

SUPPLEMENTAL DATA

A bacterial antirepressor with SH3 domain topology mimics operator DNA in sequestering the repressor DNA recognition helix

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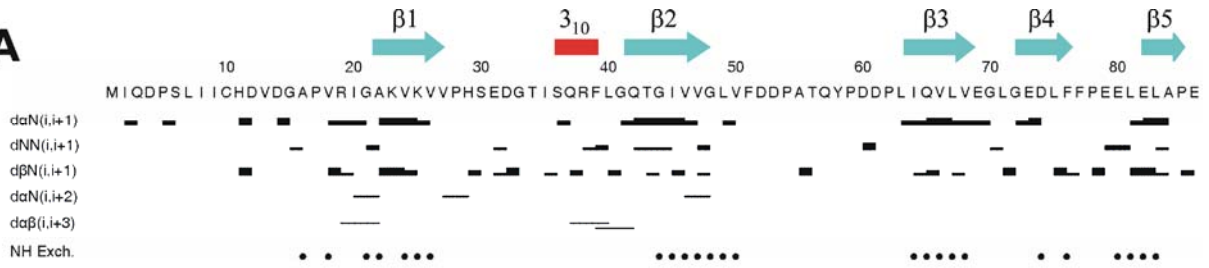
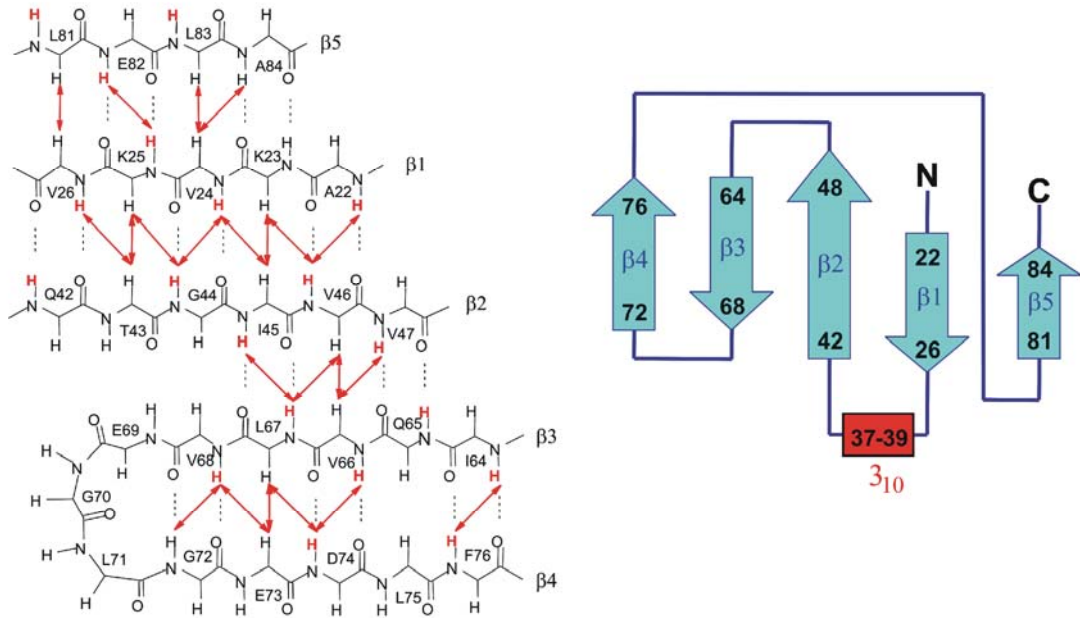
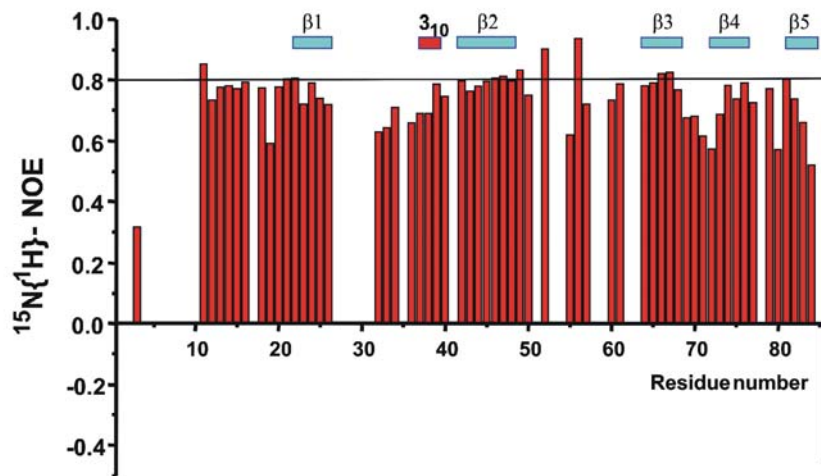
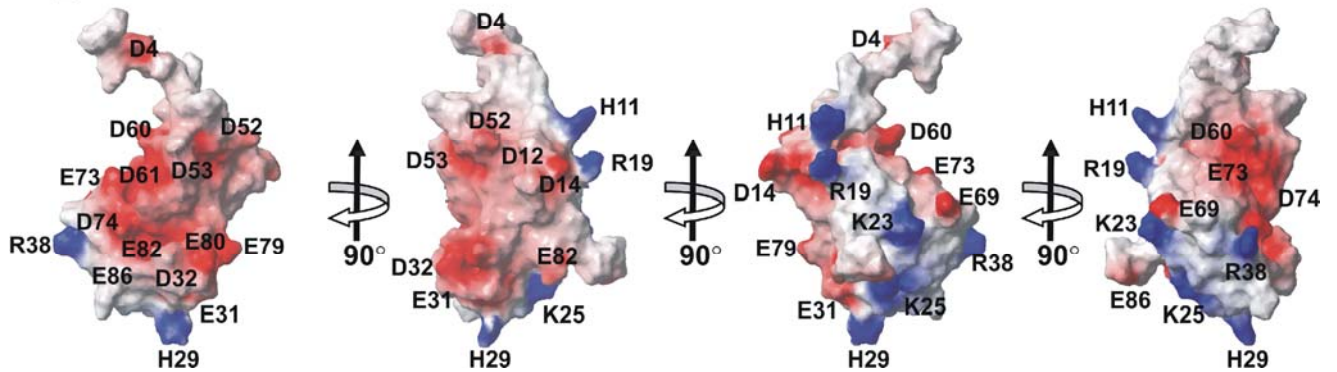
Supplementary Table S1. Strains and plasmids used in this work

Strain or plasmid	Relevant phenotype	Genotype or description	Source or reference
<i>M. xanthus</i> strains			
DK1050	Car ⁺	Wild-type	(1)
MR1776*	Car ⁻	$\Delta carS$	pMR3184 x DK1050
MR1935*	Car ⁺ , Km ^R	$carS'$ (encodes CarS with non-native R-I-L following M1)/ $\Delta carS$	pMR3233 x MR1776
MR1936*	Car ⁻ , Km ^R	$carS'(D52A/D53A) / \Delta carS$	pMR3234 x MR1776
MR1937*	Car ⁻ , Km ^R	$carS'(E79A/E80A) / \Delta carS$	pMR3235 x MR1776
MR1938*	Car ⁻ , Km ^R	$carS'(L63A) / \Delta carS$	pMR3267 x MR1776
MR1939*	Car ⁻ , Km ^R	$carS'(L75A) / \Delta carS$	pMR3264 x MR1776
MR1940*	Car ⁻ , Km ^R	$carS'(F77A) / \Delta carS$	pMR3268 x MR1776
Plasmids			
pET15b	Amp ^R	Vector for overexpression of H ₆ -tagged protein.	Novagen
pBJ114	Km ^R Gal ^S	Vector to generate gene deletions/mutations in <i>M. xanthus</i> .	(2)
pEG202	Amp ^R HIS3	Vector for expressing N-terminal fusions to the LexA DNA-binding domain (DBD) under the control of ADH1 promoter for yeast two-hybrid analysis.	(3)
pJG4-5	Amp ^R TRP1	Vector for expressing N-terminal fusions to the B42 transcriptional activation domain (TAD) under control of the galactose-inducible GAL1 promoter for yeast two-hybrid analysis.	(3)
pSH18-34	Amp ^R URA3 lacZ	<i>lacZ</i> under the control of a GAL10-8 LexA-GAL promoter region. Reporter for yeast two-hybrid analysis.	(3)
pMAR975	Km ^R Gal ^S	pBJ114 derivative lacking an <i>EcoRI</i> site	(4)
pMR2606	Amp ^R TRP1	pJG4-5 derivative for expressing CarS fused to B42-TAD	(5)
pMR2609	Amp ^R HIS3	pEG202 derivative for expressing CarA fused to LexA DBD	(5)
pMR3125	Amp ^R	pET15b derivative for overexpression of H ₆ -CarS(D52A/D53A)	This work
pMR3126	Amp ^R	pET15b derivative for overexpression of H ₆ -CarH(Nter)(R27A)	This work
pMR3138	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(E31A/D32A) fused to B42-TAD	This work
pMR3139	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(D52A/D53A) fused to B42-TAD	This work
pMR3140	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(D60A/D61A) fused to B42-TAD	This work
pMR3141	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(E73A/D74A) fused to B42-TAD	This work
pMR3142	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(E79A/E80A) fused to B42-TAD	This work
pMR3155	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(D12A/D14A) fused to B42-TAD	This work
pMR3156	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(E69A) fused to B42-TAD	This work
pMR3179	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(D4A) fused to B42-TAD	This work
pMR3180	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(E82A) fused to B42-TAD	This work
pMR3181	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(E86A) fused to B42-TAD	This work
pMR3184	Km ^R Gal ^S	pMAR975 with the $\Delta carS$ allele used to generate $carS$ -deleted <i>M. xanthus</i>	This work
pMR3233	Km ^R Gal ^S	pMR3184 with $carS'$ introduced into the <i>EcoRI</i> site.	This work
pMR3234	Km ^R Gal ^S	pMR3184 with $carS'(D52A/D53A)$ introduced into the <i>EcoRI</i> site.	This work
pMR3235	Km ^R Gal ^S	pMR3184 derivative with $carS'(E79A/E80A)$ introduced into <i>EcoRI</i> site.	This work
pMR3249	Amp ^R	pET15b derivative for overexpression of H ₆ -CarS(F77A)	This work
pMR3261	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(F77A) fused to B42-TAD	This work
pMR3262	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(L75A) fused to B42-TAD	This work
pMR3263	Amp ^R TRP1	pJG4-5 derivative for expressing CarS(L63A) fused to B42-TAD	This work
pMR3264	Km ^R Gal ^S	pMR3184 derivative with $carS'(L75A)$ introduced into the <i>EcoRI</i> site.	This work
pMR3267	Km ^R Gal ^S	pMR3184 derivative with $carS'(L63A)$ introduced into the <i>EcoRI</i> site.	This work
pMR3268	Km ^R Gal ^S	pMR3184 derivative with $carS'(F77A)$ introduced into the <i>EcoRI</i> site.	This work

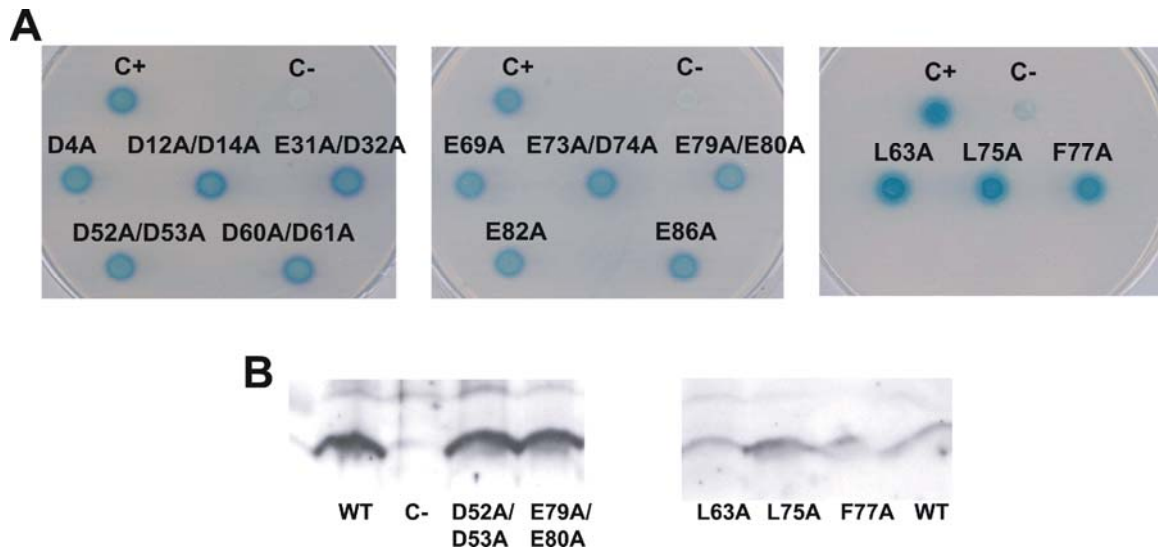
* Generated in this study as described in SI Materials and methods. Car⁺: wild-type phenotype for light-induced carotenogenesis; Car⁻: null phenotype for light-induced carotenogenesis.

Supplementary Table S2. NMR data and structural statistics for the final structures of CarS1

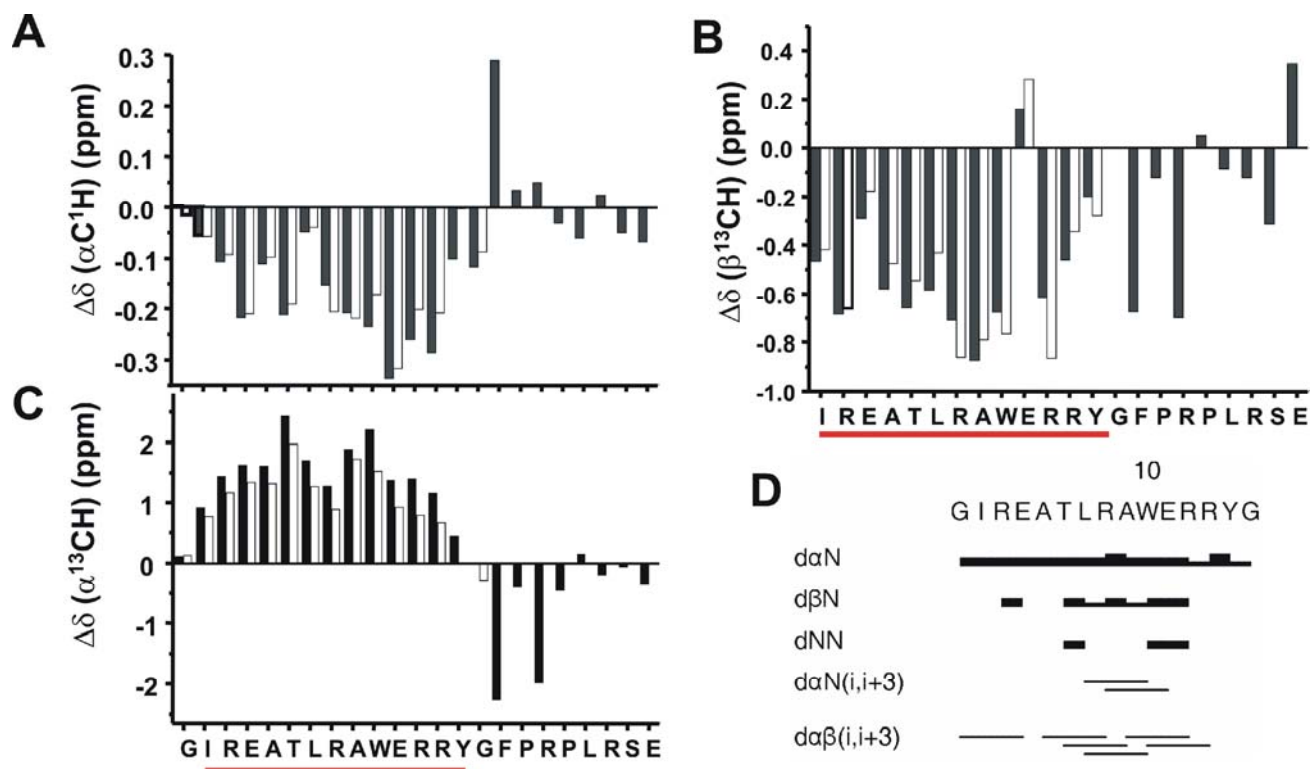
Total number of NOE distance constraints	968
Intraresidue ($i-j=0$)	287
Sequential ($ i-j =1$)	246
Medium-range ($2 \leq i-j \leq 1$)	105
Long-range ($ i-j \geq 5$)	330
Dihedral angle constraints (TALOS)	
Total (ϕ, ψ)	141
Pairwise root mean square deviations (Å)	
Backbone atoms (residues 1-86)	3.4 ± 1.0
Backbone atoms (residues 10-86)	0.8 ± 0.1
Heavy atoms (residues 1-86)	3.8 ± 1.0
Heavy atoms (residues 10-86)	1.4 ± 0.1
Maximum constraint violations	
Distance (Å)	0.15 ± 0.05
Dihedral angles (°)	2.3 ± 0.7
Structural quality (Ramachandran plot analysis)	
Residues in the most favourable regions (%)	82.3
Residues in additional allowed regions (%)	15.8
Residues in generously allowed regions (%)	0.4
Residues in disallowed regions (%)	1.5

A**B****C****D**

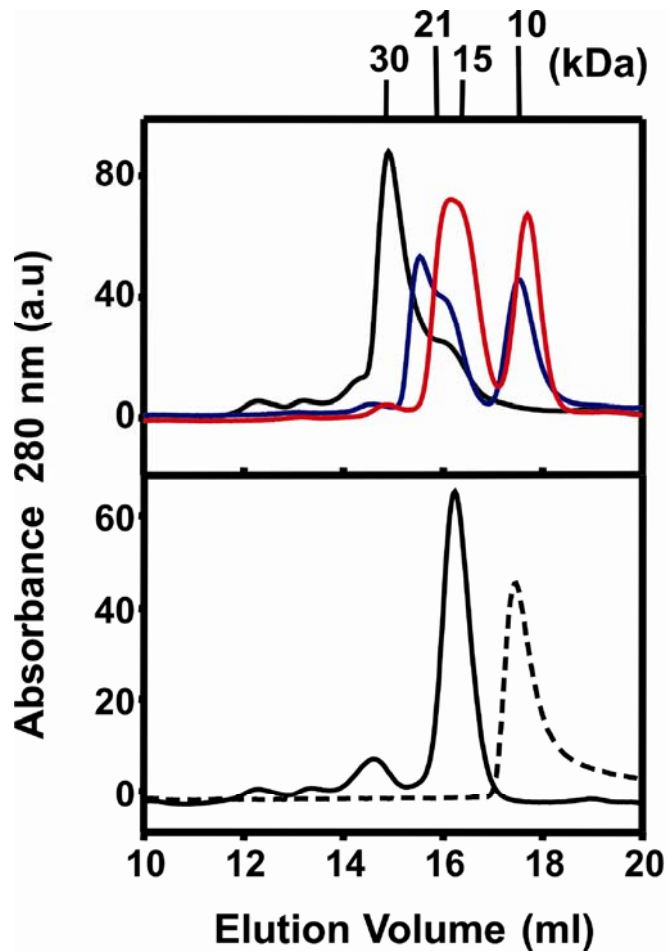
Supplementary Figure S1. (A) Summary of NMR data showing sequential and medium range NOEs involving backbone NH and C_αH, and side chain C_βH protons relative to residue i. The thickness of the bars scales with weak, medium, and strong NOE intensities. Elements of secondary structure deduced from NMR data are indicated with β-strands as blue arrows and the 3₁₀-helix as a red bar. Black dots indicate residues with the slowest amide ¹H exchange (NH/ND) with solvent. (B) Schematics showing the anti-parallel β-sheet topology in CarS1. On the left, non-sequential NOEs are indicated by the red two-headed arrows. Amide ¹H in red are those that exchange slowest with solvent. The arrangement of the anti-parallel β-strands and the 3₁₀-helix is shown on the right with delimiting residues numbered. (C) Steady-state heteronuclear ¹⁵N{¹H}-NOEs for the backbone amides plotted against residue number. Secondary structure elements from the NMR structure are indicated on top as in (A). (D) Four views at 90° clockwise rotations of the electrostatic surface representation of CarS1 showing regions of positive and negative charge, respectively, in blue and red, the color intensity scaling with electrostatic potential, and the charged residues labeled.



Supplementary Figure S2. Analysis of CarS mutants *in vivo*. **(A)** Yeast two-hybrid analysis of the interactions of each CarS mutant with CarA on galactose plates. C+: LexA-CarA and B42-CarS; C-: LexA-CarA only; the rest express LexA-CarA and the indicated B42-CarS*. **(B)** Western blot analysis using mouse monoclonal anti-CarS antibodies of whole cell extracts of the indicated strains obtained after cell growth in the light. C- is the $\Delta carS$ strain; the others are derived from introducing the indicated allele into the $\Delta carS$ strain.



Supplementary Figure S3. NMR analysis of peptides corresponding to $\alpha 2$, the CarA DNA recognition helix. Deviation ($\Delta\delta$) of the observed chemical shift (δ_{obs}) for peptides P01 (black bars) and P02 (open bars) from the corresponding random coil value (δ_{rc}) shown for the $\alpha\text{C}^1\text{H}$ (A), $\alpha^{13}\text{CH}$ (B), and $\beta^{13}\text{CH}$ (C). The helical segment, as inferred from the NMR data, is underlined in red and coincides with $\alpha 2$ in the high resolution NMR structure determined for CarANt. (D) Summary of medium and short range NOEs for the shorter P02 peptide. NMR spectra were recorded at 5 °C and at pH 6.5.



Supplementary Figure S4. Effect of ionic strength on stable CarS-CarANt complex formation. The bottom panel shows the elution profiles for pure CarS (solid curve) or pure CarNt (dashed curve) off a Superdex-200 analytical gel filtration column in buffer containing 150 mM NaCl as described in the main text. The top panel shows the elution profiles for mixtures of CarS and CarNt in buffer with NaCl concentration at 150 mM (black curve), 300 mM (blue curve), and 1 M (red curve). M_r (in kDa) for each peak maximum is indicated. Note that increasing the ionic strength shifts the 30 kDa peak corresponding to the complex towards lower M_r , with a concomitant rise in the intensities of peaks corresponding to free proteins, indicating that stable complex formation is increasingly hindered by higher ionic strength.

Supplementary Figure S5. Sequence alignments of CarS, CarA, and CarH proteins in *M. xanthus* and *S. aurantiaca*. **(A)** Alignment of CarS proteins. Arrows in magenta point to residues important for CarS interaction with CarA in *M. xanthus* identified in this study. **(B)** Alignment of CarA proteins. **(C)** Alignment of CarH proteins. In **(B)** and **(C)** the helix $\alpha 2$ segment is boxed in red. Identical residues are shaded black and have an asterisk in the “consensus” line below. Similar residues are shaded gray. The *S. aurantiaca* protein sequences are based on our annotation of the genome sequence. We encountered errors in the corresponding genome annotation deposited at NCBI and have commented on it elsewhere (see 6).

SUPPLEMENTARY REFERENCES

1. Ruiz-Vázquez,R.M.and Murillo,F.J. (1984) Abnormal motility and fruiting behaviour of *Myxococcus xanthus* bacteriophage resistant strains induced by a clear plaque mutant of bacteriophage Mx8. *J. Bacteriol.*, **160**, 818-821.
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4. Cayuela,M.L., Elías-Arnanz,M., Peñalver-Mellado,M., Padmanabhan,S. and Murillo,F.J. (2003) The *Stigmatella auriantica* homolog of *Myxococcus xanthus* HMGA-type transcription factor CarD: insights into the functional modules of CarD and their distribution in bacteria. *J. Bacteriol.*, **185**, 3527-3537.
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6. Pérez-Marín,M.C., Padmanabhan,S., Polanco,M.C., Murillo,F.J. and Elías-Arnanz,M. (2008) Vitamin B₁₂ partners the CarH repressor to downregulate a photoinducible promoter in *Myxococcus xanthus*. *Mol. Microbiol.*, **67**, 804-819.