

Supporting Information

Gonzalez et al. 10.1073/pnas.0911854107

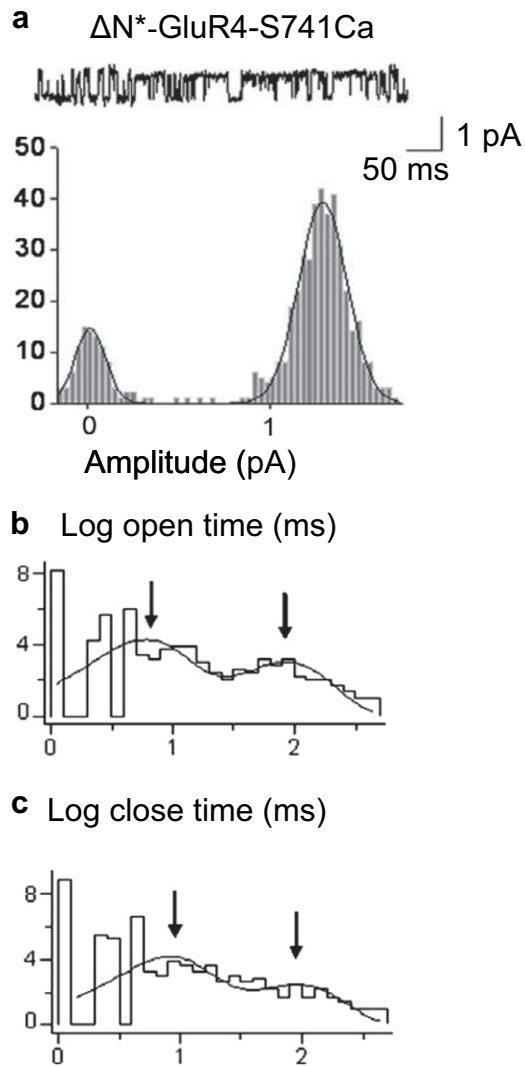


Fig. S1. Single-channel currents from bilayers of membrane preparations of modified glutamate receptors expressed in oocytes obtained using the same protocol as for the LRET investigations. (A) Amplitude histograms and representative single-channel traces are shown for terbium chelate and ATTO465-labeled ΔN^* -GluR4-S741C receptors. Currents were recorded at 120 mV in the presence of 1 mM glutamate and 100 μ M cyclothiazide. The primary conductance state is 9–12 pS. (B) Open and (C) close time distributions are shown for ΔN^* -GluR4-S741C; two distinct close times and two distinct open times were obtained.

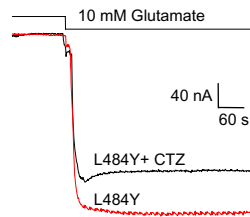


Fig. S2. Currents from GluR4-S742C-L484Y receptors expressed in oocytes. Currents were recorded in saturating concentrations of glutamate (10 mM) and in the presence and absence of cyclothiazide (CTZ) (100 μ M).

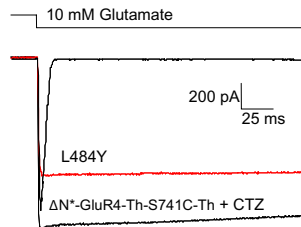


Fig. S3. Currents from Δ N*-GluR4-Th-S741C-Th receptors expressed in HEK-293 cells. Currents were recorded in the presence of saturating concentrations (10 mM) of glutamate, in the presence of 10 mM glutamate and 100 μ M cyclothiazide (CTZ), and for the L484Y mutation.

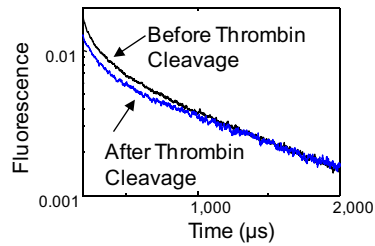


Fig. S4. The LRET lifetimes measured at 510 nm for donor:acceptor-tagged Δ N*-GluR4-Th-S741-Th before and after thrombin cleavage shown in the logarithmic y axis.

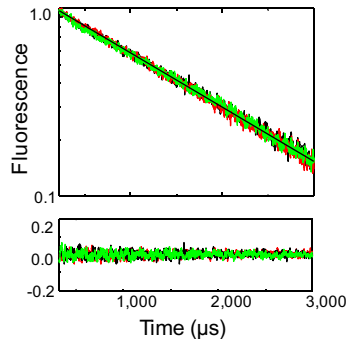


Fig. S5. LRET lifetimes shown in the logarithmic y axis for donor-only-tagged Δ N*-GluR4-S741 expressed in oocytes. Lifetimes were measured at 488 nm in the Apo (black), open (10 mM glutamate in the presence of 100 μ M cyclothiazide; red), and desensitized (10 mM glutamate; green) states. Residuals for the LRET lifetime fits are shown in linear y axis.

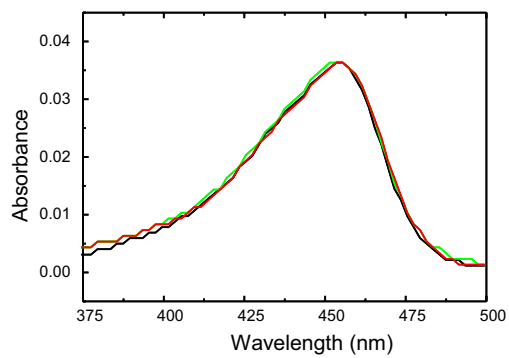


Fig. S9. Absorption spectrum of ATTO 465 (black) is not significantly affected by the addition of 10 mM glutamate (green) or 100 μ M cyclothiazide (red).