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Supplemental Information

E-Cadherin Marks a Subset

of Inflammatory Dendritic Cells

that Promote T Cell-Mediated Colitis

Karima RR Siddiqui, Sophie Laffont, and Fiona Powrie



Figure S1. Related to figure 1. Phenotypic characterisation of E-cadherin⁺ DCs.

Tissues from BALB/c WT mice were harvested and the distribution and phenotype of E-cadherin⁺ DCs was assessed by FACS. (a) Representative FACS-analysis gating strategies applied to all experiments unless otherwise stated. Steady state, but not colitic, MLN cells were CD11c-enriched. Cells were gated on the high FSC and SSC cell population in order to remove the majority of CD11c⁻ cells, 7-AAD⁺ dead cells were excluded and E-cadherin expression amongst the CD11c^{hi}MHC class II^{hi} cells was examined. (b) Staining of E-cadherin, CD11c and the relevant isotype control of E-cadherin on cells from MLN, spleen and ALN (axillary lymph nodes). (c) Expression of MHC class II and co-stimulatory molecules CD80, CD86 and CD40 on E-cadherin⁺ and E-cadherin⁻ CD11c^{hi} cells. Black solid lines, E-cadherin⁺ DCs; grey solid line, E-cadherin, Gr1, CD8 α , CD11b and CD103 gated on CD11c^{hi} DCs from MLN cell preparations. Positioning of the quadrants reflects isotype controls. In all cases, representative plots from 5-10 individual analyses are shown. Experiments have been repeated more than 3 times.



Figure S2. Related to figure 2. E-cadherin expression during T cell mediated colitis

(a) Tissues from colitic B6 Rag1^{-/-} and steady state B6 WT mice were harvested and the distribution of E-cadherin⁺ DCs was assessed by FACS. Staining of E-cadherin and CD11c gated on MHC class II⁺ cells from colon, MLN, spleen and blood. Positioning of the quadrants reflects the isotype control. Representative plots from 4 individual mice per group are shown. (b) CD11c^{hi}MHC class II^{hi}7-AAD⁻CD4⁻CD45⁺ DCs were FACS-sorted from pooled cell preparations of colon, MLN and spleen of colitic BALB/c Rag2^{-/-} mice (n = 6). *Cdh1 (E-Cadherin)* gene expression was assayed in triplicate by qPCR and each sample was normalized relative to HPRT expression. Mean \pm S.E.M expression from the triplicate samples is shown. Representative data from 1 of 2 experiments are shown, a further experiment gave similar results.



Figure S3. Related to figure 3. Characterisation of Gr1⁺ and Gr1⁻ monocytes

(a) Phenotypic characterisation of FACS-sorted $Gr1^+$ and $Gr1^-$ CD115⁺ monocytes harvested from pooled blood and BM from congenic CD45.1 mice (n = 8). Representative staining of CD11b, ckit and CD11c. Positioning of the quadrants reflects the isotype controls. (b) Mean frequency of CD45.1⁺ cells expressing CD11c^{hi} derived from $Gr1^+$ monocytes (n = 3). Error bars represent s.d. Representative data from 1 of 2 independent experiments.



Figure S4. Related to figure 5. MLN E-cadherin⁺ CD11c^{hi} cells from colitic mice display characteristics associated with DCs

(a) Representative staining of MHC class II, CD80, CD86 and CD40 by E-cadherin⁺ and E-cadherin⁻ CD11c^{hi} cells from the MLN of colitic Rag2^{-/-} mice (n = 4). Black solid lines, E-cadherin⁺ DCs; grey solid line, E-cadherin⁻ DCs; and dashed lines, isotype control antibodies. (b-c) 2 x 10⁵ CFSE-labelled CD4⁺ T cells from DO11.10 SCID mice were cultured in triplicate with 3 x 10⁴ E-cadherin⁺ or E-cadherin⁻ CD11c^{hi}CD103⁻7-AAD⁻CD4⁻ cells, or myeloid MHC class II⁺CD11c⁻F4/80⁺ cells, taken from the MLN of colitic BALB/c Rag2^{-/-} mice in the presence or absence of OVA protein (5µg/ml). At day 4 of culture, the cells were counted, and T cell CFSE dilutions were assessed by FACS analysis. The percentage of CD4⁺ T cells present in each cell division was determined. (b) Mean percentage of CD4⁺ T cells present in Each of the CFSE divisions. Black bar, T cell proliferation driven by E-cadherin⁺ DCs; open bar, T cell

proliferation driven by myeloid MHC class II⁺CD11c⁻F4/80⁺ cells (c) Number of cells in E-cadherin⁺ (black bar), E-cadherin⁻ (dark grey bar) or myeloid (light grey bar) DC-T cell cultures containing OVA protein, or in E-cadherin⁺ DC-T cell cultures without OVA protein (open bar). Error bar represents S.E.M of triplicate cultures. Data from 1 of 2 independent experiments are shown.



Figure S5. Related to figure 7. Phenotypic characterisation of E-cadherin⁺ BM-DCs

(a-d) BM-DCs were generated in the presence of GM-CSF or Flt3L. At day 8 of culture, cells were harvested, pooled, and the distribution and phenotype of E-cadherin⁺CD11c⁺ cells were analysed by FACS. (a) Mean percentage of CD11c⁺ cells

expressing E-cadherin following culture in GM-CSF or Flt3L. (b) Expression of MHC class II and co-stimulatory molecules CD80, CD86 and CD40 on E-cadherin⁺ and E-cadherin⁻ CD11c⁺ BM-DCs cultured in GM-CSF. Black solid lines, E-cadherin⁺ BM-DCs; grey solid line, E-cadherin⁻ BM-DCs; and dashed lines, isotype control antibodies. (c) Representative staining of E-cadherin, CD103 and the two relevant isotype control antibodies gated on CD11c⁺ BM-DCs cultured in GM-CSF. (d). Representative staining of E-cadherin, CD8 α and CD11b gated on CD11c⁺ BM-DCs cultured in GM-CSF. In all cases, representative plots from more than 3 independent experiments are shown.