

Combinatorial profiling of chromatin-binding modules reveals multi-site discrimination

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Supplementary Methods

Library Construction

The combinatorial histone H3 peptide library was synthesized on TentaGel NH₂ resin (90 μm, 0.26 mmol/g loading, 2.86 x 10⁶ beads/g) using the split-pool approach¹ for sites of posttranslational modification. Standard amino acid building blocks were purchased from Peptides International. All modified amino acids are commercially available from Novabiochem and peptide reagents were obtained from Sigma Aldrich. A linker composed of methionine, arginine and a polyethylene glycol (PEG) segment was installed with standard Fmoc/tBu chemistry². Sites of modification included positions 2 (R, Rme1, Rme2s, Rme2a, Cit), 3 (T, Tph), 4 (K, Kme1, Kme2, Kme3, Kac), 6 (T, Tph), 8 (R, Rme1, Rme2s, Rme2a, Cit), 9 (K, Kme1, Kme2, Kme3, Kac) and 10 (S, Sph). Varying amounts of capping reagents were added to the coupling reactions at lysine and arginine residues to generate a mass ladder for peptide sequencing (Supplementary Fig. 1)³. The capping scheme was as follows: 2.5 % Ac-Ala for K, Kme1, Kme2 and Kme3 and 5 % Boc-Ala for Kac at position 9; 1.5 % Ac-Ala for R, Rme1, Rme2a and 1.5 %

Ac-Ala + 3 % Boc-Ala for Rme2s and Cit at position 8; 2.5 % Ac-Ala for K, Kme1, Kme2 and Kme3 and 5 % Boc-Ala for Kac at position 4; 1.5 % Ac-Ala for R, Rme1, Rme2a and 1.5 % Ac-Ala + 3 % Boc-Ala for Rme2s and citrulline at position 8. A 5 mg portion of the library was deprotected 2 x 2 hours with Reagent K (TFA/EDT/thioanisole/water/phenol: 82.5%, 2.5%, 5%, 5%, 5%)⁴ prior to use.

Immunoblotting and Immunoabsorption Assay

Lyophilized untreated histones purified by acid extraction from HeLa cells (Millipore 17-306) were reconstituted in 1x NEBuffer 3 (50 mM Tris-HCl, 10 mM MgCl₂, 100 mM NaCl, 1 mM DTT, pH 7.9) and phosphatase inhibitors (10 mM NaF and 1 mM NaVO₄). Histones (10 µg, in duplicate) were separated by SDS-PAGE (15%) and transferred to PVDF. Membranes were blocked overnight in 5% BSA-TBST (0.05%) at 4°C. Membranes were incubated for 3 hours at room temperature with 1:7,500 anti-histone H3 phospho T6 (Abcam ab14102) in blocking solution +/- 1 µM histone H3 (residues 1-10) phospho T6 peptide (peptide was preincubated with antibody solution for 1 hour prior to application to membranes, and all antibody solutions were pre-equilibrated in batch to insure uniform solutions across membranes). Membranes were washed 5 minutes in TBST (0.05%) three times. Membranes were incubated for 1 hour at room temperature with 1:5,000 HRP-conjugated goat anti-rabbit (Santa Cruz) in blocking solution. Membranes were then washed 5 minutes in TBST (0.05%) four times. Detection was performed using 1:1 SuperSignal West Dura Substrate (Pierce) and membranes were exposed to film.

Following detection with anti-histone H3 phospho T6, the membranes were stripped in 0.8 M Tris, 2% SDS, 0.01 M 2-mercaptoethanol for 30 minutes at 42°C, followed by extensive washing in TBST (0.05%). Membranes were blocked overnight at 4°C in 5% BSA-TBST (0.05%). Membranes were incubated for 1 hour with 1:10,000 anti-histone H3 (Abcam ab46765) in blocking solution, followed by three five-minute washes with TBST (0.05%). Membranes were incubated with 1:600 HRP-conjugated goat anti-rabbit (Pierce) in blocking solution, followed by three five-minute washes. Detection was performed using 1:1 SuperSignal West Dura Substrate (Pierce) and membranes were exposed to film. All antibody solutions were pre-equilibrated in batch to insure uniformity.

Mass Spectrometry for H3T6ph identification

Acid extracted histones from HeLa cells were separated by reverse-phase HPLC using a C18 column (2.1 mm i.d. ×250 mm, Vydac, Hesperia, CA) on an Beckman Coulter System Gold HPLC (Fullerton, CA). The gradient consisted of 30-60% B in 100min, followed by 60-100%B in 20min (A = 5% MeCN in 0.2% TFA, B = 90% acetonitrile in 0.188% TFA). Fractions corresponding to histone H3 were pooled and dried by speed vacuum. The H3 sample was then diluted in deionized water. An aliquot corresponding to approximately 10 µg was then taken and treated with propionic anhydride reagent as previously described⁵. The sample was then digested by trypsin at a protein:enzyme ratio of 10:1 for 6 hours at 37°C, with the reaction being quenched by addition of glacial acetic acid to drop the pH to ~3. The H3 sample was then subjected to methyl esterification and immobilized metal affinity chromatography (IMAC) as

previously described⁶. H3 peptides were desalted using STAGE tips as previously reported⁷ and loaded onto a C18 packed fused silica microcapillary column (75µm) constructed with an integrated ESI tip through an Eksigent AS-2 autosampler (Eksigent Technologies Inc., Dublin, CA) for nanoLC-MS/MS on an Orbitrap mass spectrometer (ThermoFisher Scientific, San Jose, CA). A full mass spectrum at 30,000 resolution was acquired in the Orbitrap prior to obtaining 10 data-dependent MS/MS spectra in the ion trap. All spectra were manually inspected.

Isothermal Titration Calorimetry (ITC)

Peptides for ITC analysis were synthesized using solid phase peptide synthesis (SPPS) on the Intavis ResPep SL robot (Intavis, Koeln, Germany). Standard Fmoc/tBu amino acid couplings were used and premodified amino acid building blocks (Tph, Kme3, Rme2s; Novabiochem) were incorporated at positions 2,3,4 and 6 depending on the peptide. Peptide concentration was determined by weight of purified dry product. All H3 peptides were normalized to each other by HPLC analysis. In order to confirm the concentration of several peptide stock solutions amino acid analysis was performed (UC-Davis; Proteomics Core Facility). Thermodynamic analyses were performed at 25 °C using a VP-ITC isothermal titration calorimeter (MicroCal LLC, Northampton, MA, USA). Histone peptides (0.5- 2.8 mM) were injected into solutions containing GST-fusion protein (20-350 µM) . In a typical experiment 37 injections of peptide were delivered at 200s intervals to a 1.4 mL solution of protein. Different volumes were injected during the course of the experiment (1×1 µL , 6 × 4 µL, and 33 × 8 µL). For the titration of H3K4me3T3ph into the JMJD2A-DTD injections were: 1×1 µL , 6 × 4 µL,

$15 \times 8 \mu\text{L}$, and $9 \times 16 \mu\text{L}$. The initial point was routinely discarded, a common practice in ITC. Curve fitting was done by using Origin 7.0 with a standard one-site model provided by MicroCal Lavenberg-Marquardt nonlinear regression. A peptide into buffer control corrected for dilution heats of injection in each experiment. All experiments were performed in 25mM Tris buffer (pH 7.2) containing 50mM NaCl and 2mM BME.

Supplementary Results

Supplementary Dataset 1

SI Table 1. Sequences of 100 randomly chosen beads from the H3 library

SI Table 2. Screening results for BHC80-PHD finger

SI Table 3. Screening results for AIRE-PHD finger 1

SI Table 4. Screening results for ING2-PHD finger

SI Table 5. Screening results for RAG2-PHD finger

SI Table 6. Screening results for for JMJD2A DTD

Supplementary Table 1

Random Beads

	1	2	3	4	5	6	7	8	9	10
1	A	Rme2s	Tph	Kme1	Q	T	A	Rme2a	Kme2	Sph
2	A	Cit	Tph	K	Q	Tph	A	Rme2a	Kme3	S
3	A	R	Tph	Kme1	Q	T	A	Cit	Kme3	Sph
4	A	R	Tph	Kme1	Q	Tph	A	R	Kme1	S
5	A	R	Tph	Kme3	Q	T	A	Rme1	Kme1	S
6	A	Rme1	Tph	Kme3	Q	T	A	Rme2a	Kme3	Sph
7	A	Rme2a	Tph	Kac	Q	T	A	Rme2a	Kme3	S
8	A	R	Tph	Kme2	Q	T	A	Cit	K	Sph
9	A	Rme2a	Tph	Kme1	Q	T	A	Rme2a	K	S
10	A	Rme2a	T	Kme1	Q	T	A	R	Kme2	S
11	A	Rme2s	T	K	Q	Tph	A	Rme2s	Kme1	Sph
12	A	Rme2s	T	Kme2	Q	Tph	A	Rme2s	Kme1	Sph
13	A	Cit	T	Kac	Q	T	A	Rme2s	Kme3	Sph
14	A	Rme2s	T	Kac	Q	Tph	A	Rme2a	K	Sph
15	A	Rme2s	T	Kme3	Q	T	A	Rme2s	Kme2	Sph
16	A	Cit	Tph	Kme1	Q	T	A	Rme2s	Kme2	Sph
17	A	Cit	T	Kac	Q	Tph	A	Cit	Kme1	S
18	A	R	T	Kme1	Q	T	A	Cit	Kac	Sph
19	A	Rme2s	T	K	Q	T	A	R	K	S
20	A	Rme1	T	Kme2	Q	T	A	Rme2s	K	Sph
21	A	Cit	Tph	Kme3	Q	T	A	Rme1	Kme2	Sph
22	A	R	T	Kme3	Q	Tph	A	Rme2a	Kme1	S
23	A	Rme2s	T	Kme3	Q	Tph	A	Rme2s	Kme1	S
24	A	Rme2s	Tph	Kme3	Q	T	A	Rme2a	Kme3	S
25	A	Rme2s	T	Kme1	Q	T	A	Rme1	Kme2	Sph
26	A	Rme1	Tph	Kme3	Q	Tph	A	Rme2a	K	S
27	A	Rme2s	Tph	Kac	Q	Tph	A	Rme2s	Kme2	Sph
28	A	Rme2a	Tph	Kme1	Q	T	A	Rme1	K	Sph
29	A	R	Tph	Kme3	Q	T	A	Rme2a	Kme2	Sph
30	A	Rme1	T	Kac	Q	T	A	Rme1	Kme2	S
31	A	R	Tph	Kme1	Q	Tph	A	Rme1	Kme2	Sph
32	A	Cit	T	Kac	Q	T	A	Rme2s	K	Sph
33	A	Rme1	T	Kme1	Q	Tph	A	Rme2s	K	S
34	A	Cit	T	K	Q	T	A	Cit	Kme1	Sph
35	A	R	Tph	Kme3	Q	Tph	A	Cit	K	S
36	A	Rme2a	Tph	Kme1	Q	T	A	Rme2a	Kme3	Sph
37	A	Cit	Tph	Kme2	Q	T	A	Rme1	Kme3	Sph
38	A	Rme2s	Tph	K	Q	Tph	A	R	K	Sph
39	A	R	Tph	Kme1	Q	Tph	A	Rme2a	Kme1	S
40	A	Cit	Tph	Kme3	Q	T	A	Cit	K	S
41	A	Rme2s	Tph	Kme1	Q	T	A	R	Kme1	Sph
42	A	Rme1	Tph	Kac	Q	T	A	Cit	Kme3	Sph
43	A	Rme2a	Tph	Kme1	Q	Tph	A	Rme2a	K	Sph
44	A	Rme2a	T	Kme1	Q	Tph	A	Cit	Kme1	S

45	A	Rme2s	T	Kac	Q	T	A	Rme1	K	Sph
46	A	R	T	K	Q	Tph	A	Rme1	Kme3	S
47	A	R	T	Kme2	Q	Tph	A	R	Kme1	Sph
48	A	R	T	Kme1	Q	T	A	Cit	K	S
49	A	Rme2s	T	Kme1	Q	T	A	Cit	Kme3	S
50	A	Rme2a	Tph	Kme3	Q	T	A	R	Kac	S
51	A	Rme	T	K	Q	Tph	A	Rme	Kme3	Sph
52	A	Rme	Tph	Kme3	Q	Tph	A	R	Kme2	Sph
53	A	Cit	Tph	Kme2	Q	T	A	R	K	Sph
54	A	Rme	T	Kme2	Q	T	A	Rme2a	Kme2	S
55	A	R	T	Kme2	Q	Tph	A	R	Kac	Sph
56	A	Rme	T	Kme3	Q	Tph	A	Rme	Kme2	S
57	A	Rme2s	Tph	Kme3	Q	Tph	A	Rme2a	Kac	S
58	A	Cit	T	Kme2	Q	Tph	A	Rme2s	Kme2	Sph
59	A	R	Tph	Kme3	Q	T	A	R	Kme3	S
60	A	Cit	Tph	Kme	Q	Tph	A	Rme2s	Kme2	Sph
61	A	Cit	T	Kme2	Q	T	A	Rme2s	Kac	S
62	A	R	T	K	Q	Tph	A	R	Kme2	S
63	A	Rme2a	T	K	Q	T	A	Rme	Kac	S
64	A	Rme2s	Tph	Kac	Q	T	A	Cit	Kme3	Sph
65	A	Rme2s	Tph	Kac	Q	Tph	A	Cit	Kme3	S
66	A	Rme	T	K	Q	Tph	A	R	Kme	S
67	A	Rme	T	K	Q	T	A	Rme2a	Kac	Sph
68	A	Rme	T	K	Q	T	A	R	K	S
69	A	Rme2a	T	K	Q	T	A	Rme2a	Kme2	Sph
70	A	Rme2a	T	K	Q	T	A	Rme	Kme3	Sph
71	A	Cit	T	Kme3	Q	Tph	A	Rme2a	Kac	S
72	A	R	T	Kme	Q	T	A	Rme	Kme	S
73	A	Rme2s	Tph	K	Q	Tph	A	Rme2a	Kme	S
74	A	Rme2a	T	K	Q	Tph	A	Cit	Kme	Sph
75	A	Cit	T	Kac	Q	Tph	A	Rme	Kme2	Sph
76	A	Rme2s	T	Kme3	Q	T	A	R	K	S
77	A	Cit	Tph	Kme	Q	T	A	Rme2a	Kac	Sph
78	A	R	T	Kme2	Q	Tph	A	R	K	S
79	A	Rme2a	T	K	Q	T	A	Cit	Kme3	S
80	A	Cit	T	K	Q	T	A	Rme	Kme	Sph
81	A	Rme2a	Tph	Kme2	Q	T	A	Rme2a	Kac	Sph
82	A	Rme2s	Tph	Kac	Q	T	A	Rme	Kme	S
83	A	Rme2s	Tph	Kme3	Q	T	A	R	Kac	Sph
84	A	R	Tph	Kme3	Q	Tph	A	Rme2a	Kac	S
85	A	Rme2s	T	Kme	Q	T	A	Rme2s	Kme3	Sph
86	A	Rme2s	T	Kme3	Q	T	A	Rme	Kme2	Sph
87	A	Rme	Tph	Kme	Q	T	A	Cit	K	Sph
88	A	R	T	K	Q	Tph	A	Rme	Kac	S
89	A	Rme	T	Kac	Q	Tph	A	Rme2a	Kme2	Sph
90	A	R	T	Kac	Q	T	A	Cit	Kac	S
91	A	Rme2a	Tph	K	Q	Tph	A	R	Kme2	Sph

92	A	Rme	Tph	Kme2	Q	Tph	A	Rme2s	K	Sph
93	A	Rme2s	Tph	Kac	Q	T	A	Rme2s	Kme3	Sph
94	A	Rme2a	Tph	Kme	Q	Tph	A	Cit	Kme2	S
95	A	Cit	Tph	Kme	Q	Tph	A	Rme	Kme3	Sph
96	A	R	T	Kme3	Q	Tph	A	Rme	Kme2	Sph
97	A	Rme2a	Tph	Kme2	Q	T	A	R	Kme	Sph
98	A	Rme2a	T	Kme2	Q	Tph	A	Rme2s	Kme	S
99	A	Rme2s	Tph	K	Q	T	A	Cit	Kme	S
100	A	Cit	T	Kme2	Q	Tph	A	Rme2a	Kme3	Sph

Totals

R= 22	T=52	K= 21
Rme1= 16	Tph=48	Kme1= 25
Rme2s= 25		Kme2= 16
Rme2a= 18		Kme3= 22
Cit=19		Kac= 16

T= 56
Tph = 44

R= 19	K= 21	S= 45
Rme1= 21	Kme1 = 21	Sph = 55
Rme2s= 17	Kme2 = 23	
Rme2a = 24	Kme3 = 21	
Cit= 19	Kac = 14	

Supplementary Table 2

BHC80 Positive Hits

	1	2	3	4	5	6	7	8	9	10
1	A	R	T	K	Q	T	A	R	Kme3	Sph
2	A	R	T	K	Q	T	A	Rme1	Kac	S
3	A	R	T	K	Q	T	A	Rme1	Kac	Sph
4	A	R	T	K	Q	T	A	Rme1	Kac	Sph
5	A	R	T	K	Q	T	A	Rme1	Kme2	S
6	A	R	T	K	Q	T	A	Rme2s	Kac	Sph
7	A	R	T	K	Q	T	A	Cit	K	S
8	A	R	T	K	Q	T	A	Cit	K	S
9	A	R	T	K	Q	T	A	Cit	K	S
10	A	R	T	K	Q	T	A	Cit	Kme1	S
11	A	R	T	K	Q	T	A	Cit	Kme1	S
12	A	R	T	K	Q	T	A	Cit	Kme1	Sph
13	A	R	T	K	Q	T	A	Cit	Kme2	S
14	A	R	T	K	Q	T	A	Cit	Kme3	Sph
15	A	R	T	K	Q	T	A	Cit	Kme3	Sph
16	A	Rme1	T	K	Q	T	A	R	Kac	S
17	A	Rme1	T	K	Q	T	A	R	Kac	Sph
18	A	Rme1	T	K	Q	T	A	Rme1	K	S
19	A	Rme1	T	K	Q	T	A	Rme1	Kme2	S
20	A	Rme1	T	K	Q	T	A	Rme1	Kme2	S

21	A	Rme1	T	K	Q	T	A	Rme1	Kme3	Sph
22	A	Rme1	T	K	Q	T	A	Rme1	Kme3	Sph
23	A	Rme1	T	K	Q	T	A	Rme2s	Kac	Sph
24	A	Rme1	T	K	Q	T	A	Rme2a	Kme1	Sph
25	A	Rme1	T	K	Q	T	A	Rme2a	Kac	Sph
26	A	Rme1	T	K	Q	T	A	Cit	K	Sph
27	A	Rme1	T	K	Q	T	A	Cit	Kac	Sph
28	A	Rme2s	T	K	Q	T	A	Rme2a	Kac	Sph
29	A	Rme2a	T	K	Q	T	A	R	Kme2	Sph
30	A	Rme2a	T	K	Q	T	A	Rme1	Kme1	Sph
31	A	Rme2a	T	K	Q	T	A	Rme2a	Kac	Sph
32	A	Cit	T	K	Q	T	A	Rme1	K	S
33	A	Rme2a	T	Kme2	Q	T	A	Rme2s	K	Sph

BHC80 Negative

Hits

	1	2	3	4	5	6	7	8	9	10
1	A	Rme1	T	K	Q	Tph	A	Rme2a	Kme3	Sph
2	A	Rme2s	T	K	Q	T	A	Rme1	Kme1	Sph
3	A	Rme2s	Tph	K	Q	T	A	R	Kme2	S
4	A	Rme2s	Tph	K	Q	T	A	Cit	Kme3	Sph
5	A	Rme2s	Tph	K	Q	Tph	A	Rme2s	Kme2	Sph
6	A	Cit	T	K	Q	T	A	R	Kac	Sph
7	A	Cit	T	K	Q	Tph	A	Rme1	Kme2	Sph
8	A	R	T	Kme1	Q	T	A	Rme1	Kac	S
9	A	Rme2a	T	Kme1	Q	Tph	A	Rme2a	K	Sph
10	A	Cit	T	Kme1	Q	Tph	A	R	Kme3	S
11	A	Rme2a	Tph	Kme1	Q	T	A	R	Kme3	Sph
12	A	Rme1	Tph	Kme1	Q	Tph	A	Rme2s	Kac	Sph
13	A	R	Tph	Kme2	Q	Tph	A	R	Kme2	Sph
14	A	R	Tph	Kme2	Q	Tph	A	Rme1	Kme1	Sph
15	A	Rme2s	T	Kme2	Q	T	A	R	Kac	Sph
16	A	Rme2a	Tph	Kme2	Q	Tph	A	Rme2s	Kac	S
17	A	Cit	Tph	Kme2	Q	Tph	A	Rme2a	Kme1	S
18	A	R	T	Kme3	Q	Tph	A	Rme2a	Kme3	Sph
19	A	R	Tph	Kme3	Q	Tph	A	Rme2a	Kme1	S
20	A	Rme1	T	Kme3	Q	Tph	A	Rme2a	K	Sph
21	A	Rme1	Tph	Kme3	Q	T	A	Rme2a	Kme1	S
22	A	Rme2s	Tph	Kme3	Q	Tph	A	R	K	Sph
23	A	Rme2s	Tph	Kme3	Q	Tph	A	R	Kac	S
24	A	Rme2s	Tph	Kme3	Q	Tph	A	R	Kac	S
25	A	Rme1	Tph	Kme3	Q	Tph	A	Rme2a	Kac	S
26	A	R	Tph	Kac	Q	Tph	A	R	Kme2	Sph
27	A	Rme2s	Tph	Kac	Q	T	A	Rme2s	Kme2	S
28	A	Rme2a	Tph	Kac	Q	Tph	A	Rme1	Kme1	S

**Supplementary
Table 3**

AIRE-PHD1 Positive

Hits

	1	2	3	4	5	6	7	8	9	10
1	A	R	T	K	Q	T	A	Rme1	Kme1	S
2	A	R	T	K	Q	T	A	Rme1	Kme2	S
3	A	R	T	K	Q	T	A	Rme2a	Kme3	S
4	A	R	T	K	Q	T	A	Cit	Kme2	Sph
5	A	R	T	K	Q	T	A	Cit	Kme3	S
6	A	Rme1	T	K	Q	T	A	R	Kme1	Sph
7	A	Rme1	T	K	Q	T	A	R	Kme2	S
8	A	Rme1	T	K	Q	T	A	Rme1	Kme1	Sph
9	A	Rme1	T	K	Q	T	A	Rme1	Kac	S
10	A	Rme1	T	K	Q	T	A	Rme2s	K	Sph
11	A	Rme1	T	K	Q	T	A	Rme2a	Kme3	S
12	A	Rme1	Tph	K	Q	T	A	R	Kme2	Sph
13	A	Rme2s	T	K	Q	T	A	Rme1	Kme3	S
14	A	Rme2a	T	K	Q	T	A	R	K	Sph
15	A	Rme2a	T	K	Q	T	A	Rme1	Kme1	S
16	A	Rme2a	T	K	Q	T	A	Rme1	Kme1	S
17	A	Rme2a	T	K	Q	T	A	Rme1	Kme3	S
18	A	Cit	T	K	Q	T	A	R	Kme2	S
19	A	R	T	Kme1	Q	T	A	R	Kme1	S
20	A	R	T	Kme1	Q	T	A	Rme1	K	S
21	A	R	T	Kme1	Q	T	A	Rme1	K	Sph
22	A	R	T	Kme1	Q	T	A	Rme1	Kac	S
23	A	R	T	Kme1	Q	T	A	Rme2s	K	Sph
24	A	R	T	Kme1	Q	T	A	Rme2s	K	Sph
25	A	R	T	Kme1	Q	T	A	Rme2s	Kme1	S
26	A	R	T	Kme1	Q	T	A	Rme2a	Kme3	S
27	A	R	T	Kme1	Q	T	A	Rme2a	Kac	S
28	A	R	T	Kme1	Q	T	A	Cit	K	S
29	A	Rme1	T	Kme1	Q	T	A	Rme2a	K	S
30	A	Rme2a	T	Kme1	Q	T	A	Rme2a	K	S

AIRE-PHD1

Negative Hits

	1	2	3	4	5	6	7	8	9	10
1	A	R	Tph	K	Q	T	A	Rme2s	Kme3	Sph
2	A	R	Tph	K	Q	T	A	Cit	Kme1	Sph
3	A	Rme2s	Tph	K	Q	T	A	R	Kme3	Sph
4	A	Rme2s	Tph	K	Q	T	A	Rme2a	Kme1	Sph
5	A	Rme2s	Tph	K	Q	Tph	A	Rme2s	Kme2	Sph
6	A	Rme2a	Tph	K	Q	T	A	Rme2s	Kme1	S
7	A	Cit	Tph	K	Q	Tph	A	Rme1	Kme1	Sph
8	A	R	Tph	Kme1	Q	Tph	A	Rme2a	Kme3	Sph
9	A	Rme1	Tph	Kme1	Q	T	A	Cit	K	Sph
10	A	Rme2s	T	Kme1	Q	Tph	A	Rme2a	Kme1	S
11	A	Rme2s	Tph	Kme1	Q	T	A	Rme2s	Kme3	Sph
12	A	R	Tph	Kme2	Q	Tph	A	Rme1	Kac	Sph

13	A	R	Tph	Kme2	Q	Tph	A	Cit	Kme1	Sph
14	A	Rme1	T	Kme2	Q	Tph	A	Rme2s	K	S
15	A	Rme2s	T	Kme2	Q	Tph	A	Rme2s	Kac	S
16	A	Cit	Tph	Kme2	Q	Tph	A	Rme2a	Kme2	Sph
17	A	Cit	Tph	Kme2	Q	Tph	A	Rme2s	Kac	S
18	A	R	Tph	Kme3	Q	Tph	A	Rme2a	Kme1	S
19	A	Rme1	Tph	Kme3	Q	Tph	A	R	Kme2	Sph
20	A	Rme1	Tph	Kme3	Q	Tph	A	Rme2a	Kac	Sph
21	A	Cit	T	Kme3	Q	Tph	A	Rme2a	Kme3	S
22	A	R	Tph	Kac	Q	Tph	A	R	Kme1	Sph
23	A	R	Tph	Kac	Q	Tph	A	Cit	K	Sph
24	A	Rme1	T	Kac	Q	T	A	Rme2a	Kac	S
25	A	Rme1	Tph	Kac	Q	T	A	Rme1	Kme2	Sph
26	A	Rme1	Tph	Kac	Q	Tph	A	R	K	Sph
27	A	Rme2s	Tph	Kac	Q	T	A	Rme2s	Kme2	Sph
28	A	Rme2s	Tph	Kac	Q	T	A	Rme2s	Kme3	Sph
29	A	Rme2s	Tph	Kac	Q	Tph	A	R	Kac	S
30	A	Rme2s	Tph	Kac	Q	Tph	A	Cit	Kme2	Sph
31	A	Cit	Tph	Kac	Q	T	A	Rme2a	K	Sph
32	A	Cit	Tph	Kac	Q	Tph	A	Rme1	K	Sph
33	A	Cit	Tph	Kac	Q	Tph	A	Rme2s	Kme3	Sph

**Supplementary
Table 4**

ING2-PHD Positive
Hits

	1	2	3	4	5	6	7	8	9	10
1	A	R	T	Kme3	Q	T	A	R	Kme2	S
2	A	R	T	Kme3	Q	T	A	Rme1	K	Sph
3	A	R	T	Kme3	Q	T	A	Rme1	Kme2	S
4	A	R	T	Kme3	Q	T	A	Rme1	Kac	Sph
5	A	R	T	Kme3	Q	T	A	Rme2s	K	S
6	A	R	T	Kme3	Q	T	A	Rme2s	Kme3	Sph
7	A	R	T	Kme3	Q	T	A	Rme2a	Kme2	S
8	A	R	T	Kme3	Q	T	A	Rme2a	Kme2	Sph
9	A	R	T	Kme3	Q	T	A	Rme2a	Kac	S
10	A	R	T	Kme3	Q	T	A	Cit	Kme1	Sph
11	A	R	T	Kme3	Q	T	A	Cit	Kme2	S
12	A	R	T	Kme3	Q	Tph	A	R	Kme3	S
13	A	R	T	Kme3	Q	Tph	A	Rme1	K	S
14	A	R	T	Kme3	Q	Tph	A	Rme2s	Kme1	S
15	A	R	T	Kme3	Q	Tph	A	Rme2s	K	Sph
16	A	R	T	Kme3	Q	Tph	A	Rme2a	K	S
17	A	R	T	Kme3	Q	Tph	A	Rme2a	Kme2	S
18	A	R	Tph	Kme3	Q	T	A	R	Kme2	Sph
19	A	R	Tph	Kme3	Q	T	A	Rme2s	Kme2	S
20	A	Rme1	T	Kme3	Q	T	A	Rme2s	Kme1	S

21	A	Rme1	T	Kme3	Q	T	A	Rme2a	Kme1	Sph
22	A	Rme1	T	Kme3	Q	T	A	Rme2a	Kme2	S
23	A	Rme1	T	Kme3	Q	T	A	Cit	Kme2	Sph
24	A	Rme1	T	Kme3	Q	T	A	Cit	Kme2	Sph
25	A	Rme1	T	Kme3	Q	T	A	Cit	K	Sph
26	A	Rme2s	T	Kme3	Q	T	A	R	Kme3	Sph
27	A	Rme2s	T	Kme3	Q	T	A	Rme1	Kme2	Sph
28	A	Rme2a	T	Kme3	Q	T	A	Rme1	Kme2	S
29	A	Rme2a	T	Kme3	Q	T	A	Cit	Kme3	Sph
30	A	R	T	Kme2	Q	T	A	Rme1	Kac	S
31	A	R	T	Kme2	Q	Tph	A	Cit	K	S
32	A	Rme1	Tph	Kme2	Q	T	A	R	Kme2	S

ING2-PHD Negative

Hits

	1	2	3	4	5	6	7	8	9	10
1	A	R	Tph	Kme3	Q	Tph	A	Cit	K	Sph
2	A	Rme1	Tph	Kme3	Q	Tph	A	Cit	K	Sph
3	A	Cit	Tph	Kme3	Q	Tph	A	Rme2s	Kme1	Sph
4	A	R	Tph	Kme2	Q	Tph	A	Rme2s	Kac	Sph
5	A	Rme1	Tph	Kme2	Q	Tph	A	Cit	K	S
6	A	Rme2s	Tph	Kme2	Q	T	A	Rme2a	Kac	Sph
7	A	Rme2s	Tph	Kme2	Q	Tph	A	Rme2s	K	Sph
8	A	Cit	T	Kme2	Q	T	A	Rme2a	Kac	Sph
9	A	Cit	Tph	Kme2	Q	Tph	A	R	Kme1	Sph
10	A	R	Tph	Kme1	Q	Tph	A	Cit	Kme3	Sph
11	A	Rme1	Tph	Kme1	Q	T	A	Cit	Kac	Sph
12	A	Rme1	Tph	Kme1	Q	Tph	A	Cit	Kme1	Sph
13	A	Rme2s	Tph	Kme1	Q	Tph	A	R	Kac	Sph
14	A	Rme2a	Tph	Kme1	Q	Tph	A	R	Kac	S
15	A	Cit	Tph	Kme1	Q	T	A	R	Kme2	Sph
16	A	Cit	Tph	Kme1	Q	Tph	A	R	Kac	S
17	A	Cit	Tph	Kme1	Q	Tph	A	Rme2a	Kac	S
18	A	Rme1	T	K	Q	Tph	A	Rme1	Kme2	Sph
19	A	R	Tph	Kac	Q	Tph	A	Rme2s	Kac	Sph
20	A	Rme1	T	Kac	Q	Tph	A	Cit	Kme2	Sph
21	A	Rme1	Tph	Kac	Q	Tph	A	Cit	Kme2	S
22	A	Rme2s	Tph	Kac	Q	Tph	A	Rme2s	Kme2	Sph
23	A	Rme2s	Tph	Kac	Q	Tph	A	Cit	Kme3	Sph
24	A	Cit	T	Kac	Q	Tph	A	Rme1	Kme1	S
25	A	Cit	T	Kac	Q	Tph	A	Rme1	Kme2	Sph
26	A	Cit	T	Kac	Q	Tph	A	Cit	Kac	Sph
27	A	Cit	Tph	Kac	Q	T	A	Rme2a	Kac	Sph
28	A	Cit	Tph	Kac	Q	Tph	A	Rme1	Kme1	Sph
29	A	Cit	Tph	Kac	Q	Tph	A	Rme2s	Kac	S
30	A	Cit	Tph	Kac	Q	Tph	A	Rme2s	Kac	Sph
31	A	Cit	Tph	Kac	Q	Tph	A	Rme2s	Kac	Sph

**Supplementary
Table 5**

RAG2-PHD Positive
Hits

	1	2	3	4	5	6	7	8	9	10
1	A	R	T	Kme3	Q	T	A	Rme1	Kme3	Sph
2	A	R	T	Kme3	Q	T	A	Cit	Kme1	S
3	A	R	Tph	Kme3	Q	Tph	A	Rme2a	Kme3	Sph
4	A	Rme1	T	Kme3	Q	T	A	R	K	S
5	A	Rme1	T	Kme3	Q	T	A	R	Kme1	S
6	A	Rme1	T	Kme3	Q	T	A	R	Kme1	S
7	A	Rme1	T	Kme3	Q	T	A	R	Kme2	S
8	A	Rme1	T	Kme3	Q	T	A	R	Kac	S
9	A	Rme1	T	Kme3	Q	T	A	Rme1	K	S
10	A	Rme1	T	Kme3	Q	T	A	Rme1	K	S
11	A	Rme1	T	Kme3	Q	T	A	Rme1	Kme1	S
12	A	Rme1	T	Kme3	Q	T	A	Rme1	Kme1	Sph
13	A	Rme1	T	Kme3	Q	T	A	Rme2s	Kme1	S
14	A	Rme1	T	Kme3	Q	T	A	Cit	K	S
15	A	Rme2s	T	Kme3	Q	T	A	R	Kme1	S
16	A	Rme2s	T	Kme3	Q	T	A	R	Kme1	Sph
17	A	Rme2s	T	Kme3	Q	T	A	Rme2a	Kme2	S
18	A	Rme2a	T	Kme3	Q	T	A	R	K	S
19	A	Rme2a	T	Kme3	Q	T	A	R	Kme1	S
20	A	Rme2a	T	Kme3	Q	T	A	R	Kme2	S
21	A	Rme2a	T	Kme3	Q	T	A	Rme1	Kme1	S
22	A	Rme2a	T	Kme3	Q	T	A	Rme1	Kme2	S
23	A	Rme2a	T	Kme3	Q	T	A	Rme2s	K	S
24	A	Rme2a	T	Kme3	Q	T	A	Rme2s	K	S
25	A	Rme2a	T	Kme3	Q	T	A	Rme2a	K	S
26	A	Rme2a	T	Kme3	Q	T	A	Rme2a	Kac	S
27	A	Rme2a	T	Kme3	Q	T	A	Cit	K	S
28	A	Cit	T	Kme3	Q	T	A	R	K	Sph
29	A	Cit	T	Kme3	Q	T	A	R	Kme2	Sph
30	A	Cit	T	Kme3	Q	T	A	Rme1	Kac	S
31	A	Cit	T	Kme3	Q	T	A	Rme2s	Kme1	S
32	A	Rme1	T	Kme2	Q	T	A	Rme1	Kme2	S

RAG2-PHD Negative
Hits

	1	2	3	4	5	6	7	8	9	10
1	A	R	T	Kme3	Q	T	A	Rme2s	Kac	S
2	A	R	Tph	Kme3	Q	Tph	A	Rme1	Kac	Sph
3	A	Rme2s	Tph	Kme3	Q	Tph	A	Rme2s	Kac	S
4	A	Rme2a	Tph	Kme3	Q	T	A	Rme2s	K	Sph
5	A	Rme2a	Tph	Kme3	Q	T	A	Cit	Kme3	S
6	A	Cit	T	Kme3	Q	Tph	A	Rme2a	Kac	S
7	A	R	Tph	Kme2	Q	T	A	Rme1	Kme1	Sph
8	A	R	Tph	Kme2	Q	Tph	A	Rme1	Kac	Sph
9	A	R	Tph	Kme2	Q	Tph	A	Cit	K	Sph

10	A	Rme2s	Tph	Kme2	Q	T	A	Cit	K	Sph
11	A	Rme2a	T	Kme2	Q	Tph	A	Rme1	Kac	S
12	A	Rme2a	Tph	Kme2	Q	T	A	Rme2a	Kac	Sph
13	A	Cit	Tph	Kme2	Q	Tph	A	Rme2a	Kme3	S
14	A	R	Tph	Kme1	Q	Tph	A	R	K	Sph
15	A	Rme2s	T	Kme1	Q	T	A	Rme2s	K	Sph
16	A	Rme2s	Tph	Kme1	Q	T	A	Rme2a	Kme2	Sph
17	A	Rme2a	Tph	Kme1	Q	Tph	A	Rme2a	Kme3	S
18	A	Cit	Tph	Kme1	Q	T	A	Rme2s	Kme2	Sph
19	A	Cit	Tph	Kme1	Q	T	A	Cit	Kac	S
20	A	R	T	K	Q	Tph	A	Cit	Kac	S
21	A	Rme1	Tph	K	Q	T	A	Rme2s	Kac	Sph
22	A	Rme1	Tph	K	Q	Tph	A	Rme1	K	Sph
23	A	Rme1	Tph	K	Q	Tph	A	Rme1	Kac	S
24	A	Rme2a	Tph	K	Q	Tph	A	Rme2a	K	S
25	A	R	T	Kac	Q	T	A	Rme2s	Kac	Sph
26	A	Rme1	Tph	Kac	Q	T	A	Rme1	Kme3	S
27	A	Rme2s	Tph	Kac	Q	Tph	A	Cit	Kme3	Sph
28	A	Rme2a	Tph	Kac	Q	T	A	Rme2s	Kme1	S
29	A	Rme2a	Tph	Kac	Q	Tph	A	Rme2a	Kme1	S
30	A	Cit	T	Kac	Q	Tph	A	Rme2a	Kme1	Sph
31	A	Cit	Tph	Kac	Q	T	A	R	Kac	S

**Supplementary
Table 6**

JMJD2A-DTD
Positive Hits

	1	2	3	4	5	6	7	8	9	10
1	A	R	T	Kme3	Q	T	A	R	K	S
2	A	R	T	Kme3	Q	T	A	R	Kac	Sph
3	A	R	T	Kme3	Q	Tph	A	R	Kme1	Sph
4	A	R	T	Kme3	Q	Tph	A	R	Kac	Sph
5	A	R	T	Kme3	Q	Tph	A	Rme1	Kme2	Sph
6	A	R	T	Kme3	Q	Tph	A	Rme2s	Kme2	S
7	A	R	T	Kme3	Q	Tph	A	Rme2s	Kme3	Sph
8	A	R	T	Kme3	Q	Tph	A	Rme2s	Kac	S
9	A	Rme1	T	Kme3	Q	T	A	R	Kme2	Sph
10	A	Rme1	T	Kme3	Q	T	A	Rme1	K	S
11	A	Rme1	T	Kme3	Q	T	A	Rme2a	K	Sph
12	A	Rme1	T	Kme3	Q	T	A	Rme2a	Kme3	Sph
13	A	Rme1	T	Kme3	Q	Tph	A	R	Kme1	Sph
14	A	Rme1	T	Kme3	Q	Tph	A	R	Kme2	S
15	A	Rme1	T	Kme3	Q	Tph	A	Rme2s	K	Sph
16	A	Rme1	T	Kme3	Q	Tph	A	Rme2s	K	Sph
17	A	Rme1	Tph	Kme3	Q	T	A	R	Kme3	S
18	A	Rme2s	T	Kme3	Q	T	A	Rme1	Kac	S
19	A	Rme2s	T	Kme3	Q	T	A	Rme2a	Kme2	Sph

20	A	Rme2s	T	Kme3	Q	Tph	A	R	Kme1	S
21	A	Rme2s	T	Kme3	Q	Tph	A	R	Kme2	S
22	A	Rme2a	T	Kme3	Q	Tph	A	Rme2a	Kme3	S
23	A	Cit	T	Kme3	Q	T	A	R	K	Sph
24	A	Cit	T	Kme3	Q	T	A	Rme1	Kme2	Sph
25	A	Cit	T	Kme3	Q	T	A	Rme1	Kme2	Sph
26	A	Cit	T	Kme3	Q	Tph	A	Rme2a	Kme1	S
27	A	R	T	Kme2	Q	Tph	A	R	Kme3	S
28	A	Rme2a	T	Kme2	Q	T	A	R	Kme2	Sph

JMJD2A-DTD Negative Hits

	1	2	3	4	5	6	7	8	9	10
1	A	Rme2s	Tph	Kme2	Q	Tph	A	R	Kme3	Sph
2	A	Rme2s	Tph	Kme2	Q	Tph	A	Cit	Kme2	S
3	A	Rme2a	T	Kme2	Q	T	A	Cit	K	S
4	A	Rme2a	Tph	Kme2	Q	T	A	Cit	Kme1	S
5	A	R	Tph	Kme1	Q	Tph	A	R	Kme3	Sph
6	A	Rme1	Tph	Kme1	Q	Tph	A	R	Kme3	Sph
7	A	Rme1	Tph	Kme1	Q	Tph	A	Rme2s	K	Sph
8	A	Rme2s	Tph	Kme1	Q	T	A	Rme1	Kme1	Sph
9	A	Rme2s	Tph	Kme1	Q	T	A	Cit	Kme1	Sph
10	A	Rme2s	Tph	Kme1	Q	Tph	A	R	Kac	Sph
11	A	Rme2a	Tph	Kme1	Q	T	A	R	Kme1	S
12	A	Rme2a	Tph	Kme1	Q	Tph	A	Rme2s	Kme3	Sph
13	A	Rme2a	Tph	Kme1	Q	Tph	A	Cit	Kac	Sph
14	A	R	Tph	K	Q	T	A	R	Kac	S
15	A	R	Tph	K	Q	Tph	A	Rme2s	Kac	Sph
16	A	Rme1	T	K	Q	T	A	R	Kac	Sph
17	A	Rme2s	T	K	Q	Tph	A	Rme2a	Kme2	Sph
18	A	Rme2s	Tph	K	Q	Tph	A	R	Kme1	Sph
19	A	Rme2s	Tph	K	Q	Tph	A	R	Kme3	Sph
20	A	R	Tph	Kac	Q	Tph	A	Rme2a	Kme1	Sph
21	A	Rme1	Tph	Kac	Q	Tph	A	Rme1	Kme1	S
22	A	Rme2s	Tph	Kac	Q	Tph	A	Cit	Kac	Sph
23	A	Cit	T	Kac	Q	T	A	R	K	Sph
24	A	Cit	T	Kac	Q	Tph	A	R	Kac	Sph
25	A	Cit	T	Kac	Q	Tph	A	Rme2s	Kme2	Sph
26	A	Cit	Tph	Kac	Q	T	A	Rme1	Kme3	Sph
27	A	Cit	Tph	Kac	Q	T	A	Rme1	Kac	Sph

SI Table 7**K_d Values for H3K4unmod Binding Modules determined by ITC**

Peptide	BHC80- PHD	AIRE- PHD1
H3K4unmod.	19.6 ± 3.6 μM	1.71 ± 0.2 μM
H3T3ph	N.D.B (480 μM) ^a	N.D.B (480 μM) ^a
H3T6ph	N.D.B (1 mM) ^a	N.D.B (1 mM) ^a
H3R2me2s	57.2 ± 4.9 μM	

K_d Values for H3K4me3 Binding Modules determined by ITC

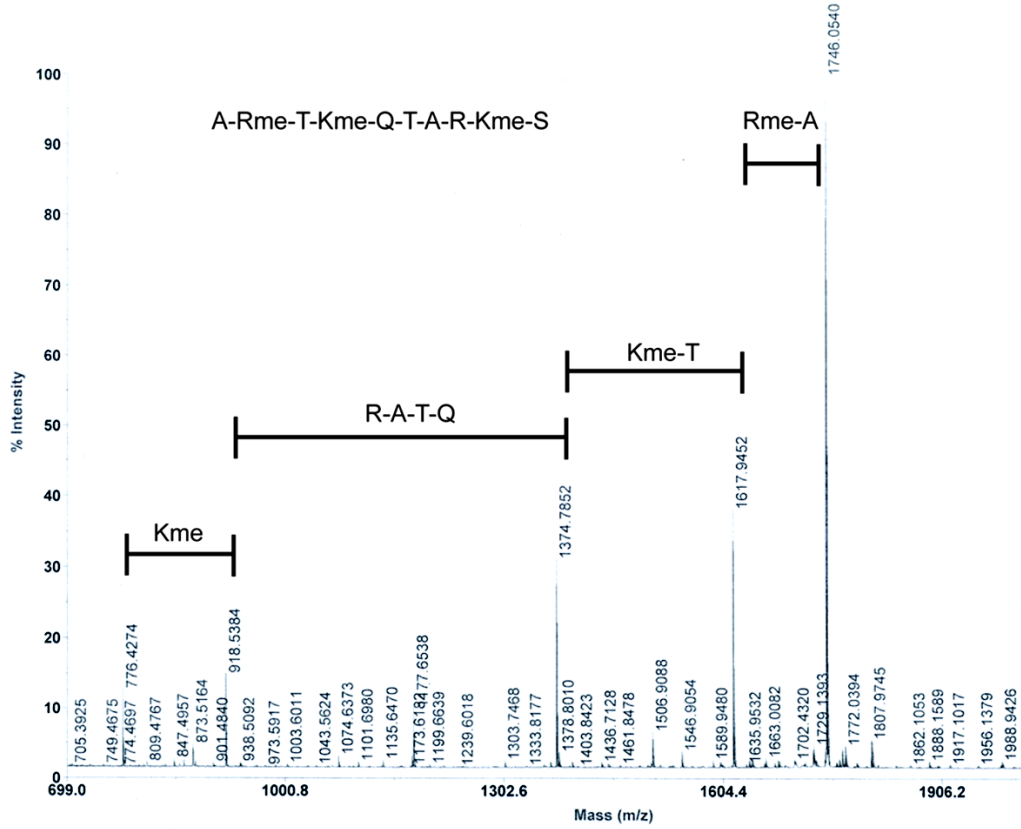
Peptide	ING2- PHD finger	RAG2- PHD finger	JMJD2A-DTD
H3K4me3	0.98 ± 0.1 μM	17.8 ± 2.1 μM	1.1 ± 0.1 μM
H3K4me3T3ph	N.D.B (500 μM) ^a	N.D.B (500 μM) ^a	N.D.B (2.9 mM) ^a
H3K4me3T6ph	19.9 ± 4.9 μM	N.D.B (750 μM) ^a	1.8 ± 0.2 μM
H3K4me3R2me2s	17.3 ± 2.3 μM		

N.D.B = No Detectable Binding

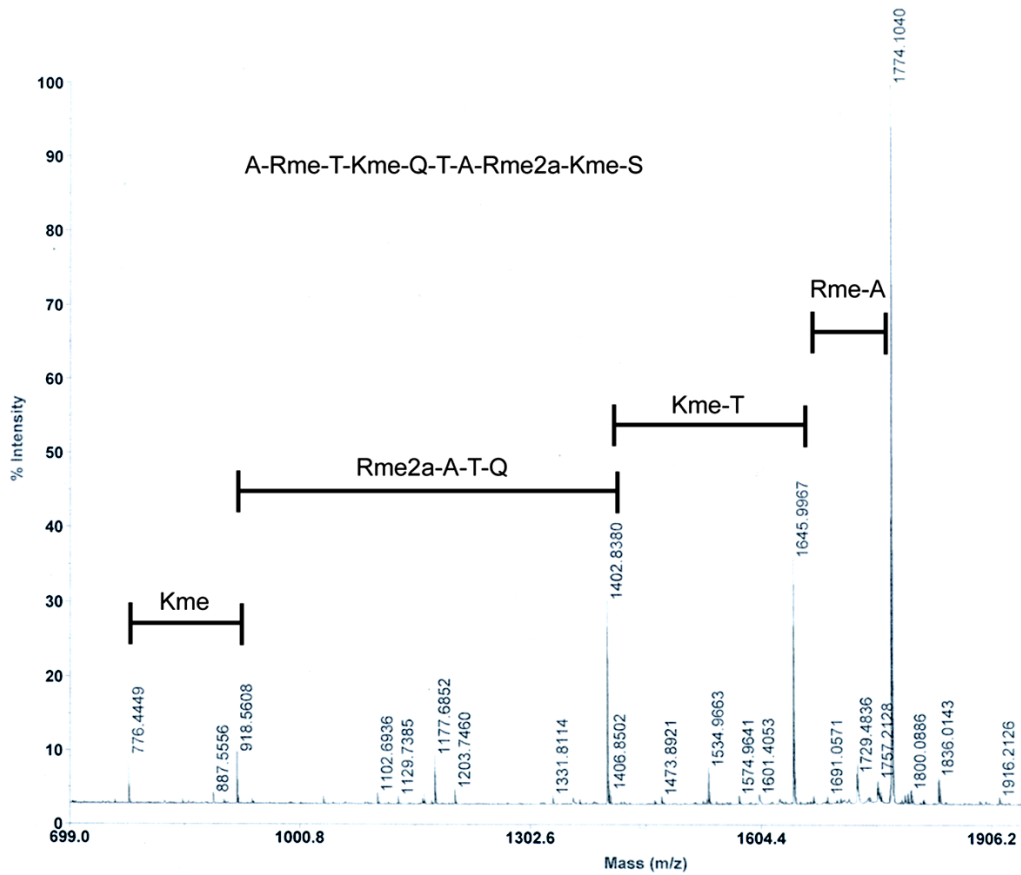
^a highest concentration of peptide tested

SI Fig. 1

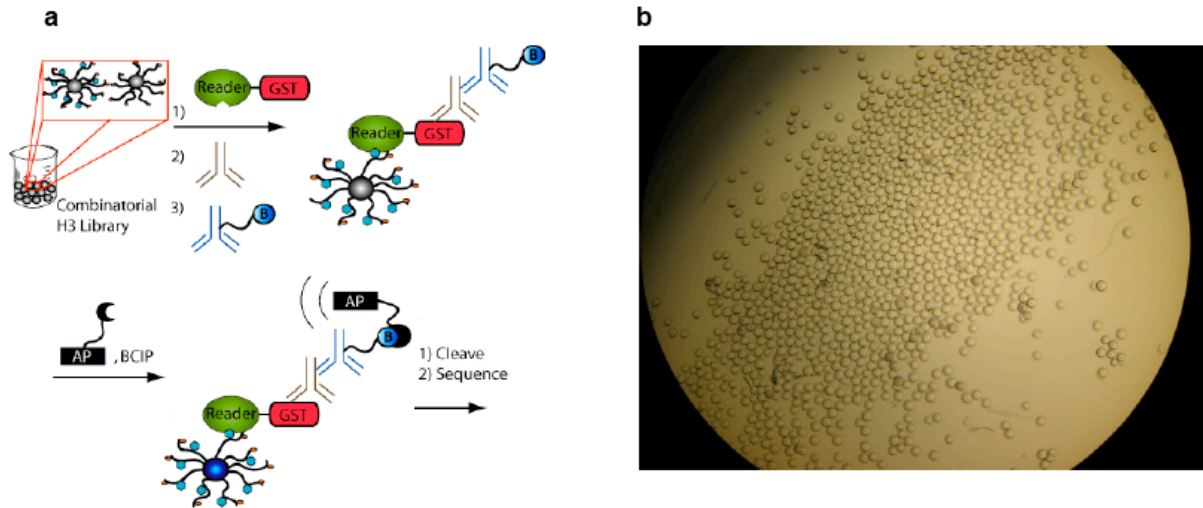
a



b

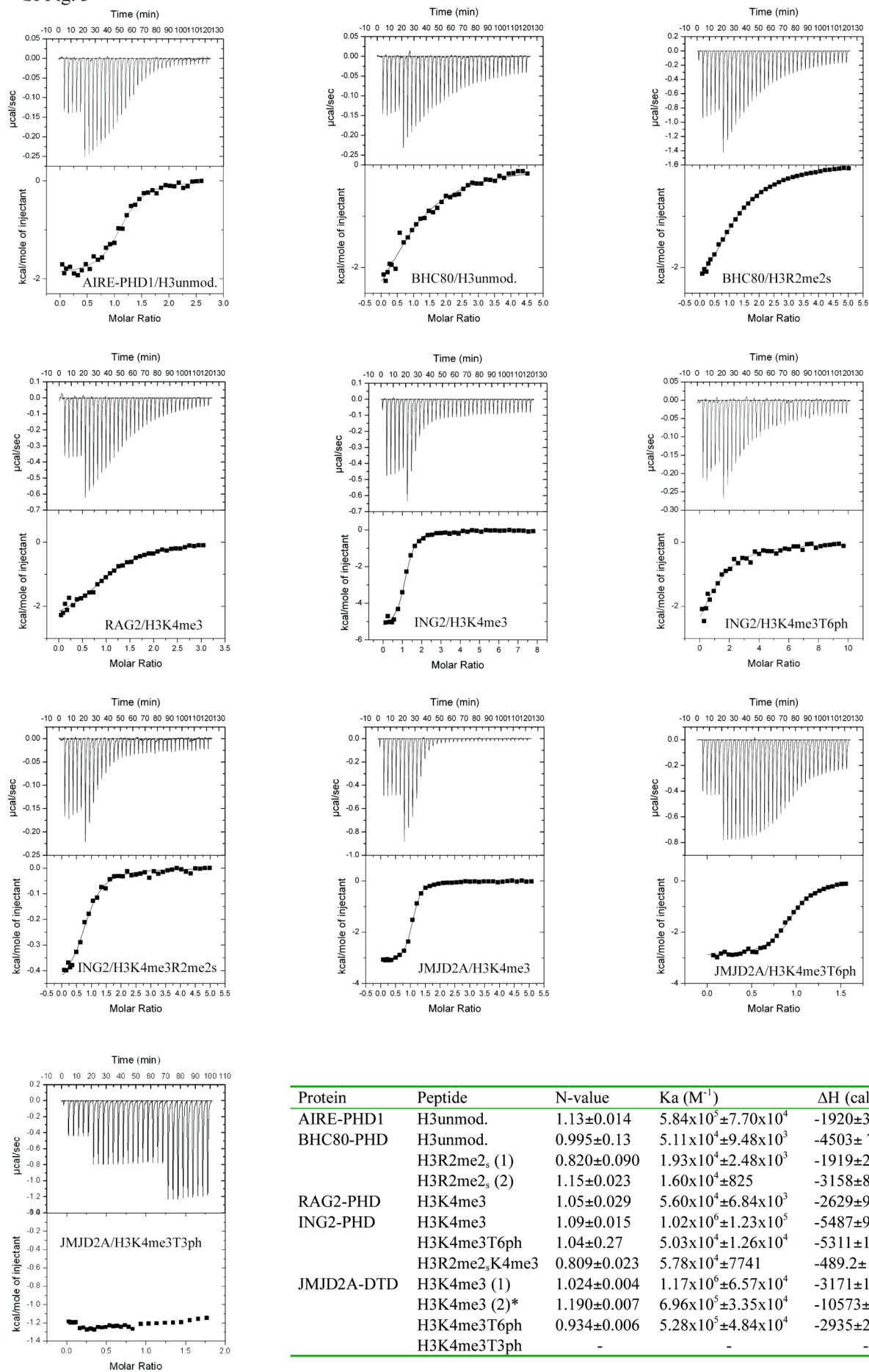


SI Fig 1. MS mass ladder created by the library capping strategy. (a) The peak at 1746.054 m/z represents the full length H3 tail from this particular bead. The difference amino acid Ala (A1) and monomethylated-arginine (R2me). The difference between 1617.95 m/z and the next capped species (1374.78 m/z) shows unphosphorylated threonine (T3) and monomethylated lysine (K4me). The next two capped species in this spectrum reveal unmodified threonine (T6), unmodified arginine (R8) and monomethylated-lysine (K9). (b) This spectrum represents a peptide with similar identity to that in *a* except that R8 is asymmetrically dimethylated, as opposed to unmodified. The addition of this PTM is reflected in a different m/z for two of the capped species, (1402.83 m/z and 1645.99 m/z) and a full-length product that differs in mass by two methyl groups (28.05 m/z).



SI Fig 2. On-bead assay for H3 library screening. (a) After blocking with bovine serum albumin for nonspecific interactions, the H3 library is incubated with a GST-fusion of the protein of interest (reader). Afterwards, the library is treated with a GST-specific primary antibody followed by a biotinylated secondary antibody. Subsequent colorimetric development with SAAP and BCIP results in turquoise staining of beads bearing peptides that interact with the target protein. (b) A portion of the library screened with GST to ensure minimal non-specific binding incurs no color change after 40 minutes of BCIP development.

SI Fig. 3

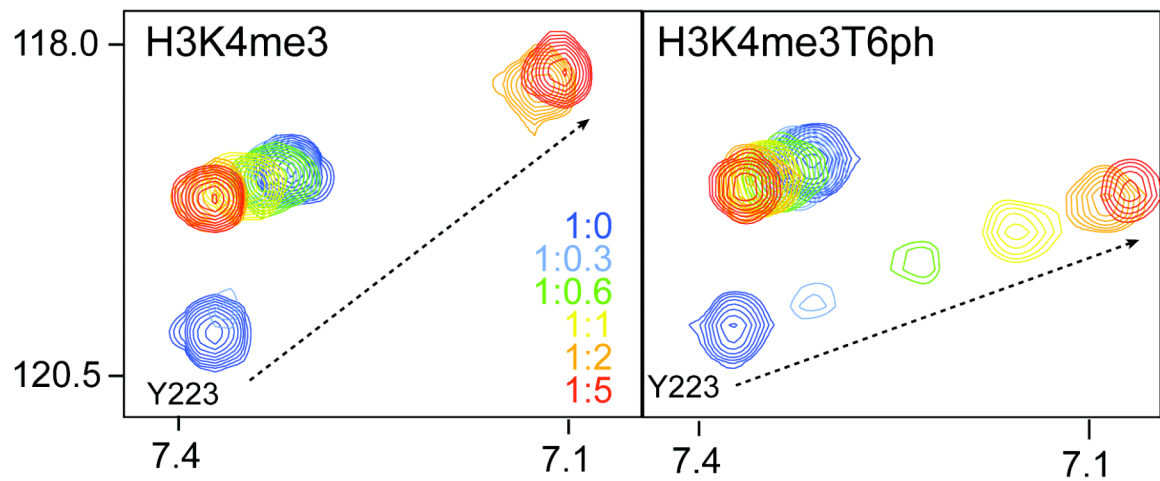


Protein	Peptide	N-value	K_a (M^{-1})	ΔH (cal/mol)
AIRE-PHD1	H3unmod.	1.13 ± 0.014	$5.84 \times 10^5 \pm 7.70 \times 10^4$	-1920 ± 31.4
BHC80-PHD	H3unmod.	0.995 ± 0.13	$5.11 \times 10^4 \pm 9.48 \times 10^3$	-4503 ± 744.9
	H3R2me _{2s} (1)	0.820 ± 0.090	$1.93 \times 10^4 \pm 2.48 \times 10^3$	-1919 ± 258.8
	H3R2me _{2s} (2)	1.15 ± 0.023	$1.60 \times 10^4 \pm 825$	-3158 ± 81.7
RAG2-PHD	H3K4me3	1.05 ± 0.029	$5.60 \times 10^4 \pm 6.84 \times 10^3$	-2629 ± 96.5
ING2-PHD	H3K4me3	1.09 ± 0.015	$1.02 \times 10^6 \pm 1.23 \times 10^5$	-5487 ± 90.4
	H3K4me3T6ph	1.04 ± 0.27	$5.03 \times 10^4 \pm 1.26 \times 10^4$	-5311 ± 1630
	H3R2me _{2s} K4me3	0.809 ± 0.023	$5.78 \times 10^4 \pm 7741$	-489.2 ± 17.2
JMJD2A-DTD	H3K4me3 (1)	1.024 ± 0.004	$1.17 \times 10^6 \pm 6.57 \times 10^4$	-3171 ± 14.7
	H3K4me3 (2)*	1.190 ± 0.007	$6.96 \times 10^5 \pm 3.35 \times 10^4$	-10573 ± 109.0
	H3K4me3T6ph	0.934 ± 0.006	$5.28 \times 10^5 \pm 4.84 \times 10^4$	-2935 ± 25.2
	H3K4me3T3ph	-	-	-

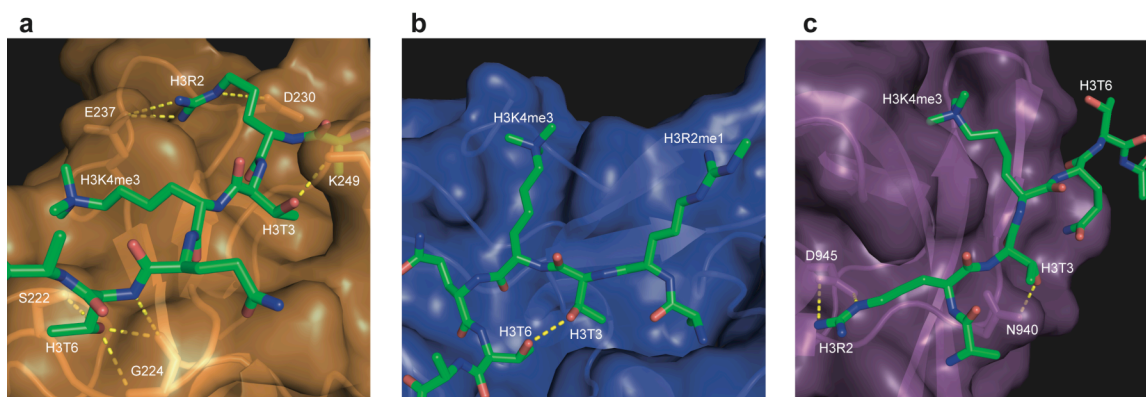
SI Fig 3. Isothermal titration calorimetry (ITC) for modules bound to H3 peptides.

The experimental curves are represented in the top panel and the corresponding enthalpy plots are shown in the bottom panel for each individual experiment. The protein and peptide are labeled in the lower panel. A summary of the thermodynamic and curve fitting parameters are listed in the bottom table. Titrations of H3R2me2s into the BHC80-PHD and the H3K4me3 peptide into JMJD2A-DTD are shown for two separate experiments (labeled as (1) and (2)) in the table and a representative curve/enthalpy plot is displayed for each. The K_d values for these titrations are averaged using both experiments (Table 1 and 2 in the text). Titrations were performed at protein concentrations from 20-350 μ M and peptide concentrations from 0.5-2.8 mM.

* The variability in these separate experiments is likely due to the fact that these two experiments were performed with different preparations of JMJD2A and a different synthetic preparation of H3K4me3 peptide. Although the purity was similar between preparations, we observed 3-fold higher heats of dilution with the peptide that yielded \sim 10,000 cal/mol, suggesting that accompanying salts/counterions may have contributed to the differences in molar heats.



SI Fig 4. ^1H , ^{15}N HSQC NMR titrations. Six superimposed ^1H , ^{15}N HSQC spectra of the ING2 PHD finger (0.2 mM) collected during addition of the H3K4me3T6ph (*a*) or H3K4me3 (*b*) peptides are color-coded according to the peptide concentrations (inset).



SI Fig 5. H3 binding-module cocrystal structures. Representations of previously solved co-crystal structures of a panel of H3K4me3 binding modules illustrate the hydrogen bonding and electrostatic contacts made between H3 side chain and the various effector domains. Space filling models of the ING2-PHD finger (A, orange, 2G6Q)⁸, the RAG2-PHD finger (B, blue, 2V85)⁹, and the JMJD2A DTD (C, purple, 2GFA)¹⁰ are shown binding H3 peptides. Carbon, nitrogen and oxygen atoms are shown in green, blue and red, respectively for the H3 peptides.

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