## **Supporting Information**

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**Fig. S1.** Relative *preproinsulin* expression in  $\beta$ ERKO<sup>-/-</sup> islets (*A*) and GPERKO<sup>-/-</sup> islets (*B*) after E2 (10<sup>-8</sup> M) treatment in vitro (*n* = 3–12 mice/group). Results are expressed as mean ± SEM.



**Fig. S2.** Metabolic characterization of  $PER\alpha KO^{-/-}$  mice. (*A*–*D*) Glucose tolerance (*A*) and corresponding area under the curve (*B*), fasting blood glucose (*C*), and fed blood glucose (*D*) at 8 wk in  $PER\alpha KO^{-/-}$  and  $ER\alpha lox/lox$  mice. (*E*–*H*) Glucose tolerance (*E*) and corresponding area under the curve (*F*), fasting blood glucose (*G*), and fed blood glucose (*H*) at 24 wk in  $PER\alpha KO^{-/-}$  and  $ER\alpha lox/lox$  mice (*n* = 6–20/group). Results are expressed as mean ± SEM.



**Fig. S3.** ER $\alpha$ -mediated ERK1/2 phosphorylation and nuclear translocation. (A and C) MIN6 cells were treated with E2 (10<sup>-8</sup> M) as indicated (A) or with E2 (10<sup>-8</sup> M), EDC (10<sup>-8</sup> M), PPT (10<sup>-8</sup> M), or PD 98059 (10<sup>-5</sup> M) (C) for 5 min. Phosphorylation of ERK1/2 was determined by Western blot analysis with an antibody against phosphorylated ERK1/2. (*B*) Localization of phosphorylated ERK1/2 in MIN6 cells after treatment with E2 (10<sup>-8</sup> M), PPT (10<sup>-8</sup> M), EDC (10<sup>-8</sup> M), or E2 + PD98059 (10<sup>-5</sup> M) for 5 min.



**Fig. S4.** Inhibition of E2-induced ERK1/2 phosphorylation by PP1. (*A*) Localization of phosphorylated ERK1/2 in MIN6 cells treated with either E2 ( $10^{-8}$  M) or E2 + PP1 ( $10^{-5}$  M). (*B*) Inhibition of E2-induced ERK1/2 phosphorylation by PP1 in MIN6 cells. Cells were treated with E2 ( $10^{-8}$  M) or PP1 ( $10^{-5}$  M) for 5 min before Western blot analysis. (*C* and *D*) Inhibition of E2-induced preproinsulin gene expression (*C*) and rise in insulin concentration (*D*) by PP1 ( $10^{-5}$  M) in WT islets. Mouse islets were treated with PP1 for 48 h before measurements of *preproinsulin* expression and insulin concentration. Results are expressed as mean ± SEM.



**Fig. S5.** ER $\alpha$ -induced NeuroD1 nuclear localization. NeuroD1 predominant nuclear localization in the presence of E2 (10<sup>-8</sup> M) or PPT (10<sup>-8</sup> M) at 2.8 mM and 11 mM glucose in MIN6 cells (n = 3-6/group). Representative images of four independent experiments are shown. Arrows indicate cells with predominant nuclear localization of NeuroD1.



**Fig. S6.** E2-induced *preproinsulin* transcription does not involve direct binding of ER $\alpha$  to the insulin promoter. ChIP showing the recruitment of ER $\alpha$  to the IGF1 gene sequence containing an ERE (*A*) or insulin promoter (*B*) after 24-h treatment with E2 (10<sup>-8</sup> M) or vehicle (V) at 11 mM glucose in MIN6 cells. After immunoprecipitation of ER $\alpha$  (HC20; Santa Cruz Biotechnology), real-time qPCR amplification of the IGF1 gene sequence containing an ERE and insulin promoter were performed using Sybr Green (BioRad), as described in *Materials and Methods*. Results were normalized to vehicle 11 mM glucose and represent mean  $\pm$  SEM (n = 4-5/group). \*P < 0.05.

## Table S1. PCR primer sequences

Gene	Forward primer	Reverse primer	Ref.
mERα	5'-TTGCCCGATAACAATAACAT-3'	5'-GGCATTACCACTTCTCCTGGGAGTCT-3'	(1)
$\beta$ -actin	5'-AGGTCATCACTATTGGCAAC-3'	5'-ACTCATCGTACTCCTGCTTG-3'	(2)
mInsulin promoter	5'-GAAGGTCTCACCTTCTGG-3'	5'-GGGGGTTACTGGATGCC-3'	(3)
mIGF-1	5'GCAGATAGAGCCTGCGCAATGGA-3'	5'-GGCTGCTGATTTTCCCCATCGCT-3'	(4)

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4. Hewitt SC, Li Y, Li L, Korach KS (2010) Estrogen-mediated regulation of Igf1 transcription and uterine growth involves direct binding of estrogen receptor alpha to estrogen-responsive elements. J Biol Chem 285:2676–2685.