

SUPPLEMENTAL DATA

The following materials are available in the online version of this article.

Supplemental Figure S1. RPL10 sequences are highly conserved between different organisms at nucleotide and amino acid levels. Alignment of the nucleotide (A), coding (B) and amino acid (C) sequences of *Zea mays* and *A. thaliana* RPL10s. Start and stop codons are bold letters. D, Multiple sequence alignment of RPL10 proteins from different species. For protein accession numbers, see Methods. At, *A. thaliana*; Zm, *Zea mays*. The sequences were aligned using the Clustal W2 program. Dashes (-) indicate spaces introduced to promote optimal alignment, perfect matches are represented by an asterisk (*), high amino-acid similarities by double dots (:), and weak similarities by a single dot (.). Symbols are indicated below the sequences. Signature sequences (motifs) specific to Amidation are shaded grey, specific to Glycosylation pink, specific to Myristylation yellow, specific to protein kinase C light blue and specific to casein kinase II green. Putative zif domain is bold-underlined.

Supplemental Figure S2. Coimmunoprecipitation of RPL10 proteins in *A. thaliana*. A, Immunoblot analysis of *A. thaliana* recombinant RPL10 proteins. Partially purified recombinant RPL10 proteins were run on 12% SDS-PAGE and subjected to immunoblot analysis for RPL10. Ten micrograms of total proteins were loaded in all lanes. B, SDS-PAGE (10%) of RPL10-associated proteins. C, Classification of RPL10-associated proteins based on their cell functions. Proteins with percentage of coverage higher than 10% or at least two tryptic peptides were included in the diagram. Clustering was performed according to Usadel et al. (2006). D, Immunoblot analysis of RPL10-associated proteins. RPL10 proteins were immunoprecipitated from *A. thaliana* crude extracts with antibodies against *H. sapiens* QM protein. The immunocomplexes were solubilized, run on 12% SDS-PAGE and subjected to immunoblot analysis for RPL10, eukaryotic translation initiation factor 2 alpha (eIF2 alpha) and eukaryotic translation initiation factor 2 beta (eIF2 beta). The numbers indicate the molecular mass in kDa. CE: crude extract, IP: immunoprecipitate.

Supplemental Figure S3. Complementation of *A. thaliana* homozygous *rpl10B* mutants with WT At *RPL10B*. A, Presence of WT *RPL10B* transcript in transformed *A. thaliana rpl10B* mutant plants analyzed by PCR on genomic DNA. Lanes 1: negative control (without DNA); lane 2-4: genomic DNA from leaves of transformed plants; lanes 5: positive control (pCHF3-

RPL10B). B, At *RPL10B* expression level in Arabidopsis WT, *rpl10B* homozygous and complemented plants analyzed by RT-qPCR. Each reaction was normalized using the C_t values corresponding to the *POLYUBIQUITIN10* mRNA. The means of the results obtained using three independent biological experiments are shown, the error bars indicate the S.D. of the samples. WT levels were set at 1. C, 15-day-old WT (left), *rpl10B* mutant (middle) and complemented plants. Scale bar: 1 cm.

Supplemental Figure S4. Inhibition of protein synthesis by UV-B in *A. thaliana* WT and *rpl10* mutant plants. Forty micrograms of total proteins were resolved by 12% SDS-PAGE after *in vivo* [³⁵S]Met labeling, visualized by autoradiography (A) and staining with Coomassie Blue (B) following the UV-B treatment and recovery period indicated.

Supplemental Figure S5. UV-B treatment is not lethal to *A. thaliana* plants. Chlorophyll a (A), Chlorophyll b (B), Flavonoids (C), Maximum Efficiency of PSII (D) and Total proteins (E) were measured after 4 h UV-B (4 h UV-B), 16 h post-treatment (16 h recovery) and in untreated controls (no UV-B). Measurements are the average of six adult leaves from four different plants. Statistical differences from the control are marked with an asterisk (P<0.05).

Supplemental Figure S6. Typical 2D gels of leaves from heterozygous *rpl10A-1* mutant and WT plants after a 4 h UV-B treatment. As examples of proteins with differential expression, the relative abundances of some but not all spots annotated by the number that appears in Supplemental Table S3 are shown. The graphs represent one example from at least three different gels used for the differential analysis. The first dimension was carried out using 17 cm immobilized pH gradient strips (pH 3–10); acidic side to the left; and the second dimension was on 12.5% (w/v) SDS-PAGE. The relative abundance of proteins was determined. The protein spots with changes in intensities (least 1.5-fold, P <0.05) were considered to be different.

Supplemental Figure S7. Hierarchical cluster analysis of proteins showing different levels in *rpl10A* mutant plants in comparison to WT plants under control conditions and after a 4 h UV-B treatment identified by MS. A, Proteins included show different levels in *rpl10A* mutants (at least 1.5-fold) in comparison to WT plants under control or UV-B conditions. B,

Proteins included show differential abundance (at least 1.5-fold) after a UV-B treatment; these proteins changed differentially in WT plants than in the *rpl10A* mutant. Red indicates higher protein levels than the reference, green indicates lower protein levels than the reference, and black indicates no significant change.

Supplemental Figure S8. Classification of proteins showing different levels in the *rpl10A* mutant in comparison to WT plants based on their cell functions. Proteins were identified by 2D Gel electrophoresis and those showing changes in abundances of at least 1.5-fold were included. A, Proteins changed in the *rpl10A* mutant under control (no UV-B) conditions. B, Proteins changed in the *rpl10A* mutant after 4 h of UV-B.

Supplemental Figure S9. *RPL10s* promoter sequences with predicted cis-elements. The transcription initiation site (referred to as +1) is indicated in bold letter and the ATG start codon is shown in bold and underlined letters. Numbers at the left refer to the positions of nucleotides relative to the putative transcription initiation site.

Supplemental Figure 1. RPL10 sequences are highly conserved between different organisms at nucleotide and amino acid levels. Alignment of the nucleotide (A), coding (B) and amino acid (C) sequences of *Zea mays* and *A. thaliana* RPL10s. D, Multiple sequence alignment of RPL10 proteins from different species.

A

AtRPL10B	TTGCTTGTAAACA-----	279
AtRPL10C	TTGCTTGCACAA-----	263
AtRPL10A	TTGCTTGCACAA-----	276
ZmRPL10-1	TTGCCTGCAACAAAGTACATGACCAAGTCTGCAGGAAGGATGCCTCACCTTAGGGTCC	599
ZmRPL10-2	TTGCCTGCAACAA-----	321
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AtRPL10B	-----AGTACATGGTGAAGTCTGCCGGAAAGATGCGTTCATCTCCGTATTA	327
AtRPL10C	-----AGTATATGGTCAAATCTGCTGGAAAAGATGCTTTCATTTGAGGATTA	311
AtRPL10A	-----AGTACATGGTGAAGTCTGCTGGAAAAGATGCTTTCATTTGAGGATTA	324
ZmRPL10-1	GTGCCTGCAACAAAGTACATGACCAAGTCTGCAGGAAGGATGCCTCACCTTAGGGTCC	659
ZmRPL10-2	-----AGTACATGGTGAAGTCTGCTGGAAAAGGATGCCTCACCTCGGGTCC	369
	***** *	
AtRPL10B	GAGTTCATCCTTCCATGTTCTTAGGATCAAATAAGATGCTTCTTGTCGGAGCTGATA	387
AtRPL10C	GGGTTCATCCTTCCATGTTCTCAGGATTAACAAGATGCTTCTGTCGGAGCTGATA	371
AtRPL10A	GGGTTCATCCTTCCATGTTCTCAGGATTAACAAGATGCTTCTGTCGGAGCTGATA	384
ZmRPL10-1	GGGTTCACCCGTTCCATGTCCTCCGATCAACAAGATGCTTCTGTCGGGGCTGATA	719
ZmRPL10-2	GGGTTCACCCGTTCCATGTCCTTCGATCAACAAGATGCTTCTGTCGGGGCTGATA	429
	***** *	
AtRPL10B	GACTTCAGACTGGTATGAGAGGTGCTTGGCAAAGCTTGGTACTTGTCTAGAGTTG	447
AtRPL10C	GGCTTCAGACTGGAATGAGAGGTGCTTGGTAAAGCTTGGTACTTGTCTAGAGTTG	431
AtRPL10A	GGCTTCAGACTGGTATGAGAGGTGCTTGGTAAAGCTTGGTACTTGTCTGCTGTTG	444
ZmRPL10-1	GGCTCCAGACTGGAATGAGGGGTGCTTGGCAAGGCCCTAGGGCACCTGTGCTAGGGTGG	779
ZmRPL10-2	GGCTCCAGACTGGAATGAGGGGTGCTTGGCAAGGCCCTAGGGCACCTGTGCTAGGGTGG	489
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AtRPL10B	CTATTGGACAGGTTCTTGTCTGTGAGGTGCAAAGATGCTCATGGTCATCATGTCAGG	507
AtRPL10C	CGATTGGACAGGTTCTTGTCTGTAGGTGAAAGGATAATCATGGAGTTCATGTCAGG	491
AtRPL10A	CTATTGGACAGGTTCTTGTCTGTGTCAGGATGCCCATGGTCACCAGTCAGG	504
ZmRPL10-1	ACATTGGTCAGGTCCTCCCTGGTCAAGGACAACAAATGCTGCCATGCCAGCG	839
ZmRPL10-2	ACATTGGTCAGGTCCTCCCTGGTCAAGGACAACAAATGCTGCACATGCCAGTG	549
	***** *	
AtRPL10B	AGGCTCTCGTCGCTAAAGTTAACGTCGCTAAAGATCATTGTTAGCAGGA	567
AtRPL10C	AAGCTCTCGTAGAGCTAAAGTTAACGTCGCTAAAGATCATTGTTAGCAGGA	551
AtRPL10A	AGGCTCTCGTCGCTAAAGTTAACGTCGCTAAAGATCATTGTCAGCAGGA	564
ZmRPL10-1	AAGCTCTGCGTCGCGCTAAAGTTAACGTCGCTAAAGATCATTGAGAGCAGAA	899
ZmRPL10-2	AAGCTCTGCGTCGCGCAAGTTAACGTCGCTAAAGATCATTGAGAGCAGAA	609
	***** *	
AtRPL10B	AATGGGGGTTCCAAGTTAACCGTCTGATTACACAAAGCTAACGAGAGAGAGGA	627
AtRPL10C	AATGGGGATTCACTAAATTCAACCGTCTGAGTACACGAAGCTGAGAGCGATGAAGAGGA	611
AtRPL10A	AATGGGGCTTCACGAAGTTAACAGAGCTGACTTCACCAAGTTGAGGAAAGAGCGTG	624
ZmRPL10-1	AGTGGGGCTTCACCAAGTTAACAGCGCGCTGACTACCTGAAGTACAAGAGCGAGGGCAGAA	959
ZmRPL10-2	AGTGGGGCTTCACCAAGTTAACAGCGCGACTACCTGAAGTACAAGAGTGAGGGTAGAA	669
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AtRPL10B	TTGTCCTGTGCTAAATCCAAGTCTATGTCATGTCGTTGGCTAACCGTC	687
AtRPL10C	TTGTCCTGTGCTGTCAGCTAAAGTCTATGTCATGTCGTTGGCTAACCGTC	671
AtRPL10A	TTGTCCTGTGCTGTCAGCTAAAGTCTATGTCATGTCGTTGGCTAACCGTC	684
ZmRPL10-1	TTGTCCTGTGCTGTCACCCAAAGCTGCTGCCAACCCAGCGCAGACTTGAAGACCGTG	1019
ZmRPL10-2	TTGTCCTGTGCTGTCACCGCAAGCTGCTCGTAACCATGGAAGACTTGAAGACCGTG	729
	***** *	
AtRPL10B	AGCCCCGAAAGTGCCTTGTCAAGTCTGCTGAG---CTGGTCA-----CAGTGATGCAG	730
AtRPL10C	AACCTGGAAGTGCCTTCATATCAGCC---ACTAGCGAA-----TAAGAATGAAG	717
AtRPL10A	AGCCGGGAAGTGCCTTTGCCAGCCCACACTACTGAAGAGAT-----CAGAACTGAAG	737
ZmRPL10-1	CTCCCTGGAAAGGTTCTCGATGCC-GTTGCTTAAGTGC-----GGATGCGA	1066
ZmRPL10-2	CTCCCTGGAAAGGTTCTCGAGGCC-GTTGCTTAAGTCCAATCTACAGTTGAATGCGA	788
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AtRPL10B	A-----TGACTTGTGATG-----TGGAGTTGATATCCTAGTTT	765
AtRPL10C	AA-----GATGATGATGATTGTTGTTG-----TAGAACCGATAATGTTGTT	761
AtRPL10A	TATCCTCTCATCCGGTAAGAAGAAATTATAATCAGCCTGAATCTTTTACTTATCGTT	797
ZmRPL10-1	ATCCTGACGTTTGCTTACCGTATCTACTTGTCTCGTGGAACATGAATTCAAGTGT	1126
ZmRPL10-2	ATCCAGACGTTGGTTAGCCTATCTTATTTGCTTGTGCAACATGAATTAAAGTGT	848
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AtRPL10C	TGCTCTT-TTCTGTTCAATTATTGT-AACAGTTGAG--ACAAG-GATCCTCGTATG	816
AtRPL10A	ATCTCTGGTGTGTTAAGTTTGTGACAGTATTCTGAATCTTTGTG	857

ZmRPL10-1	TTTGAGGGTATTACAGTCGCTTATGTGAACCTGCCTATCT-TGTGCTGAACATCGGAATG	1185
ZmRPL10-2	AT--GGGTATTACAGTCGCTTGTGTGAACCTGCTTATCT-CGTGCTGAACATCGTTATG	905
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AtRPL10C	ATTCCAAGACAATTGTTAACATGTGTTCTTACTTTATAATT	873
AtRPL10A	ACTCTTTGTTAACAGCTGAAATGATTGTTCTCGTT	912
ZmRPL10-1	TATCCTC--CGAGTATGTTAACGCAATTATTATTGGAAATTG	1239
ZmRPL10-2	CAT-----GGAAGTACTTATCT---TTGTTCTGCTAAAATTAAAGTGTCTGCGAC	955
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AtRPL10B	-----	
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AtRPL10A	TTTCATTGCAAGTTTATCATGGCTTAGCTTAAATTTAATTGAAAGATTGCTG	972
ZmRPL10-1	AATGTCCAATTACTCGAACATTGATTTCACACGATCTTCTTATCCTTAAATTGTA	1299
ZmRPL10-2	TAAATATA-TCTTCTCAATCTGGATTGTCTC---TTAATTATTGGAGATGGTTACA	1010
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AtRPL10A	-----	
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ZmRPL10-2	CAAA-----	1014
AtRPL10B	-----	
AtRPL10C	-----	
AtRPL10A	-----	
ZmRPL10-1	CAGCGTAAACACTGGACTTCAGAGAGAATTGGGACGAGA	1399
ZmRPL10-2	-----	

B

AtRPL10A	ATGGGAAGAACCTGCGAGGTGTTACCGTCAGATCAAGGGTAAGCCATACCCAAAGTCT	60
AtRPL10C	ATGGGACGAAGAACCTGCGAGATGTTACCGTCAGATTAAGGGAAAGCCATACCCGAAATCA	60
AtRPL10B	ATGGGACGAAGAACCTGCGAGATGTTACCGCAAATTAAAGGGAAAGCCATACCCCTAAATCA	60
ZmRPL10-1	ATGGGCAGAACGGCTGCTAGATGCTATGCCAGATCAAGAACAGCCATACCCCTAAAGTCC	60
ZmRPL10-2	ATGGGGAGAACGGCTGCGAGATGCTATGCCAGATCAAGAACAGCCATACCCAAAGTCC	60
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AtRPL10B	AGATACTGTCGTGGTGTCCCAGATCTAAAGATCAGGATTACGATGTTGGTATGAAGAGG	120
ZmRPL10-1	AGGTACTGCCGTGGTGTCCCAGACCCAAGATCAGGATCTACGATGTCGGATGAAGAGG	120
ZmRPL10-2	AGGTACTGCCGTGGTGTCCCAGACCCAAGATCAGGATCTACGATGTTGGCATGAAGAGA	120
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AtRPL10B	AAAGGGTGTGATGAGTTCCCTACTGTGTCTATTGGTTCATGGGAGAACGGAGAATGTG	180
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ZmRPL10-2	AAGGGTGTGATGAGTTCCCTACTGTGTGCACCTTGCTCTGGGAGAACGGAGAATGTG	180
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ZmRPL10-2	TCCAGTGAGGCCTTGAGGCTGCCGTATT-----	210
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AtRPL10C	-----GCTTGCAACAAGTATATGGTGAATCTGCT	240
AtRPL10B	-----GCTTGTAACAAGTACATGGTGAAGTCTGCC	240
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ZmRPL10-2	-----GCTTGCAACAAGTACATGACCAAGTCTGCA	240
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AtRPL10B	GGGAAAGATGCTTTCATCTCCGTATTAGAGTTCATCCTTTCCATGTTCTTAGGATCAAT	300
ZmRPL10-1	GGAAAGGATGCCCTCACCTAGGGTCCGGTTACCCCTTCATGCTCCGTATCAAC	360
ZmRPL10-2	GGAAAGGATGCCCTCACCTCGGGTCCGGGTTACCCCTTCATGCTCCGTATCAAC	300
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AtRPL10C	AAGATGCTTCGTCGTGGAGCTGATAGGCTTCAGACTGGAATGAGAGGTGCTTTGGT	360
AtRPL10B	AAGATGCTTCCTGTGCTGGAGCTGATAGACTTCAGACTGGTATGAGAGGTGCTTTGGC	360
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ZmRPL10-2	AAGATGCTTCGTCGTGGGCTGATAGGCTCCAGACTGGAATGAGGGTGCCTTGGC	360
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AtRPL10B	AAAGCTTGGTACTTGTGTCAGAGTTGCTATTGGACAGGGTCTTTGCTGTCGTGAGGTG	420
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ZmRPL10-2	AAGCCTCAGGGCACCTGTCTAGGGTCCGGACATTGGTCAGGTCTCCCTCTGGATGC	420
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AtRPL10C	AAGGATAATCATGGAGTTCATGCTCAGGAAGCTTCGTAGAGCTAAAGTTAACGTCCT	480
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ZmRPL10-2	AAGGACAACAATGTCGCCAGTGAAGCTGCGTCCGGCCAAGTCAAGTCCCT	480
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AtRPL10C	GGTCGTAAAAGATCATTGTTAGCAGGAATGGGGATTCACTAAATTCAACCGTGTGAG	540
AtRPL10B	GGTCGTAAAAGATCATTGTTAGCAGGAATGGGGATTCACTAAATTCAACCGTGTGAT	540
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ZmRPL10-2	GGCCGCCAAAAGATTATTGAGAGCAGAAAGTGGGGCTTCACCAAGTTAGCCGCGCTGAC	540
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AtRPL10C	TACACGAAGCTGAGAGCGATGAAGAGGATTGTCCTGATGGTGTCAATGCTAAGTTCTA	600

AtRPL10B	TACACAAAGCTAAGGCAAGAGAACAGGATTGTCCCTGATGGTGTAAATGCCAAGTTCTA	600
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ZmRPL10-2	TACCTGAAGTACAAGAGTGAGGGTAGAATTGTTCTGATGGTGTCAACGCAAAGCTGCTC	600
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AtRPL10C	TCAAACCATGGTCCATTGGCTAACCGTCACCTGGAAAGTGCTTCAATCAGCC- ACTAG	659
AtRPL10B	TCTTGCCATGGTCCGGTGGCTAACCGTCAGCCGGAAAGTGCTTCTTCAGCT-GGTGC	659
ZmRPL10-1	GGCAACCACGGCAGACTTGAGAAGCGTGCCTGGAAAGGCTTCCTGATGCC-GTTGC	719
ZmRPL10-2	GGTAACCATGGAAGACTTGAGAAGCGTGCCTGGAAAGGCTTCCTCGAGGCC-GTTGC	659
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AtRPL10A	TGA --- 663	
AtRPL10C	CGA ATAA 666	
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ZmRPL10-2	TTAA --- 663	
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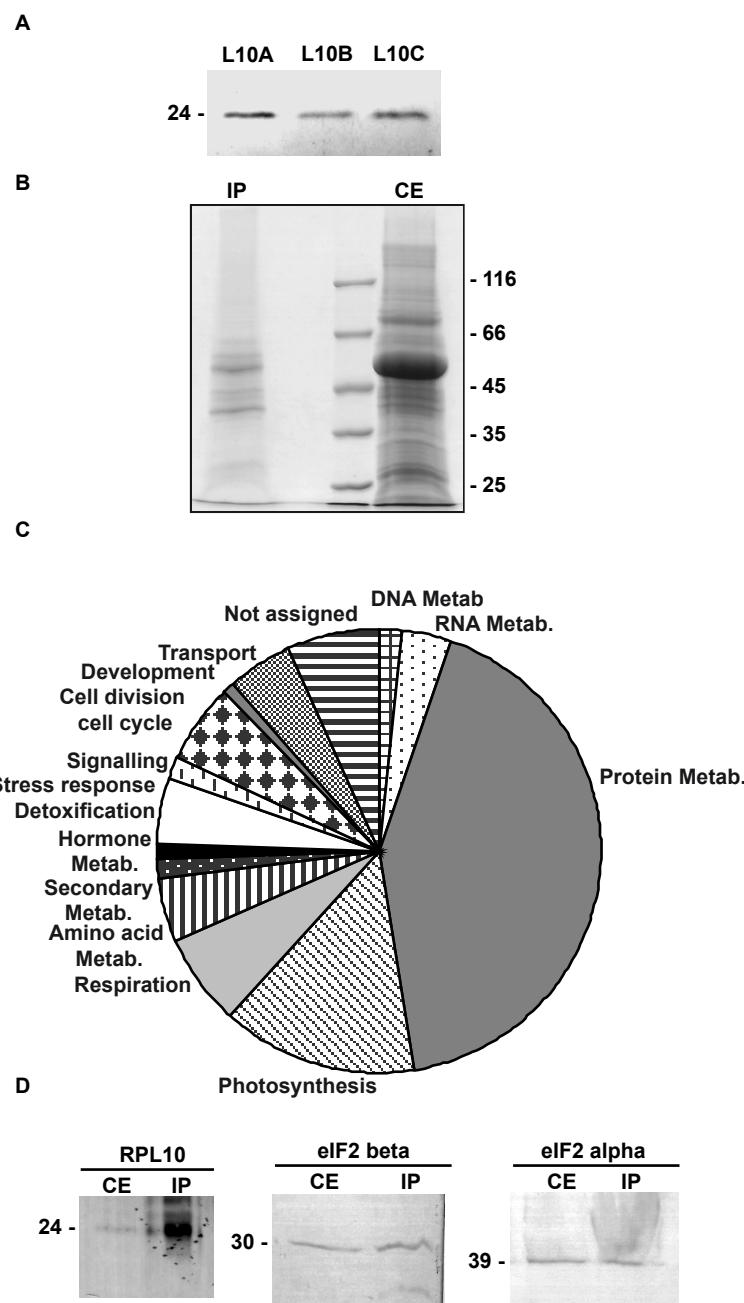
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AtRPL10C	MGRRPARCYRQIKGKPYPKSRYCRGVPDFPKIRIYDVGMRKGVD EFPFCVHLVSWEKENV	60
ZmRPL10-1	MGRRPARCYRQIKNKPYPKSRYCRGVPDFPKIRIYDVGMRKGVD EFPFCVHLVSWEKENV	60
ZmRPL10-2	MGRRPARCYRQIKNKPYPKSRYCRGVPDFPKIRIYDVGMRKGVD EFPFCVHLVSWEKENV	60
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AtRPL10A	SSEALEAAARIACNKYMVKSA GKD H L R I V H P F H V L R INKMLSCAGADRLQTGMRGAFG	120
AtRPL10B	SSEALEAAARIACNKYMVKSA GKD H L R I V H P F H V L R INKMLSCAGADRLQTGMRGAFG	120
AtRPL10C	SSEALEAAARIACNKYMVKSA GKD H L R I V H P F H V L R INKMLSCAGADRLQTGMRGAFG	120
ZmRPL10-1	SSEALEAAARIACNKYMTKSA GKD H L R V R H P F H V L R INKMLSCAGADRLQTGMRGAFG	120
ZmRPL10-2	SSEALEAAARIACNKYMTKSA GKD H L R V R H P F H V L R INKMLSCAGADRLQTGMRGAFG	120
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AtRPL10B	KALGTCARVAIGQVLLSVRCKDAHGHQA EALRRAFKF PGRQKIIIVSRKWGFTKFNRAD	180
AtRPL10C	KALGTCARVAIGQVLLSVRCKDNHGVA EALRRAFKF PGRQKIIIVSRKWGFTKFNRAD	180
ZmRPL10-1	KPQGT CARV DIGQVLLSVRCKDNNAAHASE A ALRRAFKF PGRQKIIIESRKW GFTKF SRAD	180
ZmRPL10-2	KPQGT CARV DIGQVLLSVRCKDNNAAHASE A ALRRAFKF PGRQKIIIESRKW GFTKF SRAD	180
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AtRPL10A	FTKLRQEKR V PDGVNAKF L SCHG P LANRQPGSAFLPAHY-	220
AtRPL10B	YTKLRQEKR V PDGVNAKF L SCHG P LANRQPGSAFLSAGAQ	221
AtRPL10C	YTKLRAMKR V PDGVNAKF L SNHG P LANRQPGSAFISATSE	221
ZmRPL10-1	YLKYKSEGRI V PDGVNAKLLGNHGRLEKAPGKAFLDAVA-	220
ZmRPL10-2	YLKYKSEGRI V PDGVNAKLLGNHGRLEKAPGKAFL EA VA-	220
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D

<i>H. sapiens</i>	MGRRP PARCYRYCKNKPYPKSRCRGVPDAKIRIFDLGRKKAKVDEFPLCGHMSDEYEQL 60
<i>M. musculus</i>	MGRRP PARCYRYCKNKPYPKSRCRGVPDAKIRIFDLGRKKAKVDEFPLCGHMSDEYEQL 60
<i>G. gallus</i>	----- PRCYRYCKNKPYPKSRCRGVPDPKIRIFDLGRKKAKVDEFPLCGHMSDEYEQL 55
<i>C. elegans</i>	MGRR PARCYRYIKNKPYPKSRCRGVPDAKIRIFDLGNKRANVDTFPACVHMMNSNEREHL 60
<i>S. cerevisiae</i>	MARR PARCYRYQKNKPYPKSRYNPAPDSKIRIYDGLGKKATVDEFPLCVHLVSNELEQL 60
<i>Z. mays-1</i>	MGR RPARCYRQIKNKYPKSRSCRGVPDPKIRIYDVGMRKGVDEFPYCVHLVSWEKENV 60
<i>Z. mays-2</i>	MGRR PARCYRQIKNKYPKSRSCRGVPDPKIRIYDVGMRKGVDEFPYCVHLVSWEKENV 60
<i>S. melongena</i>	MGRR PARCYRQIKNKYPKSRSCRGVPDPKIRIYDVGMRKGVDEFPFCVHLVSWEKENV 60
<i>L. esculentum</i>	MGRR PARCYRQIKNKYPKSRSCRGVPDPKIRIYDVGMRKGVDEFPFCVHLVSWEKENV 60
<i>A. thaliana-A</i>	MGRR PARCYRQIKGKPYPKSRYCRGVPDPKIRIYDVGMRKGVDEFPFCVHLVSWEKENV 60
<i>A. thaliana-B</i>	MGRR PARCYRQIKGKPYPKSRYCRGVPDPKIRIYDVGMRKGVDEFPFCVHLVSWEKENV 60
<i>A. thaliana-C</i>	MGRR PARCYRQIKGKPYPKSRYCRGVPDPKIRIYDVGMRKGVDEFPFCVHLVSWEKENV 60
<i>P. taeda</i>	MGRR PARCYRQIKNKYPKSRSCRGVPDPKIRIYDVGMRKGVDEFPFCVHLVSWEKENV 60
<i>T. brucei</i>	MARR PARCYFCKNKPYPKSRCRGVPDPRIRTFDIGKRAPDEFPVCHHVSRELEQI 60
<i>E. histolytica</i>	MGRRP RCYRLVRGHPSKYCRGVPDPRIKLFDIGNRSAPCDFPCCCHIVGLERENI 60
	***** : .:*****: *.*.*: * : * * * * : . * * :
<i>H. sapiens</i>	SSEALEA ARICANKYMKSCGKDGFH IRVLHPFH V IRINKMLSCAGADRLQTGMRGAFG 120
<i>M. musculus</i>	SSEALEA ARICANKYMKSCGKDGFH IRVLHPFH V IRINKMLSCAGADRLQTGMRGAFG 120
<i>G. gallus</i>	SSEALEA ARICANKYMKSCGKDGFH IRVLHPFH V IRINKMLSCAGADRLQTGMRGAFG 115
<i>C. elegans</i>	SSEALEA ARICANKYMKSCGKDGFH IRVRKHPFH V TRINKMLSCAGADRLQTGMRAYG 120
<i>S. cerevisiae</i>	SSEALEA ARICANKYMTTSGRDAFHL RVRVHPFH V LRINKMLSCAGADRLQQGMRGAWG 120
<i>Z. mays-1</i>	SSEALEA ARICANKYMTKSAGKDAFHL RVRVHPFH V LRINKMLSCAGADRLQTGMRGAFG 120
<i>Z. mays-2</i>	SSEALEA ARICANKYMTKSAGKDAFHL RVRVHPFH V LRINKMLSCAGADRLQTGMRGAFG 120
<i>S. melongena</i>	SSEALEA ARICANKYMTKSAGKDAFHL RVRVHPFH V LRINKMLSCAGADRLQTGMRGAFG 120
<i>L. esculentum</i>	SSEALEA ARICANKYMTKSAGKDAFHL RVRVHPFH V LRINKMLSCAGADRLQTGMRGAFG 120
<i>A. thaliana-A</i>	SSEALEA ARICANKYMTKSAGKDAFHL RVRVHPFH V LRINKMLSCAGADRLQTGMRGAFG 120
<i>A. thaliana-B</i>	SSEALEA ARICANKYMTKSAGKDAFHL RVRVHPFH V LRINKMLSCAGADRLQTGMRGAFG 120
<i>A. thaliana-C</i>	SSEALEA ARICANKYMTKSAGKDAFHL RVRVHPFH V LRINKMLSCAGADRLQTGMRGAFG 120
<i>P. taeda</i>	SSEALEA AGRIACANKYMKVFKAGKDGFH RVRVHPFH V LRINKMLSCAGADRLQTGMRGAFG 120
<i>T. brucei</i>	SSEALEA ARIQANKYMKRANKECFHM IRIHPFH V LRINKMLSCAGADRLQTGMRQSYG 120
<i>E. histolytica</i>	SSEAMEA ARISINKNLKYAGKDGFH IRIHPFH V LRINKMLSCAGADRLQTGMRGAWG 120
	*****:***:*** * * . .: * : * * * * * *****:*****: *** : :
<i>H. sapiens</i>	KPQGT VARVHIGQVIMSIRTKLQNKEHVIEALRRAFKFPGRQKIHI SKK WGFTKFNADE 180
<i>M. musculus</i>	KPQGT VARVHIGQVIMSIRTKLQNKEHVIEALRRAFKFPGRQKIHI SKK WGFTKFNADE 180
<i>G. gallus</i>	KPQGT VARVHMGQVIMSIRTKAQNKEHVIEALRRAFKFPGRQKIHI SKK WGFTKFNADE 175
<i>C. elegans</i>	KPQGL VARVDIGDILFSMIRKEGNVKAIEAFRRAFKFPGRQKIIVSSRKWGFTKWDRED 180
<i>S. cerevisiae</i>	KPHGLA VARVIDGQIIFSVRTKDSNKDVVECLRARRYKFPGQQKII SKK WGFTTNLDRPE 180
<i>Z. mays-1</i>	KPQGT CARVDIGQVLLSVRVCDNNAAHASEALRRAFKFPGRQKIIE SRK WGFTKFSRAD 180
<i>Z. mays-2</i>	KPQGT CARVDIGQVLLSVRVCDNNAAHASEALRRAFKFPGRQKIIE SRK WGFTKFSRAD 180
<i>S. melongena</i>	KPQGV CARVAIGQVLLSVRVCDNNSHAQEALRRAFKFPGRQKIIV SRK WGFTKFSRTD 180
<i>L. esculentum</i>	KPQGV CARVAIGQVLLSVRVCDKGNANHAQEALRRAFKFPGRQKIIV SRK WGFTKFSRTD 180
<i>A. thaliana-A</i>	KALGT CARVAIGQVLLSVRVCDAHGHHQAQEALRRAFKFPGRQKIIV SRK WGFTKFNRAD 180
<i>A. thaliana-B</i>	KALGT CARVAIGQVLLSVRVCDAHGHHQAQEALRRAFKFPGRQKIIV SRK WGFTKFNRAD 180
<i>A. thaliana-C</i>	KALGT CARVAIGQVLLSVRVCDHNHGVAQEALRRAFKFPGRQKIIV SRK WGFTKFNRAE 180
<i>P. taeda</i>	KPQGT CARVAIGQVLLSVSRDNHSNAQEALRRAFKFPGREKIIVNRKWGFTKYTRAD 180
<i>T. brucei</i>	KPNGT CARVRIGQILLSMRTKDTYVPQALESLRRAKMFPGRQIIVISKYWGFTNIRNE 180
<i>E. histolytica</i>	KSYGSCARV QVQVLI SGRC KEQHLPAMIK CFRLAC YK FG R QKLVISNK WGFTKYKREE 180
	*. * * * : * : * : * : : : * * . * * . : * . * * * :
<i>H. sapiens</i>	FEDMVAEKRLIPDGCGVKY I PNRGPLDK-WRALHS----- 214
<i>M. musculus</i>	FEDMVAEKRLIPDGCGVKY I PNRGPLDK-WRALHS----- 214
<i>G. gallus</i>	FEEMVAQKRLIPDGCGVKY I PNRGPLDK-WRALHAA----- 210
<i>C. elegans</i>	YERMRAEGLRSDG V QLQREHG P LT K -WIENPI----- 214
<i>S. cerevisiae</i>	YLKK REAGEVKDDGAFVKFLSKKGSLENNIREFPEYFAAQA ----- 221
<i>Z. mays-1</i>	YLKYKSEGR I VPDG V NAK L LG N HGR L EK-RAPGKAFLDAV----- 220
<i>Z. mays-2</i>	YLKYKSEGR I VPDG V NAK L LG N HGR L EK-RAPGKAFL A VA----- 220
<i>S. melongena</i>	YLKYKSEN R IVPDG V NAK L LG N HGP L AA-RQPGRAFLSSS----- 219
<i>L. esculentum</i>	YLKYKSEN R IVPDG V NAK L LG N HGP L AA-RQPGRAFL A AN----- 220
<i>A. thaliana-A</i>	FTKLRQEKR V VPDG V NAK L LG N HGP L AN-RQPGSAFLPAHY----- 220
<i>A. thaliana-B</i>	YTKLRQEKR V VPDG V NAK L LG N HGP L AN-RQPGSAFLSAGAQ----- 221
<i>A. thaliana-C</i>	YTKLRA M KR V VPDG V NAK L LG N HGP L AN-RQPGSAFISATSE----- 221
<i>P. taeda</i>	YLKWKTEN R IVPDG V NAK L LG N HGP L AN-RKPGQAFLKPAV V ILSSLVA 228
<i>T. brucei</i>	YEELRDAGKLQQRGLHV K LT P KG K ITP--YNIMA----- 213
<i>E. histolytica</i>	YQQLNKDGKIIADG C YFKLATT K GPLPK--VN----- 210
	: . * : * : * : :

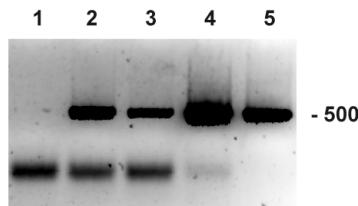
Supplemental Figure 2



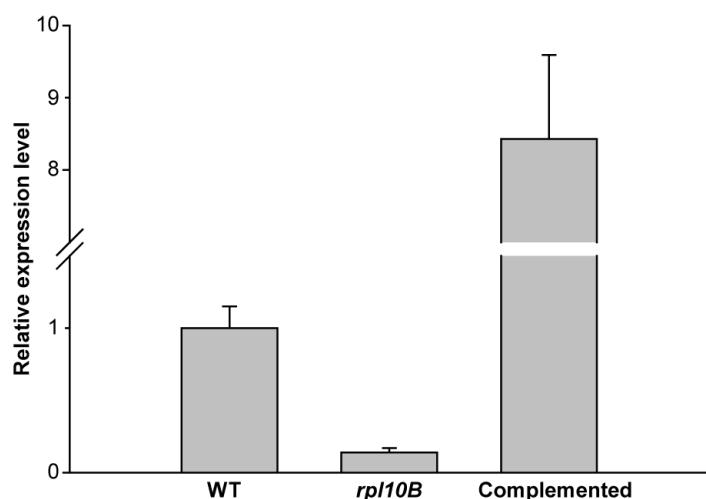
Coimmunoprecipitation of RPL10 proteins in *A. thaliana*. **A, Immunoblot analysis of *A. thaliana* recombinant RPL10 proteins.** Partially purified recombinant RPL10 proteins were run on 12% SDS-PAGE and subjected to immunoblot analysis for RPL10. Ten micrograms of total proteins were loaded in all lanes. **B, SDS-PAGE (10%) of RPL10-associated proteins.** **C, Classification of RPL10-associated proteins based on their cell functions.** Proteins with percentage of coverage higher than 10% or at least two tryptic peptides were included in the diagram. Clustering was performed according to Usadel et al. (2006). **D, Immunoblot analysis of RPL10-associated proteins.** RPL10 proteins were immunoprecipitated from *A. thaliana* crude extracts with antibodies against *H. sapiens* QM protein. The immunocomplexes were solubilized, run on 12% SDS-PAGE and subjected to immunoblot analysis for RPL10, eukaryotic translation initiation factor 2 alpha (eIF2 alpha) and eukaryotic translation initiation factor 2 beta (eIF2 beta). The numbers indicate the molecular mass in kDa. CE: crude extract, IP: immunoprecipitate.

Supplemental Figure 3

A



B



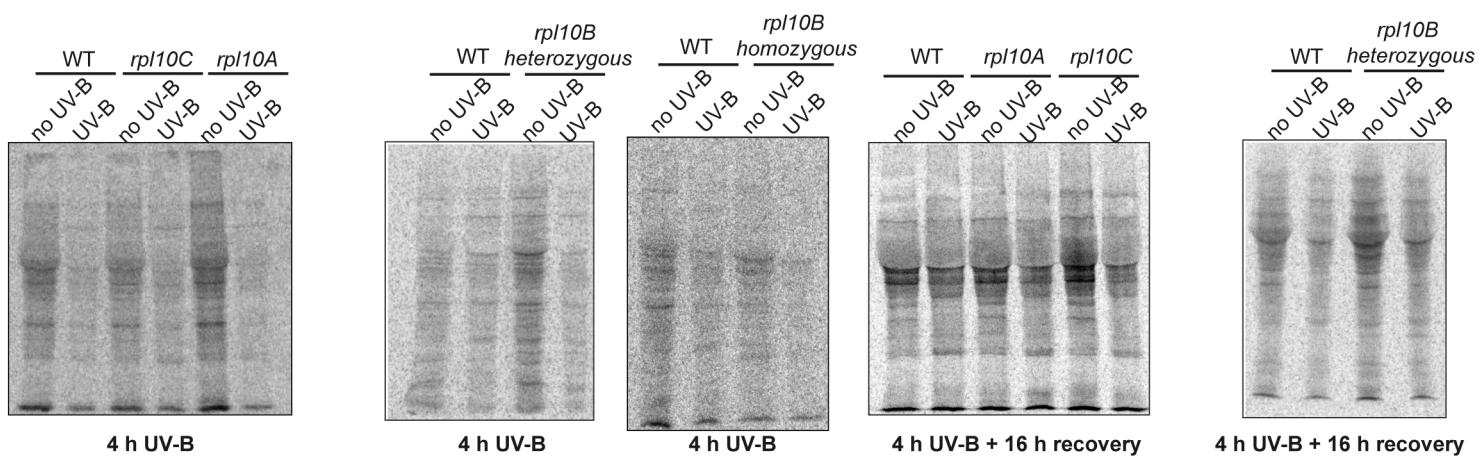
C



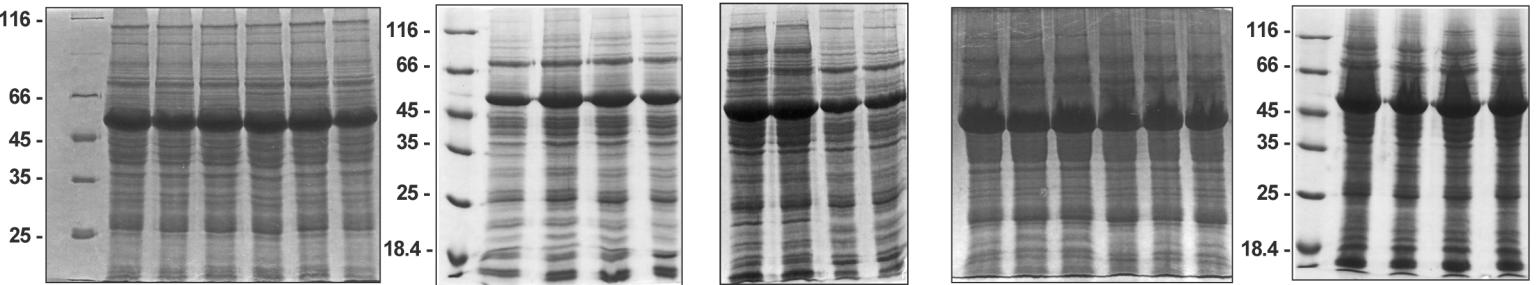
Complementation of *A. thaliana* homozygous *rpl10B* mutants with WT At *RPL10B*. A, Presence of WT *RPL10B* transcript in transformed *A. thaliana* *rpl10B* mutant plants analyzed by PCR on genomic DNA. Lanes 1: negative control (without DNA); lane 2-4: genomic DNA from leaves of transformed plants; lanes 5: positive control (pCHF3-*RPL10B*). B, At *RPL10B* expression level in Arabidopsis WT, *rpl10B* homozygous and complemented plants analyzed by RT-qPCR. Each reaction was normalized using the C_t values corresponding to the *POLYUBIQUITIN10* mRNA. The means of the results obtained using three independent biological experiments are shown, the error bars indicate the S.D. of the samples. WT levels were set at 1. C, 15-day-old WT (left), *rpl10B* mutant (middle) and complemented plants. Scale bar: 1 cm.

Supplemental Figure 4

A



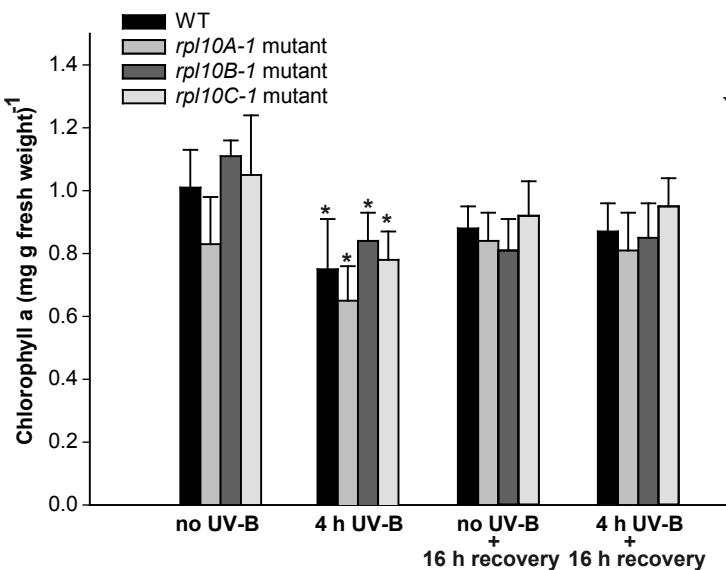
B



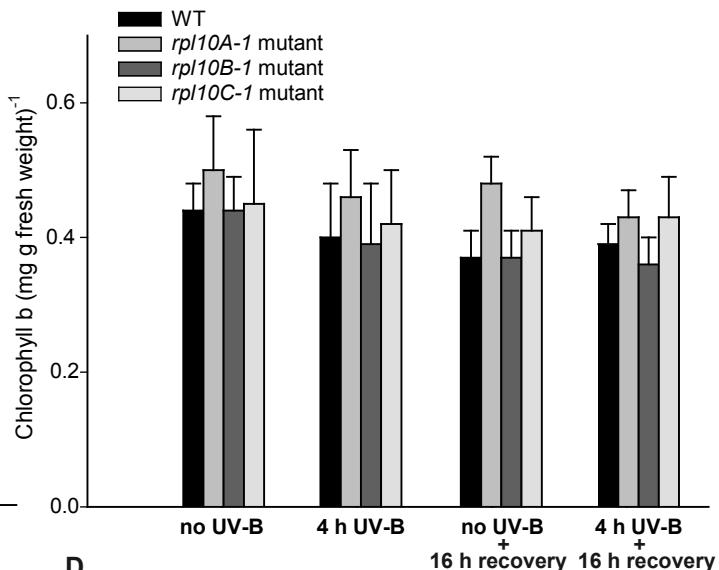
Inhibition of protein synthesis by UV-B in *A. thaliana* WT and *rpl10* mutant plants. Forty micrograms of total proteins were resolved by 12% SDS-PAGE after *in vivo* [35 S]Met labeling, visualized by autoradiography (A) and staining with Coomassie Blue (B) following the UV-B treatment and recovery period indicated.

Supplemental Figure 5

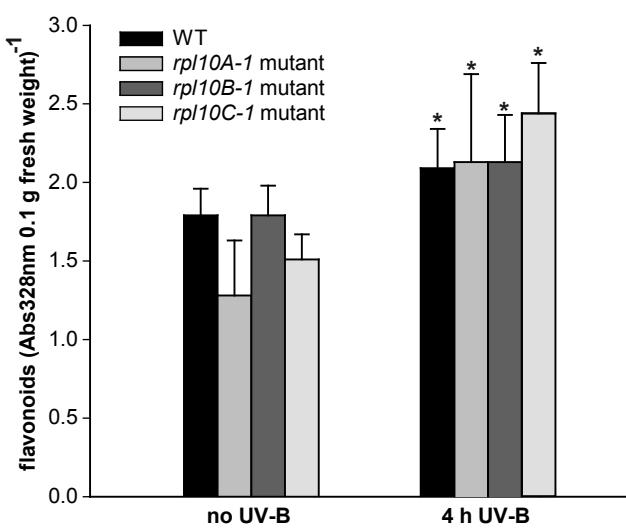
A



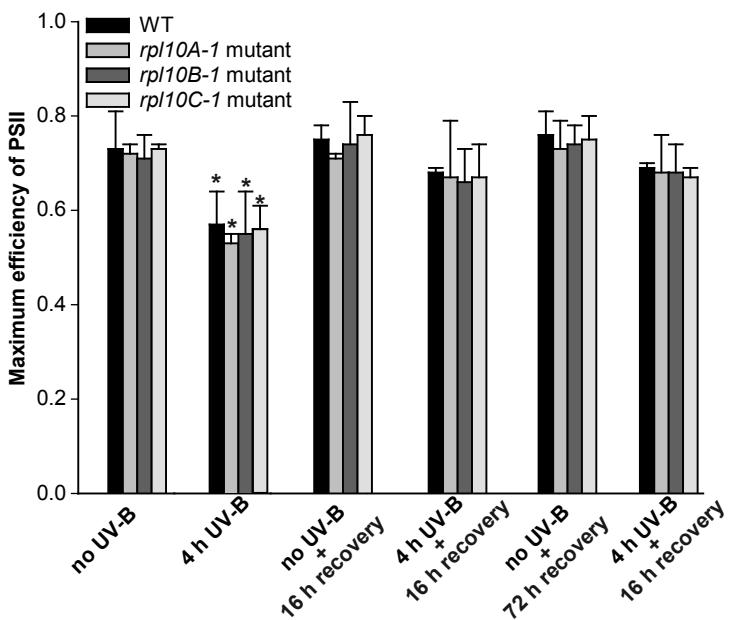
B



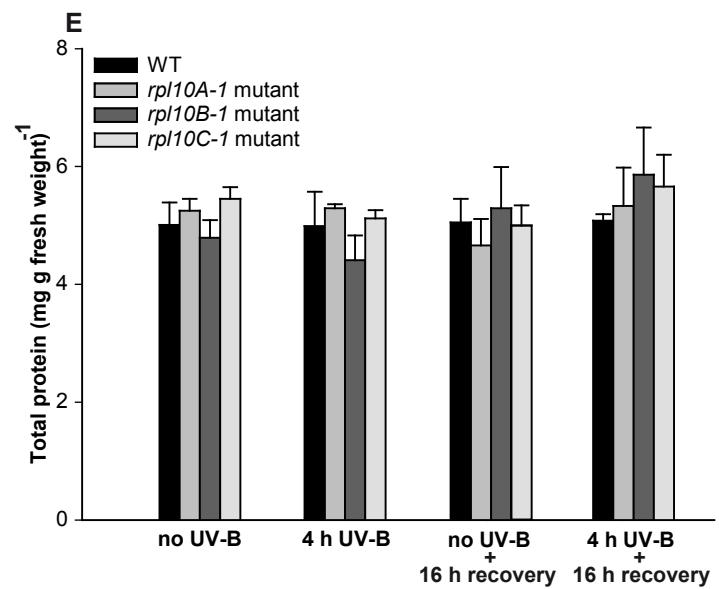
C



D

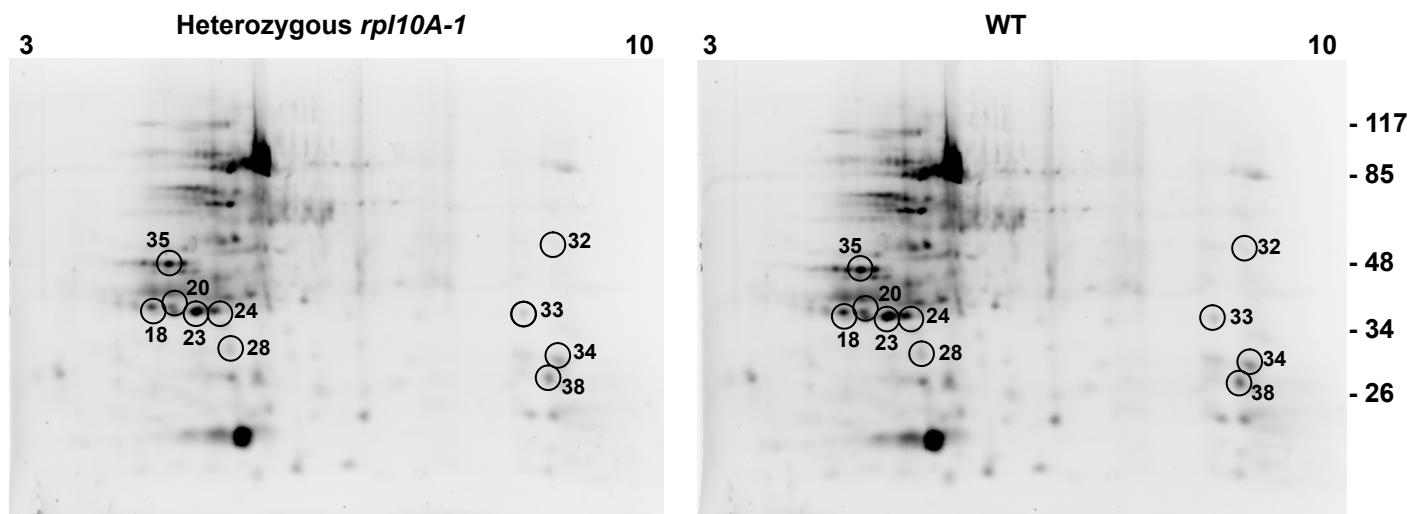


E



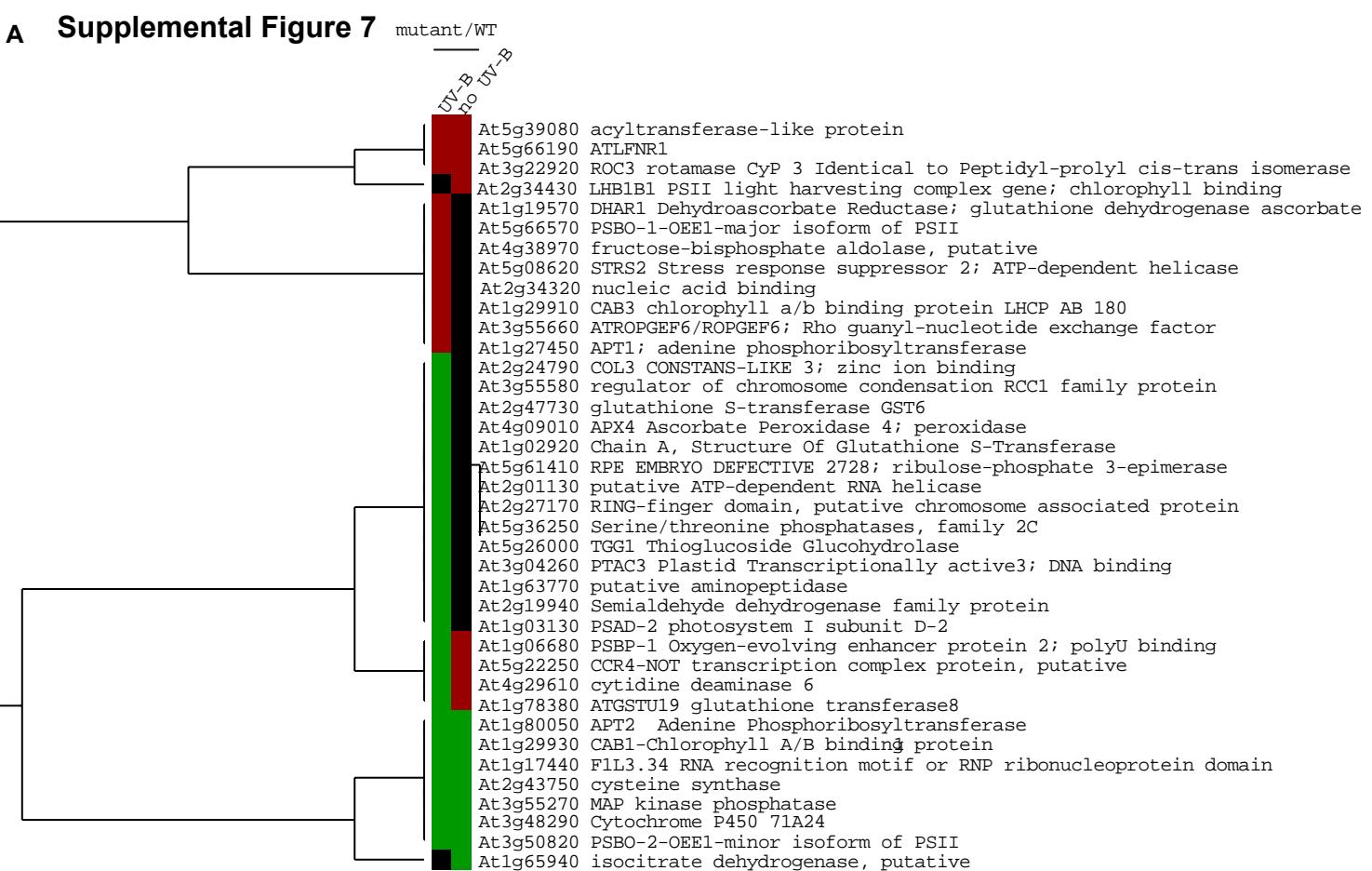
UV-B treatment is not lethal to *A. thaliana* plants. Chlorophyll a (A), Chlorophyll b (B), Flavonoids (C), Maximum Efficiency of PSII (D) and Total proteins (E) were measured after 4 h UV-B (4 h UV-B), 16 h post-treatment (16 h recovery) and in untreated controls (no UV-B). Measurements are the average of six adult leaves from four different plants. Statistical differences from the control are marked with an asterisk ($P<0.05$).

Supplemental Figure 6

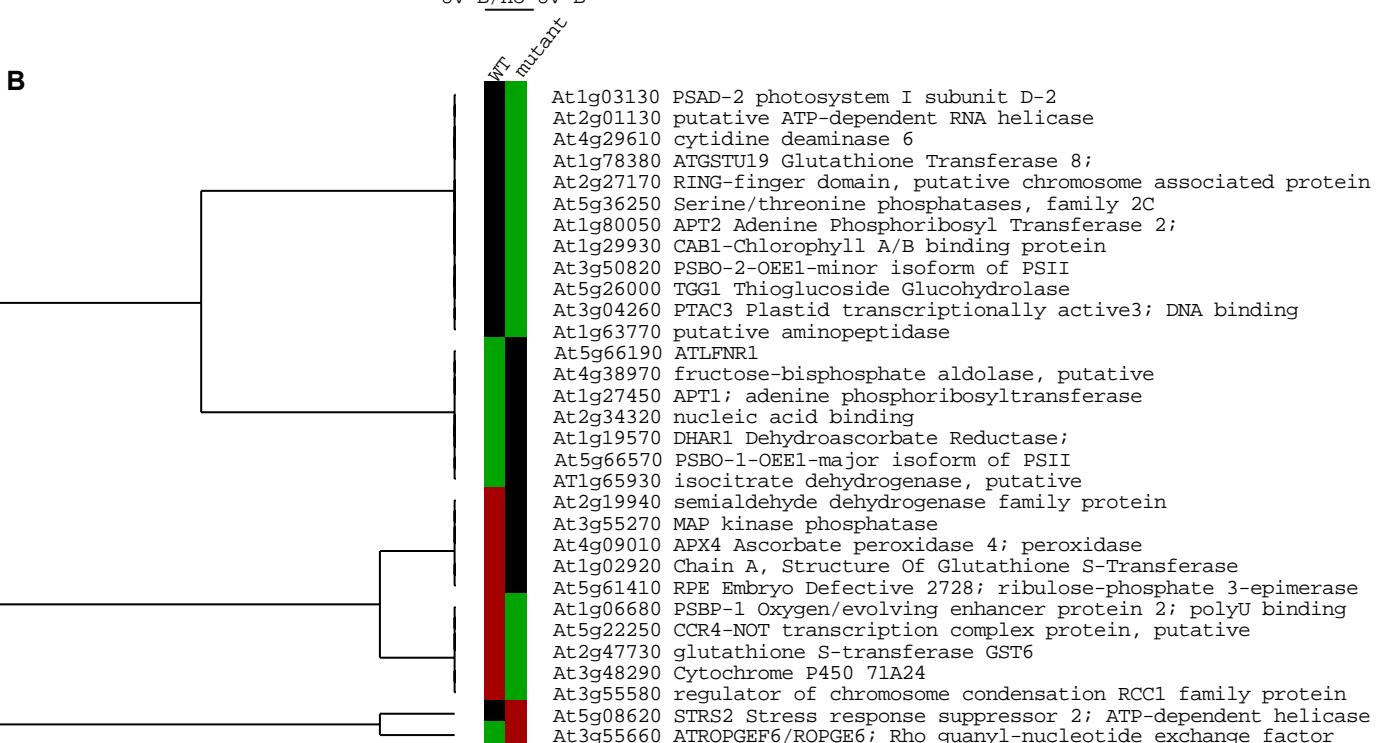


Typical 2D gels of leaves from heterozygous *rpl10A-1* mutant and WT plants after a 4 h UV-B treatment. As examples of proteins with differential expression, the relative abundances of some but not all spots annotated by the number that appears in Supplemental Table S3 are shown. The graphs represent one example from at least three different gels used for the differential analysis. The first dimension was carried out using 17 cm immobilized pH gradient strips (pH 3–10); acidic side to the left; and the second dimension was on 12.5% (w/v) SDS–PAGE. The relative abundance of proteins was determined. The protein spots with changes in intensities (least 1.5-fold, P <0.05) were considered to be different.

A Supplemental Figure 7

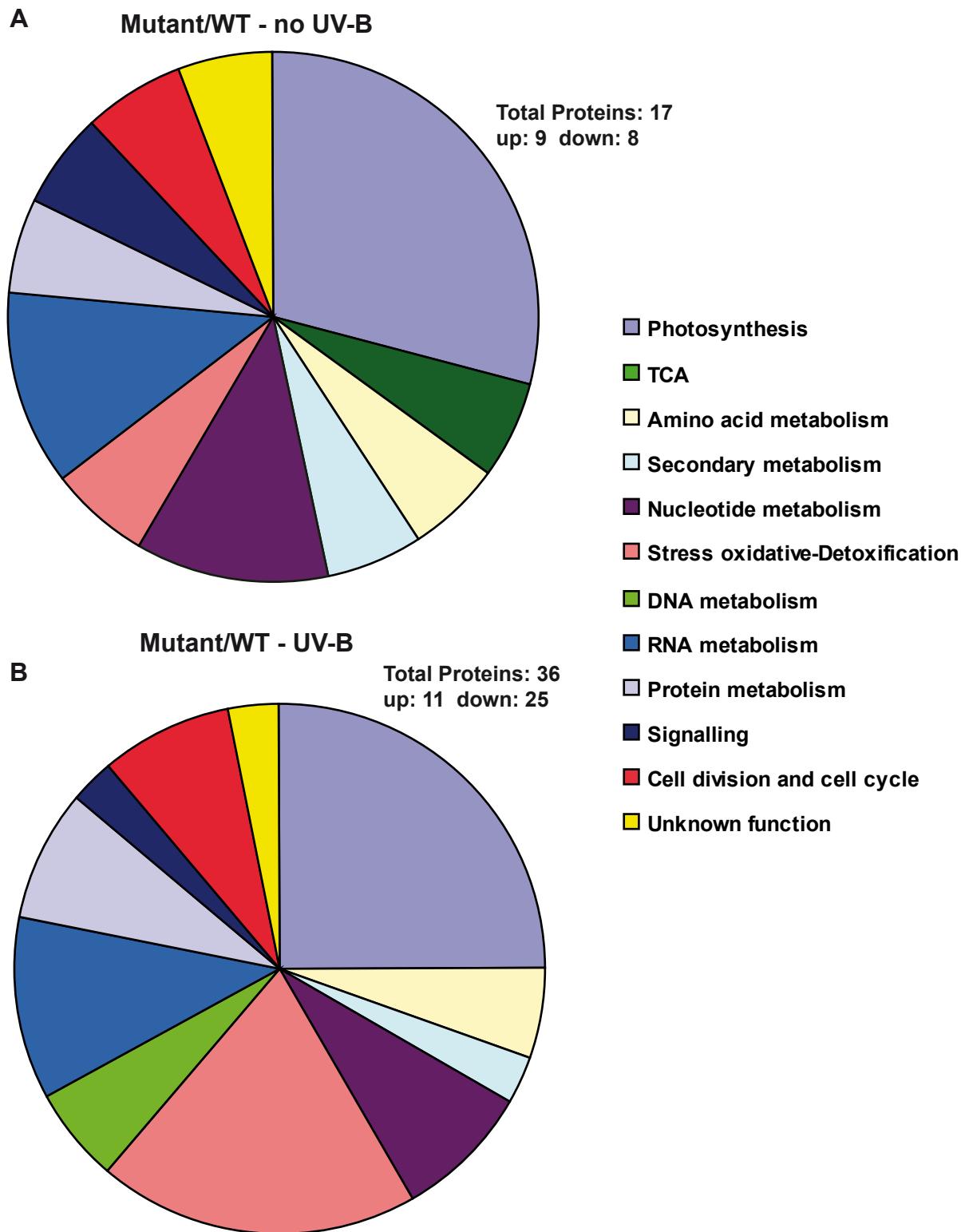


B



Hierarchical cluster analysis of proteins showing different levels in *rpl10A* mutant plants in comparison to WT plants under control conditions and after a 4h UV-B treatment identified by MS. A, Proteins included show different levels in *rpl10A* mutants (at least 1.5-fold) in comparison to WT plants under control or UV-B conditions. B, Proteins included show differential abundance (at least 1.5-fold) after a UV-B treatment; these proteins changed differentially in WT plants than in the *rpl10A* mutant. Red indicates higher protein levels than the reference, green indicates lower protein levels than the reference, and black indicates no significant change.

Supplemental Figure 8



Classification of proteins showing different levels in the *rpl10A* mutant in comparison to WT plants based on their cell functions. Proteins were identified by 2D Gel electrophoresis and those showing changes in abundances of at least 1.5-fold were included. A, Proteins changed in the *rpl10A* mutant under control (no UV-B) conditions. B, Proteins changed in the *rpl10A* mutant after 4 h of UV-B.

Supplemental Figure 9. *RPL10s* promoter sequences with predicted cis-elements. The transcription initiation site (referred to as +1) is indicated in bold letter and the ATG start codon is shown in bold and underlined letters. Numbers at the left refer to the positions of nucleotides relative to the putative transcription initiation site.

RPL10A

```

-1000 TAACTTGAAAAGTTCGGTGGAGATCTAACGCTAAACTTAATTTCTTCCCGGTTA
                                         Homeobox
-937 ACCAATAAGCGATCCATCTACATACAGAGCATGCCCGAGACGAGGAAGTATTAATCCGAT
                                         CCAAT-BOX
-874 AAGGAGAAGAAGGATTGATATACCTCCAAGTGTTGGTGCTAAATCAAAACTTCACATGTAG
                                         UVBox           MYC2
-811 TAGCGTTCTAGGCCAAGTTCGGAAGAGTTATACATAACCAAACCGGTTGTATATGCCACT
                                         UVBox           ARF1
-748 GATTTTGCTTGCCAAATCCAATTAAACGTGACTAAATAACTTGGCTGCTCAAGACAGAT
                                         ACE             UVBox
-685 TTGTTGCAACCTGGAAACAGGGAAACGTCGATGCCATCGAGTGGCGGGATTATAAACATGTT
                                         ACE
-622 GTTAAGGTTGGTAATCAAAGAGGCAAACAGACCGTCACAACACTATTGGGAAAAGTTGGTA
                                         MRE
-559 AATATGATATCGTTCTGATGATATCAACAAACACGTTAGTTAAAGTGGAGGAGAATCAGCAGT
                                         ACE
-496 AACATGATGGGCAACACCAACCGTACTGGGTACTTCAGACACCAATACAAGATTTAGATCTT
                                         ACE             CCAAT-BOX
-433 CCCGCCAGCTGAGCAGATCAACTGTTCGCCTGGAAATTGAGATTGATTGTCACCTCCA
-370 TTGTTGCAAGCAGACTTGAATCTGAGCAGAGATTACCGGAACTCTCTCAAGAATATCCTC
-307 AACGGTGTCGTGGGAAGCAATTGCATTATTCTCTGTCTATTGAGAGGATTGTTCTGAGT
-244 GATGGATAACATGAAAGATATGCTTATTGTATCAATTCAATCCAATGTTGATTTTCCTTG
                                         CCAAT-BOX
-181 AGGAGGAAGATAAAAAAAAAAAACGTATATACAATCGATGGGCCCTAACCTATCCCTAACAA
                                         GATA-BOX        ACE           SORLIP2
-118 AATCTTTAATATGTAATGCGCTTAATAGTTAAAGCCCATTAGTTAAAAACCCAGAGCTAT
-55 ATTGTTGACCTAGCAAATTTCGGATCTATAAATTGAAGCCATTTCAGGTCATTAGTTTTT
                                         W-BOX           +1
                                         CGTCGAGCAGCCCGCTTTTGCCGAGGAAGGATAAGAGAGACGCCATG
                                         I-BOX

```

RPL10B

```

-1000 AGTTTGCCGTACCCCTATCAAAGCTGTGATTAATGCAAGTCTAAATCAAGGCCAATCTT
                                         CCAAT-BOX
-937 GATGCATAACACATGAGCAGATTCTGCTCCAGACAAATCTCTAGATCACTAATAGCCGAAAG
                                         MYC2/DPBF1
-874 AAAAGCTAACGTAACGTACTGACATTAACGACCACCTCACTTTCTCATAATCCTAAACAAA
                                         ACE
-811 TCCAAAGCCAACGCTTCTTAGAAACCTGAAGATAACCATTCAAAAGTACTAAACGTAACA
                                         ACE
-748 GCATCAACACTCTGGCATTTCGCGAACACTTCCGAGCATTCAATCTTCCACAAACA
-685 CAGTAAAAATGTATAAGAGCATTCTCAAATCAGTAAAACCATAAACCCAGATCTCAAAGCA
-622 ATCCCCATGCAATCCCTCACCAATTGAAACACACAATTCACGAGAACATGATTTAAGAGTCGTG
                                         CCAAT-BOX
-559 ATAAACGAGAACGTTCCAAGGTCAAACCTTAGCTCTCAATTGGTTGAAACAGAAAATGCT
                                         GATA-BOX        UVBox           MRE

```

-496 CGCTCCGGCTCATCGCTAACGAGTAACCTCTGATCAGTGTGAACATAAAGAGATTAGTG
GT1-motif
 -433 TTGGAGACATGCTCGAAGATCGACGAAGCGTATCGAATGTCAAGTACAGAGGAAAAGCAAGA
 -370 AGCTTGCTCACTCGCAAATCGCTTGTGAGTCCCCTTCACCATGTAACCATGAATTCGA
 -307 GAAACTTCGACGGTGTCTCTGCAAGACCTAGATCGTTGATA**AG**TTCTGACATTGTGGAGAC
GATA-BOX
 -244 AACAAACGATTCAAGATT CCTACGACGGAACACGAAATTGCGCTTGAGCCGTGACGTAATGCC
ACE
 -181 AATGTCATCAGAAAGAAAAAAAGTCGCCGGAAATAAACACGGATTGTTTTAAGCTTAAAAA
 -118 TATCAAATTGGGCTTAGTCCTTA**ATGGGCTT**TATTTGGTCAAATCCAGT**TACGTGGCAA**
SORLIP2 **ABRE/ACE**
 -55 AGAGAATTAGGGCTTTGTTCTTCTTCTTCAATTCTAGGATTGAAACAACA**ACCGCAGT**
I-BOX
⁺¹
 CTTCTTCTTCTTCTTCTTCAATTCTAGGATTGAAACAACA**ACCGCAGT**

RPL10C

-1000 GATGCTATCACGACAAATTAGTCCAAATGGGCAGCGTATATTTCTTAT**GGGCCTAAACA**
SORLIP2
 -937 AAGGAAACTAACTAATTATAATCAACTTATGATCATCAGTCATTATGAA**AGGTT**TATATTGA
MRE
 -874 CAGGAGATTGTGTTAAAACCATTGAGTATATCTGTTGAATCATTGAGACAATTGT
 -811 GATTTTGCTTCTGATCCCATTCTTTAGTCGTGCATATGTGATCTGTATTGTCTAA
 -748 TAAGGATTATGATCTGCCCCCTAAACTC AAAATTGGAGGCCATGATCTGATTTGAATGT
 -685 GGTTCTTATAGTTTGCTTGATATTGAGCAGACGAATGATGGTGAATATCCATCTAAGTCA
 -622 AATACAGATTCCATTCTCTTTAATACAAATCAAATAAGAAA**ACTGAA**GTTGCAAGCTCGC
 -559 TTGTACTAAAGTTCTGAAAGTTTATTCTCGACTAAATAATGT**CCAAG**TGGAAGCAAGACA
UVBox
 -496 TAAGCTCCATTGTTGATAGAATGGAGCTTATGTTGGCTTGTGTTCAAAATGGTTT
 -433 TAACTTATGTGATTGTTAGCCAAATGTGGACTCTGAAGATGGTTTCAGTTTGTT
 -370 AATGTCCTTGTACTATTGTTACAGTCACACAGAGTTACAAAGATCATTGATTGTT
 -307 TATAAGAGAACAAATT**GATA**ACATATCTTTGATGTGGATGGATTGTTAACCTCTAGA
GATA-BOX
 -244 AGAACACCCAAATTGGCTACAAAGTTGTCAACACATTGTTCTCGTTCTTGA**ACTTGA**
T-BOX
 -181 CTAAAAATAAAACCTCGGCCTAAGAAAATAGCGTGTTCACATTGGATGATTAAGGCCT
 -118 GGACCGAGTATCAAATAATCTAA**ATGGGCT**AGCTAAGCAGTTACGTAAGCTAAATTAGG
SORLIP2 **ACE**
 -55 GTTTGTTTGCCGCACCTATAAAACAAACACAGCTTCCATTGTAATTCTAGGGTTT
⁺¹
 GCAAAACCAACACCGAAGATCCAACAC**ATG**

Supplemental Table S2. Segregation of *RPL10A* alleles

SALK 010170 line (n=47)

Heterozygote	WT
35 (74%)	12 (26%)

Progeny of *rpl10A* heterozygous plant (n=104)

Heterozygote	WT
68 (65%)	36 (35%)

SALK 106656 line (n=67)

Heterozygote	WT
57 (85%)	10 (15%)

Progeny of *rpl10A* heterozygous plant (n=32)

Heterozygote	WT
20 (63%)	12 (37%)

Supplemental Table S4. 5'UTR sequences in *A. thaliana* *RPL10* transcripts

<i>RPL10</i>	AGI number	Sequence of the 5'UTR	Sequence of the putative 5'TOP
<i>RPL10A</i>	At1g14320	AGTTTTTCGTCGAGCAGCCGCCCTTTT GGCCGAGGAAGGATAAAGAGAGACGCC	-
<i>RPL10B</i>	At1g26910	CTTCTTCTTCTTCTTCTTCTTCTTCATT TCTAGGATTCGAAACAACAATCAACCGCG	CTTCTTCTTCTTCTTCTTCTTCTTC
<i>RPL10C</i>	At1g66580	AATTCTAGGGTTTGCAAAACCAACACC GAAGATCCAACACG	-

Supplemental Table S5. Primer Sequences used for PCR

Name	Sequence	Purpose
<i>Zm thioredoxine-like-for</i>	5'-GGACCAGAAGATTGCAGAAG-3'	qRT-PCR
<i>Zm thioredoxine-like-rev</i>	5'-ACGGATGTCCCCATGAAGA-3'	qRT-PCR
<i>Zm actine1-for</i>	5'-CTTCGAATGCCAGCAAT-3'	qRT-PCR
<i>Zm actine1-rev</i>	5'-CGGAGAACATGCATGAGGAAG-3'	qRT-PCR
<i>Zm RPL10-1-for</i>	5'-TGCAAGGACAACAATGC-3'	qRT-PCR
<i>Zm RPL10-1-rev</i>	5'-TCTGCCCTCGCTTTGT-3'	qRT-PCR
<i>Zm RPL10-2-for</i>	5'-GCTCGGTAACCATGGAAGA-3'	qRT-PCR
<i>Zm RPL10-2-rev</i>	5'-GAGATAAGCAGGTTCACACA-3'	qRT-PCR
<i>At UBQ10-for</i>	5'-AAGCAGCTTGAGGATGGAC-3'	qRT-PCR
<i>At UBQ10-rev</i>	5'-AGATAACAGGAACGGAAACATAGT-3'	qRT-PCR
<i>At CDPK3-for</i>	5'-CGCTGAGAACCTTCTGAAG-3'	qRT-PCR
<i>At CDPK3-rev</i>	5'-CCATCTCCATCCATATCAGC-3'	qRT-PCR
<i>At RPL10A-for-1</i>	5'-TCCTTCTCATCCGGTGA-3'	qRT-PCR
<i>At RPL10A-rev-1</i>	5'-GCCAAGAACGAAGGAACA-3'	qRT-PCR screening
<i>At RPL10B-for-1</i>	5'-TGGTGTTCCCGATCCTAA-3'	qRT-PCR
<i>At RPL10B-rev-1</i>	5'-ATCTTCCGGCAGACTT-3'	qRT-PCR screening
<i>At RPL10C-for-1</i>	5'-CCAACACCGAAGATCCAA-3'	qRT-PCR screening
<i>At RPL10C-rev-1</i>	5'-TTTGGAATCTGGCACAC-3'	qRT-PCR screening
<i>At RPL10A-for-2</i>	5'-GCTTGGAATTTGGTACTTGCTC-3'	screening
<i>At RPL10B-for-2</i>	5'-GTCGCCGGAATAAACACG-3'	screening
<i>At RPL10C-for-2</i>	5'-GCCAAATGTGGACTCTGAAG-3'	screening
<i>At RPL10B-for-3</i>	5'-TAAATTGGTACCAACAA TCAACCGCGATGGGAC-3'	cloning
<i>At RPL10B-rev-2</i>	5'-ATTTAAGTCGACCTGTGCACCAGCTG	cloning

	ACAAGAAC-3'	
<i>prom35-for</i>	5'-CTATCCTTCGCAAGACCCTTC-3'	screening
<i>LB-SALK</i>	5'-GTCCGCAATGTGTTATTAAGTTGTC-3'	screening
<i>LB3-SAIL</i>	5'-TTCATAACCAATCTCGATACAC-3'	screening