

Specific sides to multifaceted glycosaminoglycans are observed in embryonic development

Kenneth L. Kramer

Overview of Glycosaminoglycan Biosynthesis

A short summary of GAG polymerization and modification follows. For more detailed discussion, several excellent reviews should be consulted [1-3].

The longest vertebrate polysaccharide, HA is composed of *N*-acetylglucosamine (GlcNAc) and glucuronic acid (GlcA) repeated to about 10,000 disaccharides (Fig 1). The structure is characterized by a 4-fold helix that is stabilized by weak intramolecular hydrogen bonds, giving HA its well-characterized viscoelastic properties that are critical to its function in eyes and joints [4]. HA synthesis is accomplished by one of three co-polymerases, glycosyltransferases which transfer both monosaccharides to the elongating chain (Supplemental Table). In contrast to the other GAGs, HA is not modified by sulfation, suggesting that it might represent an early evolutionary GAG. However, invertebrates like *Drosophila* or *C. elegans* do not contain homologs of the vertebrate HA co-polymerases.

Galactose (Gal) and GlcNAc are repeated in KS for about 10-50 disaccharide units and are sulfated at the 6-*O* position on both saccharides. KS is not found in invertebrates; a single copy of each polymerizing and modifying enzyme is found in vertebrates. KS can be distinguished from other sulfated GAGs based on its linkages to the core protein, elongation method, and additional modifications. There are two types of KS core protein linkages: KS I (or corneal KS) is found on an *N*-glycan core structure while KS II (or skeletal KS) links via an *O*-glycan core structure, both of which are commonly observed in secreted and membrane-bound glycoproteins. KS polymerization is blocked when one of two closely related sulfotransferases is absent, *Chst2* in brain [5, 6] and *Chst6* in cornea [7], indicating that KS synthesis requires alternating polymerization with 6-*O* sulfation [8]. Finally, the KS chain is not just modified by sulfation, but fucose and sialic acid as well [9].

Invertebrates synthesize CS, but it is not sulfated in *Drosophila* and is undersulfated in *C. elegans* [10]. Addition of *N*-acetylgalactosamine (GalNAc) initiates synthesis of CS on a linker tetrasaccharide that is shared with HS. In vertebrates, CS/DS chain initiation and elongation involves two GalNAc glycosyltransferases and a family of four co-polymerases that each possess GalNAc- and GluA-transferase activity, yet each co-polymerase interacts with another co-polymerase to achieve optimal polymerization [11]. Sulfation at 6-*O* GalNAc occurs in the medial/trans golgi apparatus; epimerizing GlcA to IdoA is dependent on sulfation at the 4-*O*-position and likely takes place later in the golgi [12, 13]. The 2-*O* sulfotransferase prefers 6-*O* and 4-*O* modified CS as substrates, indicating that it is likely the last enzyme in the pathway [14].

HS is the only GAG sulfated in *Drosophila* or *C. elegans* at the same positions as it is in vertebrates [10]. After addition of GlcNAc to the linker tetrasaccharide, GlcA and GlcNAc are polymerized by alternately adding to the nonreducing end by a family of Ext copolymerases. After

elongation or as the chain is elongating, the *Ndst* family of enzymes remove the *N*-acetyl group from selected GlcNAc residues and substitute most of the free amines with sulfates. Subsequent epimerization and sulfation occurs primarily in the *N*-sulfated regions. While *Drosophila* and *C. elegans* only have one gene for each enzymatic step, a similar array of disaccharides is produced [10].

Gene	Description	Human	Mouse	Zebrafish	<i>Drosophila</i>	<i>C. Elegans</i>		
HA								
<i>HAS1</i>	Polymerizes the alternate addition of GlcA & GlcNAc	3036	15116	403130				
<i>HAS2</i>		3037	15117	260350				
<i>HAS3</i>		3038	15118	282555				
KS								
<i>B4GALT4</i>	Adds Gal to KS	8702	56375	492760				
<i>B3GNT7</i>	Adds GlcNAc to KS	93010	227327	286748				
<i>CHST6</i>	Sulfates GlcNAc at the nonreducing end at the C6 hydroxy	4166	56773	100170782				
<i>CHST2</i>		9435	54371	561896				
<i>CHST1</i>		8534	76969	445124				
CSIDS								
Chain initiation and elongation								
<i>CSGALNACT1</i>	Adds GalNAc to linkage oligosaccharide	55790	234356	793671	36079			
<i>CSGALNACT2</i>		55454	78752	560826				
<i>CHSY1</i>	Polymerizes the alternate addition of GlcA & GalNAc	22856	269941	324407	32497	172851		
<i>CHSY3</i>		337876	78923	368477				
<i>CHPF</i>		Cooperates with CHSY for chondroitin polymerization	79586	74241	794660	32259	176231	
<i>CHPF2</i>	54480		100910	100000654				
Chain modification								
<i>CHST3</i>	Sulfates GalNAc at the C6 hydroxy	9469	53374	556800				
				559721				
<i>CHST7</i>	Sulfates GalNAc at the C6 hydroxy	56548	60322	563748				
<i>CHST15</i>	Sulfates GalNAc4S at the C6 hydroxy	51363	77590	565523				
<i>DSE</i>	Epimerizes GlcA to IdoA	29940	212898	568713				
<i>DSEL</i>		92126	319901	568099				
<i>CHST14</i>	Sulfates GalNAc in DS at the C4 hydroxy	113189	72136	404732				
<i>CHST11</i>	Sulfates GalNAc at the C4 hydroxy	50515	58250	404232	326156			
<i>CHST12</i>		55501	59031	407076				
<i>CHST13</i>		166012	71797	565671				
<i>UST</i>	Sulfates GlcA at the C2 hydroxy	10090	338362	557218				
HS								
Chain initiation and elongation								
<i>EXTL3</i>	Adds GlcNAc to linkage oligosaccharide	2137	54616	493783	37198	176502		
<i>EXTL2</i>		2135	58193	558373				
<i>EXTL1</i>		2134	56219					
<i>EXT1</i>	Polymerizes the alternate addition of GlcA & GlcNAc	2131	14042	497279	36614	178080		
				497280				
<i>EXT2</i>		2132	14043	497281				
<i>EXT2</i>				493780	3772101			
Chain modification								
<i>NDST1</i>	Deacetylates and sulfates GlcNAc	3340	15531	62397*	38736	177675		
				74936*				
<i>NDST2</i>				8509			17423	798226
<i>NDST3</i>		9348	83398	100317129				
<i>NDST4</i>		64579	64580					
<i>GLCE</i>	Epimerizes GlcA to IdoA	26035	93683	405776	35569	175562		
				100007670				
<i>HS2ST1</i>	Sulfates GlcA and IdoA at the C2 hydroxy	9653	23908	791188	44433	181309		
<i>HS6ST1</i>	Sulfates GlcNAc at the C6 hydroxy	9394	50785	553162	42380	189586		
				791107				
<i>HS6ST2</i>		90161	50786	378450				
<i>HS6ST3</i>		266722	50787	569353				
<i>HS3ST1</i>	Sulfates GlcNAc at the C3 hydroxy	9957	15476	792524				
<i>HS3ST2</i>				557111				
<i>HS3ST3</i>				9956	195646	571246		
				9953	15478	558084	32918	3564879
				9955	54710	558084		
<i>HS3ST4</i>		9951	628779	563570				
<i>HS3ST5</i>		222537	319415	562344	37161	185573		
				565201				
<i>HS3ST6</i>		64711	328779	100008592				

Supplemental Table: Diversity of GAG biosynthetic enzymes from 4 model organisms compared to humans. Gene IDs from NCBI's Entrez Gene database were used to consistently identify glycosyltransferases and sulfotransferases as there are numerous inconsistencies between and within online databases. While the enzymatic activity for many gene products has not been fully characterized,

identification is straightforward as the open reading frame covers few exons and significant homologies exist, particularly in the active sites. The vast majority of genes listed here are specific to GAG synthesis, but there are exceptions. For example, the glycosyltransferase B3GAT1 recognizes a wide substrate specificity that includes branched N- and O-glycans [15] while the sulfotransferase CHST3 has been shown to modify GalNAc in CS along with Gal in KS and in the HS-CS linker tetrasaccharide [16].

* Ensembl gene ID.

References (cited in the supplemental material)

- [1] Prabhakar V, Sasisekharan R. The biosynthesis and catabolism of galactosaminoglycans. *Adv Pharmacol* 2006;53:69-115.
- [2] Esko JD, Selleck SB. ORDER OUT OF CHAOS: Assembly of Ligand Binding Sites in Heparan Sulfate1. *Annual Review of Biochemistry* 2002;71:435-71.
- [3] Laurent TC, Fraser JR. Hyaluronan. *Faseb J* 1992;6:2397-404.
- [4] Almond A, Deangelis PL, Blundell CD. Hyaluronan: the local solution conformation determined by NMR and computer modeling is close to a contracted left-handed 4-fold helix. *J Mol Biol* 2006;358:1256-69.
- [5] Zhang H, Muramatsu T, Murase A, Yuasa S, Uchimura K, Kadomatsu K. N-Acetylglucosamine 6-O-sulfotransferase-1 is required for brain keratan sulfate biosynthesis and glial scar formation after brain injury. *Glycobiology* 2006;16:702-10.
- [6] Ito Z, Sakamoto K, Imagama S, Matsuyama Y, Zhang H, Hirano K, et al. N-Acetylglucosamine 6-O-Sulfotransferase-1-Deficient Mice Show Better Functional Recovery after Spinal Cord Injury. *J Neurosci* 2010;30:5937-47.
- [7] Kitayama K, Hayashida Y, Nishida K, Akama TO. Enzymes responsible for synthesis of corneal keratan sulfate glycosaminoglycans. *J Biol Chem* 2007;282:30085-96.
- [8] Seko A, Yamashita K. Biosynthesis of Keratan Sulfate. *Experimental Glycoscience* 2008. p. 67-9.
- [9] Fosang AJ, Last K, Poon CJ, Plaas AH. Keratan sulphate in the interglobular domain has a microstructure that is distinct from keratan sulphate elsewhere on pig aggrecan. *Matrix Biol* 2009;28:53-61.
- [10] Lawrence R, Olson SK, Steele RE, Wang L, Warrior R, Cummings RD, et al. Evolutionary differences in glycosaminoglycan fine structure detected by quantitative glycan reductive isotope labeling. *J Biol Chem* 2008;283:33674-84.
- [11] Izumikawa T, Koike T, Shiozawa S, Sugahara K, Tamura J-i, Kitagawa H. Identification of Chondroitin Sulfate Glucuronyltransferase as Chondroitin Synthase-3 Involved in Chondroitin Polymerization. *Journal of Biological Chemistry* 2008;283:11396-406.
- [12] Hoppe W, Glossl J, Kresse H. Influence of monensin on biosynthesis, processing and secretion of proteodermatan sulfate by skin fibroblasts. *Eur J Biochem* 1985;152:91-7.
- [13] Pacheco B, Maccarana M, Malmstrom A. Dermatan 4-O-sulfotransferase 1 is pivotal in the formation of iduronic acid blocks in dermatan sulfate. *Glycobiology* 2009;19:1197-203.
- [14] Ohtake S, Kimata K, Habuchi O. Recognition of sulfation pattern of chondroitin sulfate by uronosyl 2-O-sulfotransferase. *J Biol Chem* 2005;280:39115-23.
- [15] Fondeur-Gelinotte M, Lattard V, Gulberti S, Oriol R, Mulliert G, Coughtrie MW, et al. Molecular basis for acceptor substrate specificity of the human beta1,3-glucuronosyltransferases GlcAT-I and GlcAT-P involved in glycosaminoglycan and HNK-1 carbohydrate epitope biosynthesis, respectively. *Glycobiology* 2007;17:857-67.
- [16] Kitagawa H, Tsutsumi K, Ikegami-Kuzuhara A, Nadanaka S, Goto F, Ogawa T, et al. Sulfation of the Galactose Residues in the Glycosaminoglycan-Protein Linkage Region by Recombinant Human Chondroitin 6-O-Sulfotransferase-1. *Journal of Biological Chemistry* 2008;283:27438-43.