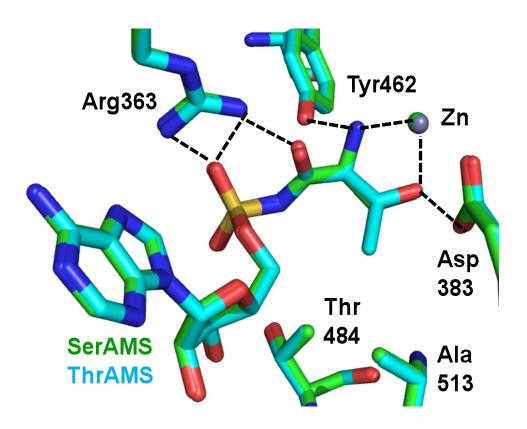
Supplemental data for

AMINOACYL TRANSFER RATE DICTATES CHOICE OF EDITING PATHWAY IN THREONYL-tRNA SYNTHETASE

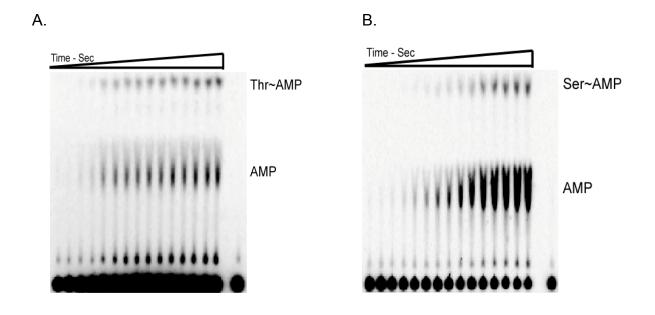
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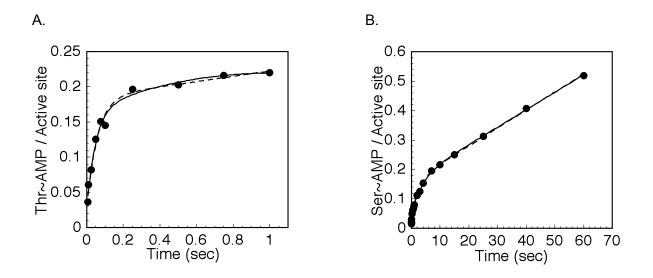
Supplemental Figure 1.



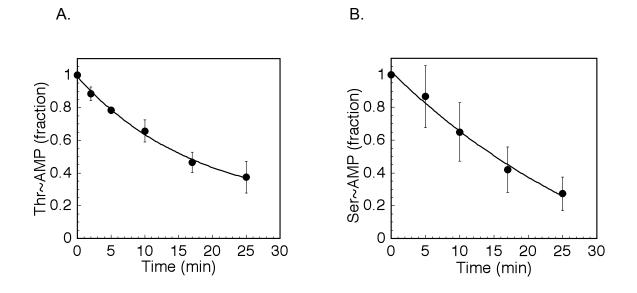
Overlay of the ThrRS:ThrAMS and ThrRS:SerAMS active site complexes (PDB id's 1EVL and 1FYF). The cognate adenylate is shown in cyan and the seryl~adenylate is in green. Active site residues that make contact to aminoacyl moieties of adenylates are shown.



Asymmetric amino acid activation by ThrRS. The rapid quench experiments were performed at pH 8.0 and 37 °C in the absence of tRNA as described in Experimental Procedures, using 10 μ M ThRS active sites, 100 μ M [α -³²P] ATP, and 5 mM threonine or 150 mM serine. (A), TLC chromatogram showing the formation of [α -³²P] AMP and Thr~[α -³²P] AMP, and (B) TLC chromatogram showing the formation of [α -³²P] AMP and Ser~[α -³²P] AMP.



Formation of adenylate under pre-steady state conditions: The experimental conditions are same as described in supplemental Fig 2. The solid line represents a double exponential with a linear scale fit, while fit to a burst equation is presented as dashed line.



Solvent-mediated adenylate hydrolysis is slower than the steady state rate of adenylate turnover. (A), decay curve for threonyl-adenylate solvolysis in the absence of ThrRS. (B) decay curve for seryl-adenylate solvolysis. The adenylate formation reactions were carried out for five minutes in Buffer A at pH 8.0 and 37 °C as described in Experimental Procedures, employing 2 μ M wild type ThrRS, 100 μ M [α -³²P] ATP, and 5 mM threonine or 300 mM serine. The reaction was terminated by adding EDTA (pH 8.0) to 500 mM and SDS to 0.2% final. Hydrolysis of the adenylates was monitored by chromatography on PEI-TLC plates. The decrease in the adenylate fraction was plotted and analyzed by fitting to exponential decay equation as described in Experimental Procedures. Error bars represent mean ± S.D from three independent experiments.