

Figure W1. LC/MS/MS measurement of PGE₂ and PGE₃ in HCA-7 human CRC cell–conditioned medium. (A) Chromatogram of authentic PGE₂ (100 ng/ml) and PGE₃ (50 ng/ml) running at 5.7 and 5.0 minutes, respectively. (B) Chromatograms of extracted cell culture medium. The lower traces are PGE₃ and PGE₂ channels for unconditioned medium. The top two traces are PGE₃ and PGE₂ channels for medium conditioned by cells cultured in the presence of 95% (vol./vol.) ethanol carrier alone for 24 hours. Note the absence of a PGE₃ peak and a large PGE₂ peak. (C) Chromatograms of extracted conditioned medium from cells cultured in the presence of EPA-FFA (0.2 μ mol/10⁶ cells [equivalent to a 50- μ M concentration]) for 24 hours. PGE₃ is now present in conditioned medium along with lesser amounts of PGE₂ compared with medium conditioned in the absence of EPA-FFA (compare with B).



Figure W2. EP4 receptor signaling in human CRC cells. (A) Human CRC cell lines that generated a PGE₂-induced increase in intracellular cAMP at 5 minutes were identified. (B) cAMP induction by PGE₂ (1 μ M) was inhibited by the EP4 receptor antagonist (EP4 antg) ONO-AE3-208 (10 μ M) but not mimicked by the EP2 receptor agonist butaprost (but; 1 μ M) in LoVo, HCT116, and HCA-7 cells, confirming dominant PGE₂-EP4 receptor signaling in these human CRC cell lines (but not HRT18). (C) Immunofluorescence detection of EP4 receptor protein in i) HT-29-EP4 cells (membranous colocalization [yellow] of anti-V5 tag [red] and EP4 receptor [green]), ii) and iii) LoVo cells demonstrating cytoplasmic and membranous localization (arrowheads), and iv) PGE₂ (1 μ M)-treated LoVo cells demonstrating ligand-induced receptor internalization (inset—no primary antibody control).