



Fig. S1. A. Enzyme activities of CYP93G2-expressing yeast microsomes toward different flavonoid substrates. Data are mean \pm SD of triplicate measurements and expressed in nmol product min⁻¹ mg protein⁻¹. N.D. = not detected. B. Production of 2-hydroxyeriodictyol from eriodictyol in reactions catalyzed by CYP93G2-expressing yeast microsomes.



Fig S2. Accumulation of *C*-glucosyl-2-hydroxyneriodictyol in transgenic Arabidopsis co-expressing CYP93G2 and OsCGT. (A) Detection of a distinct peak producing a [M-H]⁻ ion at m/z 465 in the double transformant sample. (B) MS/MS spectrum of the m/z 465 ion revealed the fragmentation pattern consistent with that reported for *C*-glucosyl-2-hydroxyflavanone in the open chain form as shown in (C).



Fig. S3. Representative extracted ion chromatograms (XICs) for the *C*-hexosylflavone derivatives (M4 – M19, Table 1) in the rice wild-type (WT) and the CYP93G2 mutant (MT) non-hydrolyzed methanol samples.



Fig. S4. MS/MS spectra for M4-M13 (*C*-hexosyl-flavone *O*-glycosylated derivatives). Fragmentation pattern for selected ions observed in the spectra is shown based on the structure of a 6*C*-hexosyl-flavone.



Fig. S5. MS/MS spectra and fragmentation patterns for M14 – M19 (*C*-hexosyl-*C*-pentosyl flavones). Fragmentation pattern for selected ions observed in the spectra is shown based on the structure of a 6*C*-heoxyl-8*C*-pentosyl flavone.

Precursor ion [M+H] ⁺	Retention time (min)	MS/MS ions	Compound assignment
613	12.8	451, 289	2-hydroxy-naringenin di-O,O-hexosides
451	17.1,18.5	289	2-hydroxy-naringenin O-hexosides
629	12.9	467, 305	2-hydroxy-eriodictyol di-O,O-hexosides
467	16.1, 16.8	305	2-hydroxy-eriodictyol O-hexosides

Table S1. List of 2-hydroxyflavanone *O*-glycosides identified in Arabidopsis overexpressing CYP93G2. Metabolites were identified in non-acid hydrolyzed extracts by LC-MS. The identities of the $[2-hydroxyflavanone+H]^+$ ions (bold) had been confirmed by their MS³ spectra

Table S2. List of primers used in this study

Gene		Forward Primer		Reverse Primer	Purpose
CYP93G2	CL543	5'-ACTATGGAGGAAGGCGTCGTCGGTG	CL544	5'-GGAGTAGAAGGAAGGGAGCGGTTGG	Full-length cDNA
					cloning for yeast expression
	CL993	5'-GCGTCTAGAATGGAGGAAGGCGTCGT	CL994	5'-CGCGAGCTCTCAGGAGTAGAAGGAAGG	Full-length cDNA
					cloning for
					Arabidopsis
					over-expression
	CL1216	5'-GCTGGAGACCATCATCGAG	CL1256	5'-TGAAGAAGACCTGGGTGTCG	Genomic PCR
					and RT-PCR
OsCGT	CL1316	5'-GATGGATCCATGCCGAGCTCTG	CL1317	5'-ACCCTCGAGTCAATTAGTGCG	Full-length cDNA
					cloning for
					Arabidopsis
					over-expression
	CL1329	5'-CTCTCCCTCTGCGCCTACTT	CL1330	5'-GATCTTTTGCCTGGTTGCTC	Genomic PCR
					and RT-PCR