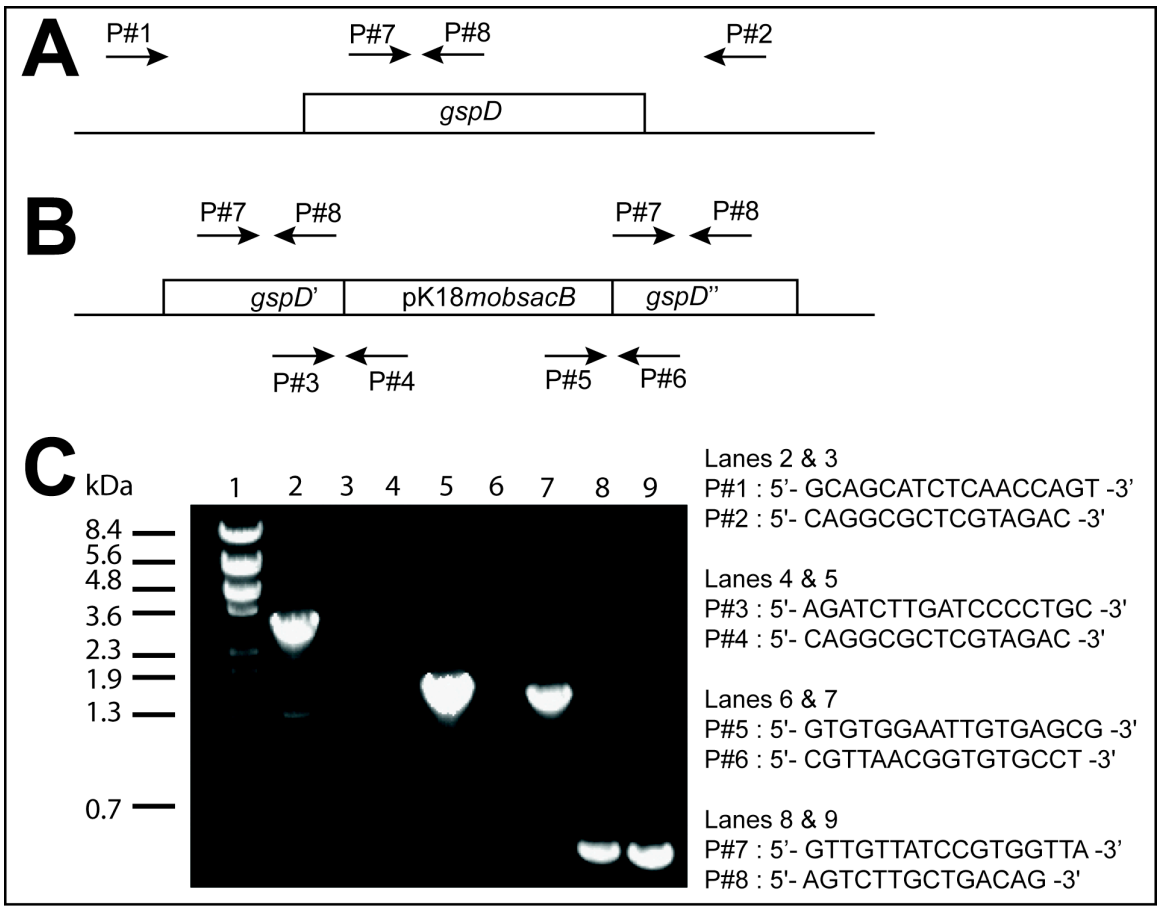

Figure S1. Confirmation of the *C. japonicus gspD*::pJGG1 insertion mutation. A liquid culture of wild type and the candidate *gspD*::pJGG1 insertion were used as template for amplification via PCR. The amplification products were then fractionated by agarose gel electrophoresis and the DNA detected by staining with ethidium bromide as described in "Materials and Methods". **Panel A & B**. Map of wild type and *gspD*::pJGG1 insertion, with locations of primers used for PCR. "P#1-P#8" refer to the primer number used for amplification (the sequences of which are shown above). **Panel C**. Agarose gel electrophoresis of PCR amplified DNA. **Lane 1**, λ BstEII standard; **Lane 2**, PCR amplification of *gspD* region from wild type *C. japonicus* with primers P#1 and P#2; **Lane 3**, PCR of *gspD* region from *gspD*::pJGG1 mutant with primers P#1 and P#2; **Lane 4**, PCR from wild type *C. japonicus* with primers P#3 and P#4; **Lane 5**, PCR from the *gspD*::pJGG1 mutant with primers P#3 and P#4; **Lane 6**, PCR from wild type *C. japonicus* with primers P#5 and P#6; **Lane 7**, PCR from the *gspD*::pJGG1 mutant with primers P#5 and P#6; **Lane 8**, PCR of internal *gpsD* region from wild type *C. japonicus* with primers P#7 and P#8; **Lane 9**, PCR of internal *gspD* region from the *gspD*::pJGG1 mutant with primers P#7 and P#8.