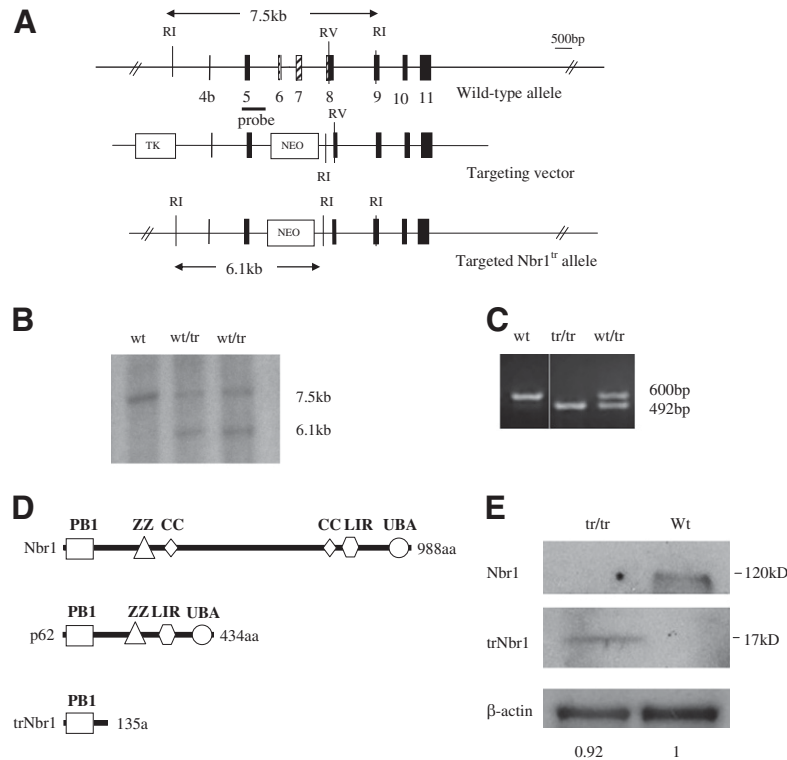
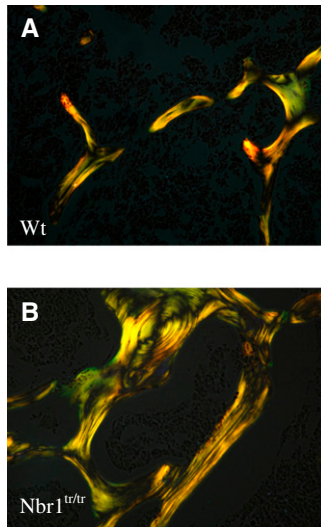


# Supporting Information

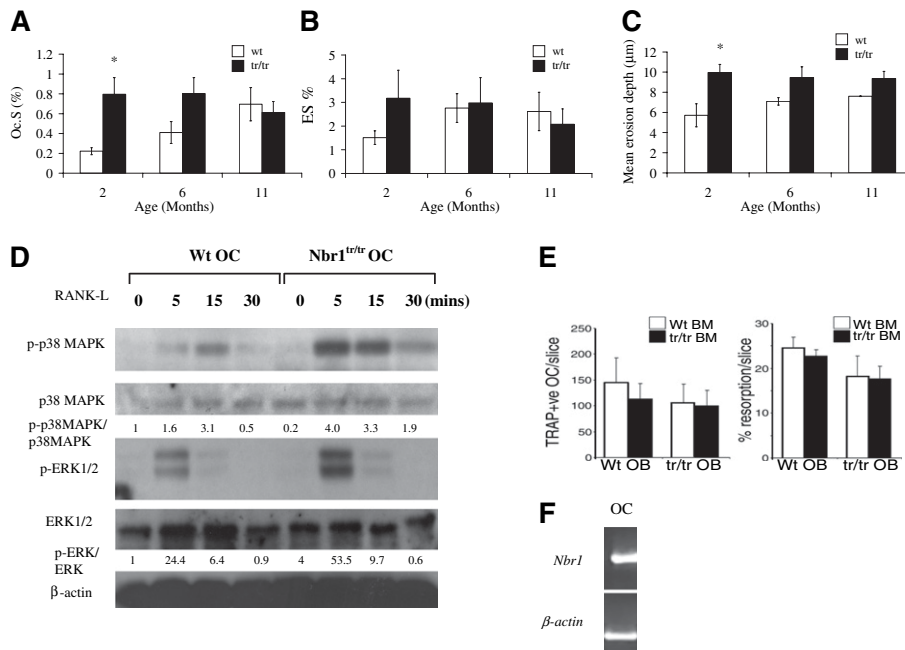
Whitehouse et al. 10.1073/pnas.0913058107



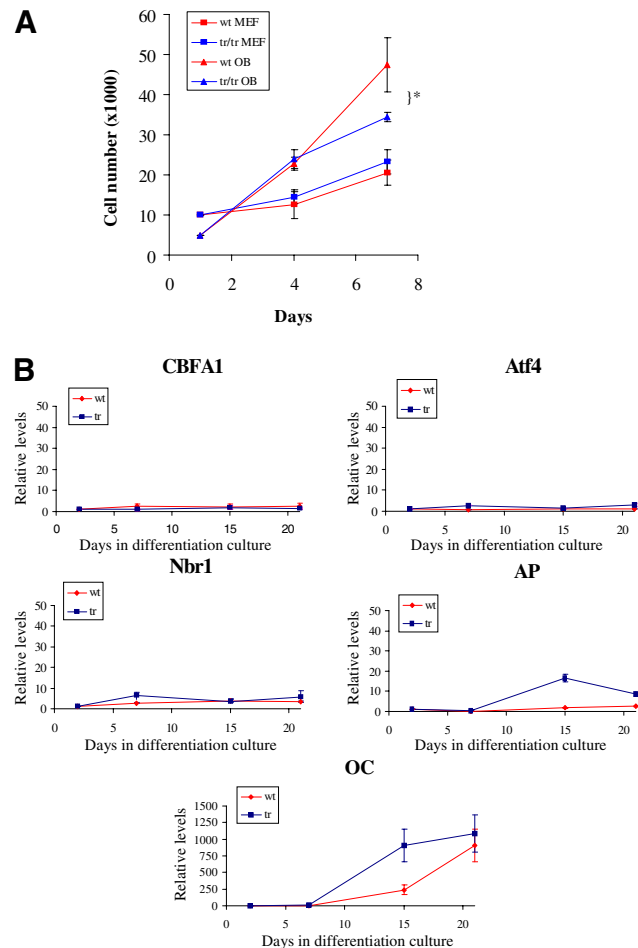
**Fig. S1.** Targeted disruption of neighbor of Breast cancer 1 (*Brca1*) gene *Nbr1*. (A) Schematic representation of *Nbr1* targeting strategy. Boxes represent exons 4b–11 of *Nbr1*. Hashed exons are lost in the targeted allele. NEO, neomycin; TK, thymidine kinase; RI, *EcoRI*; RV, *EcoRV*. (B) Correct 5' arm homologous recombination was confirmed in two embryonic stem cell clones using an exon/intron 5 probe (as shown in A) for Southern hybridization analysis of *EcoRI*-digested genomic DNA. (C) Subsequent mice were genotyped by PCR (600 bp; exon6/7 wild-type allele, 492 bp; neo). (D) Schematic representation of *Nbr1*, p62, and truncated *Nbr1* (tr*Nbr1*) protein domain structure. PB1, Phox and Bem1p domain; ZZ, zinc binding; CC, coiled coil; LIR, LC3-interacting region 1; UBA, ubiquitin-associated domain. Drawn to scale. (E) Western blot analysis of *Nbr1* and tr*Nbr1* expression in osteoblasts from homozygous *Nbr1*<sup>tr/tr</sup> mice (tr/tr) or wild-type (Wt) mice showing ~120 kD full length and 17 kD truncated protein. Actin, loading control. Quantification of Western blots shows that both proteins are expressed at similar levels.



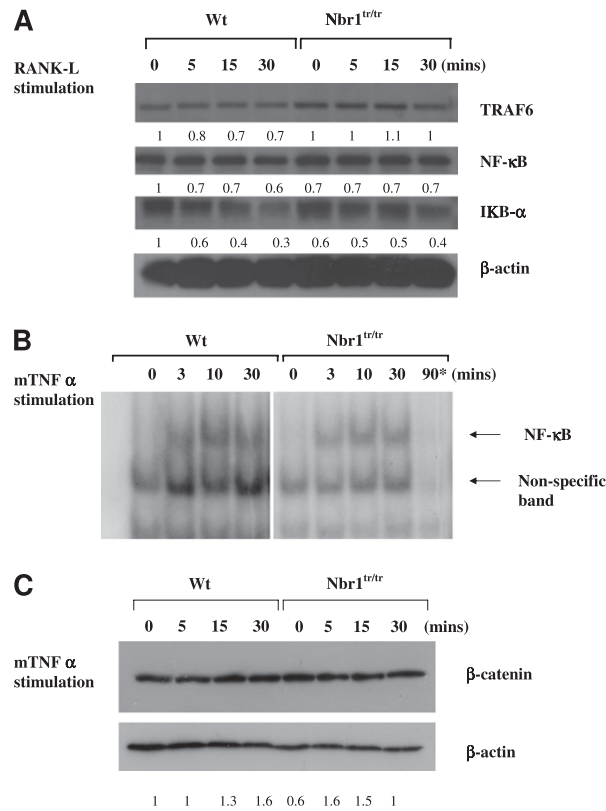
**Fig. S2.** *Nbr1*<sup>tr/tr</sup> bone shows properties of mature lamellar and not woven bone, as shown by parallel alignment of the collagen fibers. Histological sections of 8-mo-old (A) wild-type and (B) *Nbr1*<sup>tr/tr</sup> femurs photographed under polarized light. (Magnification: 20x.)



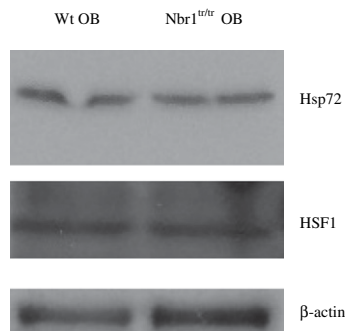
**Fig. S3.** Histomorphometry analysis of osteoclast parameters and cell-signaling pathway perturbation in osteoclast precursors from *Nbr1*<sup>tr/tr</sup> mice and age-matched controls. White bars, wild-type mice; black bars, *Nbr1*<sup>tr/tr</sup> mice; x axis, age of animals (months). (A) Oc.S, osteoclast surface. (B) ES, eroded surface. (C) Mean erosion depth. Differences between the genotypes were assessed using Student's *t* test. Error bars represent SEM (*n* = 3; \**P* < 0.05). (D) Receptor activator for nuclear factor kappa B ligand (RANK-L)-induced ERK1/2 and p38 MAPK activation in isolated wild-type (Wt) and *Nbr1*<sup>tr/tr</sup> osteoclast precursor cells (OC). (E) Osteoclast differentiation assays. Coculture of primary calvarial osteoblasts (OBs) from WT or *Nbr1*<sup>tr/tr</sup> mice with macrophage colony stimulating factor-dependent bone marrow (BM) cells from WT or *Nbr1*<sup>tr/tr</sup> mice on dentine slices as indicated. The number of tartrate resistant acid phosphatase (TRAP) +ve osteoclasts (OC; *Left*) and percent resorption (*Right*) were quantified after 7 and 10 d, respectively. The data represent the mean ± SD of four representative dentine slices. (F) RT-PCR analysis confirms endogenous *Nbr1* expression in primary murine osteoclast (OC) cultures.



**Fig. S4.** (A) Proliferation curves for calvarial-derived osteoblasts (OB) and murine embryonic fibroblasts (MEF) from wild-type and  $Nbr1^{tr/tr}$  mice. The data represents the mean  $\pm$  SD of triplicate representative wells for each time point ( $*P < 0.05$ ). (B) Quantitative RT-PCR analysis of OB marker gene expression in calvarial-derived osteoblast cultures from  $Nbr1^{tr/tr}$  mice (blue) and Wt (red) mice during a 21-d time course of in vitro osteoblast differentiation. CBFA1/Runx2; Atf4; Nbr1; AP, alkaline phosphatase; OC, osteocalcin. Relative expression of each gene was normalized to  $\beta$ -actin and levels at day 2, and wild-type expression is set as 1 (y axis). The data represent the mean  $\pm$  SD of triplicate representative wells.



**Fig. S5.** Ex vivo analysis suggests that canonical Wnt and NF-κB pathways are unaffected in Nbr1<sup>tr/tr</sup> mice. (A) Western blot analysis showed no difference in levels of NF-κB pathway components after RANK-L stimulation in osteoclasts from wild-type (wt) or homozygous Nbr1<sup>tr/tr</sup> mice. (B) NF-κB mobility shift assay of nuclear extract from MEF from wild-type (wt) or homozygous Nbr1<sup>tr/tr</sup> mice showed no difference in TNFα-induced NF-κB nuclear accumulation. (\*100-fold excess of nonradiolabeled probe shows NF-κB band specificity.) (C) β-catenin levels are equivalent in mTNFα-treated osteoblasts from wild-type (wt) or homozygous Nbr1<sup>tr/tr</sup> mice (β-actin, loading control). Quantitation shows levels of protein compared with that of untreated wild-type cells.



**Fig. S6.** Ex vivo analysis of downstream p38 MAPK effectors in osteoblasts showed no effect on protein levels by Nbr1 truncation. Wild-type (Wt) and Nbr1<sup>tr/tr</sup> osteoblasts (OB) were serum starved to activate the p38 MAPK pathway and analyzed for changes in levels of heat shock factor protein 1 (HSF1) and heat shock protein 72 (Hsp72). Levels are normalized to β-actin and are equivalent.

**Table S1. Summary of bone mineral density (BMD) and bone-area measurements from femurs and calvariae of age-matched 3- and 9-mo-old Nbr1<sup>tr/tr</sup> mice and wild-type controls**

	Genotype	3 mo old	<i>P</i> value	9 mo old	<i>P</i> value
Total BMD (mg/cm <sup>3</sup> )	Wt	515.65 ± 30.08	5.59 × 10 <sup>-5</sup>	533.17 ± 77.14	2.52 × 10 <sup>-5</sup>
	tr/tr	814.44 ± 81.03		1,055.79 ± 83.41	
Trabecular BMD (mg/cm <sup>3</sup> )	Wt	38.43 ± 24.40	2.25 × 10 <sup>-4</sup>	73.71 ± 20.42	2.07 × 10 <sup>-5</sup>
	tr/tr	416.76 ± 161.18		837.9 ± 171.54	
Cortical BMD (mg/cm <sup>3</sup> )	Wt	894.07 ± 40.68	4.22 × 10 <sup>-6</sup>	906.95 ± 122.56	1.2 × 10 <sup>-3</sup>
	tr/tr	1,125.22 ± 23.74		1,233.55 ± 15.71	
Total bone area (mm <sup>2</sup> )	Wt	2.37 ± 0.14	1.11 × 10 <sup>-4</sup>	2.80 ± 0.29	10.4 × 10 <sup>-5</sup>
	tr/tr	3.18 ± 0.22		4.00 ± 0.09	
Trabecular bone area (mm <sup>2</sup> )	Wt	1.05 ± 0.07	3.04 × 10 <sup>-4</sup>	1.27 ± 0.13	9.21 × 10 <sup>-5</sup>
	tr/tr	1.44 ± 0.10		1.80 ± 0.04	
Cortical bone area (mm <sup>2</sup> )	Wt	1.31 ± 0.07	9.45 × 10 <sup>-5</sup>	1.55 ± 0.15	9.48 × 10 <sup>-5</sup>
	tr/tr	1.77 ± 0.12		2.21 ± 0.05	

Differences between the genotypes were assessed using Student's *t* test (*n* = 5).