

## LEGANDS FOR SUPPLEMENTARY FIGURES

**FIG. S1. Alignment of protein sequences of GOGAT  $\beta$  subunit-like proteins and  $\beta$ -subunits from *E. histolytica* and other organisms.** The sequences were aligned using the CLUSTAL W program. Accession numbers of these sequences are as follows: *Escherichia coli* (P09832), *Azospirillum brasilense* (AAG38999), *Clostridium saccharobutylicum* (AAP06761), *E. histolytica* EhNO1 and 2 (XP\_656997 and XP\_653573). The cysteine residues that are proposed to participate in the formation of [4Fe-4S] clusters are highlighted in bold. The residues that match the consensus sequence implicated in the formation of an adenylate binding fold in the FAD- or NAD(P)H-binding region are marked with a plus ("+" ). A sharp ("#" ) indicates tyrosine residues that are proposed to be replaced by acidic residues in NADH-dependent enzymes, and a triangle ("▲" ) indicates the basic residue that is proposed to favour NADPH binding. Residues that match the second FAD consensus sequence are marked with an asterisk (\*). The GXGXX(G/A/P) motif described in the text is shown in bold.

**FIG. S2. Phylogenetic reconstructions of GOGAT  $\beta$  subunit and GOGAT  $\beta$  subunit-like proteins from a variety of organisms.** Phylogenetic trees of 32  $\beta$ -subunit and  $\beta$ -subunit-like proteins were constructed using the CLUSTAL W program and drawn with the MEGA4.1 program, and a representative neighbor-joining tree is shown. The number at the nodes represents the bootstrap value as a percentage of 1000 replicates. The scale bar indicates 0.1 substitutions at each amino acid position. Species names and accession numbers of the sequences are also indicated. The  $\beta$  subunit domains of the *M. sativa*, *A. thaliana*, *S. cerevisiae*, *P. falciparum*, *D. melanogaster*, and *C. elegans* polypeptides were included from residue positions 1676, 1701, 1635, 2513, 1625, and 1706, respectively.

**FIG. S3. Expression and purification of recombinant EhNOs.** Protein samples were electrophoresed on a 12% SDS-PAGE gel under reducing conditions, and then stained with Coomassie Brilliant Blue R250. Lane 1, molecular mass markers; lanes 2 and 3, total lysate of uninduced and isopropyl  $\beta$ -d-thiogalactopyranoside-induced *E. coli* cultures, respectively; lane 4, purified rEhNO1; lane 5, purified rEhNO2 (10  $\mu$ g).

Suppl. Figure S1

EhNO1 MKSFNITHVLKRNPEERINDWKEVVVGYSSKEETIEEASRCLGCKKPGCVPTC 52  
 EhNO2 MAANYNRTVPVPMRNPEERINDWKEVVVGYTKEDTKTEASRCLGCKKPGCVPTC 53  
 C. saccharobutylicum MTCNDIQLLDEADRCLLCKKPRCKENC 27  
 E. coli MSQNVYQFIDLQRVDPKPKPLKIRKIEFVEIYEPFSEGQAKAQADRCLS CGNPYCEWKC 59  
 A. brasilense MANQRMLGFVHTAQRMPPDKRPAERERRQDFAEIIYARFSDERANEQANRCSQC GVPFCQVHC 60

EhNO1 PAHINIPEYIIHLKKGEFNEATNVILKDMPLLIHICGRVCP--HYCEKTCVKAK-RGGELQ 109  
 EhNO2 PAHIDIIPGFISELKKEQYTEAVSVILKRLPFPNIICGRVCP--HYCQGKCVKAR-RGGELE 110  
 C. saccharobutylicum PIQTSIPEIISLYKENRLREAGEILFNNNPLSAVCSLVCIHEDQCKGNCKIKGI-KGEP IR 86  
 E. coli PVHNYIPNWLKLANEGRIFEAAELSHQTNTLPEVCGRVCPQDRLC EGSCTLND-EFGAVT 118  
 A. brasilense PVSNNIPDWLKL TSEGRLEEAYEVSQATNNFPEICGRICPQDRLC EGN CVIEQSTHGAVT 120

FAD-I

EhNO1 IMELKRSAITYANEEDIKIDIAP--DTSKKVAVVVGSGPAGLAAAYFLRLKKGHKVVVYEQK 167  
 EhNO2 IMELKRSAITYSTEEDIKIDIAP--DTGKKVAVVVGSGPAGLAAAYFLRLKKGHKVVVYEQK 168  
 C. saccharobutylicum FHDIEYEISM KYLEEVRFENLPK---DKDRIAIIGGGPAGITIAFILANKGYDVTIFDAH 143  
 E. coli IGNIERYINDKAFEMGWRPDMGSKVQTGKKVAIIIGAGPAGLACADVLTRNGVKAVVFD RH 178  
 A. brasilense IGSVEKYINDTAWDQGWVKPRTPSRELGLSVGVIGAGPAGLAAAEELRAKGYEVHVYDRY 180  
 ++ + + + + + + + + +

EhNO1 HKLGGMILCIPPYRLPRDKLDEDIDRIKRLGIEFRTNAKVD---NIPSLNE-YDAVFV 223  
 EhNO2 HKLGGMILCIPPYRLPRDKLDEDIDRIKRLGIEFRTNAKVD---DITTLKKE-YDAVFV 224  
 C. saccharobutylicum VKIGGVLRYGIPEYRLPKKILDTIEDRLVELGIKIRPNTLIGPVI TLDRLYEDGYKAIFI 203  
 E. coli PEIGLLTFGIPAFKLEKEVMTRREIFTGMGIEFKLNTEVGRDVLDDLLSD-YDAVFL 237  
 A. brasilense DRMGLLVYGI PGFKLEKSVVERRVKLLADAGVIYHPNFEVGRDASLP ELRRK-HVAVLV 239

EhNO1 GIGTLKRKVLGIPGEDLIGVEHVIPYLESINTFARK-----TIGKKVAVVVGAG 271  
 EhNO2 GIGTLKRKVLGIPGEDLIGVEHVIPYLESINTFARK-----TIGKKVAVVVGAG 272  
 C. saccharobutylicum GTGVWNPKTLNIKGETRGNHFAIDYLSPEAYRLG-----EKVAVI GAG 248  
 E. coli GVGTYQSMRGGLENEDADGVYAALPFLIANTKQLMG-FGETRDEPFVSM EGKRVVVLGGG 296  
 A. brasilense ATGVYKARDIKAPGSGLGNIVAALDYLTTSNKVS LGDTVEAYENGLSNAAGKHVVVVLGGG 299  
 + + + + +

NAD (P) H

EhNO1 **FSAMDAVRVARRLGS-EAFI IYR**SEHEMPASSEIEEAKEEGVTMMTLCNPTKIIGDEN 330  
 EhNO2 **FSAMDAVRVARRLGS-EAFI VYR**SENEMPAAPSEVEEAKEEGVTMMTLCNPTKIIGDEN 331  
 C. saccharobutylicum **NVAMDAARSAKRNGAKEVTVLYR**KGFDEMSATKQEIMEAKEDGVI FNLFKSPIEITEKGI 308  
 E. coli **DTAMDCVRTSVRQGAHVTCAYR**RDEENMPGSRREVKNAREEGVEFKFNVQPLGIEVNGN 356  
 A. brasilense **DTAMDCVRTAIRQGATSVKCLYR**RDRKNMPG SQREVAHAEEEGVEFIWQAAP EG--FTGD 357  
 + + + + #▲

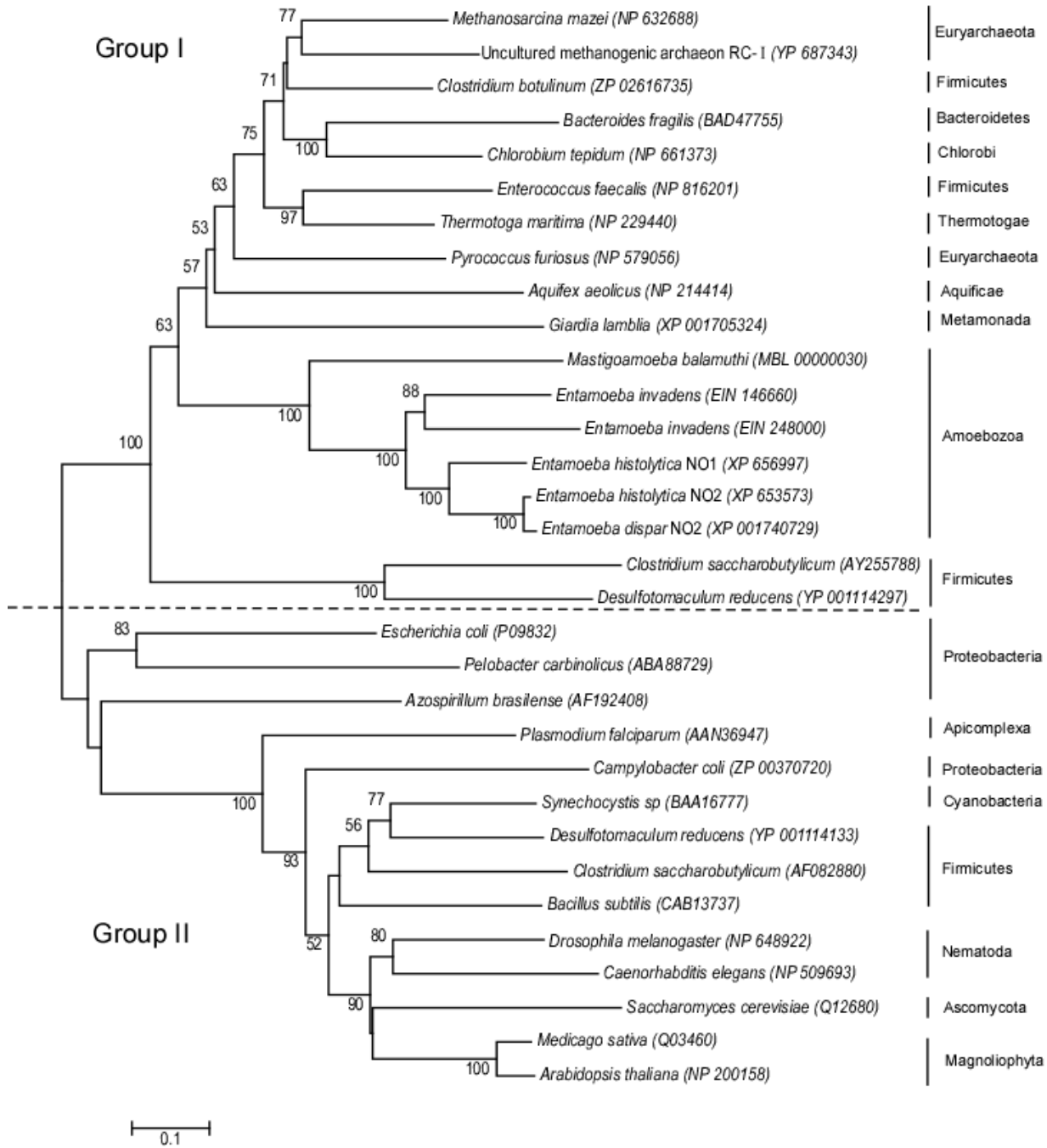
EhNO1 GKVKGIECIKMKLGE SDESGRAAPVIPGSEFVIDCDMVIQAI SQVVD----LGCCPGVE 386  
 EhNO2 GKVKGIECIKMKLGE PDASGRAAPVIPGSEFVIDCDMVIQAI SQGVE----TECCKDVE 387  
 C. saccharobutylicum R-----LSSTENVTDENGKIVTKVIEGKEEFFECDSII IAVSQTPKTN-IVSNTKELN 360  
 E. coli GKVSGVKMVRTEMGEPDAKGRRAEIVAGSEHIVPADAVIMAFGFRPHNMEWLAKHS-VE 415  
 A. brasilense TVVTGVRAVRIHLGVADATGRQTPQVIEGSEFTVQADLVIKALGFEPEDLPN AFDEPELK 417

FAD-II

EhNO1 LTQYGT LKVDENYK---TSINGLYAAGDCITGPKSIVDAVGD AHRASNAIHEYL TNLTI 442  
 EhNO2 LSKWQTLQVNDKFQ---TNVEGIYASGDCVTGPKSIVH AVGDTYVAVEAMHEYL MGKKN 443  
 C. saccharobutylicum TNKWGLIITDEKGN---TTRKGT FASGDVVTGAKTVVEAVVQAKTVANTIEEYCKNN 414  
 E. coli LDSQGRIIAPEGSDNAFQTSNPKIFAGGDIVRGS DLVVTAIAEGRKAADGIMNWLEV 472  
 A. brasilense VTRWGTLLVDHRTK---MTNMDGVFAAGDIVRGASLVVWAI RDGRDAAEGIHAYAKAKAE 474  
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EhNO1 TPMEQD 448  
 EhNO2 EE 445  
 A. brasilense APVAVAE 482

Suppl. Figure S2



Suppl. Figure S3

