

Figure S1. FACS analysis and ISRE library sorting scheme

(A) FACS analysis and gating procedure for all HEK-293 FLP-In cells. As an example, flow cytometry data from the stable NMD control is presented. Dot plots show initial gating of stable cells (P1), followed by P2 gating for cell uniformity (i.e., to remove cell aggregates) and finally the selection of live cells using 7-Amino-Actinomycin D (7AAD) staining. The P3 gate reflects the GFP positive cells and the P4 gate is drawn to indicate the upper GFP fluorescence limit of the NMD control population. P4 was used as the gate for the selection of ISS positive cells. The histogram reports the intensity of GFP fluorescence in the NMD control population. (B) FACS analysis of ISS positive stable cells after one round of sorting. Cells from gates A, B and C were sorted and the resulting histograms indicate the intensity of GFP fluorescence after 1 week in culture.

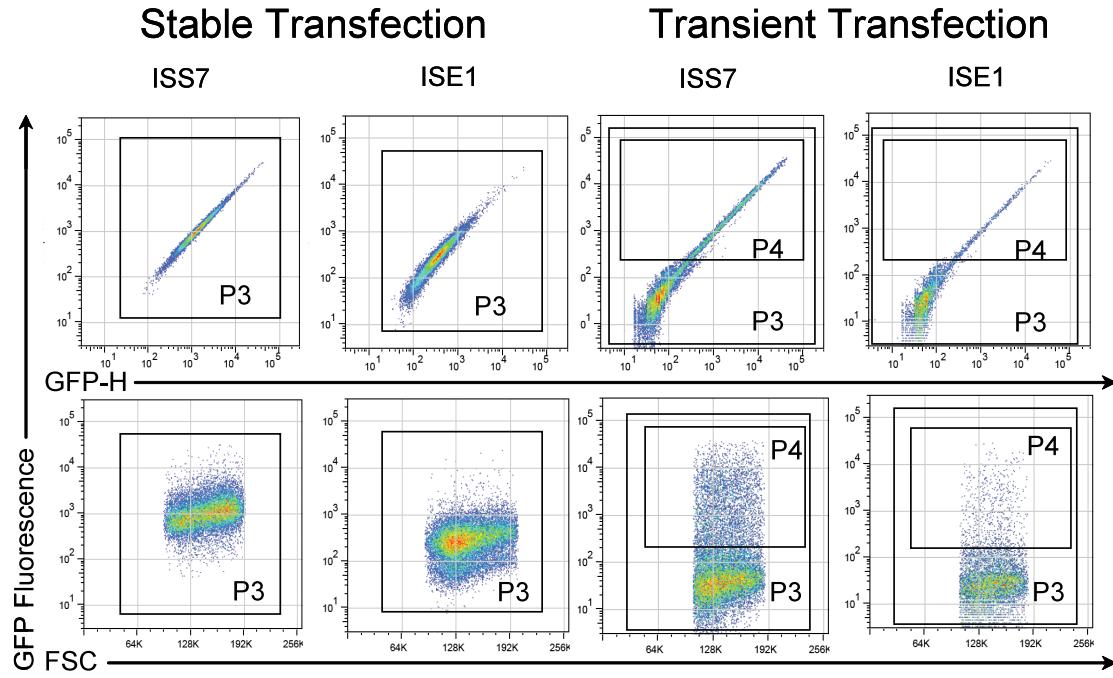
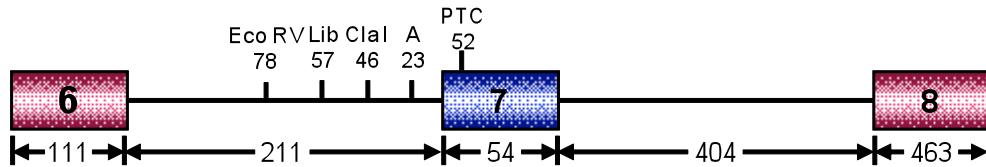


Figure S2. Assessment of splicing regulatory activity through stable and transient transfection assays

Sixteen recovered ISS sequences (ISS1-ISS16) and 1 recovered ISE sequence (ISE1) were examined for regulatory activity in both transient and stable transfection assays. Examples of assay results for two recovered sequences (ISS7, ISE1) are shown. For the stable cell line assays, mean GFP fluorescence levels were determined using gate P3. For the transient transfection assays, the P3 gate represents the untransfected cell population and the P4 gate represents the GFP-positive cells. The results of an ANOVA analysis applied to data from the transient and stable assays indicate that the two methods are not statistically similar ($P = 0.27$).

A**B**

GFP pair 1,5 5' ACTACCTGAG CACCCAGTCC GCCCTGAGCA AAGACCCCAA CGAGAACGCG GATCACATGG TCCTGCTGGA 3'
 GTTCGTGACC GCCGCCGGGA TCACTCTCGG CATGGACGAG CTGTACTAAC Exon6 5'
 pair 2 3'
 CATATGTCCA GATTCTCTTG ATGATGCTGA TGCTTTGGGA AGTATGTTAA TTTCATGGTA CATGAGTGGC
 TATCATACTG GCTATTATAT GGTAAGTAAT CACTCAGCAT CTTTCCTGAA CAATTTTTT GTAGTTATGT pair 2 3'
 GACTTTGTTT GGCTGATCAT ATTTTGTGA ATAAAATAAG TAAAATGTCT TGTGAAACAA AATGCTTTT
 Eco RV 15-mer library Cla I
 GGTACCAACA TCCATATAAA GCTATAGATA TCGATCAGTN NNNNNNNNNNN NNNNGCATCA TCGATGTCTA
 BP Exon7 PTC
 TATAGCTATT TTTTTAACT TCCTTTATTT TCCTTACAGT AATTCAAGACA AAATCAAAAAA GAAGGAAGGT
 GCTCACATTC CTTAAATTAA GGAGTAAGTC TGCCAGCATT ATGAAAGTGA ATCTTACTTT TGTAACACTT
 TATGGTTTGT GGAAAACAAA TGTTTTGAA CAGTAAAAAA GTTCAGATGT TAAAAAGTTG AAAGGTTAAT
 GTAAAACAAAT CAATATTAAA GAATTTGAT GCCAAAACCA TTAGATAAAA GTTAATCTA CATCCCTACT
 AGAATTCTCA TACTTAAC TGTTGGTTATG TGGAAGAAC ATACTTCAC AATAAAGAGC TTTAGGATAT pair 3
 5' 3'
 GATGCCATT TATATCACTA GTAGGCAGAC CAGCAGACTT TTTTTTATTG TGATATGGGA TAACCTAGGC
 ATACTGCCT GTACACTCTG ACATATGAAG TGCTCTAGTC AAGTTAACT GGTGTCCACA GAGGACATGG Exon8
 TTTAACTGGA ATTCTGCAAG CCTCTGGTTC TAATTCTCA TTTGCAGGAA ATGCTGGCAT AGAGCAGCAC
 pair 3 3' 5'
 TAAATGACAC CACTAAAGAA ACGATCAGAC AGATCTGGAA TGTGAAGCGT TATAGAAGAT AACTGGCCTC
 ATTTCTCAA AATATCAAGT GTGGGAAAG AAAAAGGAA GTGGAATGGG TAACTCTTCT TGATTAAGAAG pair 4
 3' 5'
 TTATGTAATA ACCAAATGCA ATGTGAAATA TTTTACTGGAA CTCTTTGAA AAACCATCTA GTAAAAGACT
 GGGGTGGGGG TGGGAGGCCA GCACGGTGGT GAGGCAAGTTG AGAAAATTG AATGTGGATT AGATTTGAA
 TGATATTGGA TAATTATTGG TAATTTTATG GCCTGTGAGA AGGGTGTGT AGTTTATAAA AGACTGTCTT
 AATTCGCATA CTTAACGCATT TAGGAATGAA GTGTTAGAGT GTCTTAAAT GTTCAAATG GTTTAACAAA
 ATGTATGTGA GGCGTATGTG

C**Exon included isoform:**

Exon7 Exon8
 TAATTCAAGACAAAATCAAAAGAAGGAAGGTGCTCACATTCTAAATTAAAGGAGAAATGCTGGCATAGAGCAGC...
 5' 3' pair 4

Exon excluded isoform:

Exon6 Exon8
 ...TTTCATGGTACATGAGTGGCTATCATACTGGCTATTATATG | GAAATGCTGGCATAGAGCAGCAC...
 3' 5' pair 5

Figure S3. Schematic representation of SPLICE and primer set binding sites

(A) Schematic representation of the SMN1 mini-gene. Shown below each exon and intron are their respective lengths (bp). The positions (relative to the 3' ss of exon 7) of restriction sites Eco RV and Cla I used to insert the 15-mer library are indicated. The PTC was inserted 51-nt upstream of the 5' ss of exon 7. (B) Mapping of primer set binding sites on the SMN1 mini-gene sequence. Schematic representing the locations of primer set binding for transcript isoform analysis by qRT-PCR. The locations of the branch point (BP), restrictions sites Eco RV and Cla I, the 15-mer library (plus flanking regions) and PTC are shown. (C) Schematic representing the exact locations of primer sequences spanning exon-exon junctions.

A

ISS17	GCAAGGTCCCTCTAG
ISS18	GACGGAGCCGTCTGG
ISS19	AGAGTGGCGGTGGAG
ISS20	GATATGGCGAGGGTG
ISS21	GGTGGCAGACACGAT
ISS22	AAATAGAGGCCAG
ISS23	TTATGGAGTCCTAG
ISS24	GAGGGCAGTCCGTGG
ISS25	TGGACACGTCAGTCA
ISS26	TCTGACTCAATAGTA
ISS27	AATTGGGTTGGGGG
ISS28	TATGACATGTGGGA

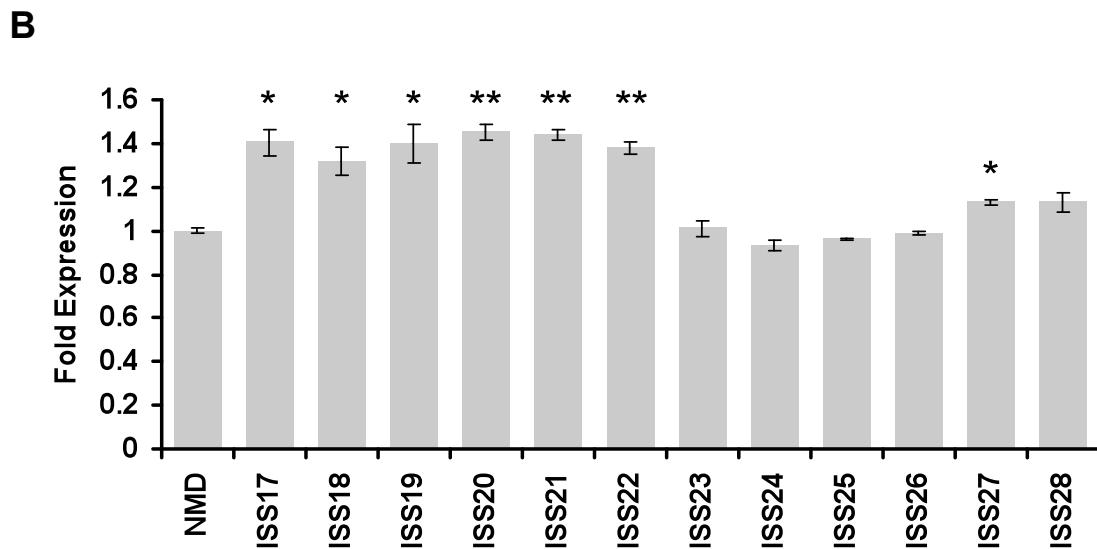


Figure S4. The activity of additional recovered ISRE sequences is validated by stable cell line assays

(A) Additional recovered ISRE sequences examined for regulatory activity. (B) Flow cytometry analysis of HEK-293 FLP-In stable cell lines generated for each recovered ISRE sequence and control construct. Mean GFP levels from two independent experiments were determined and normalized to the NMD control. The fold expression of each sample relative to NMD and average error are reported. Resulting *P*-values in comparison to the NMD control: * *P* < 0.05 and ** *P* < 0.01.

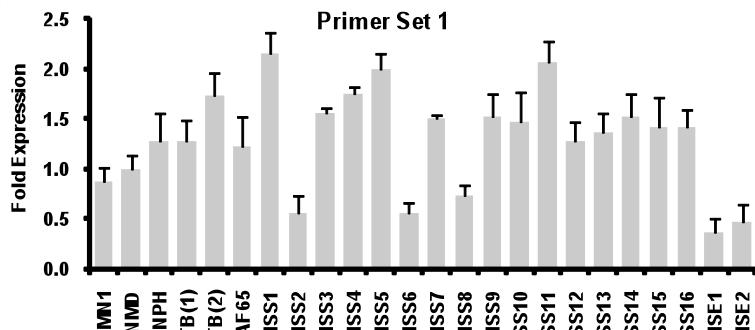
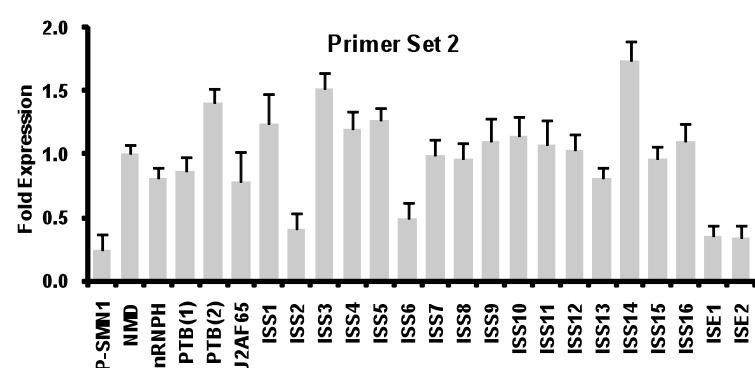
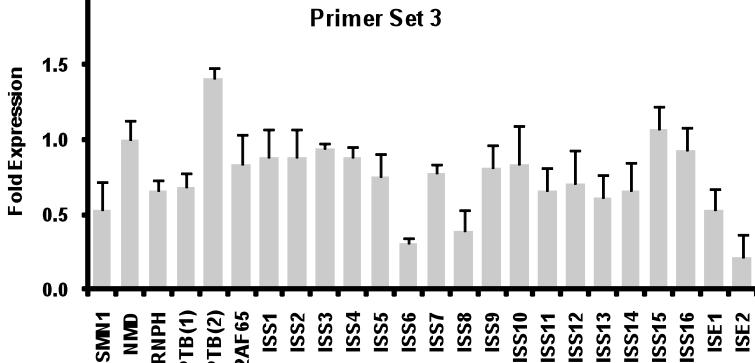
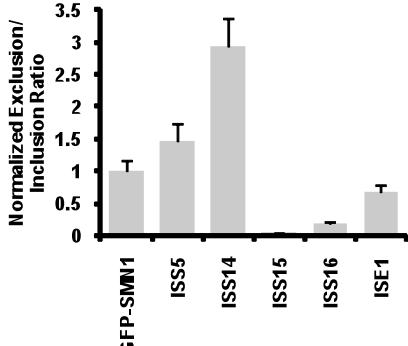
A**B****C****D**

Figure S5. Additional qRT-PCR isoform analysis of recovered ISREs and control constructs

(A) qRT-PCR analysis with primer set 1 (Figure 1, Supplementary Figure S1 and Supplementary Table S1). Results demonstrate that overall transcript levels for the GFP-SMN1, ISS controls, ISSs and ISEs did not significantly differ from the NMD control ($P = 0.2$). For all subsequent analyses, expression levels of duplicate PCR samples were normalized to the levels of *HPRT*. Fold expression data is reported as the mean expression for each sample divided by the mean NMD expression value \pm the average error. (B) qRT-PCR analysis with primer set 2. The levels of intron 6 retained in transcripts containing the selected and control ISS sequences are similar to the NMD control ($P = 0.65$). In contrast, intron 6 retention in ISE transcripts are similar to the GFP-SMN1 control ($P = 0.74$) and different from the NMD control ($P < 0.05$), suggesting that intron 6 in the GFP-SMN1 control and ISEs are processed similarly by the general splicing machinery. The retention level of intron 6 for the GFP-SMN1 control is statistically different from the NMD control ($P < 0.05$). (C) qRT-PCR analysis with primer set 3. The levels of intron 7 retention for the recovered and control ISS sequences and the GFP-SMN1 are similar to the NMD control ($P = 0.33$). The intron 7 retention levels in ISE transcripts are significantly different from the NMD control ($P < 0.05$). (D) qRT-PCR analysis with primer sets 4 and 5 on ISS5, ISS14-ISS16 and ISE1 inserted in the non-NMD-based GFP-SMN1 control construct. The transcript isoform analysis of stable cell lines demonstrates that the tested sequences maintain their regulatory activities (Figure 2B and C) in the non-NMD-based reporter. However, the transcript isoform levels of ISS15 and ISS16 displayed significant enhancer activity ($P < 0.05$), and do not correlate with measured fluorescence levels from the NMD-based reporter. The results suggest that ISS15 and ISS16 may exhibit enhanced fluorescence levels in the context of the NMD reporter due to the evasion of the NMD process. Data is reported as the expression ratio of the mean expression of the exon excluded isoform to the exon included isoform normalized to the ratio for the GFP-SMN1 control \pm the average error.

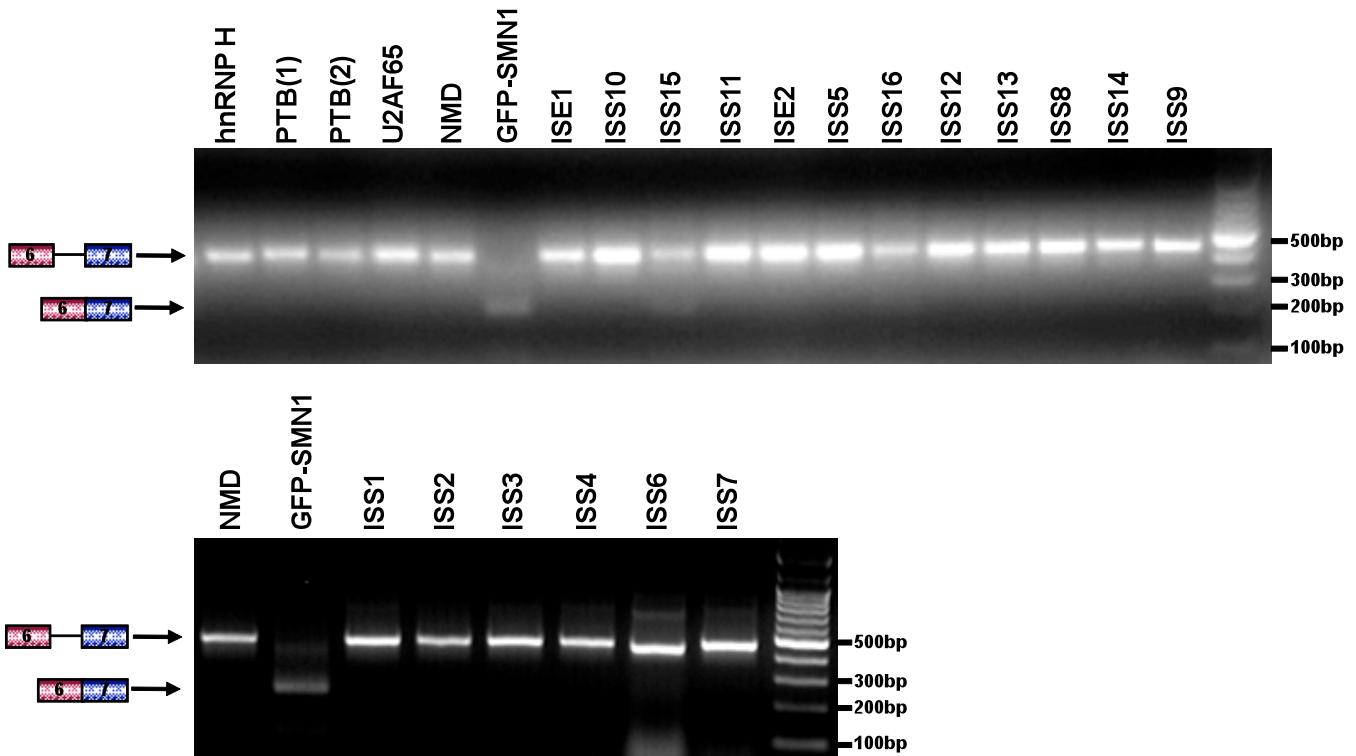


Figure S6. Examination of possible alternative 3' ss by qRT-PCR analysis of recovered ISREs and control constructs

We examined the possibility that selected ISREs and control constructs may include an alternative 3' ss by qRT-PCR analysis using the forward primer of primer set 1 (Figure 1 and Table S3) and a unique reverse primer for exon 7 (primer ex7, Table S1). The position of PCR products corresponding to the intron 6 retained and exon 7 included isoforms are indicated on the left. Using the above primer set, the expected sizes of the intron 6 retained and exon 7 included isoforms are 462 bp and 251 bp, respectively. Given the placement and length of our 15-mer library cassette (39-nt, Supplementary Figure S1), an ISRE with an alternative 3' ss would display the alternative 3' ss included isoform at a length between 297-329 bps. None of the recovered ISREs and control constructs display a PCR product within this range and therefore rule out the possibility that selected ISRE sequences may contain an alternative 3' ss. As shown above, the exon 7 included isoform was also detected in cell lines ISS15 and 16 as previously observed in our qRT-PCR analysis with primer set 4 (Figure 2C). While this data suggests that selected ISREs do not lead to alternative 3' ss processing, we cannot rule out the possibility of a minor change at the 3' ss due to aberrant splicing that would alter the reading frame of the PTC.

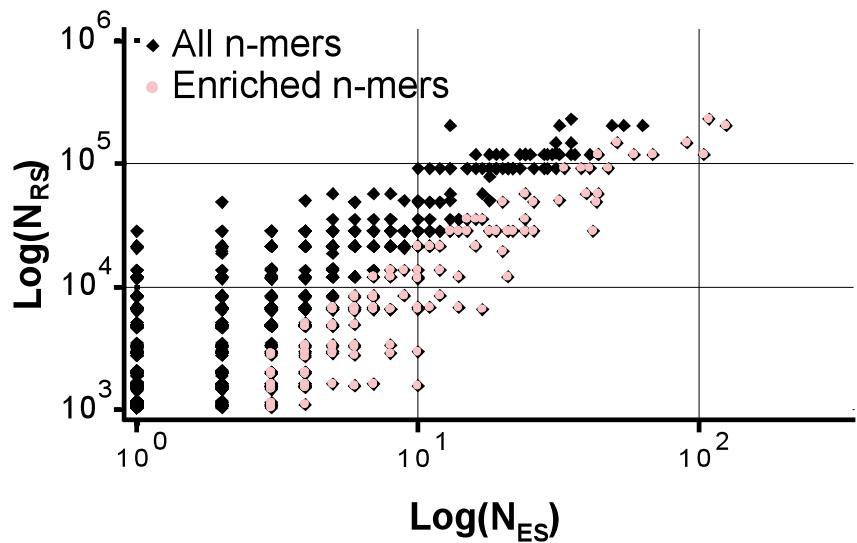


Figure S7. Scatter-plot for the occurrence frequency of all 4-6-nt n-mers in the ISRE sample set

Scatter-plot for the occurrence frequency of all 4-6-nt n-mers in the enriched sample set (N_{ES}) vs. a corresponding random sample set (N_{RS}) (black). A similar scatter-plot based on n-mers determined to be significantly enriched in the recovered ISREs is overlaid (pink).

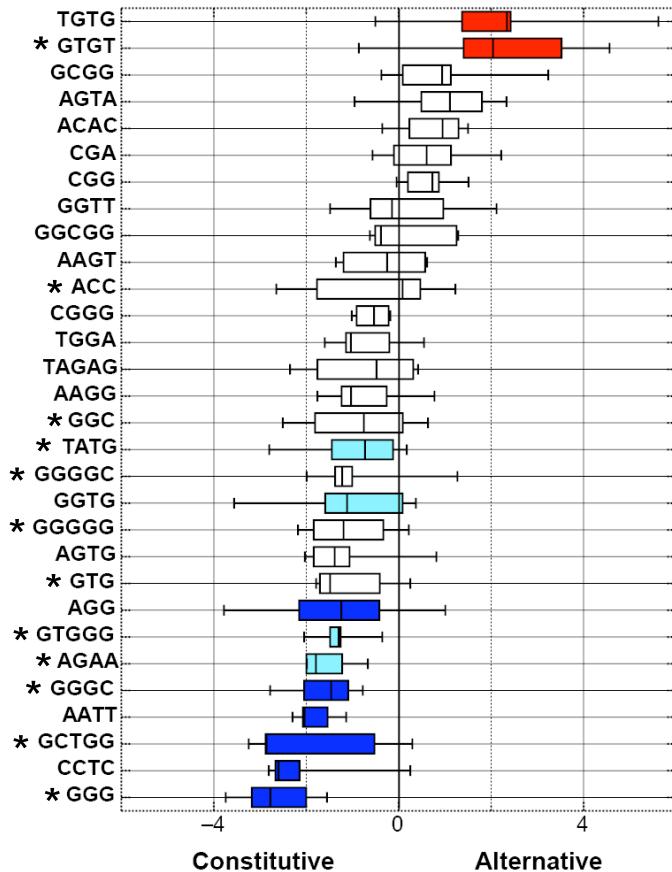


Figure S8. Enriched n-mers associate with constitutive and alternative splicing

Box-plots revealing the distribution of TA-scores for GCCS derived ISREs. The GCCS consensus motifs that are significantly associated with alternative splicing are shown in red ($P_{t-test} < 0.01$) and those that are significantly associated with constitutive splicing are shown in shades of blue (dark blue, $P_{t-test} < 0.01$; light blue, $P_{t-test} < 0.05$). In total, 9 consensus motifs are biased toward alternative splicing and 21 consensus motifs display a bias towards constitutive splicing. Elements exhibiting no significant association with either category are not shaded. Starred motifs are present in hexamers subjected to RNAi silencing studies to examine regulated splicing. The entire population of consensus n-mers significantly associates with constitutive splicing ($P_{t-test} = 1.8e^{-8}$). The stronger association with constitutively spliced exons may be a result of the selected ISREs functioning as ISSs, which have been shown to be enriched in the intronic flanks of constitutively spliced exons (1). A previous analysis of conserved intronic sequences revealed that a large number of motifs strongly associate with constitutive splicing and are more abundant than those associated with alternative splicing (2). Additional studies have also demonstrated that splice silencing may be a mechanism that represses pseudoexon inclusion (3) and that intronic sequences which repress splicing might have a fundamental role in defining real exons by silencing nearby decoy sites (4). In addition, several of our enriched motifs that associate with constitutive splicing also overlap with elements that have been previously identified upstream of constitutively spliced exons (5). Taken together, these observations are in line with results from our genome-wide analysis and suggest the utility of future computational investigations to determine the association between selected ISREs and pseudoexons.

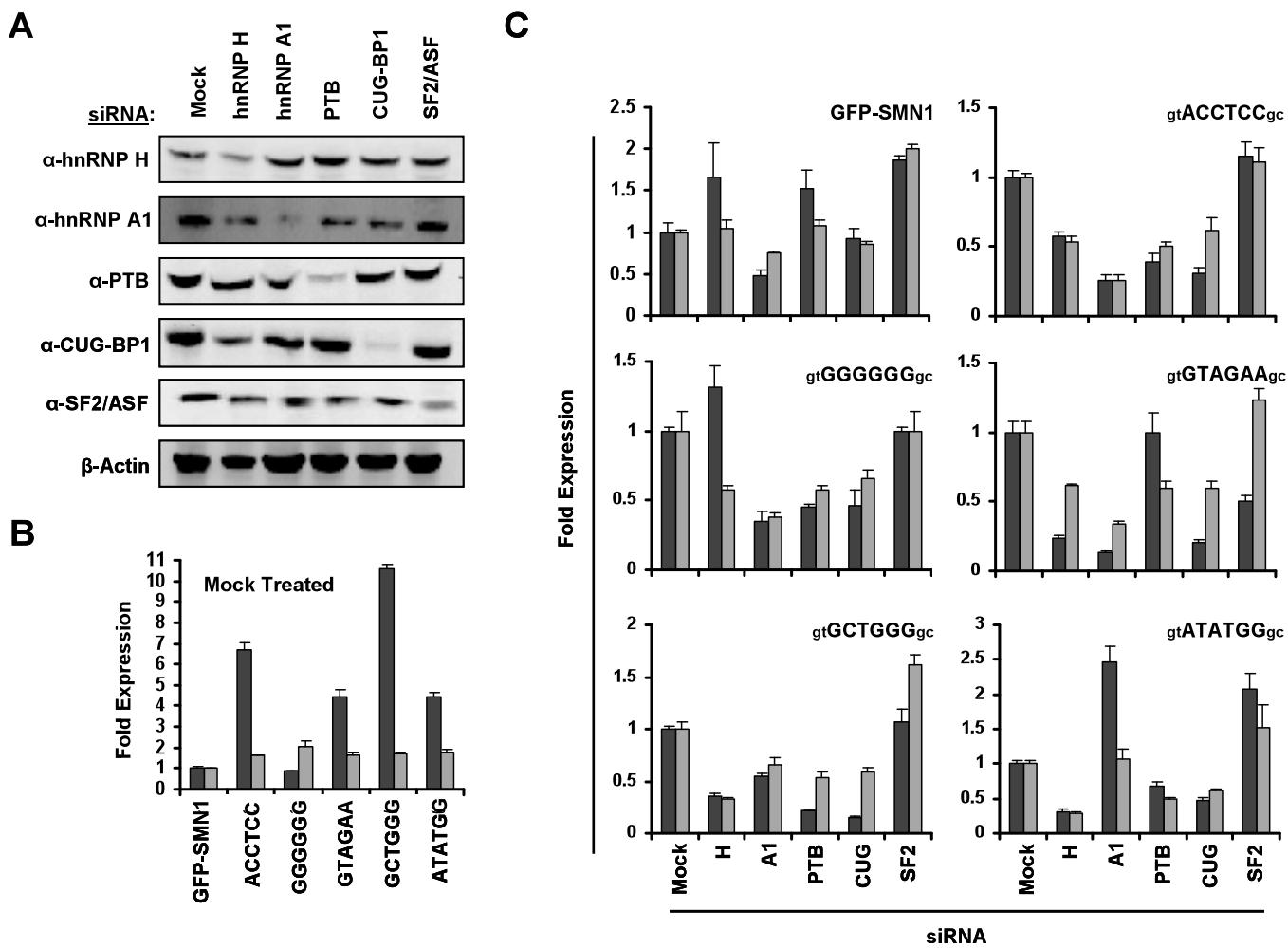


Figure S9. The effects of *in vivo* depletion of splicing factors on ISRE regulated splicing patterns

(A) Western blot analysis of total cell lysates prepared from the GFP-SMN1 control cell line treated with siRNAs targeted to trans-acting splicing factors and a mock siRNA negative control. The results demonstrate that individual siRNAs have minimal to no off-target affects. (B) qRT-PCR analysis of the mock treated ISRE hexamer and GFP-SMN1 control cell lines with primer sets specific for exon 7 excluded (black bars) and included (gray bars) products. Expression levels of duplicate PCR samples were normalized to the levels of *HPRT*. Fold expression data is reported as the mean expression for each sample divided by the mean GFP-SMN1 control expression value \pm the average error. (C) qRT-PCR analysis of the siRNA treated ISRE hexamer and GFP-SMN1 control cell lines with primer sets specific for exon 7 excluded (black bars) and included (gray bars) products. Fold expression data is reported as the mean expression for each sample divided by the mean mock siRNA treated cell line control expression value \pm the average error.

Table S1. Primer and oligonucleotide sequences

Name	Primer Sequence (5' - 3')
Ex6	CATGGACGAGCTGTACGTTAACATAATTCCCCCACCACCTC
Ex8	CGCTCG AGCACATACGCCACACATACATTG
GFP1	GCGGTACCATGGTGAGCAAGGGCG
GFP2	GGTGGTGGGGATTATGTTAACGTACAGCTCGTCCATGCC
ECmutF	CTTTTAACATCCATATAAAGCTATCGATATCTAGCTATCGAT GTCTATATAGCTATTTTTAACT
ECmutR	AGTTAAAAAAATAGCTATATAGACATCGATAGCTAGATATCG ATAGCTTATATGGATGTTAAAAAG
ISStemp	GCGCGATATCGATCAGT (N ₁₅) GCATCATCGATGC
Lib1	GCGCGATATCGATCAGT
Lib2	GCGCATCGATGATGC
Lib3	GAAACAAAATGCTTTAACATCCATA
Lib4	GGAAAATAAAAGGAAGTTAAAAAAATAGC
SMN1cDNA	TAGAAGGCACAGTCGAGG
Ex7	AAGGAATGTGAGCACCTT

Table S2. Plasmid constructs used in this work

Name	Description
pCS238	GFP-SMN1. Contains the wild-type SMN1 mini-gene fused to the N-terminus of GFP. Positive control used for all flow cytometry analysis.
pCS516	SMN1 NMD-based reporter construct. Contains the SMN1 mini-gene with a PTC in exon 7 fused to the N-terminus of GFP. Recovered ISREs as well as control ISS were inserted into this construct.
pCS517	SMN1 NMD-based containing random 15-mer. Negative control used for all flow cytometry analysis.
pCS668	U2AF65 binding site inserted into pCS516.
pCS669	hnRNP H binding site inserted into pCS516.
pCS670	PTB (1) binding site inserted into pCS516.
pCS667	PTB (2) binding site inserted into pCS516.

Table S3. Primer sequences for SMN1 transcript isoform analysis through qRT-PCR

Name	Forward Primer (5' - 3')	Reverse Primer (5' - 3')	Isoform
Pair 1	TGAGCAAAGACCCAA	TGATAGCCACTCATGTACC	GFP and Ex 6
Pair 2	CTCCCATATGTCCAGATTCT	AGCATTGTTTCACAAGACA	Ex 6 and Int 6
Pair 3	CACTAGTAGGCAGACCAG	CAGTTATCTTCTATAACGCTTCAC	Int 7 and Ex 8
Pair 4	TAAATTAAGGAGAAATGCT	GGTTTTCAAAAGAGTCCAGTAA	Ex 7/8 and Ex 8
Pair 5	TGAGCAAAGACCCAA	CCAGCATTCCATATAATAG	GFP and Ex 6/8
Pair 6	CAAAGATGGTCAAGGT CGCAAG	GGCGATGTCAATAGGACTCC	HPRT

Table S4. Primer sequences for endogenous transcript isoform analysis through qRT-PCR

Name	Gene	Hexamer	Sequence (5' – 3')	Isoform	Type of Alternative splicing
Fw.ADD3ex15_16	ADD3	ACCTCC	TGAAAAATTAGAAGAA AACCATGAGC	Exon15/16	cassette
Fw.ADD3ex14_16	ADD3	ACCTCC	GGCC TAG AAGAAA ACCATG AGC	Exon14/16	cassette
Rv.ADD3ex16	ADD3	ACCTCC	CTTCGATTTCTCTGGA GACT	ADD3 cDNA, Exon15/16, Exon 14/16	cassette
Fw.hnRNPCex1_3	HNRNPC	ACCTCC	CCC CTT CTT GTT TTC GGC TTT	Exon1/3	cassette
Fw.hnRNPCex2_3	HNRNPC	ACCTCC	CTT CAGCTACATTTC C GGCTT	Exon2/3	cassette
Rv.hnRNPCex3	HNRNPC	ACCTCC	CGAAAAGATTGCCTCC ACAT	hnRNPC cDNA and Exon1/3, Exon 2/3	cassette
Fw.CLK3ex4	CLK3	GGGGGG	CCGTGACAGCGATACA TAC	Exon 4/5, Exon4/6	cassette
Rv.CLK3ex4_5	CLK3	GGGGGG	GTTGGCTTCTCGAGGA GG	Exon 4/5	cassette
Rv.CLK3ex4_6	CLK3	GGGGGG	CCACAATCTCATCGAG GAGG	Exon 4/6	cassette
Rv.CLK3cDNA	CLK3	GGGGGG	CAAGCACTCCACCACC T	CLK3 cDNA	cassette
Fw.CADPSex16	CADPS	GGGGGG	GAAAGATATTGTTACC CCAGT	Exon 16/19, Exon16/18, Exon16/17	mutually exclusive
Rv.CADPSex16_18	CADPS	GGGGGG	CCTTTGATTCTCTTCG ATTTG	Exon16/18,	mutually exclusive
Rv.CADPSex16_19	CADPS	GGGGGG	GGCCTACATTTCTTCG ATTTG	Exon 16/19	mutually exclusive
Rv.CADPSex16_17	CADPS	GGGGGG	CTCTCTTTCCCTTCG ATTTG	Exon16/17	mutually exclusive
Rv.CADPScDNA	CADPS	GGGGGG	AAG CTT TTT GGC AGG AGT GA	CADPS cDNA	mutually exclusive
Fw.c6orf60ex15_16	C6orf60	GTAGAA	CTTACAAGTGTCAATTA GAAGAAATG	Exon 15/16	cassette
Fw.c6orf60ex14_16	C6orf60	GTAGAA	CCA ACA GAT AAG ATT AGA AGA AAT GG	Exon 14/16	cassette
Rv.c6orf60ex16	C6orf60	GTAGAA	GATCTGGTCTCTTCTG TAAGC	C6orf60 cDNA, Exon 15/16, Exon 14/16	cassette
Fw.RREB1ex11_12	RREB1	GTAGAA	GATAGCACAGACAGTC AGTCG	Exon11/12	cassette
Fw.RREB1ex10_12	RREB1	GTAGAA	ACA CAC ACT GAC AGT CAG TCG	Exon10/12	cassette
Rv.RREB1ex12	RREB1	GTAGAA	CTCCTCCTCCGGCTCAT	RREB1 cDNA, Exon11/12, Exon10/12	cassette

Fw.MADDex35	MADD	GCTGGG	AGTTCCCTGTGCGAC	Exon35/36, Exon35/37	cassette
Rv.MADDex35_36	MADD	GCTGGG	TCTATGAAAACCTGATT GTGCA	Exon35/36	cassette
Rv.MADDex35_37	MADD	GCTGGG	TAATTCAGGAAC TGAT TGTGCA	Exon35/37	cassette
Rv.MADDcDNA	MADD	GCTGGG	TAGTACAGCTCCCGAC ACTT	MADD cDNA	cassette
Fw.CAMK2Gex13_14	CAMK2G	GCTGGG	CGGGCAAGCTGCCAAA AG	Exon13/14	cassette
Fw.CAMK2Gex12_14	CAMK2G	GCTGGG	GAA CTT CTC AGC TGC CAA AAG	Exon12/14	cassette
Rv.CAMK2Gex14	CAMK2G	GCTGGG	TTGACACCGCCATCCG	CAMK2G cDNA, Exon13/14, Exon12/14	cassette
Fw.A2BP1ex15_17	A2BP1	ATATGG	GCAGACATTATGGTG GTTATG	Exon15/17	mutually exclusive
Fw.A2BP1ex16_17	A2BP1	ATATGG	TAA ATT GCT GCA GGG TGG TTA TG	Exon16/17	mutually exclusive
Rv.A2BP1ex17	A2BP1	ATATGG	CTGTCACTGTAGGCAG CG	A2BP1 cDNA, Exon15/17, Exon16/17	mutually exclusive
Fw.HNRNPA2B1ex1	HNRNPA2B1	ATATGG	CTCTAGCGGCAGTAGC A	Exon1/2, Exon1/3	cassette
Rv.HNRNPA2B1ex1_2	HNRNPA2B1	ATATGG	GT TTCTAAAGTTTCTC CATCGCG	Exon1/2	cassette
Rv.HNRNPA2B1ex1_3	HNRNPA2B1	ATATGG	GTTCCCTTTCTCTCTCC ATCGC	Exon1/3	cassette
Rv.HNRNPA2B1cDNA	HNRNPA2B1	ATATGG	CCTCAAACTTCTTCTG TGG	HNRNPA2B1 cDNA, Exon1/2, Exon1/3	cassette

Table S5. Identified ISRE regulatory sequences

ISS sequences	Name	Tested stably	Tested transiently	*Occurrence
GACGTGTGTCCTCGGG	ISE1	Y	Y	5
ATAGTGGCGGTGGAG	ISE2	Y	N	1
TACATCCCTCGGTTG	ISS1	Y	Y	2
AGAATAAGTGGGGTG	ISS2	Y	Y	1
AGTATATGGTAGGAA	ISS3	Y	Y	1
TGTTTGCGTCCAAG	ISS4	Y	Y	2
AGAATAAGTGAGGTG	ISS5	Y	Y	2
CCGAGTGCACGGGTG	ISS6	Y	Y	3
ACAGGCCAACGGGGGG	ISS7	Y	Y	1
CAAACACCTCCGATG	ISS8	Y	Y	20
GGTCGAGTCGCAAGG	ISS9	Y	Y	2
TAGGTGTGTCCTCGGG	ISS10	Y	Y	1
ACAGTGCTAAGTAGG	ISS11	Y	Y	4
AAAGACCGGGATATG	ISS12	Y	Y	7
AGTCACCTATTATAG	ISS13	Y	Y	5
TTGTAAGGTGCTGGG	ISS14	Y	Y	2
GGGGCGCGCGGGGGGG	ISS15	Y	Y	1
AGAGTGGGGCGGGTG	ISS16	Y	Y	1
GCAAGGTCCCTCTAG	ISS17	Y	N	4
GACGGAGCCGTCTGG	ISS18	Y	N	1
AGAGTGGCGGTGGAG	ISS19	Y	N	1
GATATGGCGAGGGTG	ISS20	Y	N	1
GGTGGCAGACACGAT	ISS21	Y	N	2
AAATAGAGGCCAG	ISS22	Y	N	1
TTATGGAGTTCTAG	ISS23	Y	N	8
GAGGGCAGTCAGTGG	ISS24	Y	N	1
TGGACACGTCAGTCA	ISS25	Y	N	3
TCTGACTCAATAGTA	ISS26	Y	N	1
AATTGGGTTGGGGGG	ISS27	Y	N	1
TATGACATGTGGGGA	ISS28	Y	N	1
CCGAGGAACCATAAGG				11
CCCTATGGTTCTCG				1
GACGGGTGCCTCGGG				2
GGCTGGAAGACCTGC				2
GGAGTGGCTGGTTCG				1
GGCTGGCTAGGATG				1
ACCTCAGGCTCTGAA				2
GACTGTGTTAGGC GG				1
AAAGAACGGGATATG				1
TCGAATCTCTCCAGT				1
CCTACGCTCATTATT				4
TCTTCTCTCTCTTC				1
TGTTCGCACCGCTGG				1
TGTTCGCACCACTGA				1

GTTAACCAACGATGG	1
GGTATCGAAAGTTGT	1
TACATCCAGAAGTCG	2
TGGACCAGGCGTAGC	1
CACACGTGAGAGAGA	3
GAAGGGCGACAGATA	1
AGAACGCTGGATTAA	1
TTACTTTAAGGATAA	2
ATACGGAAAGGCCTT	1
GTGCTTATATGGTT	1
TTAGTCCCATTCCGA	2
CCACTTCGTTGCCT	1
ACGTCCGTCGTGGAT	1
ACCTCGAGGTCTGAA	1
AAGGCTAGTTAGTA	1
AAGGCTAGATTAGTA	3
AGAGGAGTCGTGTCA	1
AGTGAATCGTATCA	1
ATTCCAGCTGGAGCT	3
GCCGAGTAAAGTGTA	1
CTTGAGTACCCCCGA	1
CATGCACCGACCAAG	1
AATTGTGTTGTGAT	1
AATTGTGTTGGCGG	7
TATGACGTGTGGGGG	1
TATGACATGTGGGGG	4
CAATTGAGTTGGTGT	1
CGATGGGGCAGGGGA	1
CAGTGAACCTTGCAG	1
CCTTGGTCCTGACAT	1
GAGTGGCCTAGGGAG	1
AAGTGGCACGGTTG	1
AGGTAGCCACCGTTG	1
GGGGGGGTCACTTAG	1
TGGTGGACCCGTAG	4
CTAGTAACCAGCCAG	1
CTAAGCACCCTGAG	1
CATGTCAGGACCAAG	2
CATGGACCGACCAAG	1
TATGCCTCCCCGATA	1
CGAAGAACCCCAAGG	1
CGGAGAAACCGGAGG	1
CTATCTCCTCTATG	1
TTAACACCTCCCAAG	1
CAAAGACCTGCGATG	1
CAAACACGTCCGATG	1

CTAACACCTCCGATG	2
GTGGCTAAGAATTGG	1
GTAAAGGGTGTCACT	1
ATTAATAATACTGGG	1
GTAAATAGCGCGGA	2
TGTGGTCGCGACCTG	1
GGCGGTGAGTACAG	1
GTTGTGAAAGAGGAG	1
GCGGTTGCGGGCGG	1
GCATGGCCCCGCTGG	1
GCACTAGAATCTGAG	1
GCAGTACGGGCTTAG	1
CGAGCGGCTTAGAG	1
AGAATGGACC GTGAG	1
GTACAGCGGAGAGGG	1
GTACGGTGCAGAGGG	2
GTAGTGTAGGGAGGG	1
GAAGTGTAGGGAGGG	1
ATACCGTTCA GTGGG	1
ATACCGTTCA GTGAG	3
AAAGGGGCAAGGTGG	1
AGAGTGCAGCGGG	1
GTAAATCGCGGGTG	1
GGAAATCGCGGGATG	1
GGCAATCGCGGGTG	1
CAGAGGAGTCTCTAG	1
CAAGACCGGGATATG	1
AATTATTAGTCGATG	2
GCTTAGTGAGTGATG	1
AGAAGACAAGTGGTG	1
GGTGAAAGGGGGCG	1
ACATTATGAGGGTCG	2
AGAGTAAGTGAGGTG	1
AATTGTGTTCGGTGG	1
GTGGCTATGAATTG	1

The splicing activity of the first 30 sequences was assessed by stable transfection assays. Additional sequences validated through transiently transfection assays are indicated (Y= Tested and N=Not tested). The occurrence of each sequence from the sequencing of 226 clones is also noted.*~30% of the recovered 15-mers were recovered more than once, which is likely due to assay conditions where enriched cell populations were grown for several weeks and then examined for sequence content.

TGTGTT	8	TGTG	6	'--TGTGTT	5.36891	4	5.36891	1	2	1
TTGTG	8	TGTG	5	'-TTGTG--	3.12078	7	3.12078	1	2	1
TTGTGT	8	TGTG	6	'-TTGTGT-	3.91375	3	3.91375	1	4	0.33
GGGGG	9	GGGGG	5	'-GGGGG-	11.2025	17	11.2025	1	3	1
GGGGGC	9	GGGGG	6	'-GGGGC	5.94795	6	5.94795	1	3	1
GGGGGG	9	GGGGG	6	'-GGGGGG	14.5575	10	14.5575	1	3	1
TGGGGG	9	GGGGG	6	'TGGGGG-	4.60874	4	4.60874	1	3	1
AAGTGG	10	AGTG	6	'-AAGTGG	3.88041	3	3.88041	2	2	1
AGAGTG	10	AGTG	6	'AGAGTG-	3.82024	3	3.82024	2	2	1
AGTGG	10	AGTG	5	'--AGTGG	5.97609	10	5.97609	2	2	1
GAGTG	10	AGTG	5	'-GAGTG-	3.7592	7	3.7592	2	2	1
GAGTGG	10	AGTG	6	'-GAGTGG	5.3668	4	5.3668	2	4	0.33
AGAGG	11	AGG	5	'-AGAGG--	3.84579	7	3.84579	4	13	1
AGAGGA	11	AGG	6	'-AGAGGA--	4.91846	3	4.91846	4	13	1
AGGC	11	AGG	4	'---AGGC--	5.02257	32	5.02257	4	6	1
AGGG	11	AGG	4	'---AGGG--	4.73884	21	4.73884	4	4	1
AGGGAG	11	AGG	6	'---AGGGAG	3.91151	3	3.91151	4	8	0.86
CCAGGC	11	AGG	6	'-CCAGGC--	2.5084	3	2.5084	4	6	1
CTAGGC	11	AGG	6	'-CTAGGC--	2.50462	3	2.50462	4	6	1
GAGG	11	AGG	4	'-GAGG--	6.56052	26	6.56052	4	13	1
GAGGA	11	AGG	5	'-GAGGA--	3.19377	5	3.19377	4	13	1
GAGGAG	11	AGG	6	'-GAGGAG-	5.56098	4	5.56098	4	14	0.92
GAGGC	11	AGG	5	'-GAGGC--	3.69855	10	3.69855	4	17	0.68
GAGGG	11	AGG	5	'-GAGGG--	4.57631	8	4.57631	4	15	0.83
GAGGGG	11	AGG	6	'-GAGGGG-	5.4483	4	5.4483	4	15	0.83
GGAGG	11	AGG	5	'-GGAGG--	3.8143	7	3.8143	4	14	0.92
GGAGGC	11	AGG	6	'-GGAGGC--	3.56828	4	3.56828	4	18	0.64
GGAGGG	11	AGG	6	'-GGAGGG--	3.88979	3	3.88979	4	15	0.83
GGGAGG	11	AGG	6	'GGGAGG--	4.00978	3	4.00978	4	14	0.92
TAGGC	11	AGG	5	'-TAGGC--	2.18095	8	2.18095	4	6	1
TGAGG	11	AGG	5	'-TGAGG--	4.38416	9	4.38416	4	13	1
TGAGGC	11	AGG	6	'-TGAGGC--	3.26508	4	3.26508	4	17	0.68
ATATG	12	TATG	5	'-ATATG--	3.08677	6	3.08677	4	10	1
ATATGG	12	TATG	6	'-ATATGG-	8.48791	6	8.48791	4	10	1
GATATG	12	TATG	6	'GATATG--	5.43525	4	5.43525	4	10	1
GTTATG	12	TATG	6	'GTTATG--	3.23123	4	3.23123	4	10	1
TATG	12	TATG	4	'--TATG--	1.68025	15	1.68025	4	10	1
TATGA	12	TATG	5	'--TATGA-	2.33239	5	2.33239	4	10	1
TATGAC	12	TATG	6	'-TATGAC	3.88703	3	3.88703	4	10	1
TATGG	12	TATG	5	'-TATGG-	4.37004	9	4.37004	4	10	1
TATGGC	12	TATG	6	'-TATGGC	4.25135	5	4.25135	4	10	1
TTATG	12	TATG	5	'-TTATG--	2.39546	6	2.39546	4	10	1
TTATGA	12	TATG	6	'-TTATGA-	5.42875	4	5.42875	4	10	1
AAGACC	13	ACC	6	'AAGACC---	6.67681	4	6.67681	4	6	1
ACCAA	13	ACC	5	'---ACCAA-	2.32261	4	2.32261	4	3	1
ACCAAG	13	ACC	6	'---ACCAAG	3.88703	3	3.88703	4	3	1
AGACC	13	ACC	5	'-AGACC--	2.30871	4	2.30871	4	6	1
GACC	13	ACC	4	'-GACC--	2.55586	12	2.55586	4	6	1
GACCA	13	ACC	5	'-GACCA--	2.32994	4	2.32994	4	8	0.64
GACCAA	13	ACC	6	'-GACCAA-	4.79991	3	4.79991	4	8	0.64
GACCTG	13	ACC	6	'-GACCTG-	3.91881	3	3.91881	4	6	1
GGACC	13	ACC	5	'-GGACC--	3.18228	5	3.18228	4	6	1
AATCGG	14	CGG	6	'AATCGG--	4.01211	3	4.01211	4	5	1
ATCGGC	14	CGG	6	'-ATCGGC-	2.53593	3	2.53593	4	5	1
CGGGG	14	CGG	5	'-CGGGG	2.33353	5	2.33353	4	2	1
CTCGG	14	CGG	5	'-CTCGG-	2.29696	5	2.29696	4	5	1
CTCGGG	14	CGG	6	'-CTCGGG-	3.84602	3	3.84602	4	6	0.73
TCGGCG	14	CGG	6	'-TCGGCG	3.28023	3	3.28023	4	5	1
TCGGGG	14	CGG	6	'-TCGGGG	3.32851	3	3.32851	4	6	0.73
AGAA	15	AGAA	4	'-AGAA-	2.16143	11	2.16143	4	4	1
AGAAC	15	AGAA	5	'-AGAAC-	3.21047	5	3.21047	4	4	1
GTAGAA	15	AGAA	6	'GTAGAA-	4.74249	5	4.74249	4	4	1
TAGAA	15	AGAA	5	'-TAGAA-	3.13329	6	3.13329	4	4	1
TAGAAC	15	AGAA	6	'-TAGAAC-	5.47463	4	5.47463	4	4	1
AATTG	16	AATT	5	'-AATTG-	3.14001	6	3.14001	4	4	1
AATTGT	16	AATT	6	'-AATTGT-	4.87458	3	4.87458	4	4	1
GTAATT	16	AATT	6	'GTAATT--	4.68928	5	4.68928	4	4	1
TAATT	16	AATT	5	'-TAATT--	2.33437	5	2.33437	4	4	1
TAATTG	16	AATT	6	'-TAATTG-	4.73989	4	4.73989	4	4	1
ACCTC	17	CCTC	5	'-ACCTC--	3.12256	5	3.12256	4	5	1

ACCTCC	17	CCTC	6	'-ACCTCC-	4.86105	3	4.86105	4	5	1
CACCTC	17	CCTC	6	'CACCTC--	5.06762	3	5.06762	4	5	1
CCTC	17	CCTC	4	'--CCTC--	1.66665	10	1.66665	4	5	1
CCTCC	17	CCTC	5	'--CCTCC-	2.2766	4	2.2766	4	5	1
CCTCGG	17	CCTC	6	'--CCTCGG	3.84116	3	3.84116	4	5	1
AGGGC	18	GGGC	5	'-AGGGC-	2.06499	7	2.06499	4	5	1
CTGGGC	18	GGGC	6	'CTGGGC-	3.81387	4	3.81387	4	5	1
GGGC	18	GGGC	4	'--GGGC-	8.06106	43	8.06106	4	5	1
GGGCA	18	GGGC	5	'--GGGCA	2.33091	4	2.33091	4	5	1
GGGCG	18	GGGC	5	'--GGGCG	2.32071	5	2.32071	4	5	1
TGGGC	18	GGGC	5	'-TGGGC-	3.22006	10	3.22006	4	5	1
AGTA	19	AGTA	4	'-AGTA--	2.15759	11	2.15759	4	4	1
AGTAGC	19	AGTA	6	'-AGTAGC	2.50008	3	2.50008	4	4	1
GAGTA	19	AGTA	5	'GAGTA--	2.29532	4	2.29532	4	4	1
TAGTA	19	AGTA	5	'TAGTA--	2.37311	5	2.37311	4	4	1
TAGTAG	19	AGTA	6	'TAGTAG-	3.34476	3	3.34476	4	4	1
AGTG	20	GTG	4	'-AGTG--	5.8182	24	5.8182	4	3	1
AGTGA	20	GTG	5	'-AGTGA-	4.02655	6	4.02655	4	5	0.6
AGTGAG	20	GTG	6	'-AGTGAG	5.35549	4	5.35549	4	5	0.6
CACTG	20	GTG	5	'CAGTG--	3.05845	6	3.05845	4	3	1
GTGA	20	GTG	4	'--GTGA-	1.74258	20	1.74258	4	3	1
GTGAG	20	GTG	5	'--GTGAG	2.68747	9	2.68747	4	3	1
CGGTT	21	GGTT	5	'CGGTT--	2.2958	4	2.2958	4	4	1
CGGTTG	21	GGTT	6	'CGGTTG-	3.90035	3	3.90035	4	4	1
GGTT	21	GGTT	4	'-GGTT--	1.71456	10	1.71456	4	4	1
GGTTG	21	GGTT	5	'-GGTTG-	3.06306	6	3.06306	4	4	1
GGTTGG	21	GGTT	6	'-GGTTGG	3.92331	3	3.92331	4	4	1
AGTGTA	22	GTGT	6	'AGTGTA-	4.85298	3	4.85298	4	4	1
GTGT	22	GTGT	4	'-GTGT--	3.36222	26	3.36222	4	4	1
GTGTA	22	GTGT	5	'-GTGTA-	2.6027	8	2.6027	4	4	1
GTGTAG	22	GTGT	6	'-GTGTAG	3.26747	4	3.26747	4	4	1
GTGTT	22	GTGT	5	'-GTGTT-	2.027	7	2.027	4	4	1
CGGG	23	CGGG	4	'CGGG--	3.68909	18	3.68909	4	3	1
CGGGA	23	CGGG	5	'CGGGA-	2.28819	4	2.28819	4	3	1
CGGGT	23	CGGG	5	'CGGGT-	2.35486	4	2.35486	4	3	1
CGGGTG	23	CGGG	6	'CGGGTG	5.57367	4	5.57367	4	3	1
GCGG	24	GCGG	4	'GCGG--	3.64572	18	3.64572	4	4	1
GCGGG	24	GCGG	5	'GCGGG-	6.1336	10	6.1336	4	4	1
GCGGGC	24	GCGG	6	'GCGGGC	3.67273	4	3.67273	4	4	1
GCGGGT	24	GCGG	6	'GCGGGT	5.00325	3	5.00325	4	4	1
GCGGT	24	GCGG	5	'GCGGT-	2.29151	4	2.29151	4	4	1
AACAC	25	ACAC	5	'AACAC--	2.28582	4	2.28582	4	3	1
AACACC	25	ACAC	6	'AACACC-	5.00613	3	5.00613	4	3	1
ACACCT	25	ACAC	6	'-ACACCT	4.95198	3	4.95198	4	3	1
ACACGT	25	ACAC	6	'-ACACGT	4.82098	3	4.82098	4	3	1
AAGT	26	AAGT	4	'-AAGT-	1.74801	10	1.74801	4	3	1
AAGTGT	26	AAGT	5	'-AAGTG	3.8693	7	3.8693	4	3	1
TAAGT	26	AAGT	5	'TAAGT-	2.35658	5	2.35658	4	3	1
TAAGTG	26	AAGT	6	'-TAAGTG	4.70162	4	4.70162	4	3	1
AGTGGC	27	GGC	6	'AGTGGC--	3.54163	4	3.54163	5	7	1
ATGGC	27	GGC	5	'-ATGGC---	5.93424	14	5.93424	5	7	1
GATGGC	27	GGC	6	'GATGGC--	8.14456	8	8.14456	5	7	1
GGCT	27	GGC	4	'--GGCT--	2.13292	11	2.13292	5	5	1
GGCTA	27	GGC	5	'--GGCTA-	3.17886	5	3.17886	5	5	1
GGCTAG	27	GGC	6	'--GGCTAG	3.92556	3	3.92556	5	5	1
GTGGC	27	GGC	5	'-GTGGC--	6.4307	20	6.4307	5	7	1
GTGGCT	27	GGC	6	'-GTGGCT--	4.71178	5	4.71178	5	10	0.67
TGGC	27	GGC	4	'-TGGC--	6.24044	40	6.24044	5	7	1
TGGCT	27	GGC	5	'-TGGCT--	2.34889	5	2.34889	5	10	0.67
TGGCTG	27	GGC	6	'-TGGCTG-	3.2832	3	3.2832	5	10	0.67
CCGA	28	CGA	4	'-CCGA--	2.13739	11	2.13739	5	6	1
CCGAG	28	CGA	5	'-CCGAG--	2.35878	5	2.35878	5	6	1
CCGAT	28	CGA	5	'-CCGAT--	2.30823	4	2.30823	5	8	0.71
CCGATG	28	CGA	6	'-CCGATG-	3.83631	3	3.83631	5	8	0.71
CGATG	28	CGA	5	'-CGATG-	4.61974	8	4.61974	5	4	1
CGATGG	28	CGA	6	'-CGATGG	9.90067	7	9.90067	5	4	1
TCCGA	28	CGA	5	'TCCGA--	3.05931	6	3.05931	5	6	1
TCCGAG	28	CGA	6	'TCCGAG--	3.32607	3	3.32607	5	6	1
TCCGAT	28	CGA	6	'TCCGAT--	3.83954	3	3.83954	5	8	0.71
AAGGGG	29	GGG	6	'AAGGGG	3.88538	3	3.88538	5	3	1

AGGGG	29	GGG	5	'-AGGGG	4.57194	8	4.57194	5	3	1
CTGGG	29	GGG	5	'CTGGG-	3.22319	6	3.22319	5	2	1
GGGG	29	GGG	4	'-GGGG	12.2506	42	12.2506	5	3	1
TGGG	29	GGG	4	'-TGGG-	4.60088	24	4.60088	5	2	1
TGGGG	29	GGG	5	'-TGGGG	6.39366	12	6.39366	5	5	0.4
GTTGGA	30	TGGA	6	'GTTGGA--	2.49556	3	2.49556	5	4	1
TGGA	30	TGGA	4	'-TGGA--	2.55883	15	2.55883	5	4	1
TGGAC	30	TGGA	5	'-TGGAC-	2.33125	5	2.33125	5	4	1
TGGACC	30	TGGA	6	'-TGGACC	5.45924	4	5.45924	5	4	1
TTGGAC	30	TGGA	6	'-TTGGAC-	3.93915	3	3.93915	5	4	1

Table S8. Summary of the enriched ISRE n-mers and GCCS clustering performance

	ISRE sequences	Random Sample
Total n-mers	5376	5376
Probability >Cl _{high} (2)	241	91
Clustered	193	64
% clustered	80.1%	70.33%
Number of Clusters	30	11

Table S9. Detailed comparison of GCCS clusters consensus motifs to known trans-acting factor binding sites

Class	Pictogram	Similar To
G-rich		hnRNP F/H consensus binding site (GGGGG) (6), which functions as either a splicing enhancer or silencer (7). Contains a G-triplet, a known ISE sequence (8) that is abundant in mammalian introns (9).
G-rich		High affinity hnRNPA1 binding site (TAGGG) identified by SELEX (10). Contains a G-triplet, a known ISE sequence (8) that is abundant in mammalian introns (9).
Other		hnRNP A1 binding site (TAGAGT) (11)
Other		High affinity hnRNP L binding site (CA-rich) identified by SELEX and an ISE element comprised of variable-length CA repeats (12). A/C-rich ESSs (13).
Other		CTCC and CCTCCC repeats identified by computational analysis of introns flanking skipped exons (14). CT-rich intronic sequences that act as PTB binding sites (15,16).
Other		SRp40 binding site (ACAAG) (17).
Other		SC35 binding site (AGGAGAT) (18). A purine-rich element (AGGG) identified in introns flanking skipped exons (14).
Other		Sam68 binding site (AAAA) (19,20).
Other		9G8 high-affinity binding site (GAC) identified by SELEX (18).
Other		SF2/ASF high-affinity binding site (GAAGAA) identified by SELEX (21). Tra2β high-affinity binding site (GAA) _n identified by SELEX (22).
Other		Srp30c consensus sequence (CTGGATT) (23).
GT-rich		hnRNP G binding motif (AAGT) (24).
GT-rich		CELF/Bruno-like family of proteins that bind GT repeats with high affinity (25). CUG-BP1 binding sites consisting of TGT-repeats (25). hnRNP M binding sites consisting of poly(G) and poly(T) homopolymers (26).
GT-rich		CELF/Bruno-like family of proteins that bind GT repeats with high affinity (25).
GT-rich		CELF/Bruno-like family of proteins that bind GT repeats with high affinity (25). CUG-BP1 binding sites consisting of TGT-repeats (25).
GT-rich		CELF/Bruno-like family of proteins that bind GT repeats with high affinity (25).

Table S10. ISRE pentamers that do not resemble known splicing regulatory elements

Field	Description							
n-mer	The n-mer (5mers)							
GCS	Greatest Common Substring							
clustID	ClusterID							
Both Intronic and Exonic		Intronic elements						
n-mer	GCS	clustID	n-mer	GCS	clustID	n-mer	GCS	clustID
CGATG	CGA	28	GTGGC	GGC	27	GGGGC	GGGGC	4
*TAGAG	TAGAG	7	CAAGG	AAGG	2	GCGGG	GC GG	24
AAGGC	AAGG	2	AGTGA	GTG	20	TGGGC	GGGC	18
CCGAT	CGA	28	AAGTG	AAGT	26	GGCTA	GGC	27
CGGTT	GGTT	21	AGAAT	AGAA	15	AATTG	AATT	16
GCGGT	GCGG	24	GAGGA	AGG	11	CGGGT	CGGG	23
			GGACC	ACC	13	GGGCG	GGGC	18
			ACCTC	CCTC	17	AGGGC	GGGC	18
			ATATG	TATG	12			
			TCCGA	CGA	28			
			CAGTG	GTG	20			
			GTGTA	GTGT	22			
			CGGTG	GGTG	1			
			TAGTA	AGTA	19			
			TATGA	TATG	12			
			TGGAC	TGGA	30			
			GACCA	ACC	13			
			ACCAA	ACC	13			
			AGACC	ACC	13			
			CTCGG	CGG	14			
			GAGTA	AGTA	19			
			AACAC	ACAC	25			
			CCTCC	CCTC	17			

n-mers which do not overlap with known intronic and exonic regulatory elements were placed under the ‘both intronic and exonic’ heading. Enriched ISRE pentamers that do not overlap with known intronic regulatory elements were placed under the intronic element heading and similarly for elements which do not resemble exonic regulatory elements. *The TAGAG was found to overlap with an element identified upstream of constitutively spliced exons (5).

Table S11. Overlap of enriched hexamers with extended recovered ISRE sequences

Extended ISS sequence	Enriched Hexamers
GTCGAATCTCTCCAGTGC	
GTCCTACGCTCATTATTGC	
GTTCTTCTCTCTCTTCGC	
GTTGTTCGCACCGCTGGGC	CGCTGG CTGGGC GCTGGG TGTTCG
GTTGTTCGCACCACTGAGC	TGTTCG
GTAGTCACCTATTATAGGC	
GTGTTAACCAACGATGGGC	CGATGG
GTGGTATCGAAAGTTGTGC	
GTTACATCCAGAAGTCGGC	
GTTACATCCCTCGGTTGGC	CCTCGG CGGTTG GGTTGG
GTTGGACCAGGCGTACGGC	CCAGGC GTTGA TGGACC TTGGAC
GTTGGACACGTCAGTCAGC	ACACGT GTTGA TTGGAC
GTCACACGTGAGAGAGAGC	ACACGT
GTGAAGGGCGACAGATAAGC	AAGGGC
GTAGAACGCTGGATTAAGC	CGCTGG GCTGGA GTAGAA
GTTTACTTTAAGGATAAGC	
GTATACGGAAAGGCCTTGC	GTATAC
GTGTGCTTATATGGGTTGC	ATATGG
GTTTAGTCCCATTCCGAGC	TCCGAG
GTCCACTTCGGTTGCCTGC	CGGTTG
GTACGTCCCGTCGTGGATGC	
GTACCTCGAGGTCTGAAGC	
GTACCTCAGGCTCTGAAGC	
GTAAGGCTAGTTAGTAGC	AGTAGC GGCTAG
GTAAGGCTAGATTAGTAGC	AGTAGC GGCTAG
GTAGAGGAGTCGTGTCAAGC	AGAGGA GTAGAG TAGAGG TGTCAG
GTAGTGGAATCGTATCAGC	

GTGGTCGAGTCGCAAGGGC	AAGGGC	CAAGGG	GCAAGG			
GTATTCCAGCTGGAGCTGC		GCTGGA				
GTAGTATATGGTGAGGAGC	ATATGG	GTGAGG				
GTGCCGAGTAAAGTGTAGC	AGTGTA	GTAAAG	GTGTAG			
GTTCTGACTCAATAGTAGC	AGTAGC					
GTCTTGAGTACCCCCGAGC						
GTCATGCACCGACCAAGGC	ACCAAG	CAAGGC	CCAAGG	GACCAA		
GTAATTGTGTTGTGATGC	AATTGT	ATTGTG	GTAATT	TAATTG	TGTGTT	TTGTGT
GTGACTGTGTTAGGCGGGC	GGCGGG	TGTGTT				
GTAATTGGGTTGGGGGGC	GGGGGC	GGGGGG	GTAATT	TAATTG	TGGGGG	
GTAATTGTGTTCGGTGGGC	AATTGT	ATTGTG	CGGTGG	GTAATT	GTGGGC	TAATTG
GTAATTGTGTTGGCGGGC	TGTGTT	TGTTCG	TTGTGT			
GTTATGACATGTGGGGAGC	AATTGT	ATTGTG	GGCGGG	GTAATT	TAATTG	TGGCGG
GTTATGACATGTGGGGGGC	TGTGTT	TTGTGT	GACATG	GTGGGG	GTTATG	TGACAT
GTTATGACATGTGGGGGGC	TTATGA		TATGAC		TGTGGG	
GTTATGACATGTGGGGGGC	GGGGGC	GGGGGG	GGGGGG	GTTATG	TATGAC	TGGGGG
GTTATGACATGTGGGGGGC	TGTGGG	TTATGA	GGGGGC	GTGGGG	GTTATG	TATGAC
GTTATGACATGTGGGGGGC	GACATG	GGGGGC	GGGGGG	TTATGA		
GTCACATTGAGTTGGTGTGC	TGACAT	TGGGGG	TGTGGG			
GTCGATGGGGCAGGGGAGC		CGATGG	TGGGGC			
GTCAGTGAACTTGCGAGC		TCAGTG	TTTGC			
GTCCTTGGTCCTGACATGC		GACATG	TGACAT			
GTCCGAGTGCACGGTGGC		CGGTGG	GGTGGC	TCCGAG		
GTGAGTGGCCTAGGGAGGC	AGGGAG	AGTGGC	GAGTGG	GGAGGC	GGGAGG	TAGGGA
GTGGCTGGCTAGGATGGC	TAGTAG					
GTGATATGGCGAGGGTGGC	CTGGGC	GATGGC	GCTGGG	GGCTAG	GGCTGG	GTGGCT
GTGATATGGCGAGGGTGGC	TGGCTG					
GTGATATGGCGAGGGTGGC	ATATGG	GATATG	GGGTGG	GGTGGC	TATGGC	
GTAAGTGGGCACGGTTGGC	AAGTGG	AGTGGG	CGGTTG	GGTTGG	GTGGGC	TAAGTG
GTAGGTAGCCACCGTTGGC		ACCGTT				
GTGGGGGGGTCACTTAGGC	GGGGGG	GTGGGG	TGGGGG			
GTTGGTTGGACCCGTAGGC	GGTTGG	GTTGGA	TGGACC	TTGGAC		
GTCCCTATGGTCCTCGGC		CCTCGG				
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GTAAATAGAGGCCAGGC	CCAGGC TAGAGG
GTCTAGTAACCAGCCAGGC	CCAGGC
GTCTAAGCACCAC TGAGGC	TGAGGC
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GTCATGGACCGACCAAGGC	ACCAAG CAAGGC CCAAGG GACCAA TGGACC
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GTCGGAGAAACCGGAGGGC	GGAGGG
GTCCGAGGAACCATAAGGC	TCCGAG
GTCTATCTCCTTCTATGGC	TATGGC
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GTCGAGCGGCTTAGAGGC	TAGAGG					
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	GGGGCG	GGGTGG	GGTGGC	GTAGAG	GTGGG	TAGAGT
TGGGGC						
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	TAGAGT	TGGCGG	GTAGAG			
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GTGTACGGTGCAGAGGGC	AGGGC					
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	TAGTAG	TAGGGA				
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	TAGTAG	TAGGGA				
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GTAGAGTGCAGCGGGC	AGAGTG	CGGGC	GTAGAG	TAGAGT		
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	TCGGCG	GGTGGC				
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GTGGCAATCGCGGGGTGGC	AATCGG	ATCGGC	CGGGTG	GGCGGG	GGGTGG	GGTGGC
TCGGCG						
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TATGGC						
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TATGGC						
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TATGGC	GTAAAG					
GTAATTATTAGTCGATGGC	CGATGG	GATGGC	GTAATT			
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GTAGAAGACAAGTGGTGGC	AAGTGG	GGTGGC	GTAGAA			
GTGGTTGAAGGGGGCGGC	AAGGGG	GGCGGG	GGGGCG	GGGGGC	GGGGGG	
GTACATTATGAGGGTCGGC	TTATGA					
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TAGAAT						
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TAGAAT	TAAGTG					

References

1. Blencowe, B.J. (2006) Alternative splicing: new insights from global analyses. *Cell*, **126**, 37-47.
2. Voelker, R.B. and Berglund, J.A. (2007) A comprehensive computational characterization of conserved mammalian intronic sequences reveals conserved motifs associated with constitutive and alternative splicing. *Genome Res*, **17**, 1023-1033.
3. Fairbrother, W.G. and Chasin, L.A. (2000) Human genomic sequences that inhibit splicing. *Mol Cell Biol*, **20**, 6816-6825.
4. Wang, Z., Rolish, M.E., Yeo, G., Tung, V., Mawson, M. and Burge, C.B. (2004) Systematic identification and analysis of exonic splicing silencers. *Cell*, **119**, 831-845.
5. Zhang, X.H., Heller, K.A., Hefter, I., Leslie, C.S. and Chasin, L.A. (2003) Sequence information for the splicing of human pre-mRNA identified by support vector machine classification. *Genome Res*, **13**, 2637-2650.
6. Markovtsov, V., Nikolic, J.M., Goldman, J.A., Turck, C.W., Chou, M.Y. and Black, D.L. (2000) Cooperative assembly of an hnRNP complex induced by a tissue-specific homolog of polypyrimidine tract binding protein. *Mol Cell Biol*, **20**, 7463-7479.
7. Chou, M.Y., Rooke, N., Turck, C.W. and Black, D.L. (1999) hnRNP H is a component of a splicing enhancer complex that activates a c-src alternative exon in neuronal cells. *Mol Cell Biol*, **19**, 69-77.
8. McCullough, A.J. and Berget, S.M. (1997) G triplets located throughout a class of small vertebrate introns enforce intron borders and regulate splice site selection. *Mol Cell Biol*, **17**, 4562-4571.
9. Yeo, G., Hoon, S., Venkatesh, B. and Burge, C.B. (2004) Variation in sequence and organization of splicing regulatory elements in vertebrate genes. *Proc Natl Acad Sci U S A*, **101**, 15700-15705.
10. Burd, C.G. and Dreyfuss, G. (1994) RNA binding specificity of hnRNP A1: significance of hnRNP A1 high-affinity binding sites in pre-mRNA splicing. *Embo J*, **13**, 1197-1204.
11. Hutchison, S., LeBel, C., Blanchette, M. and Chabot, B. (2002) Distinct sets of adjacent heterogeneous nuclear ribonucleoprotein (hnRNP) A1/A2 binding sites control 5' splice site selection in the hnRNP A1 mRNA precursor. *J Biol Chem*, **277**, 29745-29752.
12. Hui, J., Stangl, K., Lane, W.S. and Bindereif, A. (2003) HnRNP L stimulates splicing of the eNOS gene by binding to variable-length CA repeats. *Nat Struct Biol*, **10**, 33-37.
13. Coulter, L.R., Landree, M.A. and Cooper, T.A. (1997) Identification of a new class of exonic splicing enhancers by in vivo selection. *Mol Cell Biol*, **17**, 2143-2150.
14. Miriami, E., Margalit, H. and Sperling, R. (2003) Conserved sequence elements associated with exon skipping. *Nucleic Acids Res*, **31**, 1974-1983.
15. Chan, R.C. and Black, D.L. (1995) Conserved intron elements repress splicing of a neuron-specific c-src exon in vitro. *Mol Cell Biol*, **15**, 6377-6385.
16. Chou, M.Y., Underwood, J.G., Nikolic, J., Luu, M.H. and Black, D.L. (2000) Multisite RNA binding and release of polypyrimidine tract binding protein during the regulation of c-src neural-specific splicing. *Mol Cell*, **5**, 949-957.
17. Liu, H.X., Zhang, M. and Krainer, A.R. (1998) Identification of functional exonic splicing enhancer motifs recognized by individual SR proteins. *Genes Dev*, **12**, 1998-2012.

18. Cavaloc, Y., Bourgeois, C.F., Kister, L. and Stevenin, J. (1999) The splicing factors 9G8 and SRp20 transactivate splicing through different and specific enhancers. *Rna*, **5**, 468-483.
19. Itoh, H., Washio, T. and Tomita, M. (2004) Computational comparative analyses of alternative splicing regulation using full-length cDNA of various eukaryotes. *Rna*, **10**, 1005-1018.
20. Paronetto, M.P., Achsel, T., Massiello, A., Chalfant, C.E. and Sette, C. (2007) The RNA-binding protein Sam68 modulates the alternative splicing of Bcl-x. *J Cell Biol*, **176**, 929-939.
21. Tacke, R. and Manley, J.L. (1995) The human splicing factors ASF/SF2 and SC35 possess distinct, functionally significant RNA binding specificities. *Embo J*, **14**, 3540-3551.
22. Tacke, R., Tohyama, M., Ogawa, S. and Manley, J.L. (1998) Human Tra2 proteins are sequence-specific activators of pre-mRNA splicing. *Cell*, **93**, 139-148.
23. Simard, M.J. and Chabot, B. (2002) SRp30c is a repressor of 3' splice site utilization. *Mol Cell Biol*, **22**, 4001-4010.
24. Nasim, M.T., Chernova, T.K., Chowdhury, H.M., Yue, B.G. and Eperon, I.C. (2003) HnRNP G and Tra2beta: opposite effects on splicing matched by antagonism in RNA binding. *Hum Mol Genet*, **12**, 1337-1348.
25. Marquis, J., Paillard, L., Audic, Y., Cosson, B., Danos, O., Le Bec, C. and Osborne, H.B. (2006) CUG-BP1/CELF1 requires UGU-rich sequences for high-affinity binding. *Biochem J*, **400**, 291-301.
26. Hovhannisyan, R.H. and Carstens, R.P. (2007) Heterogeneous ribonucleoprotein m is a splicing regulatory protein that can enhance or silence splicing of alternatively spliced exons. *J Biol Chem*, **282**, 36265-36274.