

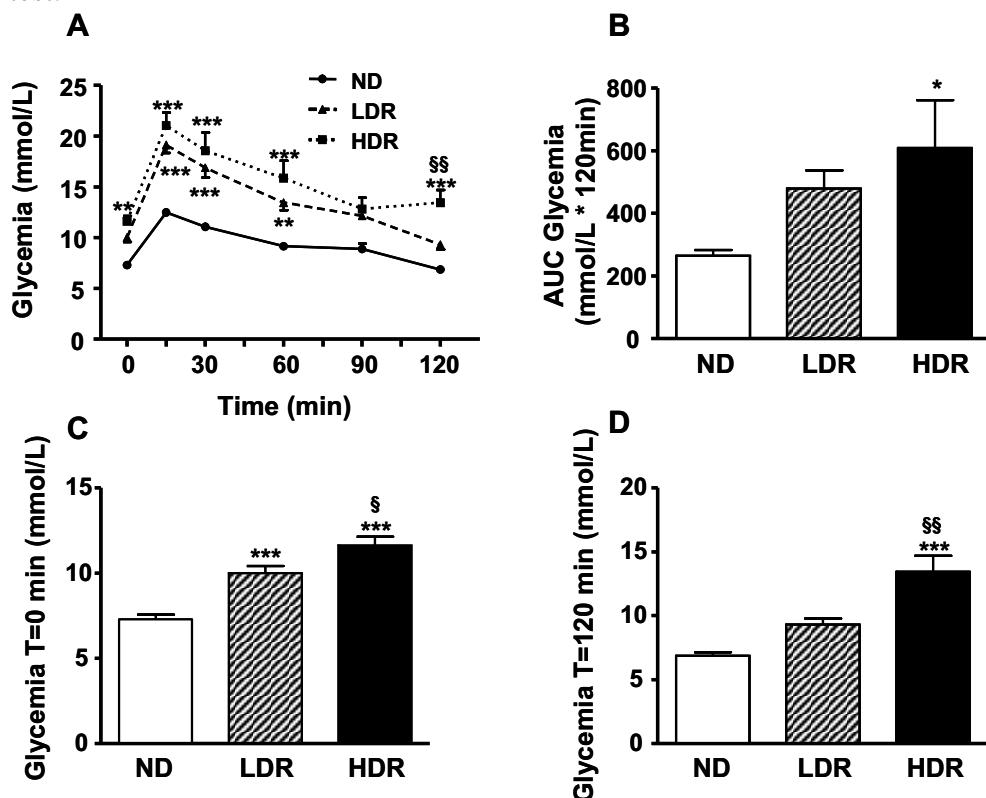
## Supplemental research design and methods

**Quantitative RT-PCR.** Total RNA was extracted from islets as previously described (1). Gene expression was determined by a standard curve method and normalized to the expression of  $\beta$ -actin. Real-time PCR analysis was performed on Rotor-Gene R3000 (Corbett Research, Mortlake, NSW, Australia) using Quantitec Sybrgreen (Qiagen, Mississauga, ON, Canada). Primers, listed in supplementary Table 1, were designed using Primer3 software. Results are expressed as the ratio of target mRNA to  $\beta$ -actin mRNA.

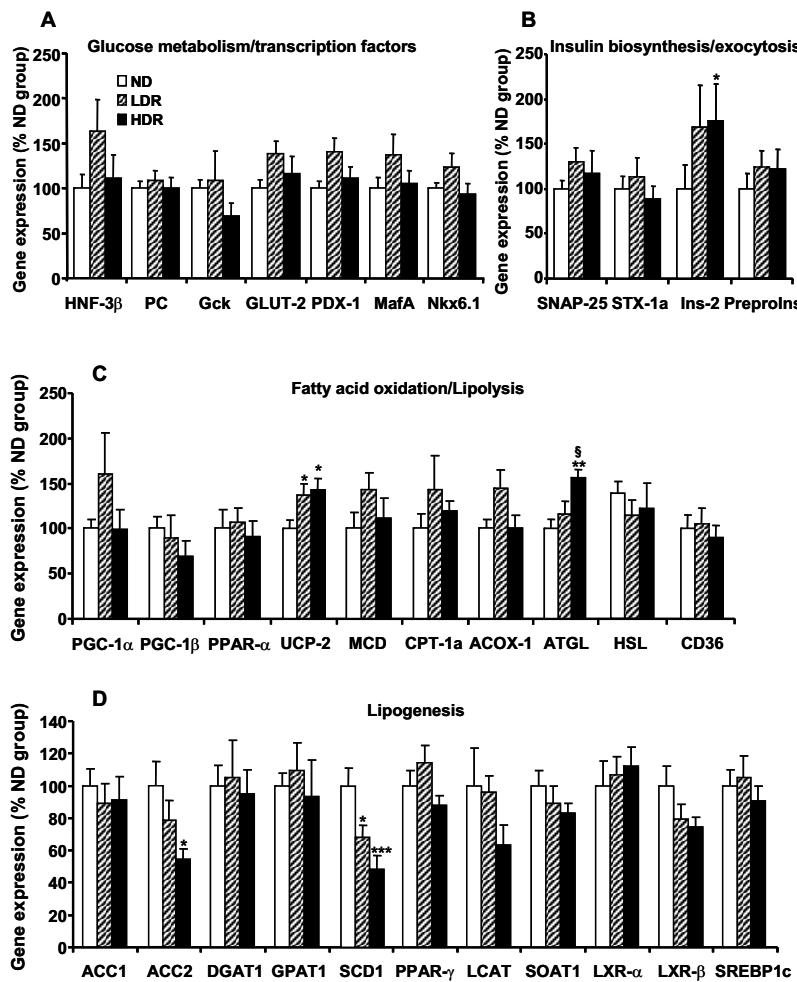
### Supplemental reference

1. Delghingaro-Augusto V, Nolan CJ, Gupta D, Jetton TL, Latour MG, Peshavaria M, Madiraju SR, Joly E, Peyot ML, Prentki M, Leahy J: Islet beta cell failure in the 60% pancreatectomised obese hyperlipidaemic Zucker fatty rat: severe dysfunction with altered glycerolipid metabolism without steatosis or a falling beta cell mass. *Diabetologia* 52:1122-1132, 2009

**Suppl. Fig.1.** Oral glucose tolerance in ND, LDR and HDR mice. (A-D) Oral glucose tolerance test (OGTT). Blood glucose (A) was measured at times 0, 15, 30, 60, 90 and 120 min in response to an oral glucose administration (1g/kg) in 6h-fasted mice. (B) Area under the curves (AUC) of the 0-120 min glycemic responses. (C-D) Blood glucose levels at time 0 (C) and a time 120 min (D) of the OGTT. Means  $\pm$  SE of 13-14 animals per group. \* $P<0.05$ , \*\* $P<0.01$ , \*\*\* $P<0.001$  ND versus LDR or HDR;  $\ddagger P<0.05$ ,  $\ddagger\ddagger P<0.01$  LDR versus HDR; one-way (B-D) or two-way (A), with Bonferroni post hoc test.



**Suppl. Fig.2.** mRNA expression of selected genes related to glucose and lipid metabolism. Gene expression was measured in islets from ND, LDR and HDR mice using real-time RT-PCR. Genes were grouped by function. (A) Glucose metabolism and transcription factors: forkhead box A2 (Foxa2/HNF-3 $\beta$ ), pyruvate carboxylase (PC), glucokinase (Gck), glucose transporter 2 (GLUT-2), pancreas-duodenum homeobox-1 (PDX-1), v-maf musculoaponeurotic fibrosarcoma oncogene homolog A (Maf-A) and NK6 homeobox-1 (Nkx6.1). (B) Insulin biosynthesis and exocytosis: preproinsulin-2 (preproins), insulin-2 (Ins-2), synaptosomal-associated protein 25 (SNAP-25) and syntaxin-1a (STX-1a). (C) Fatty acid oxidation and lipolysis: peroxisome proliferator-activated receptor- $\alpha$  (PPAR- $\alpha$ ), PPAR- $\gamma$ -coactivator-1 $\alpha$  and -1 $\beta$  (PGC-1 $\alpha$  and -1 $\beta$ ), uncoupling protein-2 (UCP-2), malonyl-CoA-decarboxylase (MCD), carnitine palmitoyltransferase-1a (CPT-1a) and acyl-CoA oxydase-1 (ACOX-1), adipose triglyceride lipase (ATGL) and hormone-sensitive lipase (HSL). (D) Lipogenesis: acetyl-CoA carboxylase-1 and -2 (ACC-1 and ACC-2), diacylglycerol-O-acyltransferase-1 (DGAT-1), glycerol-3-phosphate acyltransferase-1 (GPAT-1), stearoyl-CoA deasturase-1 (SCD-1), fatty acid transporter (FAT/CD36), lecithin-cholesterol acyltransferase (LCAT), sterol-O-acyltransferase (SOAT-1), liver-X-receptor- $\alpha$  and - $\beta$  (LXR- $\alpha$  and - $\beta$ ), sterol regulatory element binding protein-1c (SREBP-1c) and PPAR- $\gamma$ . Results are expressed as percent of the ND group values and were normalized to actin. Means  $\pm$  SE of 9–18 animals per group. LDR or HDR versus ND: \* $P$ <0.05, \*\* $P$ <0.01, \*\*\* $P$ <0.001; LDR versus HDR: § $P$ <0.05; one-way ANOVA- Bonferroni's multiple comparison test.



**Supplemental Table 1.** PCR primer sequences used for Quantitative Real Time PCR.  
Primers: S, sense; AS, antisense.

GENE	GenBank accession no	Primer sequences (5'-3')
<i>ACC-1</i>	NM_133360	S: ATGTCCGCACTGACTGTAACC AS: TCCATAGCCGACTTCCATAGC
<i>ACC-2</i>	NM_133904	S: GAAGCCTCCTCGAGTACCTG AS: GAGGAAGATGTGGTTGCAGTC
<i>ACOX-1</i>	NM_015729	S: GCGTGGAACTTGACTTCTGTC AS: ACGGCTCTGTCTTGAATCTTG
<i>ATGL</i>	NM_025802	S: TCCAAGGATGAGCTCATCCA AS: CCTCCTAAGGAATCGAAGTC
<i>β-ACTIN</i>	NM_007393	S: CATGGATGACGATATCGCTGC AS: GTACGACCAAGAGGCATAACAGG
<i>CD36</i>	NM_007643	S: AGGTCTATCTACGCTGTGTTCG AS: CAATGGTTGTCTGGATTCTGG
<i>CPT1a</i>	NM_013495	S: GGTCAAGCTGTTCAAGATAGC AS: ACCACATAGAGGCAGAAGAGG
<i>DGAT-1</i>	NM_010046	S: GAGCTATCCAGACAACCTGACC AS: AGCATCTCAAGAACCTCGT
<i>GCK</i>	NM_010292	S: ATCTTCTGTTCCACGGAGAGG AS: GATGTTAAGGATCTGCCTTCG
<i>GLUT-2</i>	NM_031197	S: CTTGGTTCATGGTTGCTGAAT AS: GCAATGTACTGGAAGCAGAGG
<i>GPAT-1</i>	NM_008149	S: CGGAACTGAACTGGAGAAGTG AS: GATGAATTGCTGGTGCCTCCTT
<i>HNF-3β</i>	NM_010446	S: CTACGCCAACATGAACACTCGAT AS: GGTGATGAGCGAGATGTACGA
<i>HSL</i>	NM_001039507	S: GGCTCACAGTTACCATCTCACC AS: GAGTACCTTGCTGTCCCTGTCC
<i>INS-2</i>	NM_008387	S: TGGAGGCTCTTACCTGGTG AS: TCTACAATGCCACGCTTCTG
<i>LCAT</i>	NM_008490	S: GGCTGCACTCTATGAAGATGG AS: TGAGGTGATCTGTCTCGTTCA
<i>LXR-α</i>	NM_013839	S: GTTCTCCAGAGCCATGAATGA AS: GTTGCAGCCTCTACTTGGAA
<i>LXR-β</i>	NM_009473	S: TCACCTACAGCAAGGACGACT AS: ATGGCGATAAGCAAGGCATAC
<i>Maf-A</i>	NM_194350	S: GTGCTGGAGGATCTGTACTGG AS: ATGGTGGTGATGGTGATGG
<i>MCD</i>	NM_019966	S: AGAAGATCAGCGAGTGTGAGG AS: AGTCAGAGCCACATGCAGAAC
<i>Nkx6.1</i>	NM_144955	S: CACGTTCTGGACAGCAAAT AS: TTGACCTGACTCTCCGTAC
<i>PC</i>	NM_008797	S: TGACTCTCTGCTCGTCAAGGT AS: AGGAACCTGCTGGTTGTGAGA
<i>PDX-1</i>	NM_008814	S: GGTATAGCCGGAGAGATGC AS: CTGGTCCGTATTGGAACG
<i>PGC-1α</i>	NM_008904	S: TAGAGTGTGCTGCTGGTTG

		AS: GATTGGTCGCTACACCACTTC
<i>PGC-1β</i>	NM_133249	S: GCTCTGGAAGGTGAAGACCTG AS: TCAAGCAGGAAGCTACTCTCG
<i>PPAR-α</i>	NM_011144	S: AGAGAATCCACGAAGCCTACC AS: GGCCATACACAAGGTCTCCAT
<i>PPAR-γ</i>	NM_011146	S: ATCAGCTCTGTGGACCTCTCC AS: GGTCAAGCTCTTGTGAATGGAA
<i>PREPROINS-2</i>	NT_039437	S: CAGTGCCAAGGTCTGAAGGT AS: TGTGTCCATCCATGACCAGT
<i>SCD-1</i>	NM_009127	S: GAAGCTGGTGATGTTCCAGAG AS: CCAGAGTGTATCGCAAGAAGG
<i>SNAP-25</i>	NM_01142	S: ATGGAGAAGGCTGATTCCAAC AS: CCATGAGAGAACATGAAGGA
<i>SOAT-1</i>	NM_009230	S: CCACGGTTACAGCAAGAGTTC AS: TGGTGGCAGTGTGTATGCTAA
<i>SREBP-1c</i>	NM_011480	S: ATGCTCCAGCTCATCAACAAAC AS: GAGGCCAGAGAACATGCAGAAGAG
<i>STX-1a</i>	NM_016801	S: GTGAGGAATTGGAAGACATGC AS: GCCTTGCTCTGGTACTTGACG
<i>UCP-2</i>	NM_011671	S: AGAGCACTGTCGAAGCCTACA AS: AGGCAGAAAGTGAAGTGGCAAG