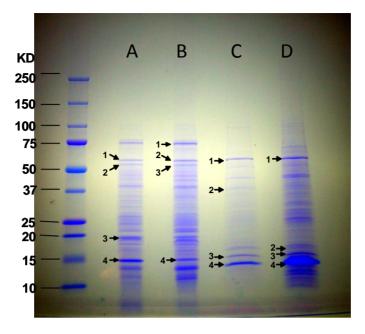
SUPPLEMENTAL DATA

Supplemental Data Table 1. Bacterial strains and plasmids used in the experiment.

	ible 1. Bacterial strains and plasmids used in the experime	
Strains or Plasmids	Description	Reference
Strains		USAMRIID, Frederick, MD
Francisella tularensis LVS	Pm^{r} , Live vaccine strain $F(a)^{0}dla = 7 \cdot Ml^{5}$ and $L = adAl = arcA06 this l had Bl7(rith)$	
<i>Esherichia coli</i> DH5α	$F'(\phi 80dlacZ vM15)$ recA1 endA1 gyrA96 thi-1 hsdR17(r'k	63
	M^+k) supE44 relA1 deoR Δ (lacZYA-argF) U169	
Shigella dysenteriae sd197	S. dysenteriae serotype 1	MGH ^a , Boston, MA
Ft.LVS::wbtA	$Pm^{r} Km^{r}$, Live vaccine strain ; $\Delta wbtA$	31
Ft.LVS::wzy	Pm^r , Live vaccine strain ; Δwzy	This study
<i>Ft</i> .LVS:: <i>wzy</i> /pTH21	Pm^{r} , Live vaccine strain ; Δwzy harboring pTH21	This study
Ft.LVS::wzy/pTH33	Pm ^{r} , Live vaccine strain ; Δwzy harboring pTH33	This study
Plasmids		
pEX18Tc	6.3-kb; Tc ^r ; $oriT^+$ sacB ⁺ , gene replacement vector with MCS from pUC18	64
pEX18Km	6.8-kb; Km^{r} ; $oriT^{+}$ sac B^{+} , gene replacement vector with MCS from pUC18	This study
pPV	Ap ¹ , Cm ¹ , <i>sacB</i> , mob	38
pFNLTP6	6.9-kb; pFNLTP1 derivative with <i>NdeI</i> , <i>Eco</i> RI, <i>SmaI</i> , <i>NotI</i> , <i>NheI</i> , and <i>XhoI</i> restriction enzyme sites (MCS2) cloned between the <i>KpnI</i> and <i>Bam</i> HI sites; km ^r Ap ^r	65
pCR2.1 TOPO	3.9-kb; plasmid for cloning PCR product, km ^r Ap ^r	Invitrogen, Carlsbad, CA
pTH30	11.4-kb; pEX18Km harboring <i>sacB</i> (from pPV) and upstream and	This study
PILLO	downstream of wzy	This study
pTH17	7.1-kb; pFNLTP6 harboring P_{groEL}	This study
pTH21	8.3-kb; pFNLTP6 harboring P_{groEL} ::wzy	This study
pTH33	8.3-kb; pFNLTP6 harboring P _{groEL} ::wzy _{shigella}	This study
pE25A	8.1-kb; pFNLTP6 harboring P_{groEL} ::wzy _{E25A}	This study
pK27A	8.1-kb; pFNLTP6 harboring P_{groEL} ::wzy _{K27A}	This study
pD52A	8.1-kb; pFNLTP6 harboring P_{groEL} ::wzy _{k27A}	This study
-	8.1 kb; pENI TD6 harboring P $_{groEL}$. w2yD52A	•
pD144A	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{D144A}	This study
pK152A	8.1-kb; pFNLTP6 harboring P_{groEL} :: <i>wzy</i> _{K152A}	This study
pK153A	8.1-kb; pFNLTP6 harboring P_{groEL} :: wzy_{K153A}	This study
pE157A	8.1-kb; pFNLTP6 harboring P_{groeL} :: wzy_{E157A}	This study
pE163A	8.1-kb; pFNLTP6 harboring P_{groeL} :: wzy_{E163A}	This study
pK165A	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{K165A}	This study
pR167A	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{R167A}	This study
pD177A	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{D177A}	This study
pE277A	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{E277A}	This study
pD287A	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{D287A}	This study
pH290A	8.1-kb; pFNLTP6 harboring PgroEL::wzyH290A	This study
pK399A	8.1-kb; pFNLTP6 harboring PgroEL::wzyK399A	This study
pE400A	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{E400A}	This study
pR406A	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{R406A}	This study
pK399A	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{K399A}	This study
pG176A	8.1-kb; pFNLTP6 harboring PgroeL::wzyG176A	This study
pG176E	8.1-kb; pFNLTP6 harboring PgroeL::wz.yG176E	This study
pG176S	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{G176S}	This study
pG176K	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{G176K}	This study
pD177S	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{D177S}	This study
pD177E	8.1-kb; pFNLTP6 harboring PgroEL::wzyD177E	This study
pD177K	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzy _{D177K}	This study
pG178A	8.1-kb; pFNLTP6 harboring P _{groEL} ::wzyG178A	This study
pG178S	8.1-kb; pFNLTP6 harboring P_{groEL} ::wzy _{G178S}	This study
pG178E	8.1-kb; pFNLTP6 harboring P_{groeL} ::wzy _{G178E}	This study
pG323A	8.1-kb; pFNLTP6 harboring P_{groEL} ::wzy _{G323A}	This study
pG323S	8.1-kb; pFNLTP6 harboring P_{eroEL} ::wzy _{G3238}	This study
pG323E	8.1-kb; pFNLTP6 harboring P_{groeL} ::: $w_{ZyG323E}$	This study
pG323R	8.1-kb; pFNLTP6 harboring P_{groEL} ::wzy _{G323R}	This study
pG323A	8.1-kb; pFNLTP6 harboring P_{groEL} ::wzy _{G323k}	This study
pG323E	8.1-kb; pFNLTP6 harboring P_{groEL} ::wzy _{G324}	This study
wzy-gfp	8.9-kb; pFNLTP6 harboring wzy::gfp	This study
P-gfp	7.8-kb; pFNLTP6 harboring P_{groeL} ::gfp	This study
P-wzy-gfp	9.1-kb; pFNLTP6 harboring P _{groEL} ::wzy::gfp	This study
		This study
P-wzy $(G_{176}E)$ -gfp P wzy $(D_{176}A)$ gfp	9.1-kb; pFNLTP6 harboring P_{groEL} ::wzy($G_{176}E$)::gfp	•
P-wzy $(D_{177}A)$ -gfp	9.1-kb; pFNLTP6 harboring P_{groEL} :: $wzy(D_{177A})$:: gfp	This study
P-wzy $(G_{323}E)$ -gfp	9.1-kb; pFNLTP6 harboring P_{groEL} ::wzy(G_{323E})::gfp	This study
P-wzy $(Y_{324}E)$ -gfp	9.1-kb; pFNLTP6 harboring P_{groEL} ::wzy($Y_{324}E$)::gfp	This study
P-wzy (K ₁₅₂ A)-gfp	9.1-kb; pFNLTP6 harboring P _{groEL} ::wzy(K ₃₅₂ E)::gfp	This study

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Identified proteins	Predicted Presence of identified proteins in each lane					
from MASS Spec. analysis	subcellular	А	В	С	D	
	location	(periplasm)	(periplasm)	(outer membrane)	(outer membrane)	
peroxidase/catalase	Periplasm ⁶⁶	○ (A-1)	○ (B-2)	× (C-1)	× (D-1)	
peroxiredoxin,AhpC-TSA family protein	Periplasm ⁶⁷	○ (A-4)	○ (B-4)	× (C-3)	× (D-3)	
outer membrane protein OmpH	Outer Membrane ⁶⁸	× (A-4)	× (B-4)	O (C-3)	○ (D-3)	
outer membrane lipoprotein blc	Outer Membrane ⁶⁹	× (A-4)	× (B-4)	○ (C-3)	O (D-3)	
17KDa outer membrane protein	Outer membrane ⁷⁰	× (A-4)	× (B-4)	○ (C-3)	○ (D-3)	

O - presence (of the protein in lane), X-no presence (of the protein in lane) A: Ft.LVS.wt (periplasm); B: Ft.LVS::wzy (periplasm); C: Ft.LVS.wt (outer membrane) ; D: Ft.LVS::wzy (outer membrane) ;

Supplemental Data Figure 1. Mass spectrometry analysis of proteins located in subcellular fractions. The isolated subcellular fractions were analyzed by SDS-PAGE gel (4-20% gradient gel) and 4 randomly chosen protein bands in each fraction was analyzed by Mass spectrometry to identify the proteins and predict their subcellular locations. The table shows the fraction-specific protein distribution in SDS-PAGE gel bands from the periplasmic and outer membrane fraction. Although they have the same MW, different proteins are found in each fraction indicating the purity of the fractions. For example, peroxidase/catalase, which is a periplasmic protein, was only detected in A-1 and B-2 (periplasmic fraction bands), but not in the C-1 and D-1 band which are collected from the outer membrane fractions even though these four bands are the same MW).

		Presence in the lane after MS spec analysis							
	Predicted	Periplasmic fraction			Outer membrane fraction				
	Subcellular Location		b	с	d	e	f	g	h
		20 µg/ml	20 µg/ml	0.2 µg/ml	0.2 µg/ml	20 µg/ml	20 µg/ml	0.2 µg/ml	0.2 µg/ml
peroxiredoxin, AhpC-TSA family protein	Periplasm	0	0	0	0	×	×	×	×
17kDa outer membrane protein	Outer membrane	×	×	×	×	0	0	0	0
20		μg/ml 2 μg/m		nl 0.2 μg/ml					
KDa WT		Δwz	y y	wт	Δw	zy	WT	Z	lwzy
Periplasmic 15 fraction 10		b	>						\bigcirc
Outer 15 membrane fraction 10	e e	f C	>	~	~	g		h	\supset

•: presence of the protein (after Mass spec. analysis)

x: *no presence of the protein (after Mass spec. analysis)*

a-h: ~15 KDa size of protein band in each fraction (All samples were ran in 4-20% gradient gell and stained with Coomasie).

20 μg/ml: original protein concentration in sample fractions (calculated by Bradford assay) **0.2 μg/ml:** 1/100 diluted concentration of sample fractions

Supplemental Data Figure 2. Mass spectrometry analysis for purity estimation. The concentrations of periplasmic or outer membrane fraction from wild type or mutant were normalized to $20 \ \mu g/ml$ and $1/100 \ dilution$ of normalized fractions (0.2 $\mu g/ml$) were prepared. In order to determine whether the specific proteins in the diluted samples can still be identified, both the diluted and undiluted samples were analyzed by SDS-PAGE and a 15KDa band in each fraction was further analyzed by Mass spectrometry. Based on this analysis, the fractions we are investigating have at least 99% purity because a fraction specific protein can still be detected in a 1/100 dilution of the membrane fraction yet not be seen in the periplasmic fractions from wild type and mutant organisms even at a 1/100 dilution. This protein was not detected in the outer membrane fraction even in the undiluted sample. Therefore the periplasmic fraction is over 99% free of outer membrane.

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