

# Supporting Information

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## SI Text

**SI Materials and Methods. 1. Cloning and sequencing of aptamers from the R3 pool.** After 3 rounds of selection, the selected pool was PCR amplified with unlabeled forward and reverse primers at the optimized PCR cycle number determined by the pilot PCR. PCR products were purified with the MinElute PCR Purification Kit (Qiagen) and cloned into *Escherichia coli* using the TOPO TA cloning kit (Invitrogen). 98 colonies were randomly picked and sequenced at the GENEWIZ San Diego Laboratory.

**2. Nitrocellulose filter binding assay.** T4 polynucleotide kinase (NEB, Ipswich, MA) was used to 5' end label 10  $\mu$ M concentrations of ssDNA aptamers with  $\gamma$  [ $^{32}$ P]-ATP. Following a 40 min, 37  $^{\circ}$ C incubation step, the reaction was heat killed by taking the end labeled samples to a temperature of 70  $^{\circ}$ C for 10 min. The end labeled oligonucleotides were purified using Sephadex G-50 microspin columns (GE Healthcare).

Following purification, a 100 pM solution of  $\gamma$  [ $^{32}$ P] 5' end labeled aptamers was incubated with 1 nM of target protein (PDGF-BB) for 1.5 h at room temperature. 50  $\mu$ L of each sample

was subsequently applied to a mixed ester filter (Millipore) that had been pretreated with 1 mL of 0.1 mM  $\text{Na}_2\text{HPO}_4$ , 1.8 mM  $\text{KH}_2\text{PO}_4$ , 137 mM NaCl, 2.7 mM KCl, and 1 mM  $\text{MgCl}_2$  (PBSM) (pH = 7.4) buffer. The filters were immediately washed with 1 mL of PBSM buffer, and then removed from the filter holders and placed in scintillation liquid. Scintillation counts were generated, and the fraction of aptamers bound to the protein,  $q$ , was calculated from the following equation

$$q = [(A + P + F) - (A + F)] / (A + P)$$

where  $A$  stands for aptamer (either the library or the #1 sequence from Quantitative Selection of Aptamers through Sequencing (QSAS) R3/R2 set),  $P$  stands for PDGF-BB protein, and  $F$  stands for filtering. In other words, the fraction of aptamers in bound form  $q$ , is equal to the signal (in cpm) generated by filters treated with a mixture of protein and aptamer minus the signal generated by aptamer nonspecifically bound to the filter in the absence of protein, divided by the total signal generated from an unfiltered mixture of aptamer plus protein.

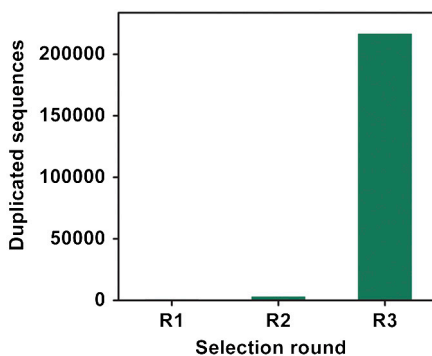


Fig. S1. Representation of the ten most frequently observed sequences from each selection pool. These had an average copy number of 594, 2,789, and 216,488 for R1, R2, and R3 pools, respectively.

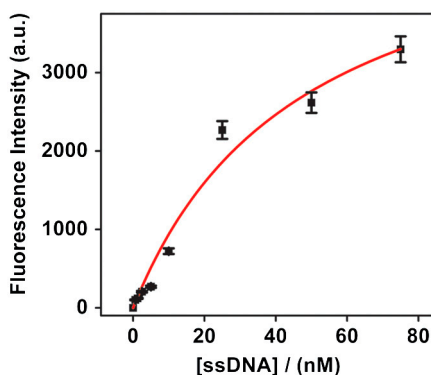
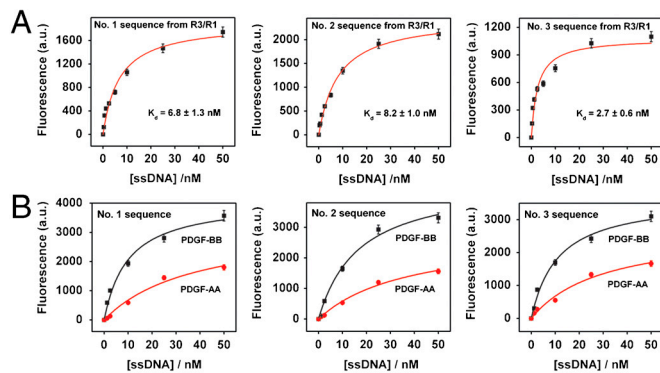
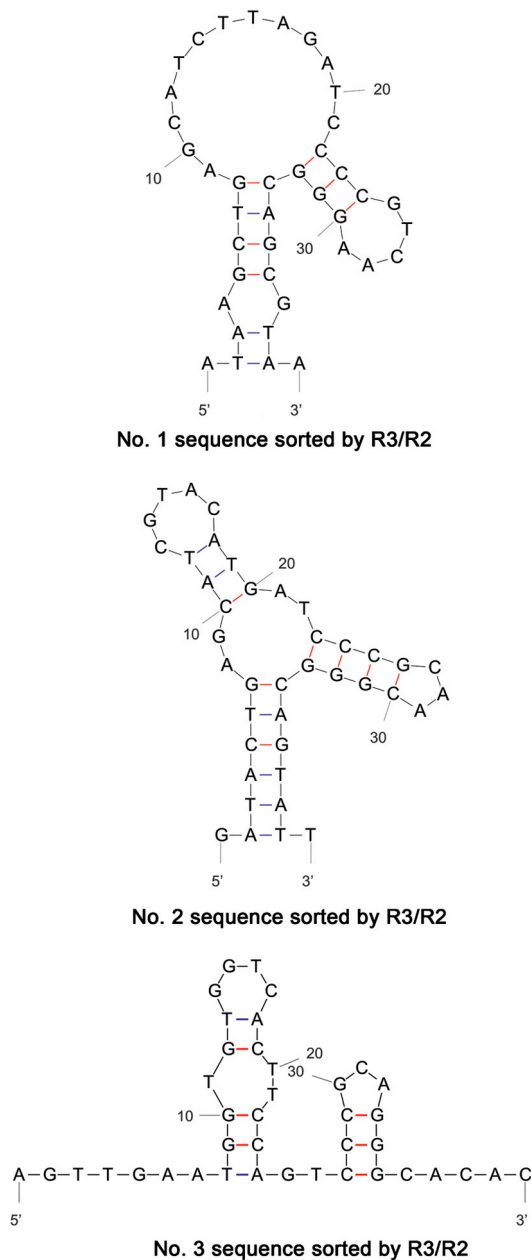


Fig. S2. The most frequently appeared sequence in the three pools was tested for its affinity for the PDGF-BB target via a fluorescence binding assay. Based on these measurements, we calculated a  $K_d$  of  $47.58 \pm 16.1$  nM, which is considerably higher (i.e., lower affinity) than other aptamers sorted by QSAS.



**Fig. S3.** The three most highly enriched sequences from QSAS R3/R1 were tested for their affinity (A) and specificity (B) for the PDGF-BB target via a fluorescence binding assay.



**Fig. S4.** Secondary structures of the three most highly enriched aptamer sequences obtained from QSAS R3/R2.



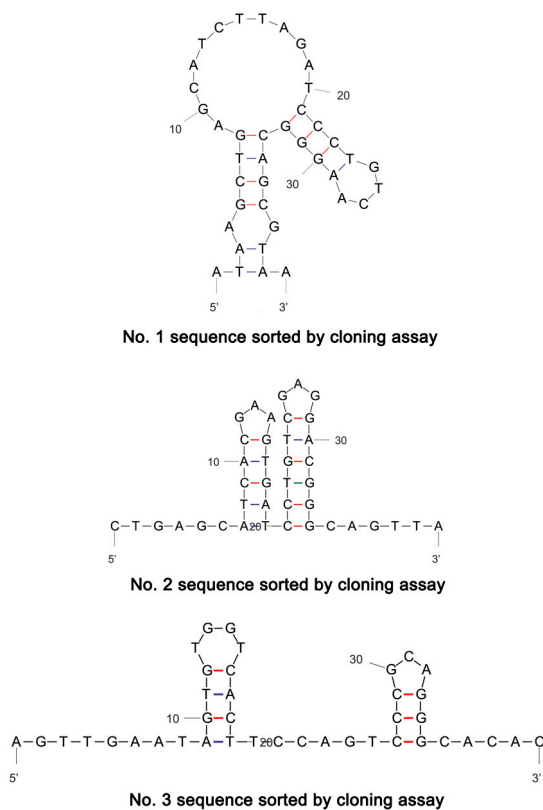


Fig. S6. Secondary structures of the top three enriched aptamer sequences obtained from randomly picked clones from the R3 pool.

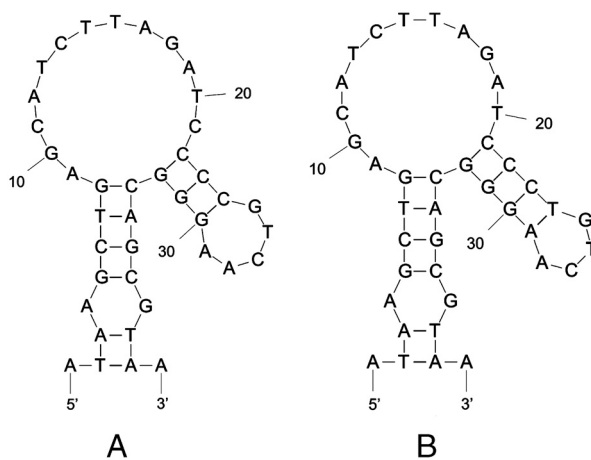


Fig. S7. The secondary structure of highly enriched PDGF-BB aptamers. (A) Sequence #1 identified by the QSAS R3/R2 method. (B) Sequence #3 identified by standard cloning and sequencing of the R3 pool. Although these two aptamers varied by only a single base, they exhibited markedly different binding affinities and specificities: the QSAS-derived sequence exhibited 6.4-fold higher binding affinity and 2.2-fold higher specificity for PDGF-BB in comparison with the cloning-derived sequence.



**Table S3. Aligned aptamer sequences obtained by traditional cloning from the selected R3 pool**

Seq. ID	Sequences (5' → 3')	
21	A—A—GTGGGGCACCAGGT—TTGGTCCCCGGATAGGGGCCACTT	Group I (16.50%)
48	GGGTCAGCGCGGAGTATCGTGTGGATCCCGTAGGGCGCCG	
32	T—CGGTGAGGTATCCATTTGGATCCTCAGTCCTCAGGCACC	
3	GC—G—CCGGTACCCTTAAGG—GTCCCGTCAGGGGGCGTCA	
58	GGCGTCACTTCTAGT—CCCGTCCACGGGGGCCAGGGTTCT	
17	T—CGCGGTCACTGCAATTAGATAGGAGTCCCGGAGGGCGCG	
26	GCTTGCTACTATA—ACATAGTCTGGAAGGGAAGCGTAC	
12	TCCTGCAGTTGGTCACTATCATTAGTCCCACAGGGCAACT	
35	CGCTAAAGCTGTAC—TCTCAGTCCCGAAGGGGAGCTTTTT	
42	CGCTAAAGCTGTAC—TCTCAGTCCCGAAGGGGAGCTTTTT	
94	A—GTTGAATAGTGTGGTCACTTCCAGTCCCGCAGGGCACAC	Group II (12.90%)
83	A—GTTGAATAGTGTGGTCACTTCCAGTCCCGCAGGGCACAC	
1	A—GTTGAATAGTGTGGTCACTTCCAGTCCCGCAGGGCACAC	
4	A—GTTGAATAGTGTGGTCACTTCCAGTCCCGCAGGGCACAC	
53	G—AGGAGTATCGGGGC—ACGATCCTATTTAAGGCCGATAACC	
62	C—GGAGTATACATTCTG—ATCCTATATAGGTAGGCCGAAACC	
81	C—GGAGTATACATTCTG—ATCCTATATAGGTAGGCCGAAACC	
85	T—CGACTATGTTGCTAC—ATCCTGTACGGTCAGGGCGAAAG	
25	CTGATTATATAACGTG—GTGATATCCGGACAAGGCAGAAAA	
67	AA—G—CTGATTACCAAACACTGC—GTGCCGCCCGACCCGGTGA	
14	GCTGAGCATCAATTCGCAT—TGATCCCACAGTTGGGCAGCTA	Group III (10.60%)
92	CTGAGCATCACGAAGT—GATCCTGTCGAGGACGGGCAGTTA	
52	CTGAGCATCACGAAGT—GATCCTGTCGAGGACGGGCAGTTA	
13	CTGAGCATCACGAAGT—GATCCTGTCGAGGACGGGCAGTTA	
16	CTGAGCATCACGAAGT—GATCCTGTCGAGGACGGGCAGTTA	
71	CG—C—ACGAGCATCGTACTTCC—GTTATGATCATGAGACACAG	
77	C—GTGGTTCCTGCGGGCAGGTTAT—CAACATGGGCAATGCGT	
27	AT—AA—GCTGAGCATCTTAG—GTCCTGTCA—AGGGGCAGCGTAA	
10	ATAAGCTGAGCATCTTAG—ATCC—CTGTCAAGGGCAGCGTAA	
11	ATAAGCTGAGCATCTTAG—ATCC—CTGTCAAGGGCAGCGTAA	
31	ATAAGCTGAGCATCTTAG—ATCC—CTGTCAAGGGCAGCGTAA	
24	ATAAGCTGAGCATCTTAG—ATCC—CTGTCAAGGGCAGCGTAA	
60	ATAAGCTGAGCATCTTAG—ATCC—CTGTCAAGGGCAGCGTAA	
78	ATAAGCTGAGCATCTTAG—ATCC—CTGTCAAGGGCAGCGTAA	
28	A—A—TTGGATTACTTCT—CAAGTCTGATGAGGGCAATTTAAA	
56	T—TGGGTACCTCGTAGG—TCCTAATGGAGGTGAGGCAACTCC	

**Table S4. Alignment of the top 36 aptamer sequences obtained from QSAS R3/R2**

Seq. ID	Sequences (5' → 3')	
11	CTGAGCATCACGAAGTGATCCTGTGCGGGACGGGCAGTTA	Group I (47.20%)
32	CTGAGCATCACGAAGTGGTCCTGTGCGAGGACGGGCAGTTA	
33	CGAATGCAAGTCCTGACGGAGGCCAGGCGCTGAAACTTGC	
21	ATAAGCTGAGCATCTTA—GATCC—CTGTCAAGGGCAGTGTA	
1	ATAAGCTGAGCATCTTA—GATCC—CCGTCAAGGGCAGCGTAA	
23	GATACTGAGAATCGTA-CATGATCC—CGCAACGGGCAGTATC	
2	GATACTGAGCATCGTA-CATGATCC—CGCAACGGGCAGTATT	
12	CCGGGAACATTGTCTAAACAATGTCATTTATGTCGGAAGC	
14	TAAGGGCACTATTGCATGGTGAGCATCCTTGATCCCGCT	
9	ATAAGCTGAGCATCTTA—GATCC—CTGTCAAGGGCAGCGTAA	
13	ATAAGCTGAGCACCTTA—GATCC—CTGTCAAGGGCAGCGTAA	
18	AATGGATGGGCACCGCTATAGTTGGTCCCAGGAGGGCATGC	
22	GATACTGAGCATCGTACA-TGATCC—CGCAACGAGCAGTATC	
29	CATACTGAGCATCGTACATGATCCC—GCAACGGGCAGTATC	
25	CGGGGCAACAACGTTGAGCATCATTATGATCGCCGCTAT	
24	AGGTGAGCATCTTTAGATCCCATTTGGGCGCCTTTCTTTT	
8	TTGGGGCGGTCCCTGTGAAAGGCAAAAATCTATTATACCGC	
16	CCGTACAGTCTAAGACTCCCATGAACATAGGCGGAAGTA	
6	CCGTACAGTCTAAGACTTCTATGAACATAGGCGGAAGTA	
30	CCGTACAGTCTAAGACTCCTATGAACATAGGCGGAAGTA	
34	CTGTTACAGTCTAAGACTCCTATGAACATAGGCGGAAGTA	
4	CCGTACAGTCTAAGACTCCTATCAACATAGGCGGAAGTA	
26	GCTCCGTACCATTCTTATGGTCCCAGGCGGCGCAAACG	
20	AGTTGAATAGTGTGGTCACTTCCAGTCCCAGGGGGCACAC	Group III (13.90%)
3	AGTTGAATGGTGTGGTCACTTCCAGTCCCAGGGGGCACAC	
5	TGTGGGTATGGTCTAATTTTTAGGCACGGAGGTACCAT	
31	GCTGAGTTAGATCCCTTTCGTAAGGGCAGCCGGGTATCTA	
7	TACGAGTTTATCCTTTTCCATTAGGCGTACAGCTCATCAA	
15	TACGAGTTGATCCTTTTTATTAGGCGTACAGCTCATCAA	
19	TACGAGTTGATCCTTTTTATTATGCGTACAGCTCATCAA	Group IV (16.70%)
36	GAGGGCTGATGCAATCCTAGTATAGGCAGCATAACATTGC	
10	GCTGAGTTAGATCCCTTTT—GTAAGGGCAGCCGGGCATCGA	
17	GCTGAGTTAGATCCCTTTT—GTAAGGGCAGCCGGGCATCTA	
35	TAAGGTGGGGATAAAGGACGTGCGGGATCGGGGGGGGGAT	
27	CTTATT—GTGCGGGCACCTCAGGTCCTAAAGTTAGGCGCAC	
28	TTTATT—GTGCGGGCACCTCAGTTCCTAAAGTTAGGCGCAC	

**Table S5. Alignment of the top 36 aptamer sequences obtained from QSAS R3/R1**

Seq. ID	Sequences (5' → 3')	
8	TGCGGGTTACTTTTAAGTCCCAAGGGGCCGTAGATCACCA	Group I (38.90%)
26	CGAGGGGTACCCTGCTGGGTCCCCGGTAGGGGCGAGTTCT	
12	TTACGGTGTGGTCACTATTTAGTCCCTACTGGGCACACC	
5	GCTGGTCACTCTAGAGTCCTTGCAGGCAGCAAACATAAT	
24	TAAAGGGTGTGGGGCACCCCTCGGGTCCCTGTGGGGCAC	
25	TTGGATCATTGGCACCTCAGTCCCTACGGGGGAATGATT	
30	CAGTGGATGGTCACTCCCAGTCCCTAACAGGCATCAACGG	
19	TCAAGTCTTTTtaggcaaccatattgTTGGTCATTTGATA	
33	CTGACGAGGCACGCAAGTGGGCACCTTTAGGTCCCTCTTT	
1	ATTTGAATAGTGTGGTCACTCCAGTCCCGCAGGGCACAC	
34	AACACGTGGGCACCTTCAGTCCCCCGCGGGGCGCGTG	
35	ATAATATCATGGTTGGGCACCTAAGGTCACGCAGTGCAAC	
23	ACGTACGTGGGCACCTCTGGTCCAGAAGGGCACGGACCTG	
9	ATGCACGGGACCATGTATGGTCTGATTATAGGGCGTGG	
31	TAGTTTATGTAATCCTGTCTTCAGGCCAGACGGTAACC	
18	GGTGGTCACTAGCTTAGTCCCATTACTTGGGCACCAGGGT	
11	GGAGCGTCACTAATTTAGTCCCAATAGGGGGATCCTTATC	
20	TGTCGAGCATCAGTGGGTGATCCTGAGGGACCAGGCGAC	
15	GAGAGCTGATGCAATCCTAGTATAGGCAGCATAACATTT	
21	GGGACGGAGCATCGTTTACGATCCCCGCGGGCCGTGGCA	
27	CCGTTACAGTCTAAGACTCCTATGAACATGGGCGGAAGTA	
36	CTGCGGTCACTGCTCAGTCTGCAATAGCAGGCGCAGACT	
3	CGGGCGCTGGGGCGGGTACCTTTGGTCCCGATATCAAATC	
6	ATTCAGATTAAGGGCTGCCTGAACTCGTTCCCGGTATCAT	
32	GTTTCCATACGGCACCTATGGTCTTGTAAGGGTATGGT	
17	TATGATCAAATCCTCGTTGTACGGGGCATAAACTGATTG	
7	TCCATCGAGGCATCCCTGTAAAGAGGATCATGAGTACGT	
29	CGTCTCAGTAGATATTACTCCTAGCTAAACTAGGCGTGAG	
4	TACTTGAGCATCGTTTTATGATCCTTTGGGCAAGTCCGT	
10	AATACCGTCACTTTTtagtctgtTGTTAAGAGCAGGGGGT	
14	CTCTCGTCACTTTAGTCCCTTTAAAGGGGAGAGCTTCG	
28	GTCGCTCGTTGTACGGTAGGTCCCTGAAGGGGAGCGATG	
13	TAGGCATGTAATGAGCATCTGTGATCCTATGGTTCTTCAT	
22	GCGTTCGAAGTTAGCGGAATTCCTCACTAGGCGCAAGGA	
16	GGCTCGGAACCGCATAGTGGTCTGGTCCAGGGGAGCC	
2	TAGTCTGTTACGGTCTCGATTGTCCAAAAGGGCAGACA	