

Discovery and Total Synthesis of a New Estrogen Receptor Heterodimerizing Actinopolymorphol A from *Actinopolymorpha rutilus*

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General experimental procedures. Optical rotations were measured in CH₃OH on a Perkin-Elmer 241 polarimeter at the sodium D line (589 nm). ¹H and ¹³C NMR spectra were recorded at 25°C on a Varian Unity Inova 500 instrument operating at 500 MHz for ¹H and 125 MHz for ¹³C nuclei. ¹H and ¹³C NMR chemical shifts were referenced to residual solvent signals. ¹H-¹H COSY, HMQC ($J_{CH}=140$ Hz), and gHMBC ($J_{XH}=8.0$ Hz) were performed using standard VARIAN pulse sequences. Electrospray ionization-mass (ESI-MS) spectra were acquired on an IonSpec HiResMALDI FT-Mass spectrometer with a 7 tesla superconducting magnet. Semipreparative HPLC was performed on a Varian liquid chromatograph system with an Atima-C18, 10 mm ×25 cm column. Silica gel chromatography applied to purification of specified synthetic compounds used 230-400 mesh size silica from Natland Int. Corp. Other chromatographic matrices included Lichroprep RP-18 gel (40–63 μm, Merck, Dramstadt, Germany) or Sephadex LH-20 (pharmacia) where indicated. Reactions were routinely run in flame-dried roundbottom flasks and under an atmosphere of Ar. All chemical reagents, unless otherwise noted, were obtained from Sigma-Aldrich and used as obtained directly from the manufacturer.

Fermentation and isolation. The strain *Actinopolymorpha rutilus* was obtained from Yunnan Institute of Microbiology, Yunnan University, Kunming, 650091, P. R. China. The inoculum was prepared by introducing the periphery of 10-day-old petri dish cultures of *A. rutilus* into 250 ml flasks containing 50 mL of the ISP2 broth, followed by shaking (250 rpm) continuously for 5 days at 28 ± 0.5°C. The follow-up fermentation was accomplished by adding the inoculum (50 mL) into 2-L flasks containing 500 mL of the culture broth [Sucrose (100 g), glucose (10.0 g), casamino acids (0.1 g), yeast extract (5.0 g), MOPS (21.0 g), trace elements (1 mL), K₂SO₄ (0.25g), MgCl₂ ·6H₂O (10.0 g), in a final volume of 1.0-L H₂O, pH 7.0], and then shaking for 7 days in the same condition. Solid phase extraction of the broth using resin (HP-20), filtration through cheesecloth, and elution of the resin with acetone afforded, after solvent removal under vacuum, a gummy extract was obtained. The extract (8.5 g) was chromatographed on silica gel column using CHCl₃-MeOH (100:0, 50:1, 20:1, 10:1, 5:1, 1:1 and 0:100, 1.50 L each) as the mobile phase to get seven fractions A-G. Fractions B and C (50:1 and 20:1, total 1.1g) were further chromatographed over RP-18 column eluted with a gradient (10% methanol, 30% methanol, 60% methanol, 90% methanol, 100% methanol) to yield five subfractions B1-B5. Subfractions B3 and B4 were combined and then subjected to Sephadex LH-20 column and eluted with methanol to yield three subfractions. These three subfractions were finally purified by semi-preparative HPLC to afford compounds **1** (1.5 mg), **2** (0.8 mg), and **3** (0.7 mg).

Physico-chemical properties of **1**, **2**, and **3**

Actinopolymorphol A (1): Colorless oil; $[\alpha]_D^{25} +20.9$ (c 0.15, CH₃OH) for natural product; $[\alpha]_D^{25} +23.5$ (c 0.28, CH₃OH) for synthesized; HR-ESIMS (positive ion) m/z 287.12567 (calcd for C₁₅H₂₀O₄Na [M+Na]⁺, 287.12538); ¹H NMR and ¹³C NMR, see Table S1.

Actinopolymorphol B (2): Colorless oil; $[\alpha]_D^{25} +7.8$ (c 0.10, CH₃OH); HR-ESIMS (positive ion) m/z 226.08290 (calcd for C₁₂H₁₃NO₂Na [M+Na]⁺, 226.08380); ¹H NMR and ¹³C NMR, see Table S1.

Actinopolymorphol C (3): Colorless oil; HR-ESIMS (positive ion) m/z 315.11250 (calcd for C₁₈H₁₆ N₂O₂Na [M+Na]⁺, 315.11039); ¹H NMR and ¹³C NMR, see Table S1.

Synthesis of **1** (*S*)-2-hydroxy-3-(4-hydroxyphenyl)-propionic acid (**4**)

(*S*)-3-(4-Hydroxyphenyl)-2-hydroxypropionyl methyl ester (5). (*S*)-3-(4-Hydroxyphenyl)-2-hydroxypropionic acid (1 g, 5.5 mmol) from Astatech, Inc. (Bristol, PA) was dissolved in 120 mL ACS grade MeOH at rt. The methanolic solution was then subjected to a constant stream of HCl gas (generated via dropwise addition of H₂SO₄ to solid NaCl) for a period of 2 h at rt followed by solvent removal *in vacuo* at 50°C. The resulting faint yellowish gum was then subjected to column chromatography over silica (2:1:0.4 Hex:EtOAc:MeOH) affording 1.08 g of methyl ester **5** as a thick transparent oil. All spectral data matched those previously reported.¹

(S)-3-(4-*O*-tert-butyldimethylsiloxyphenyl)-2-tert-butyldimethylsiloxy propionyl methyl ester (6). (S)-3-(4-Hydroxyphenyl)-2-hydroxypropionyl methyl ester (**5**) (4 g, 20.4 mmol) was dissolved in 50 mL anhydrous DMF and the solution chilled for 10 min to 0°C followed by addition of imidazole (4.16 g, 61.2 mmol). The solution was stirred at 0°C for 10 min and then TBSCl (9.22 g, 61.2 mmol) was added and the resulting mixture stirred at 0°C for 15 min. The reaction was then allowed to warm from 0°C to rt over the course of 4 h, and TLC (2:1:0.4 Hex:EtOAc:MeOH) revealed complete loss of starting material and generation of one UV active fast mobility spot. The reaction was then chilled back to 0°C and quenched by addition of cold dilute NH₄Cl (100 mL) and ~20 g NaCl. After 5 min the reaction was partitioned into an equal volume of EtOAc. Organic layer was separated and remaining aqueous phase extracted with EtOAc (3 x 50 mL). Organic layers were combined and washed with satd NH₄Cl (3 x 50) followed by washing with brine (2 x 100 mL). Organic layer was then dried over anhydrous Na₂SO₄ for 10 min. Solids were filtered off and solvent removed *in vacuo* to render 9.17 g crude **6**. Column chromatography over silica (2:1:0.4 Hex:EtOAc:MeOH) afforded 7.8 g pure **6** (90% yield). Colorless oil; HR-ESIMS (positive ion) *m/z* 447.23734 (calcd for C₂₂H₄₀ O₄Si₂Na [M+Na]⁺, 447.23573); ¹H NMR in CDCl₃: δ 6.75 (d, *J* = 8.4 Hz), 7.07 (d, *J* = 8.4 Hz), 3.01 (dd, *J* = 3.6, 12.6 Hz), 2.80 (dd, *J* = 9.2, 12.6 Hz), 4.28 (dd, *J* = 3.6, 9.2 Hz), 3.72 (OMe, s), -0.20 (s), -0.13 (s), 0.17 (s), 0.80 (s), and 0.97 (s); ¹³C NMR: δ_c 130.8 (s, C-1), 131.2 (d, C-2, 6), 120.5 (d, C-3, 5), 154.9 (s, C-4), 41.4 (t, C-7), 74.6 (d, C-8), 174.3 (s, C-9), 52.3 (q, OMe), 26.2, 26.1, 18.8, -3.9, -5.1.

(S)-3-(4-*O*-tert-butyldimethylsiloxyphenyl)-2-tert-butyldimethylsiloxy N-methoxy, N-methyl propionamide (7). To methyl ester **6** (390 mg, 0.92 mmol) was added 3 mL anhydrous THF in a flame dried roundbottom flask under Ar atmosphere. The solution was chilled to 0°C for 10 min followed by the addition of *N,O*-dimethylhydroxylamine hydrochloride (140 mg, 1.44 mmol). The slurry was then chilled to -20°C and 1.38 mL (2.8 mmol) of (CH₃)₂CHMgCl (2M in THF) added dropwise. The mixture was stirred at -20°C for 1 h followed by quenching via addition of satd NH₄Cl (10 mL) and partitioning into an equal volume of EtOAc. Aqueous layer was extracted with EtOAc (3 x 50 mL) and then organic fractions pooled and washed with saturated NH₄Cl (3 x 50 mL) and then brine (2 x 50 mL). Organic layer was dried by addition of anhydrous Na₂SO₄ for 10 min. Solids were then filtered off and solvent removed *in vacuo*. The resulting gum was subjected to column chromatography (10:1:1 Hex:EtOAc:MeOH) to afford 340 mg of **7** (81% yield). Colorless oil; HR-ESIMS (positive ion) *m/z* 476.26342 (calcd for C₂₃H₄₃ NO₄Si₂Na [M+Na]⁺, 476.26228); ¹H NMR in CDCl₃: δ_H 6.74 (d, *J* = 8.4 Hz), 7.07 (d, *J* = 8.4 Hz), 2.95 (dd, *J* = 4.4, 13.2 Hz), 2.74 (dd, *J* = 8.8, 13.2 Hz), 4.61 (dd, *J* = 4.4, 8.8 Hz), 3.61 (OMe, s), 3.29 (NMe), -0.16 (s), -0.13 (s), 0.15 (s), 0.78 (s), and 0.97 (s); ¹³C NMR: δ_c 131.2 (s, C-1), 131.0 (d, C-2, 6), 120.2 (d, C-3, 5), 154.6 (s, C-4), 32.8 (t, C-7), 72.1 (d, C-8), 174.3 (s, C-9), 61.4 (q, OMe), 40.7 (q, NMe), 26.0, 26.1, 18.6, -4.8.

(S)-8-(*O*-tert-butyldimethylsiloxy)-4-*O*-tert-butyldimethylsiloxy satabacin (8). To Weinreb amide **7** (270 mg, 0.6 mmol) was added 4 mL anhydrous THF in a flame dried roundbottom flask under Ar atmosphere. The solution was chilled to 0°C for 10 min followed by the addition of 0.7 mL 2M (CH₃)₂CHCH₂MgCl in THF (1.4 mmol) at 0°C. Reaction was stirred at 0°C for 2 h and then allowed to warm to rt; reaction was stirred at rt for 24h. Following incubation at rt, the reaction was chilled back to 0°C and then quenched via addition of cold 2N HCl (pH ~ 3) (15 mL) and partitioned into an equal volume of EtOAc. Aqueous layer was extracted with EtOAc (3 x 25 mL) and then organic fractions pooled and washed with saturated NH₄Cl (3 x 50 mL) and then brine (2 x 50 mL). Organic layer was dried by addition of anhydrous Na₂SO₄ for 10 min. Solids were then filtered off and solvent removed *in vacuo*. The resulting gum (310 mg crude) was subjected to column chromatography (15:1 Hex:EtOAc) to afford 180 mg of **8** (67% yield). Colorless oil; HR-ESIMS (positive ion) *m/z* 473.28975 (calcd for C₂₅H₄₆ O₃Si₂Na [M+Na]⁺, 473.28777); ¹H NMR in CDCl₃: δ_H 6.76 (d, *J* = 8.4 Hz), 7.05 (d, *J* = 8.4 Hz), 2.84 (dd, *J* = 4.4, 13.6 Hz), 2.69 (dd, *J* = 8.8, 13.6 Hz), 4.10 (dd, *J* = 4.4, 8.8 Hz), 2.31-2.45 (m), 2.14 (m), 0.90 (d, *J* = 6.8 Hz), 0.88 (d, *J* = 6.8 Hz), -0.30 (s), -0.11 (s), 0.17 (s), 0.85 (s), 0.89 (s); ¹³C NMR: δ_c 130.4 (s, C-1), 131.3 (d, C-2, 6), 120.3 (d, C-3, 5), 154.8 (s, C-4), 40.9 (t, C-7), 80.8 (d, C-8), 213.1 (s, C-9), 47.0 (t, C-10), 23.7 (d, C-11), 23.1 (q, C-12), 23.1 (q, C-13), 26.2, 26.1, 18.6, -5.1.

(S)-4-hydroxysattabacin (9). Diprotected ketone **8** (350 mg, 0.78 mmol) was dissolved in 8.5 mL anhydrous THF at 0°C. After 10 min 2.3 mL of tetrabutylammonium fluoride (TBAF) (1M in THF) (2.33 mmol) was added dropwise at 0°C. Reaction was stirred at 0°C for 45 min at which time TLC analysis (15:1 Hex:EtOAc) revealed surprisingly little change to starting material. The reaction was then allowed to warm to rt over the course of 1.5 h after which time it was apparent by TLC that starting material was consumed. The reaction was chilled back to 0°C and quenched via addition of cold 2N HCl (pH ~3) (15 mL) and partitioned into an equal volume of EtOAc. Aqueous layer was extracted with EtOAc (3 x 25 mL) and then organic fractions pooled and washed with satd NH₄Cl (3 x 50 mL) and then brine (2 x 50 mL). Organic layer was dried by addition of anhydrous Na₂SO₄ for 10 min. Solids were then filtered off and solvent removed *in vacuo*. The resulting 150 mg of crude diol was subjected to column chromatography (3:1 Hex:EtOAc) to afford 140 mg of **9** (81% yield). Colorless oil; ¹H NMR in CDCl₃: δ_H 6.66 (d, *J* = 8.4 Hz), 7.03 (d, *J* = 8.4 Hz), 3.06 (dd, *J* = 4.4, 14.4 Hz), 2.74 (dd, *J* = 7.6, 14.4 Hz), 4.36 (m), 2.39 (2H, d, *J* = 7.2 Hz), 2.18 (m), 0.92 (d, *J* = 6.8 Hz), 0.92 (d, *J* = 6.8 Hz), 6.18 (OH), 3.67 (OH); ¹³C NMR: δ_C 128.2 (s, C-1), 130.6 (d, C-2, 6), 115.7 (d, C-3, 5), 155.0 (s, C-4), 39.3 (t, C-7), 77.9 (d, C-8), 211.9 (s, C-9), 47.7 (t, C-10), 24.9 (d, C-11), 22.9 (q, C-12), 22.8 (q, C-13).

(S)-4-acetoxy-8-acetylsattabacin (10). Diol **9** (110 mg, 0.5 mmol) was dissolved in 2 mL anhydrous CH₂Cl₂ and chilled to 0°C for 10 min. To the chilled solution of **9** was added 500 mL of a premixed 1:1 combination of acetic anhydride and pyridine dropwise over a 5 min duration. The reaction was then allowed to warm to rt and to stir overnight. The reaction was quenched via addition of 3 mL MeOH and 3 mL EtOAc at 0°C. The solvents were removed *in vacuo* with gentle heating (~45°C) and the crude syrup subjected to immediate column chromatography (3:1 Hex:EtOAc) to afford 110 mg of diacetate **10** (72% yield). Colorless oil; [α]_D²³ +25.1 (*c* 0.18, CH₃OH); HR-ESIMS (positive ion) *m/z* 329.13618 (calcd for C₁₇H₂₂ O₅Na [M+Na]⁺, 329.13595); ¹H NMR in CDCl₃: δ_H 7.00 (d, *J* = 8.4 Hz), 7.19 (d, *J* = 8.4 Hz), 3.05 (dd, *J* = 4.4, 14.0 Hz), 2.93 (dd, *J* = 8.8, 14.0 Hz), 5.13 (dd, *J* = 4.4, 8.8), 2.32 (dd, *J* = 6.4, 17.2 Hz), 2.17 (dd, *J* = 6.8, 17.2 Hz), 2.11 (m), 0.85 (d, *J* = 6.5 Hz), 0.86 (d, *J* = 6.5 Hz), 2.25 (OAc), 2.05 (OAc); ¹³C NMR: δ_C 133.9 (s, C-1), 130.6 (d, C-2, 6), 121.9 (d, C-3, 5), 149.5 (s, C-4), 36.2 (t, C-7), 79.1 (d, C-8), 206.8 (s, C-9), 48.4 (t, C-10), 24.0 (d, C-11), 22.8 (q, C-12), 22.8 (q, C-13), 170.6 (OAc), 169.7 (OAc), 21.4 (OAc), 20.9 (OAc).

Actinopolymorphol A (1). Diacetate **10** (70 mg, 0.23 mmol) was dissolved in 1 mL pyrrolidine at rt and stirred for 1 min. The reaction was then diluted with 2 mL cold EtOAc and transferred immediately to a rapidly stirring solution of 10 mL 1:1 2N HCl:satd NH₄Cl at 0°C. The mixture was then partitioned into 25 mL EtOAc. The layers were separated and remaining aqueous layer was extracted with EtOAc (3 x 25 mL) and then organic fractions pooled and washed with satd NH₄Cl (3 x 50 mL) and then brine (2 x 50 mL). Organic layer was dried by addition of anhydrous Na₂SO₄ for 10 min. Solids were then filtered off and solvent removed *in vacuo* to afford 40 mg of monoacetate **1** in > 95 % purity based on ¹H-NMR (66% yield). Colorless oil; [α]_D²³ +20.9 (*c* 0.15, CH₃OH); HR-ESIMS (positive ion) *m/z* 287.12567 (calcd for C₁₅H₂₀O₄Na [M+Na]⁺, 287.12538); ¹H NMR and ¹³C NMR data see Table S1. See Figure 1B in manuscript for key HMBC and COSY correlations.

Table S1. Summary of NMR data for compounds **1**, **2**, and **3** in methanol-*d*₄.

No.	1		2		3	
	¹³ C	¹ H (<i>J</i> in Hz)	¹³ C	¹ H (<i>J</i> in Hz)	¹³ C	¹ H (<i>J</i> in Hz)
1	126.9				129.8	
1a			137.2			
2	130.3	7.02 (d, 8.4)	123.8	7.14 (s)	130.5	7.06 (d, 9.0)
3	115.1	6.68 (d, 8.4)	117.4		116.0	6.67 (d, 9.0)
4	156.4		118.6	7.60 (d, 9.0)	156.5	
4a			128.1			
5	115.1	6.68 (d, 8.4)	118.9	7.03 (dt, 0.8, 9.0)	116.0	6.67 (d, 9.0)
6	130.3	7.02 (d, 8.4)	121.5	7.11 (dt, 0.8, 9.0)	130.5	7.06 (d, 9.0)
7	35.7	2.98 (dd, 5.0, 14.4)	111.4	7.35 (d, 9.0)	40.2	3.96 (s)
8	79.6	5.08 (dd, 5.0, 8.2)	30.0	3.21 (dd, 5.5, 15.0) 3.09 (dd, 7.0, 15.0)	154.7	
9	207.8		77.9	4.39 (dd, 5.5, 7.0)	143.9	8.44 (s)
10	47.8	2.35 (dd, 6.6, 17.4) 2.16 (dd, 6.6, 17.4)	212.7			
11	43.7	2.03 (m)	25.5	2.14 (s)		
12	21.7	0.82 (d, 6.8)				
13	21.7	0.84 (d, 6.8)				
OAc	170.9					
OAc	19.3	2.04 (s)				

References:

Andrus, M. B.; Hicken, E. J.; Stephens, J.C.; Bedke, D. K. *J. Org. Chem.* **2006**, *71*, 8651-8654.

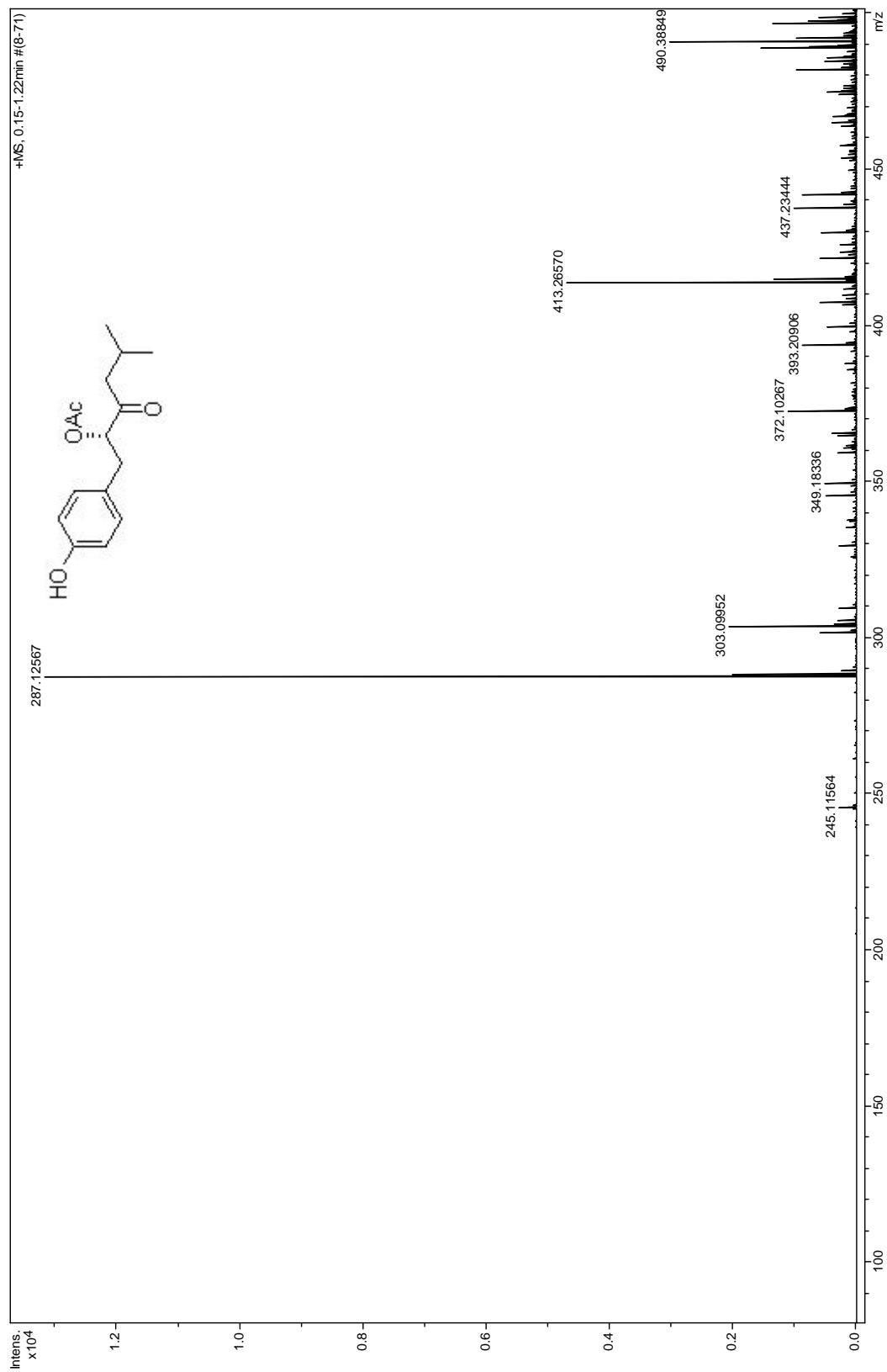


Figure S1. HRESIMS of **1**

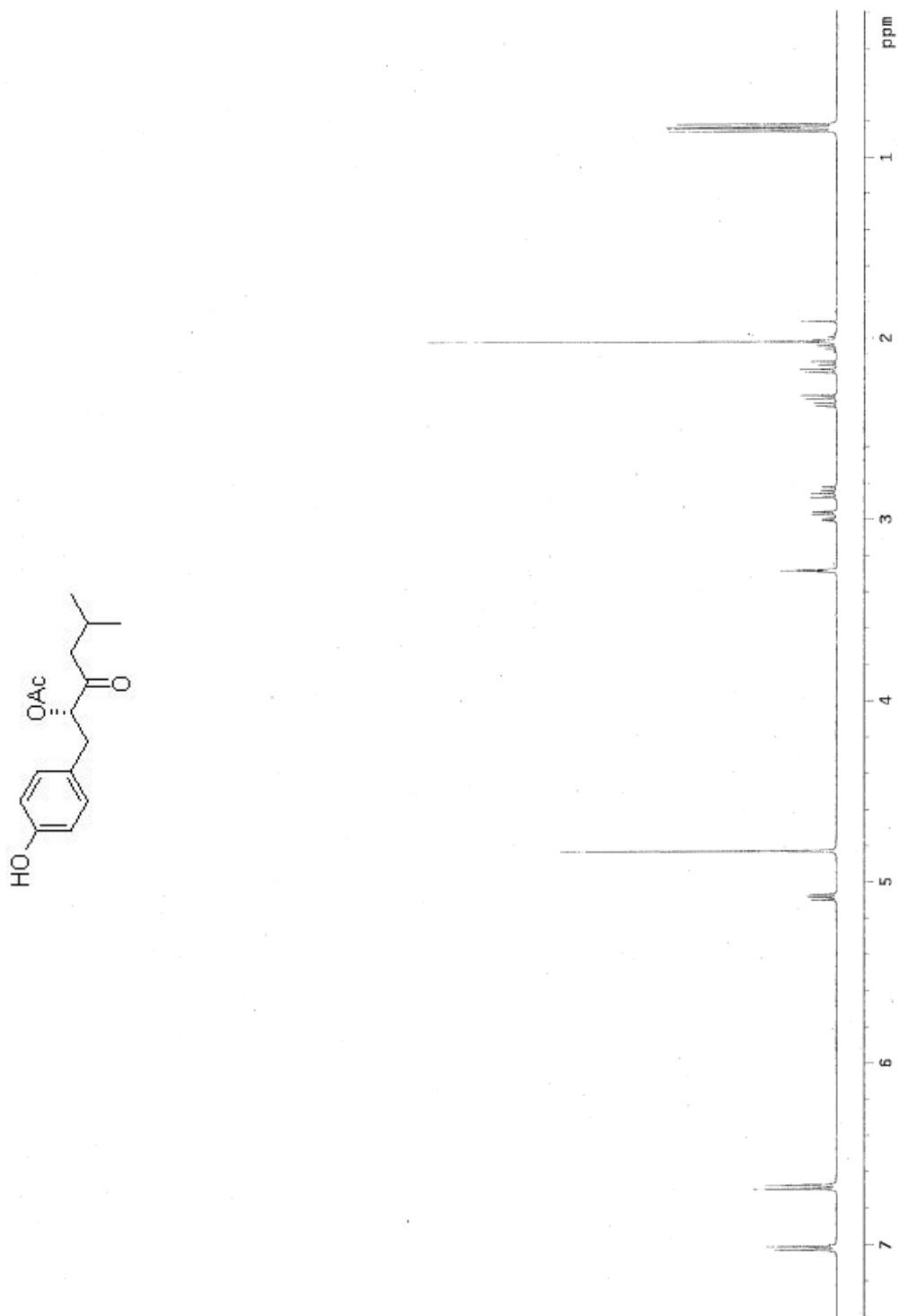


Figure S2. ¹H NMR spectrum of **1**

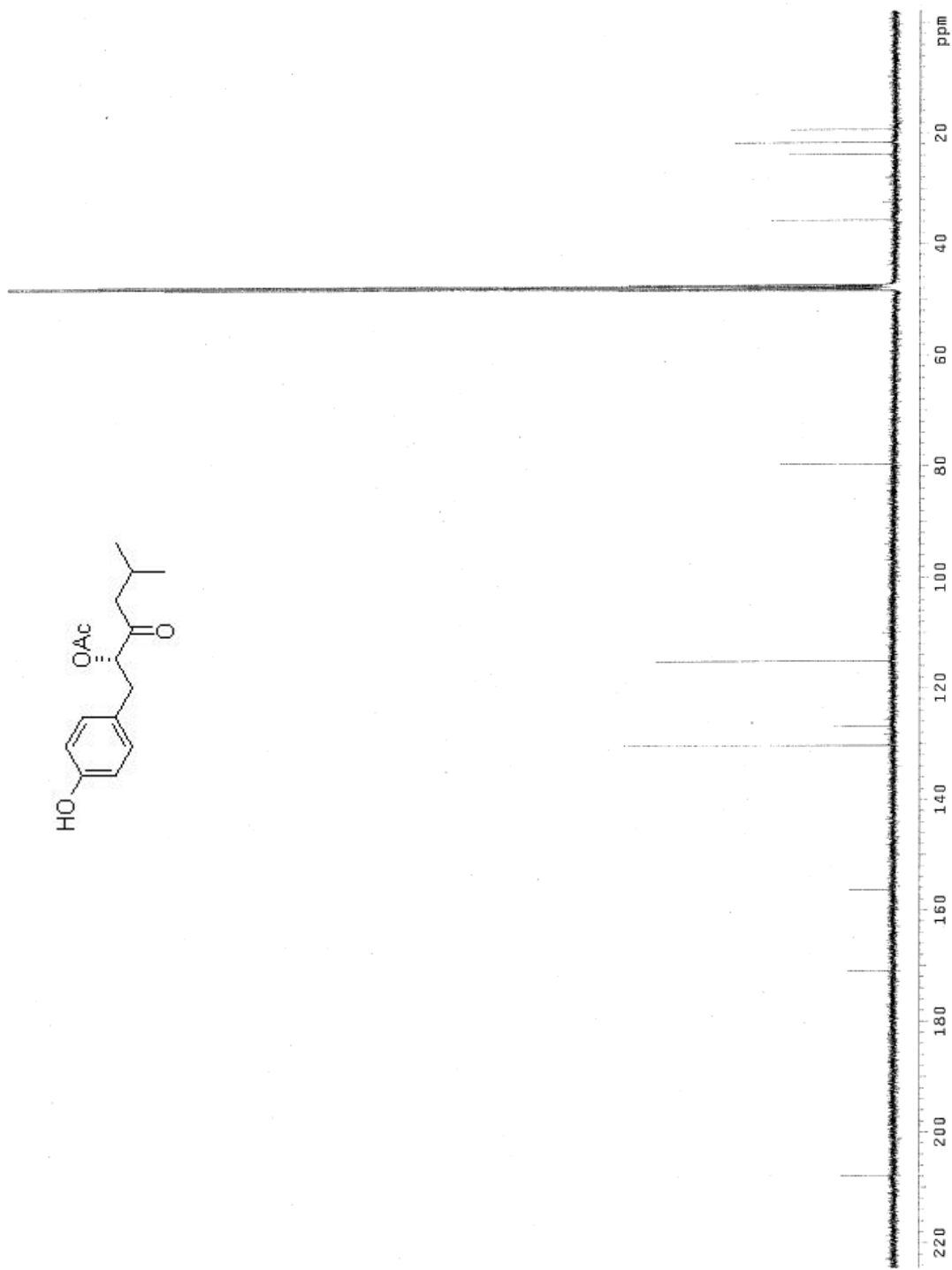


Figure S3. ^{13}C NMR spectrum of **1**

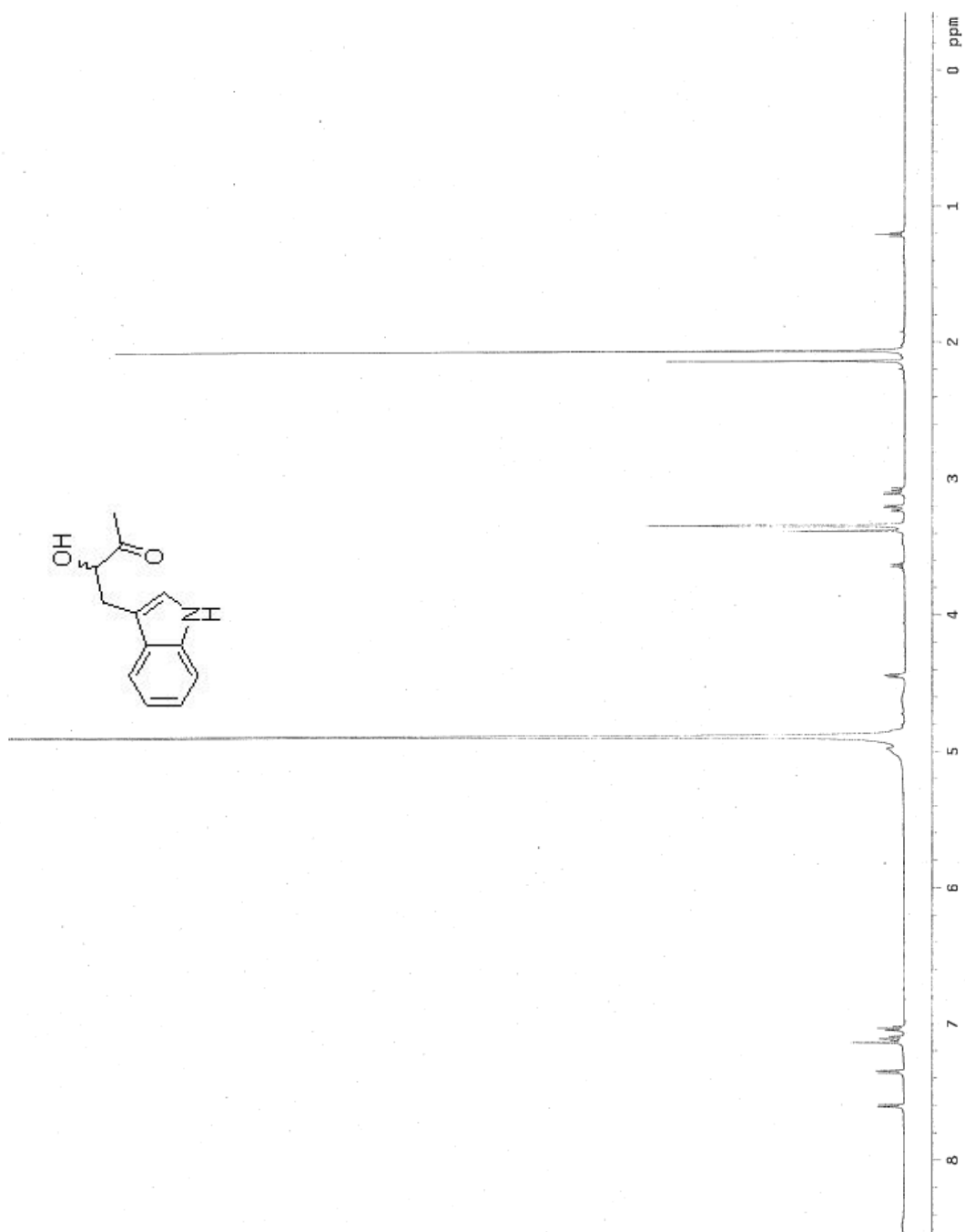


Figure S4. ¹H NMR spectrum of **2**

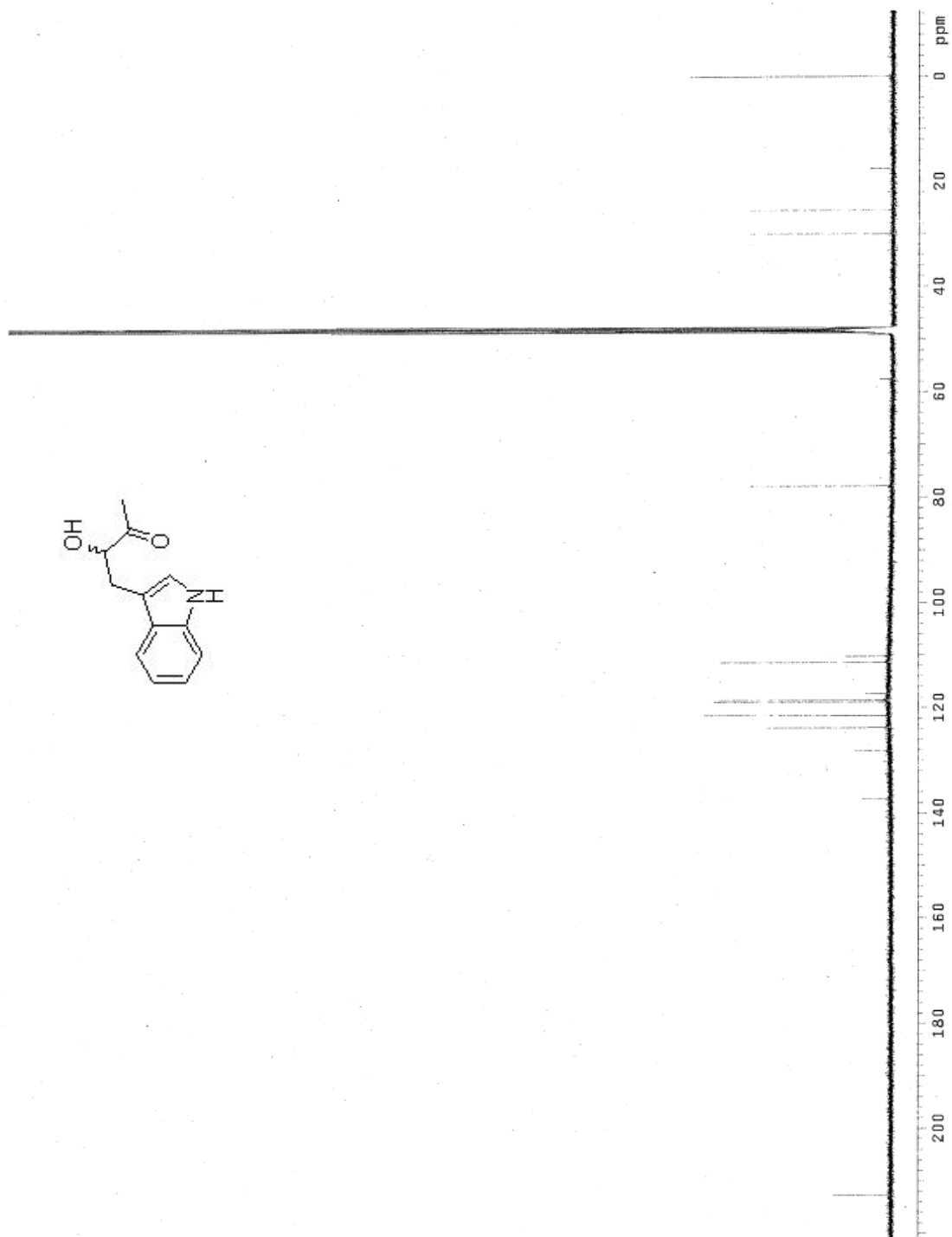


Figure S5. ^{13}C NMR spectrum of **2**

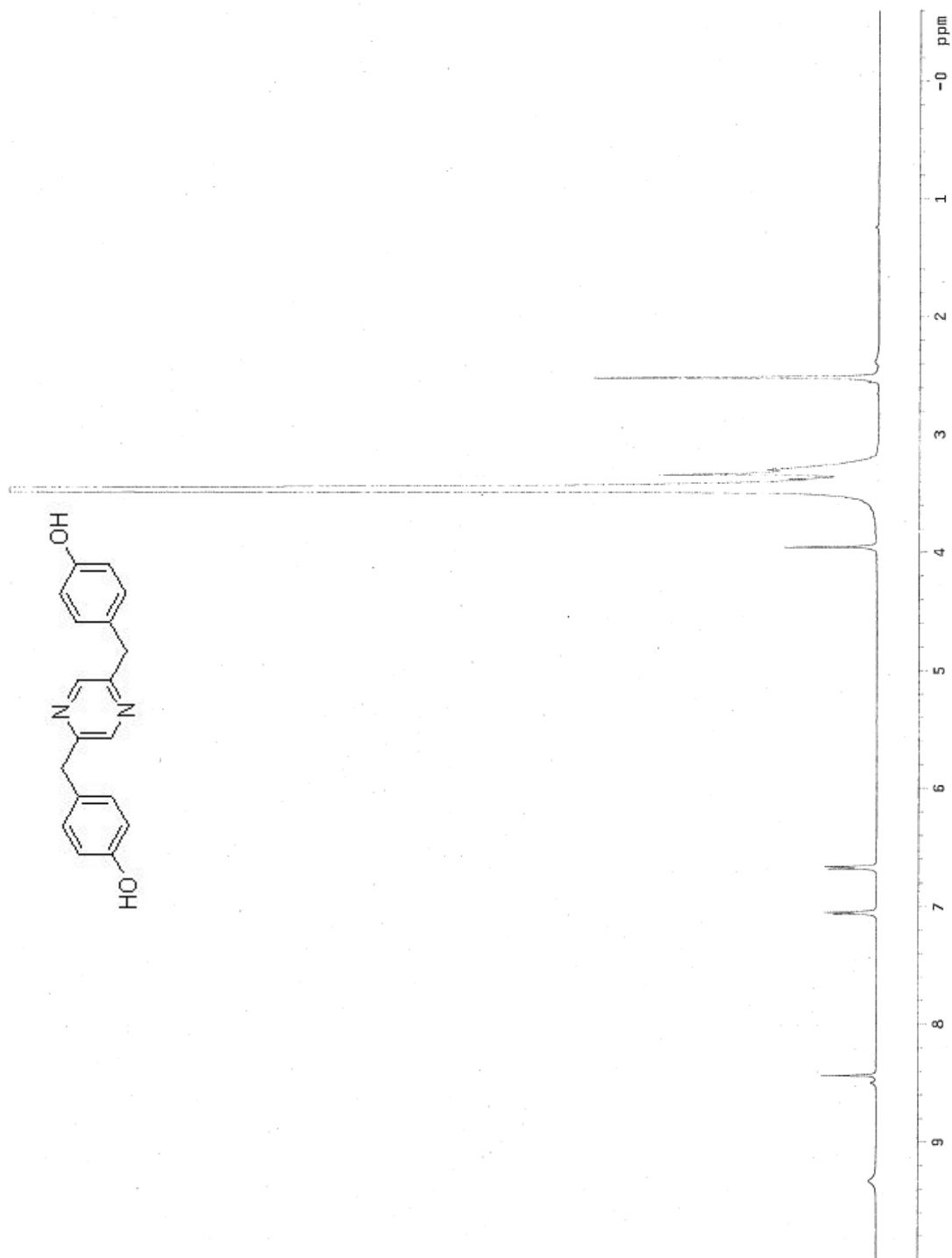


Figure S6. ^1H NMR spectrum of **3**

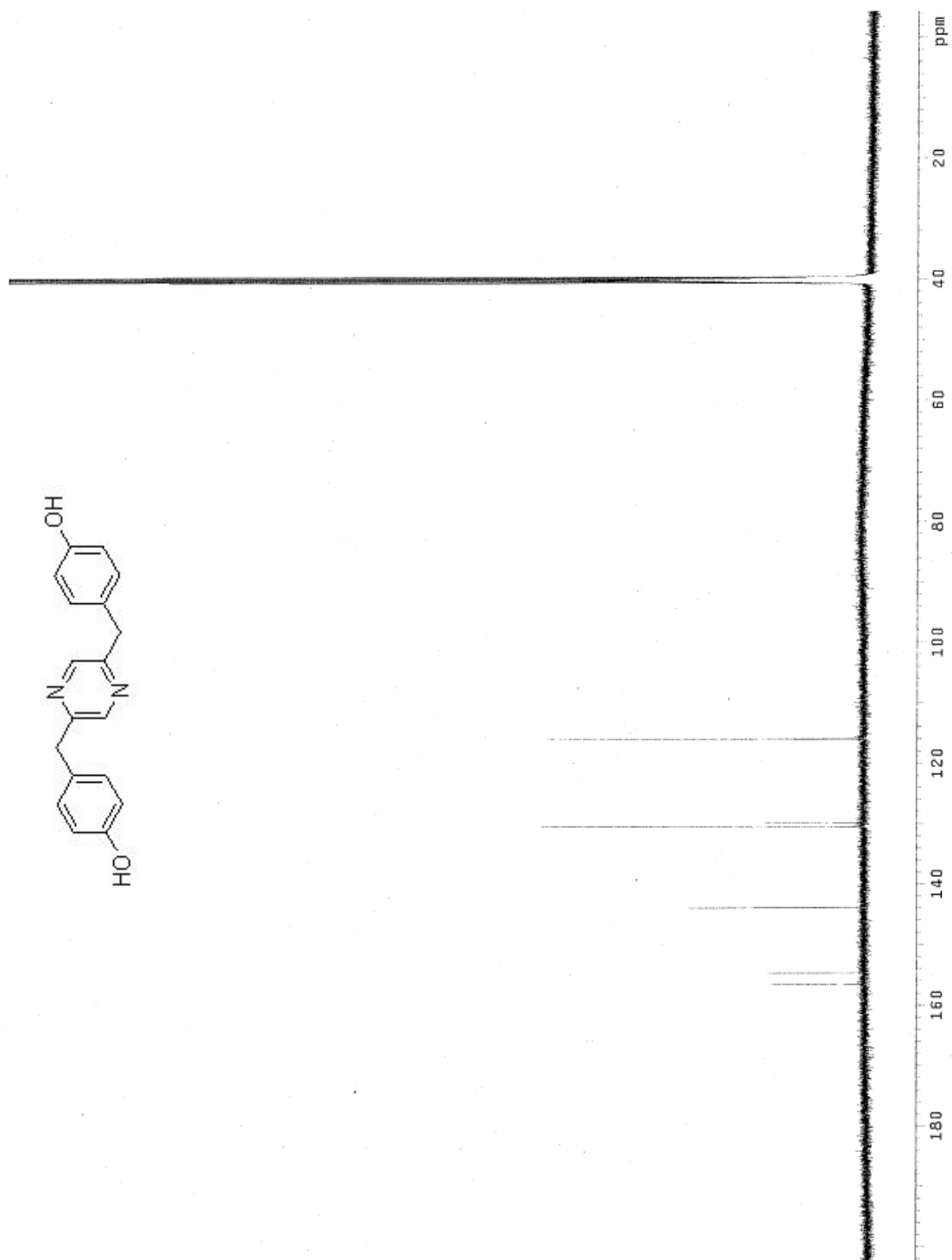


Figure S7. ^{13}C NMR spectrum of **3**

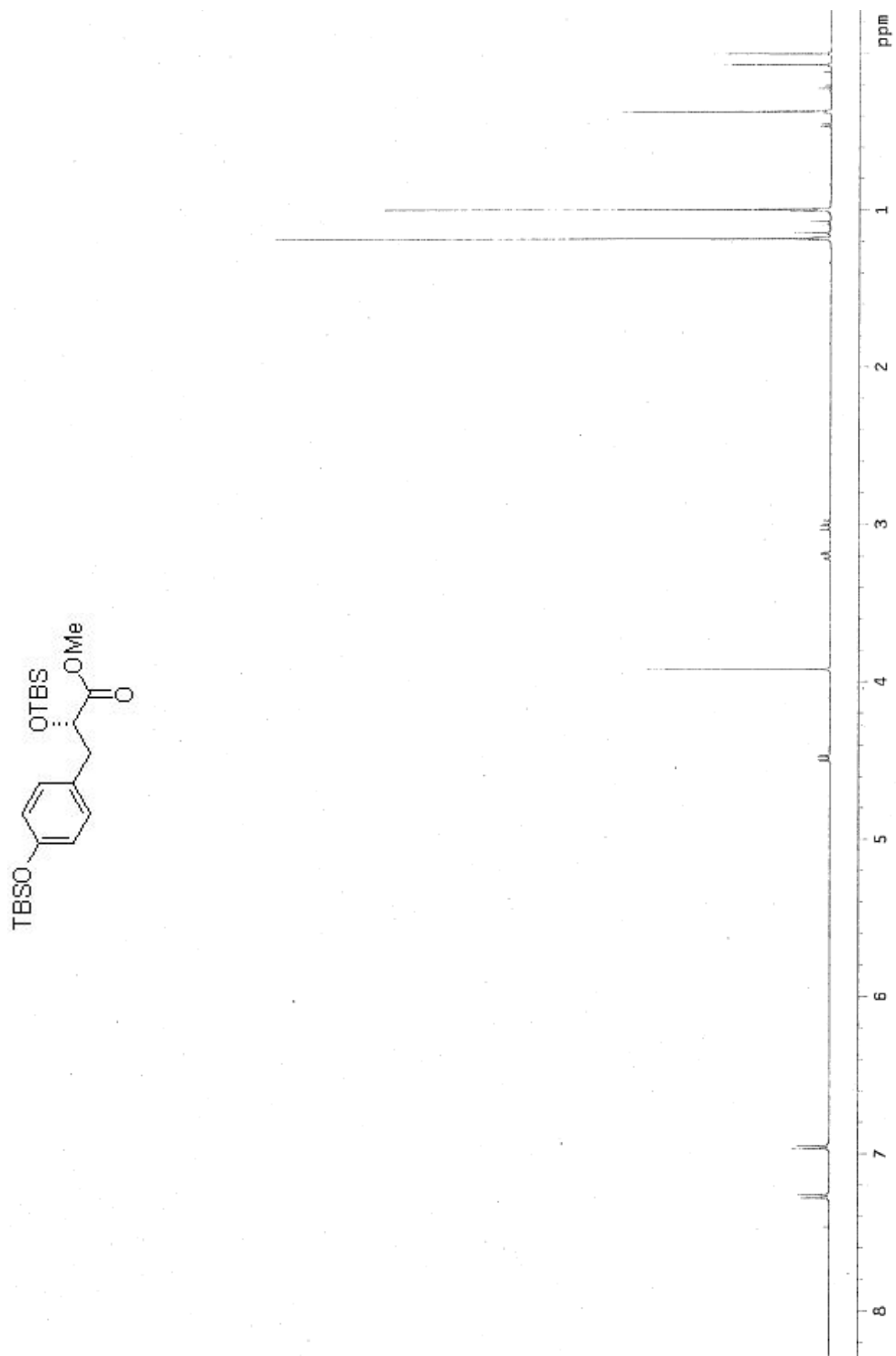


Figure S8. ¹H NMR spectrum of **6**

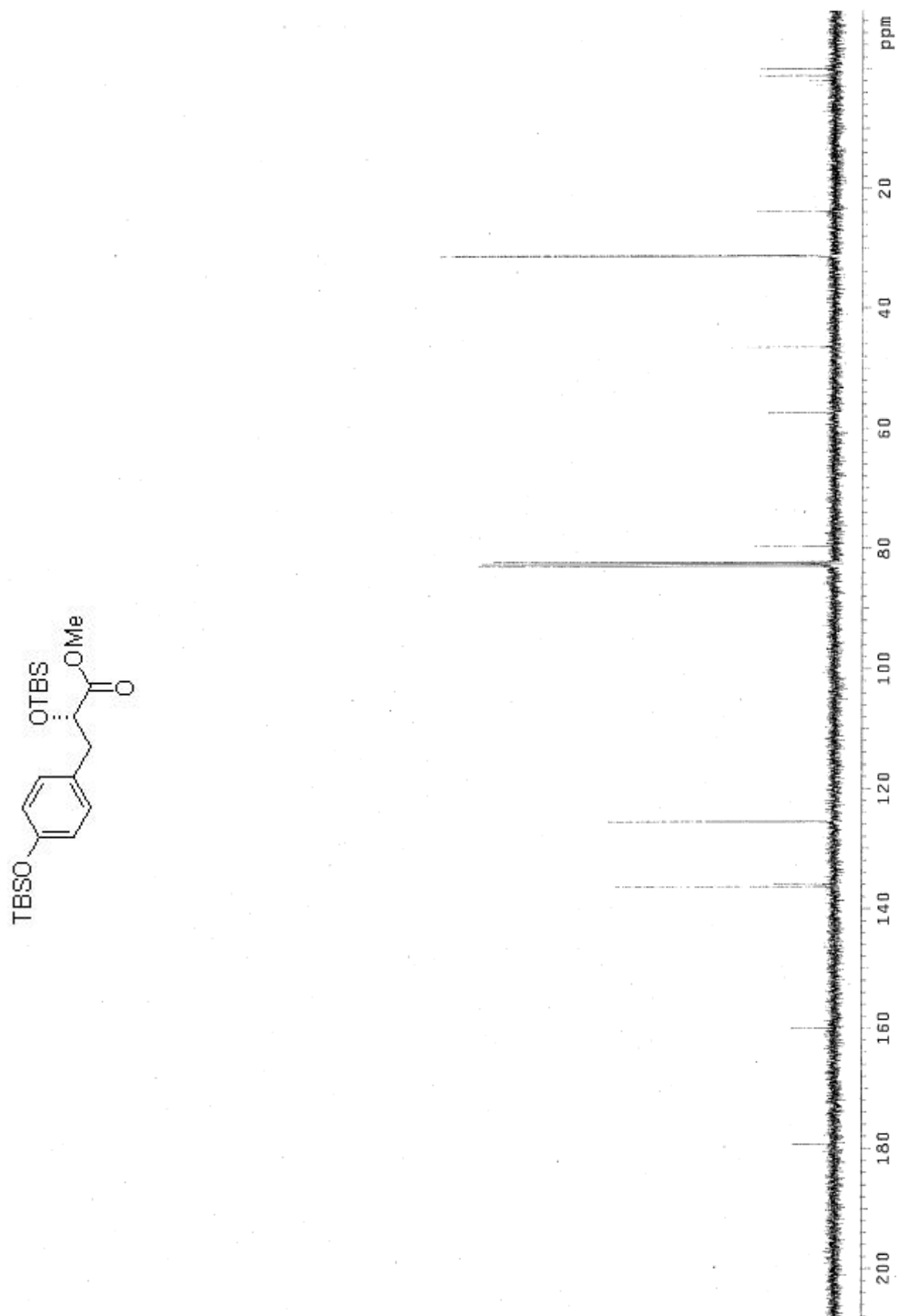


Figure S9. ^{13}C NMR spectrum of 6

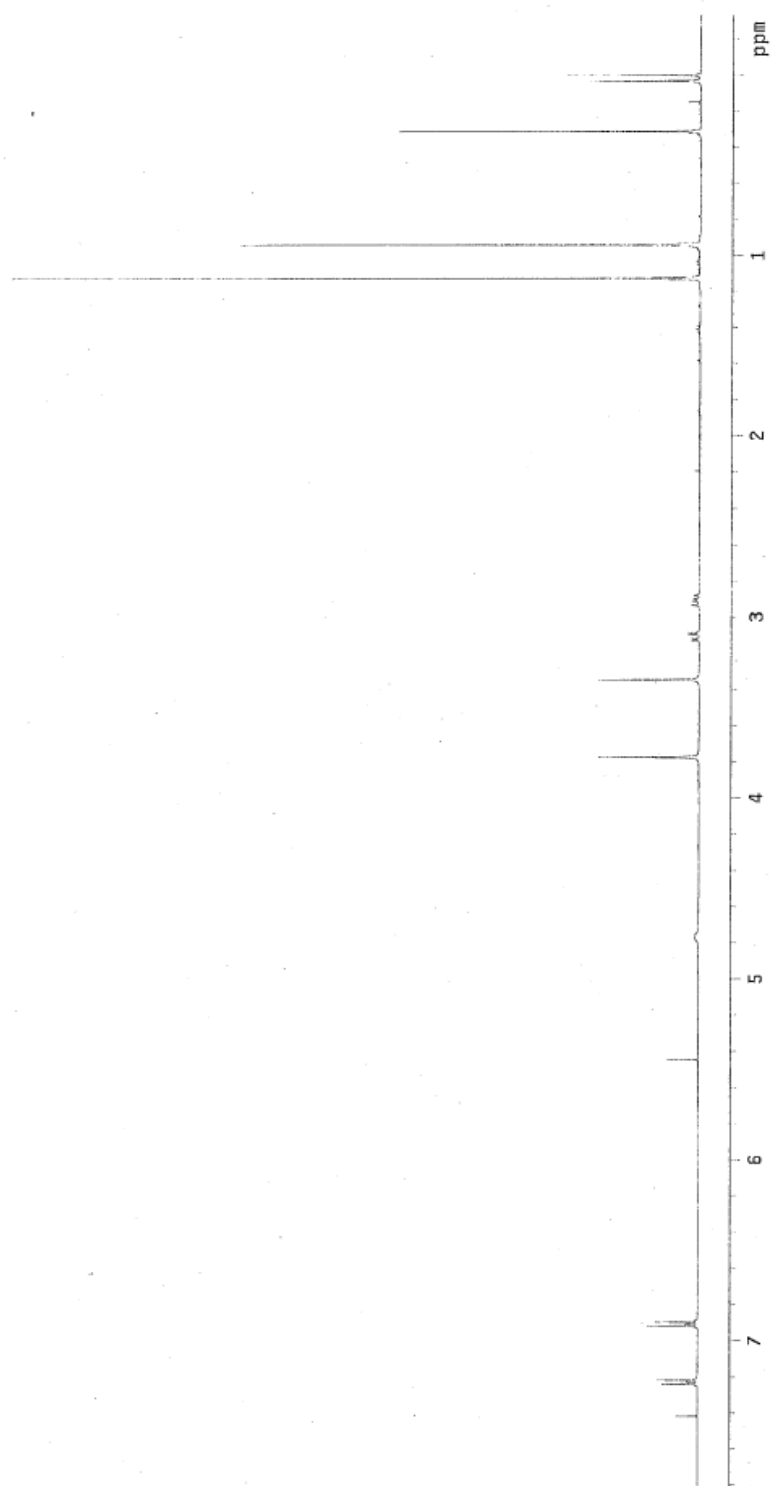
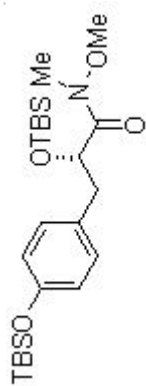


Figure S10. ^1H NMR spectrum of 7

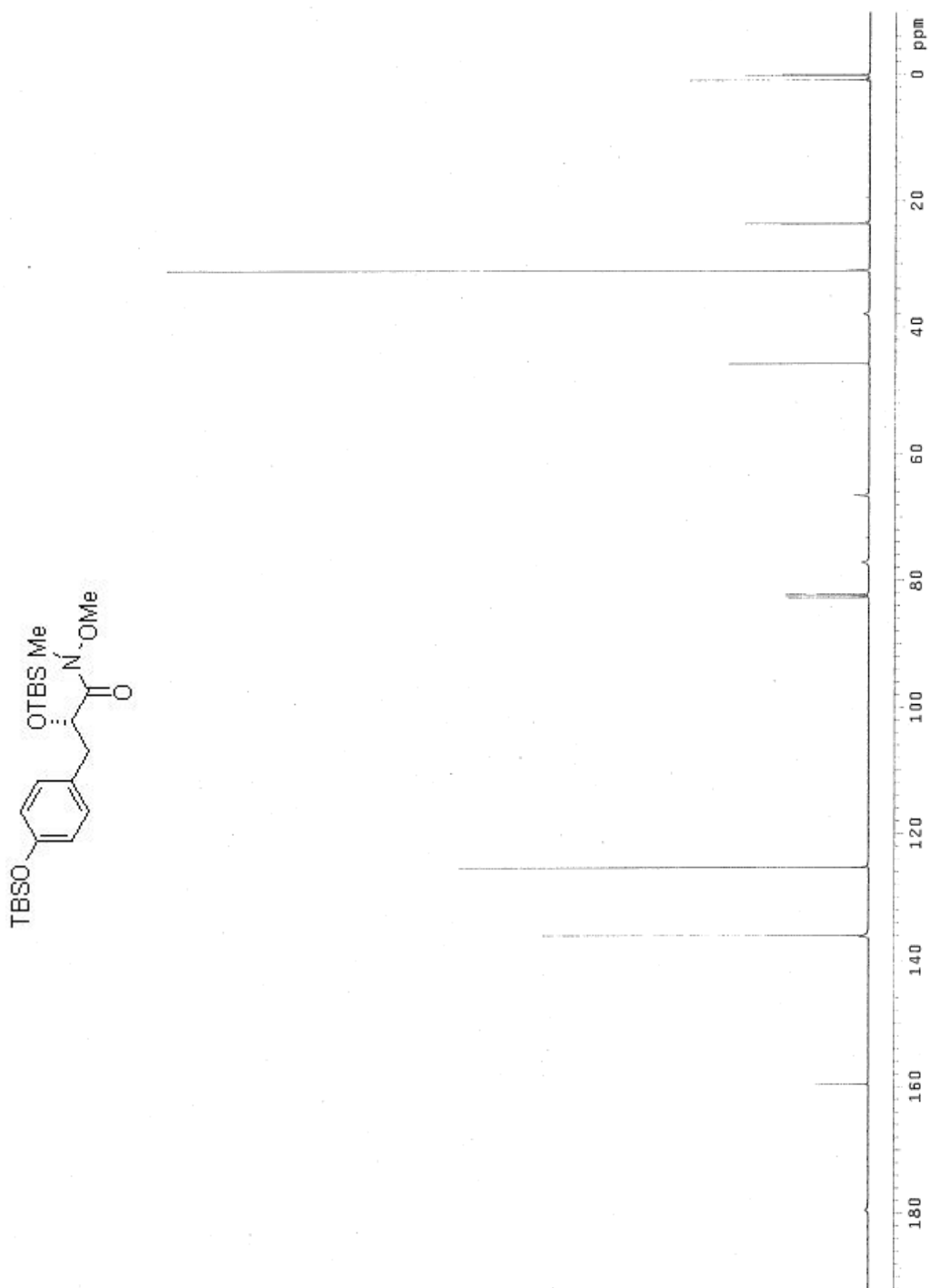


Figure S11. ^{13}C NMR spectrum of **7**

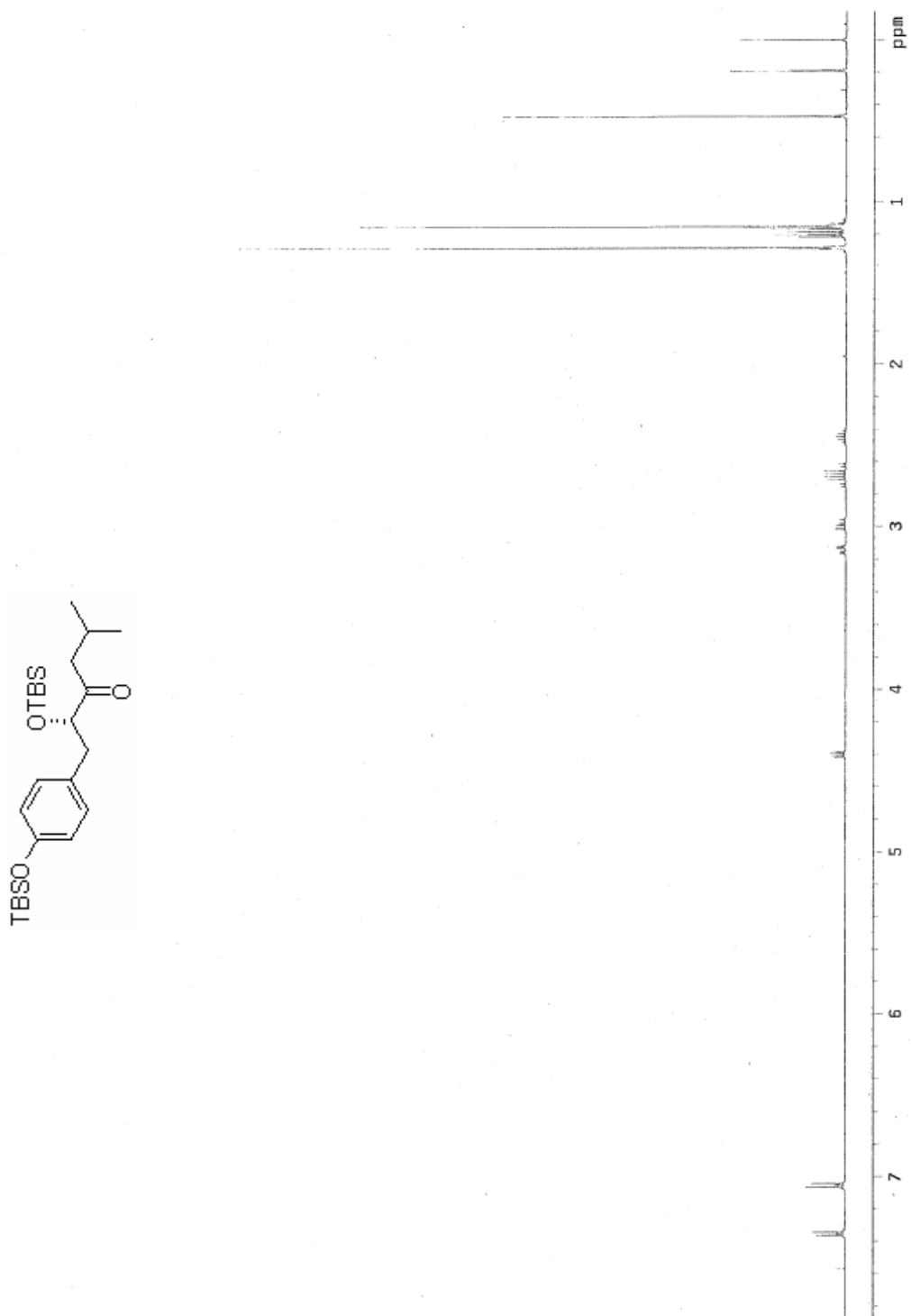


Figure S12. ¹H NMR spectrum of **8**

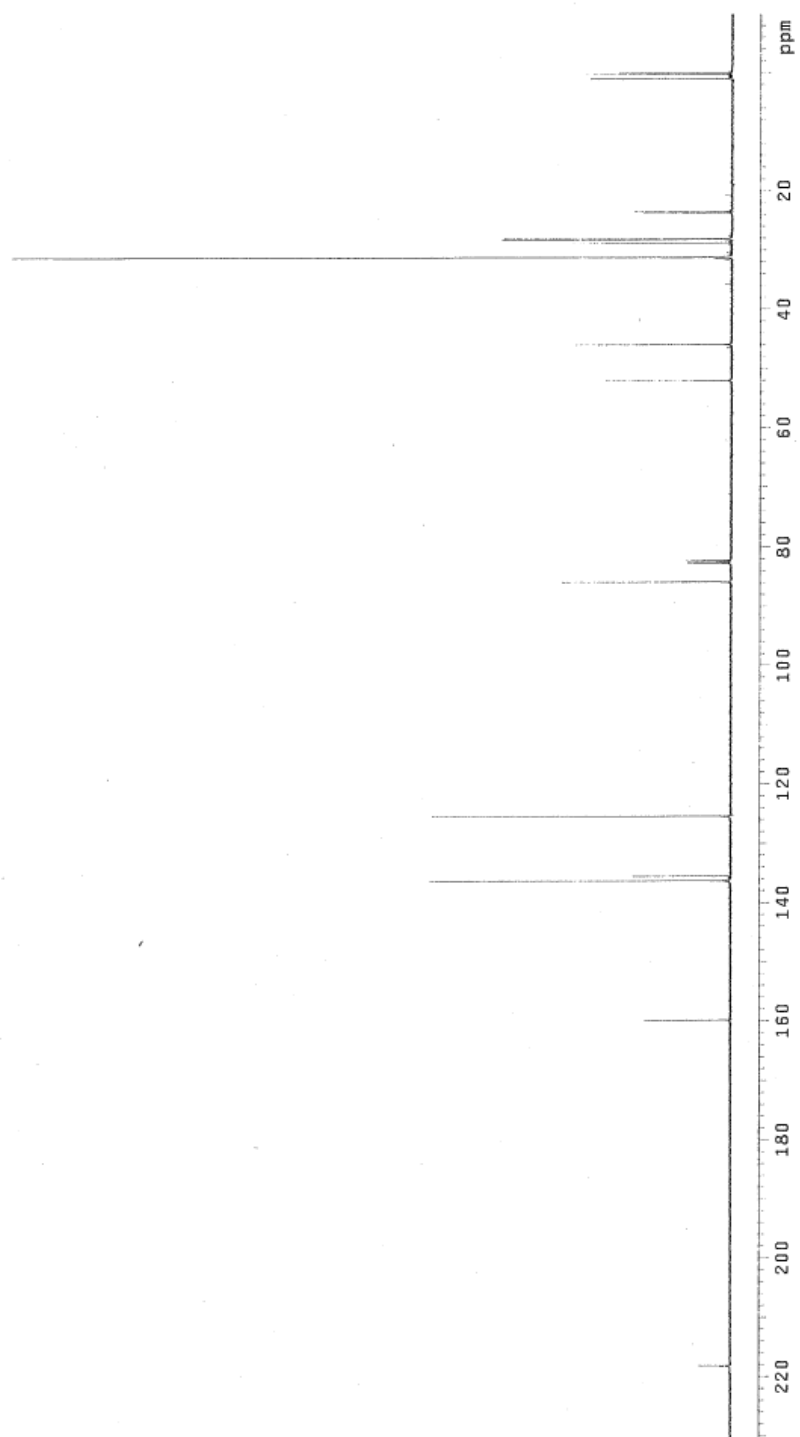
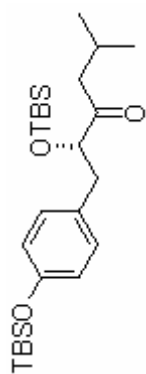


Figure S13. ^{13}C NMR spectrum of **8**

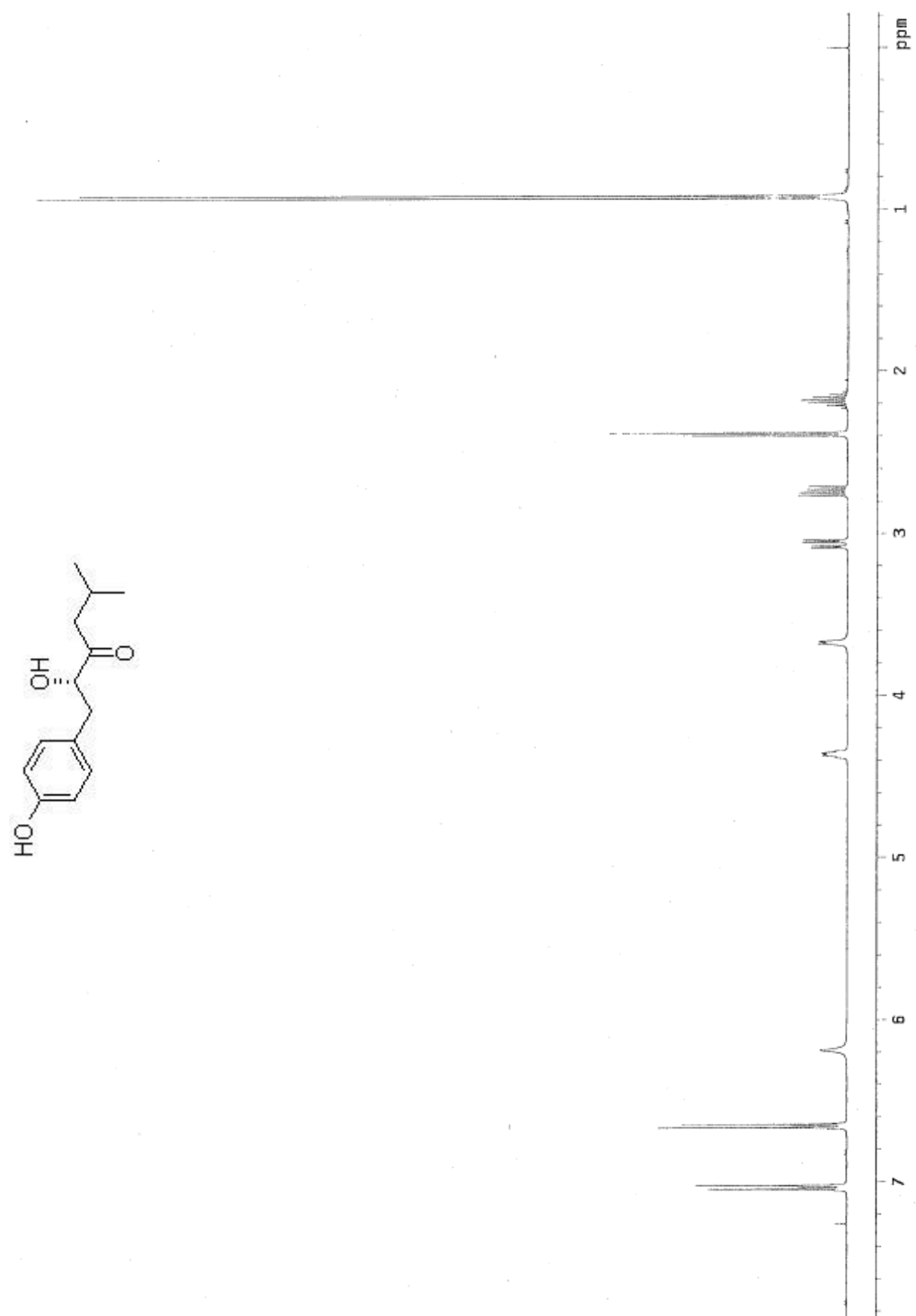


Figure S14. ¹H NMR spectrum of **9**

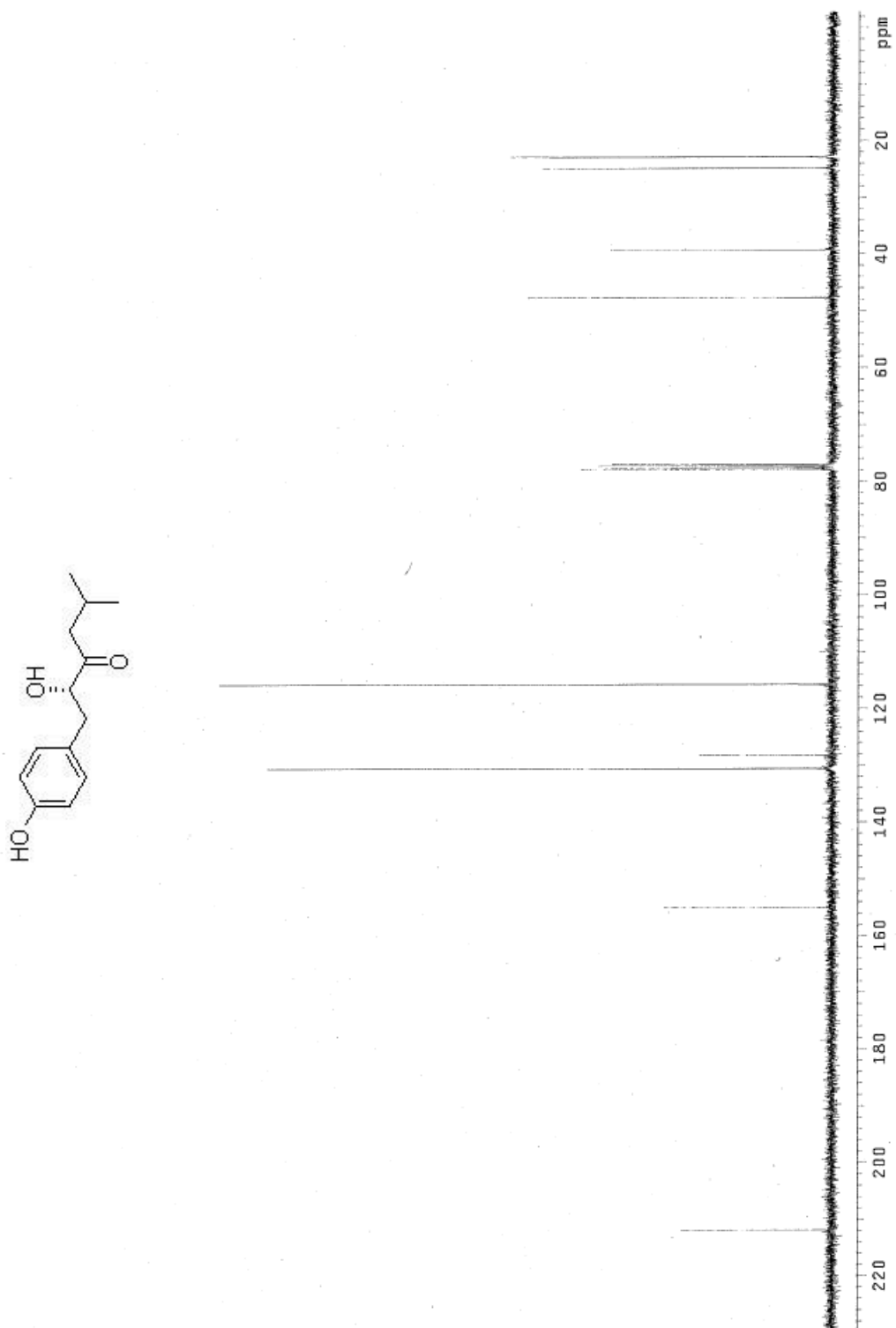


Figure S15. ^{13}C NMR spectrum of **9**

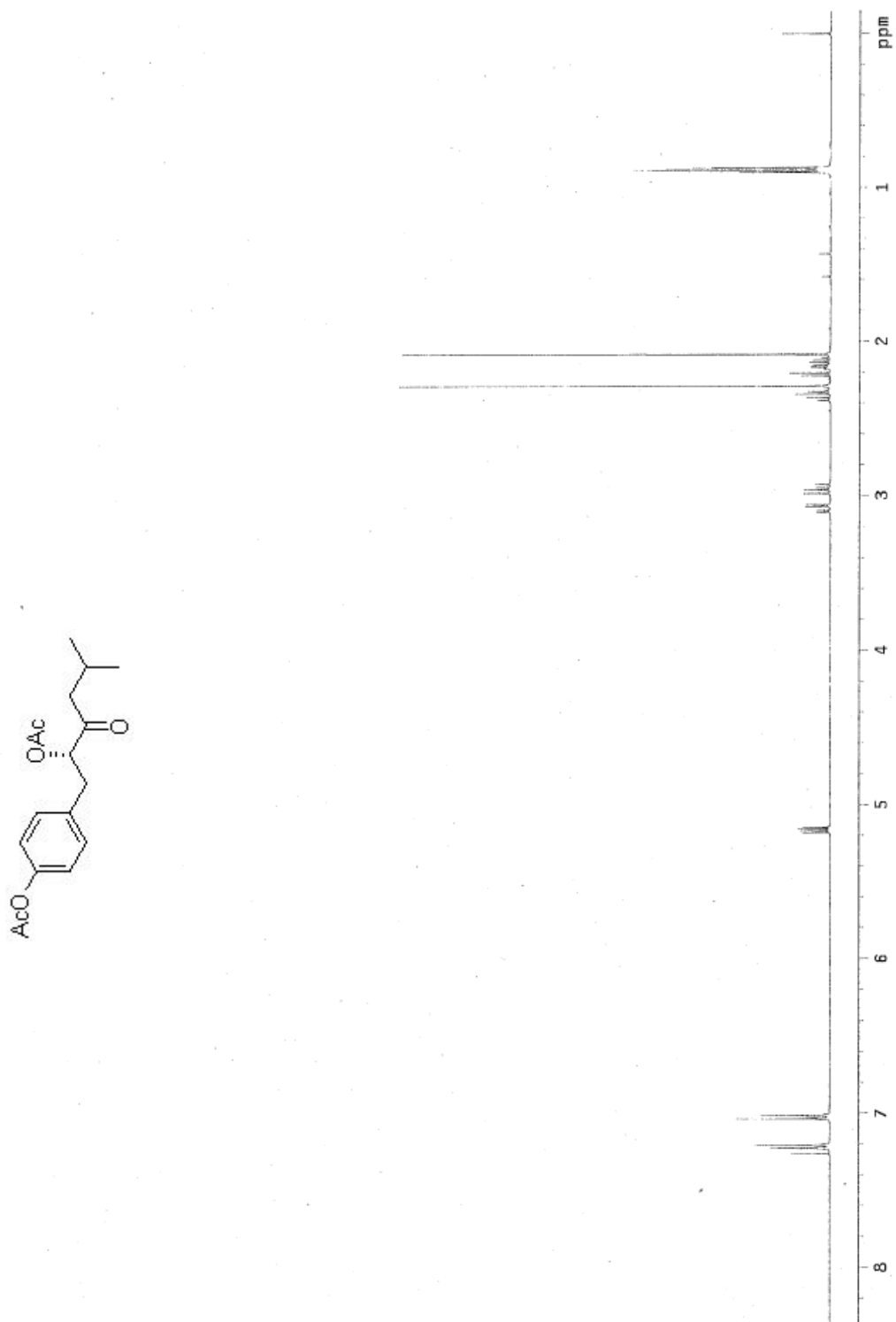


Figure S16. ¹H NMR spectrum of **10**

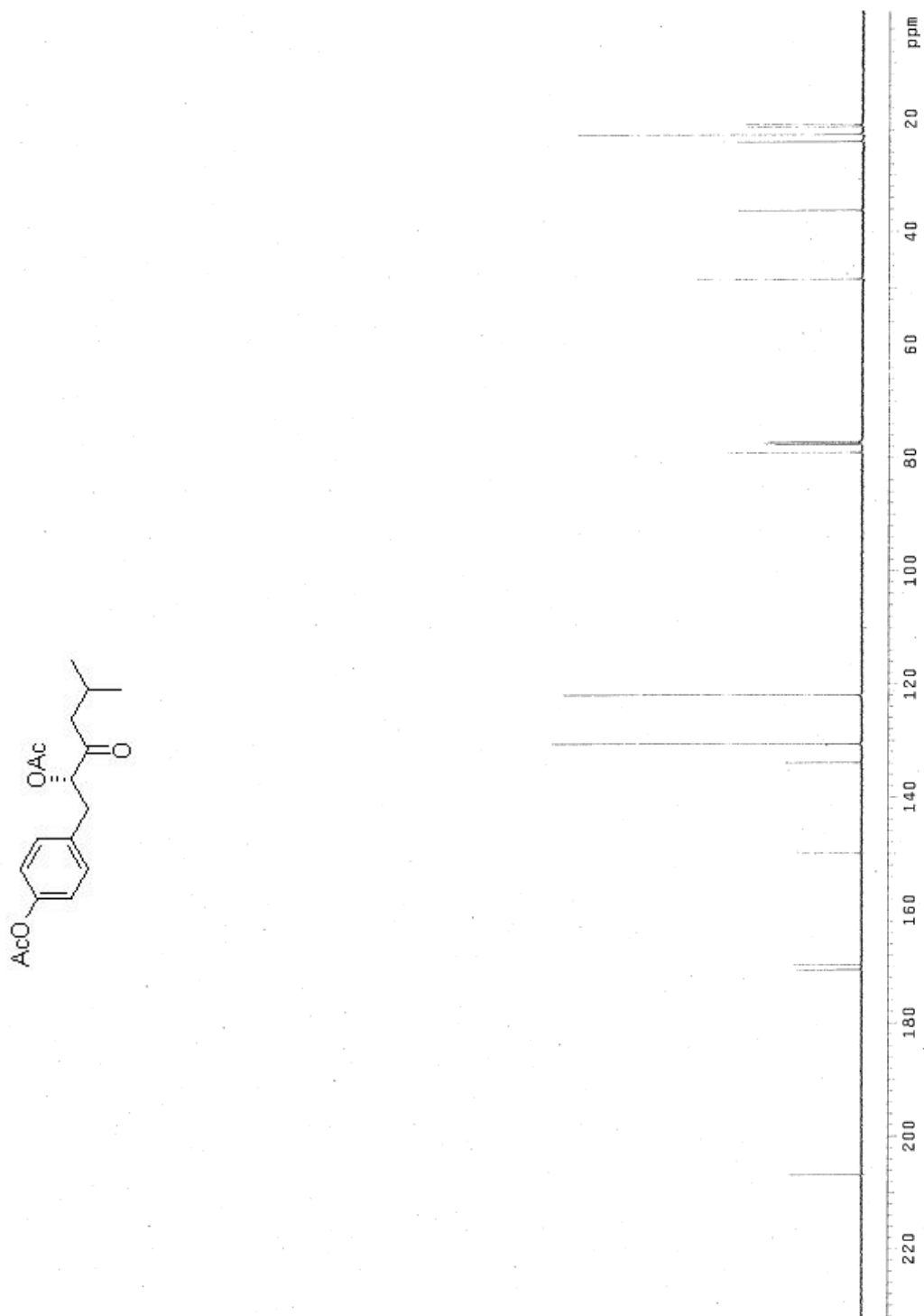


Figure S17. ^{13}C NMR spectrum of **10**