) cells. For comparison, WT cells were transfected with a scrambled siRNA control, wild type or mutant Beclin 1 encoding plasmid alone (+ WT or + *Mut. Beclin 1*, respectively), or with Beclin 1 siRNA-B. Note that expression of mutant Beclin 1 in the presence of siRNA-C (*siRNA-C + WT Beclin 1*) resulted in a 4-fold increase in Beclin 1 expression relative to the NT control. By contrast, the level of Beclin 1 following transfection of duplex and WT plasmid (*siRNA-C + WT Beclin 1*) was unchanged relative to the NT control, and 5-fold lower than in WT TLR3 cells transfected with wild type Beclin 1 encoding plasmid. *Panel B.* Rescue of TLR3-mediated type I interferon signaling by wild type Beclin 1 expression. WT TLR3-HA expressing cells were subjected to two rounds of RNAi by transfection initially with Beclin 1 siRNA-C and then grown for 96 hours prior to co-transfection of the siRNA duplex with the pISRE-LUC reporter and either WT or mutant pcDNA3-Beclin 1. Twenty-four hours after transfection the cells were stimulated with poly I:C (+ I:C: *black bars*) or vehicle (- I:C: *red bars*). The relative luciferase activity in stimulated cells transfected with reporter alone was 7.6 ± 0.8 (* $P < 0.001$ vs. unstimulated WT), whereas the luciferase activity in stimulated cells transfected with reporter plus scrambled siRNA control was 5.9 ± 0.4 (# $P = \text{NS}$ vs. stimulated WT cells). By contrast, the luciferase activity in stimulated cells treated with siRNA-C was 1.1 ± 0.1 (~85% reduction: § $P < 0.001$ vs. stimulated WT cells or stimulated WT plus scrambled siRNA). When WT TLR3 cells were transfected with WT or mutant Beclin 1 encoding plasmids the luciferase activity was significantly reduced upon poly I:C stimulation to 0.7 ± 0.1 and 3.8 ± 0.9 , respectively (∞ $P < 0.001$ vs. stimulated WT in both cases), indicating that high levels of exogenously expressed Beclin 1 inhibit TLR3 signaling. When the cells were transfected with both siRNA-C and the mutant Beclin 1 encoding plasmid there was no significant change in Beclin 1 expression relative to the amount of Beclin 1 detected in cells transfected with mutant plasmid alone (*Panel A*), and no significant change in luciferase activity (2.5 ± 0.2 : ** $P = \text{NS}$ vs. stimulated WT cells plus mutant Beclin 1). In contrast, when the cells were transfected with both siRNA-C and the wild type Beclin 1 encoding plasmid, the level of Beclin 1 expression decreased and returned to the level present in non-transfected cells (*Panel A*) and more importantly, luciferase activity was also restored and returned to the level in stimulated cells transfected with the pISRE-LUC reporter and the scrambled siRNA control (5.2 ± 0.8 : ¶ $P = \text{NS}$ vs. stimulated WT cells plus scrambled siRNA). Thus, the magnitude of type I interferon signaling was consistent with a similar level of Beclin 1 expression in the NT control and in the cells transfected with siRNA-C plus WT Beclin 1 encoding plasmid.