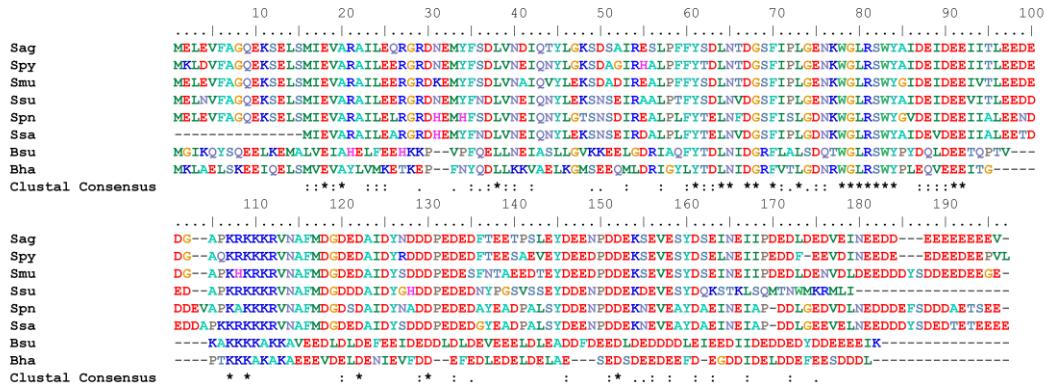


## SUPPLEMENTARY

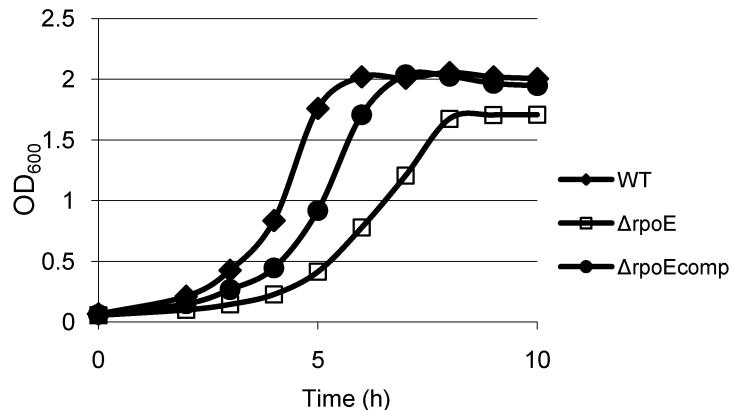
Figure S1



## S1. Amino acid sequence alignment of delta proteins using CLUSTAL W 2.0.

Identical amino acid residues are marked with a star, strongly similar with two dots, and weakly similar with one dot. The default colourscheme is used for alignments. Species abbreviations: Sag, *S. agalactiae*; Spy: *S. pyogenes* M1 GAS; Smu, *S. mutans* UA159; Ssu, *S. suis* 98HAH33; Spn, *S. pneumoniae* G54; Ssa, *S. sanguinis* SK36; Bsu: *Bacillus subtilis* ; Bha: *B. halodurans* C 125.

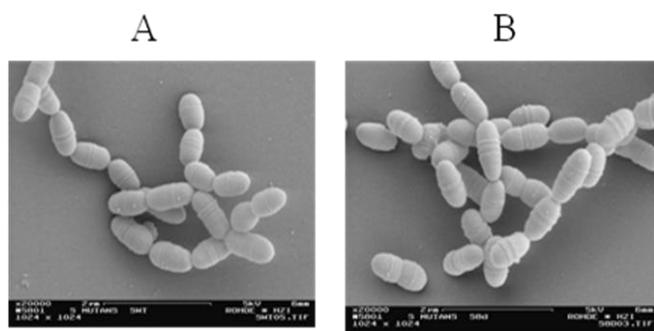
Figure S2



## S2. Growth defects of *S. mutans* $\Delta rpoE$ strain.

The growth of *S. mutans* cultures in THBY medium pH7.5 at 37 °C in an atmosphere of 5% CO<sub>2</sub> were recorded every hour by measuring the absorbance (optical density) at 600nm. Strains: WT (wild type),  $\Delta rpoE$ ,  $\Delta rpoE$ comp ( $\Delta rpoE$  complementation). The data is a representative result from at least three independent experiments.

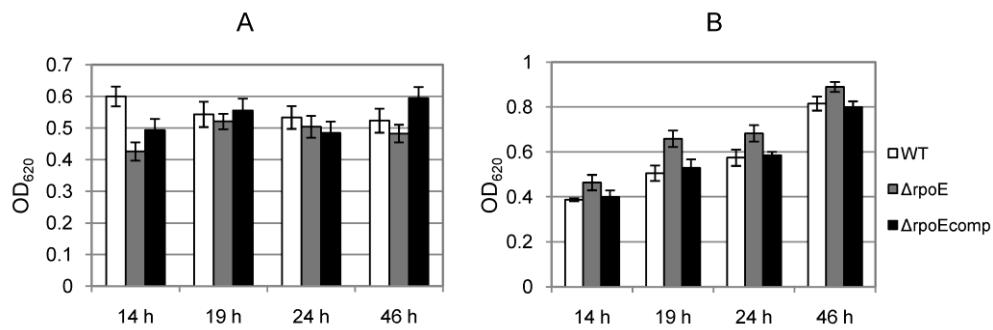
Figure S3



**S3. *S. mutans* wild type and  $\Delta rpoE$  strains cell phenotype under scanning electron microscopy.**

A: wild type strain; B:  $\Delta rpoE$  mutant.

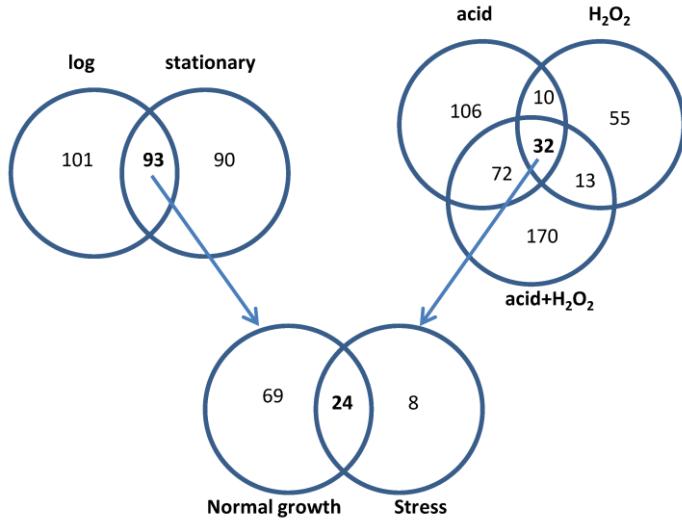
Figure S4



#### S4. Biofilm quantification by crystal violet staining.

Biofilm of *S. mutans* wild-type (blue columns),  $\Delta rpoE$  (red columns), and  $\Delta rpoE$ comp (green columns) grown in 96-well microtiter plates in THBYS (A) and BMS (B) medium were stained at different time points with 0.1% crystal violet and the absorbance of the extracted stain was measured at 620 nm. Data presented above is representative result from at least three independent experiments. Mean and standard deviation of twelve replicas of each sample are given.

Figure S5



**S5. Venn diagram showing the number of differentially expressed genes in the  $\Delta rpoE$  mutant compared to the wild type strain under 5 experimental conditions.**

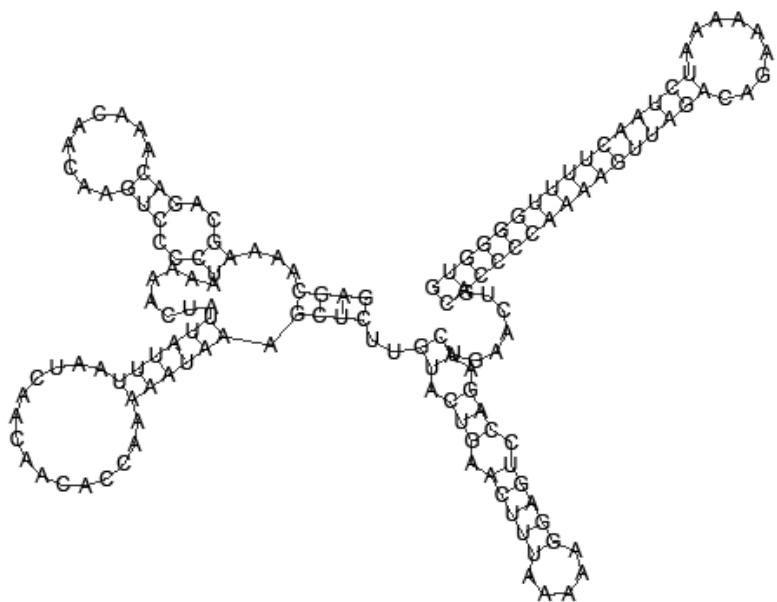
Sample collection was as following: Log, M0/W0; stationary, M1/W1; acid, M2/W2; H<sub>2</sub>O<sub>2</sub>, M3/W3; acid + H<sub>2</sub>O<sub>2</sub>, M4/W4.

Figure S6

smu-r1

A

B



C

r1.1 **SMU.948** hypothetical protein

## r1.2 SMU.1025 putative transcriptional regulator

sRNA	100	ACUGAACUGCACCCAAA         :	117
mRNA (SMU.1025)	-89	UGACUUAGACGUGGGGUUU	-106

### r1.3 **SMU.2059c** putative integral membrane protein

sRNA	1	GAGCAAA-AGCAGACAAACAACAAGUCCCCUAAAAACUAUU---AUUUAUCAACA         :   :       +	52
mRNA (SMU.2059c)	55	CUCGUUAUUCUUUGUCUCUUGUUCAGG--ACUAUAAAAACUUAAAUCGCUGU	2

### r1.4 **SMU.327** DNA repair protein RadA

sRNA	48	CAACAAACACCAA-AAAUA--AAGCUCU-UGUACUGAACUUU                       :     :           :     :	84
mRNA (SMU.327)	-119	GUAGUUGUGGUUAAAAGAUUUGAGAGACGUGGC--GAAA	-157

### r1.5 **SMU.1507c** hypothetical protein

sRNA	28	CCUAAAACU-AUU---AUUUAUCAACAACA                 :	55
mRNA (SMU.1507c)	71	GGAUUUUGAGUAGCAGUAAAAGUUGUUGU	39

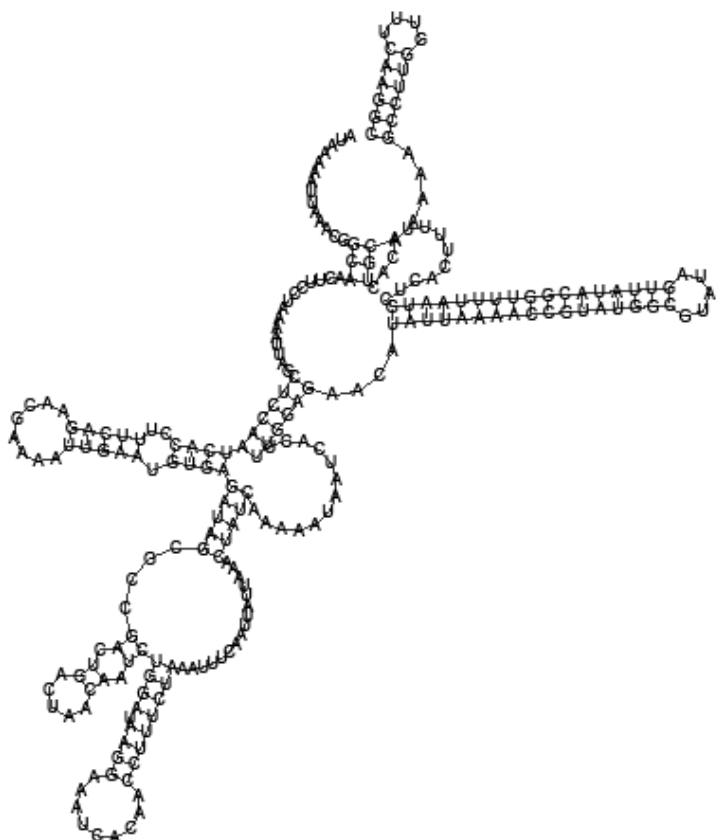
**S6. Predicted smu-r1 promoter and terminator (A), secondary structure (B), and interaction with target mRNAs (C). A: The putative -35 and -10 regions were marked by green colour. The sequence of smu-r1 was underlined. The predicted transcription start was indicated by bold, while the putative terminator was marked by red colour.**

Figure S7

smu-r2

A

B



C

r2.1      **murB**      UDP-N-acetylenolpyruvoylglucosamine reductase

sRNA	60	AUAAGGAAUACAAAC-CUUUCUAAAUAU---CAUUU-AUAAAACAUACAAAAAUAAU---C---ACUUUGGAGAACAUAAAACCGUAUUGCUGUAUGUUAUAC-GGUUUA--AUGCUCACUUUACACUGC	181
		:    :      :                :          :    :    :    :    :    :    :    :    :    :    :    :    :    :    :    :	
mRNA (mrbB)	50	UAUGCCUAUAG--UUGCGGAAGAUUUGAAAACAAGUAAGUAAAUG-UAGUAAUCGAAGAGAGAUGAAAAA---GUAAUAAAUAU---UAGC-CAU-UAAAUGGCUAAAUAUAGUUAAAUG-GAUG	-74

r2.2 **SMU.896** hypothetical protein

Score: -97 Pvalue: 0.000684607

sRNA	77	UUUCUAAUUUCAUUAAA-CUAUCAAAA   :          :      :	107
mRNA (SMU.896)	-10	AGAGACUAAAGGUAGUAGUUUGAUAGUUU	-41

## r2.3 SMU.213c hypothetical protein

Score: -88 Pvalue: 0.00236569

sRNA	75	CCUUUCUAUUUCAUUUAUAAAAC-UAUCAAAAUAAUCACUUU      :             :     : :	118
mRNA (SMU.213c)	-12	GGAAGAAGUUUAA---UAUUAGUUGUGCGGUUUAUAGU--UGAAA	-51

r2.4 SMU.1969c putative transcriptional regulator

sRNA	17	CGAAAAUUGAAUGAGAUAGCGCCGACUGACU--AACAAUCUGGUAAG--GAAAUCAC-A--ACCUUUCUAAAUAUCAAU-UA-UUUACAUCAAAAUAUACUUUGGA--GAACAUAU--UAAAA	134
		:        :       :                :                  :                         :	
mRNA (SMU.1969c)	78	GCUUUUAACUGACA-----GGUAGA--GAACUCGUAGAUGAUAGACUUUAGGGGUUUAGAAGAUU-AAAGUUUAUUAGUGGGAAAGACU--AAUAGUCAA-CCUAACUGAUUAUUGUUUU	-38

r2.5 cysK putative cysteine synthetase A; O-acetylserine lyase

Score: -83 Pvalue: 0.00470788

sRNA	59	GAUAAGGAAUACACAACCUCUUCUAAAUCUUAAUUAACUAUCAAAAAUACACUUUGGAG                :    :            :            :	122
mRNA (cysK)	-81	CUAUCCUUUAGUAUAGUGGCC--UUGAAGGUUAAAACUAUAGGUCGUUGU-ACGGAAAAUUC	-141

r2.6 SMU.375 hypothetical protein

Score: -82 Pvalue: 0.00540194

sRNA 25 GAAUGUGAGAUAGCGCCGACUGACUAACAAUCUGGUAAGGGAAUCACAACCUUUCUA 82

mRNA (SMU.375) -79 CUUCCUUUUUAUCACAG---ACUGAUUGU-AGAAGUUGUC---AGUGUUUGAAAGGAU -128

## r2.7 SMU.1392c putative acetyltransferase

Score: -80 Pvalue: 0.00711096

sRNA	8	CUUUCAGAAC--GAAAAUUGAAUGUGAGAUAGCGCCGACUGACUAACAAUCUGGAAAGGGAAUCACCUUU--CUAAAAUUCAUUUUU--AAACU---AUCAAAAA      ::        ::        :            :    :  :                    :	108
mRNA (SMU1392c)	46	GAAAGUCUUGGUCCUUUAUG--ACAUGAA-CACGGUU-A--GAUU----AUCUGUAUCUUCAGA--GGAAAAAGA--AAAUAUAGAAAUCUUGGAUCAUAGUUUU      ::        ::        :            :    :  :	-47

r2.8 **SMU.874** bifunctional homocysteine S-methyltransferase/5,10-methylenetetrahydrofolate reductase protein

Score: -80 Pvalue: 0.00711096

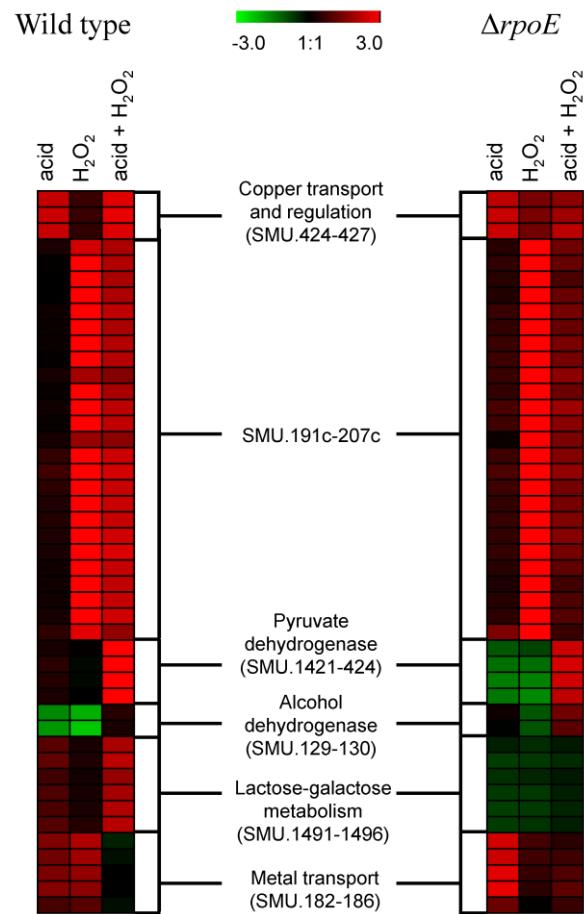
r2.9 **SMU.564** hypothetical protein

Score: -78 Pvalue: 0.00935811

sRNA	21	AAUUGAACUGUGA--GAUAG-C-GCCGACUGACUAACAAUCUGGUAAG----GAAAUACACAACCUUUCUAUUUCAA-UUAUU	95
		:   :	
mRNA (SMU.564)	-113	UUAACUUACAUUUGCACAAAAGACGGAGAACUUUUUAUCAGACCGAUACGAAAACUUUA-UGUCGUAGACAGU-AAAGUUCAGAA	-195

**S7. Predicted smu-r2 promoter and terminator (A), secondary structure (B), and interaction with target mRNAs (C).** A: The putative **-35** and **-10** regions were marked by green colour. The sequence of smu-r2 was underlined. The predicted transcription start was indicated by bold, while the putative terminator was marked by red colour.

Figure S8



**S8. Hierarchical clustering of differentially expressed genes under stresses in *S. mutans* wild type and  $\Delta rpoE$  mutant strains.**

Gene expression ratios are calculated as log2 fold changes under stresses comparing to the control (log-phase cells, before treatment). Red: up-regulated genes; green: down-regulated genes.

Table T1. Inhibition of growth of *S. mutans* wild type,  $\Delta rpoE$ , and  $\Delta rpoE$ comp strains by tetracycline and kanamycin.

A: Tetracycline ( $\mu\text{g/ml}$ )<sup>a</sup>

	1	0.8	0.6	0.4	0.2
WT	+	+	+	+	+
$\Delta rpoE$	-	-	-	+	+
$\Delta rpoE$ comp	+ <sup>c</sup>	+	+	+	+

B: Kanamycin ( $\mu\text{g/ml}$ )

	100	80	60	40	20
WT	+	+	+	+	+
$\Delta rpoE$	-	-	+	+	+
$\Delta rpoE$ comp	-	+ <sup>c</sup>	+	+	+

<sup>a</sup> +: Visible growth; -: no visible growth.

<sup>b</sup>  $\Delta rpoE$  mutant forms flocks at the presence of 0.4  $\mu\text{g/ml}$  tetracycline.

<sup>c</sup>  $\Delta rpoE$ comp strain showed weaker growth than the wild type strain.

Table T2. Differentially expressed non-coding regions in the  $\Delta rpoE$  mutant compared to the wild type strain under 5 experimental conditions, including normal growth at log-phase or early stationary-phase, growth under acid stress, H<sub>2</sub>O<sub>2</sub> stress, and acid+H<sub>2</sub>O<sub>2</sub>-stressed growth.

#	non-coding regions_start_stop / upstream gene-function / down-stream gene-function	Log2 fold change ( $\Delta rpoE$ / wild type)						
		log-phase	stationary-phase	acid-stress	H <sub>2</sub> O <sub>2</sub> -stress	Acid+H <sub>2</sub> O <sub>2</sub> -stress	Gene Dir type	Terminator start_stop
1.	NC_004350_530356_530591	1.52	1.65	1.18	1.13	1.37	<->	530364_530404
	SMU.566c-hypothetical protein	-0.30	-0.18	0.28	-0.51	-0.43		
	SMU.567-putative glutamine ABC transporter, permease	-0.50	-0.26	-0.48	-0.37	0.21		
2.	NC_004350_739911_740429	1.16	1.55	1.45	1.19	2.19	->->	739940_739955
	SMU.788-putative RNA methyltransferase	0.39	0.69	0.34	0.30	0.84		
	SMU.789-hypothetical protein	0.35	0.31	0.30	0.05	0.98		
3.	NC_004350_862809_862923	1.18	1.52	1.37	1.27	1.67	->->	862854_862874
	SMU.906-putative ABC transporter, ATP-binding protein	0.50	0.35	0.48	0.28	1.06		
	SMU.909-putative permease	0.40	0.54	0.39	0.36	0.36		
4.	NC_004350_1894335_1894524	1.25	1.58	1.82	1.17	1.66	<->	-
	SMU.2027-putative transcriptional regulator	0.22	0.50	1.28	0.14	-0.51		
	SMU.2028-levanasucrase precursor; beta-D-fructosyltransferase	0.25	0.12	-1.67	0.71	-1.12		
5.	NC_004350_1912245_1912417	1.93	1.16	1.69	1.13	1.68	<->	-
	SMU.2038-putative PTS system, trehalose-specific IIABC component	-0.23	-0.96	-0.65	-1.16	0.91		
	SMU.2040-putative transcriptional regulator; repressor of the trehalose operon	-0.03	-0.24	0.14	0.24	0.13		

6.	NC_004350_1919190_1919336	1.53	1.71	1.42	1.29	2.25	<-<-	-
	SMU.2044-putative stringent response protein, ppGpp synthetase	-0.49	-0.15	-0.14	0.03	-0.86		
	SMU.2046c-hypothetical protein	1.51	0.26	0.87	-0.30	1.23		
7.	NC_004350_368829_368962	1.83	2.31	2.35	1.51	2.31	<-->	-
	SMU.392c-hypothetical protein	0.71	1.18	1.03	0.73	0.71		
	SMU.393-hypothetical protein	-0.30	0.13	1.25	-0.57	0.92		
8.	NC_004350_381368_381675	1.03	2.46	2.48	1.54	3.31	<-->	-
	SMU.406c-hypothetical protein	-0.18	0.02	0.23	-0.29	0.36		
	SMU.407-hypothetical protein	0.22	0.13	0.15	0.54	0.71		
9.	NC_004350_401703_402053	1.34	1.48	1.32	1.29	1.64	<-->	-
	SMU.429c-hypothetical protein	-0.24	-0.04	-0.23	-0.43	-0.39		
	SMU.431-putative ABC transporter, ATP-binding protein	0.60	0.41	0.52	0.53	-0.07		

-: no terminator was found.

Table T3. Differentially expressed genes in the  $\Delta rpoE$  mutant compared to the wild type under log-phase or early stationary-phase growth conditions.

Locus Tag	Gene symbol	Gene description	Log2 fold change	
			$(\Delta rpoE / \text{wild type})$	
			Early stationary- Log-phase	phase
<b>Carbohydrate transport and metabolism, energy production and conversion</b>				
SMU.870	<i>fruR</i>	putative transcriptional regulator	-2.71	-0.70
SMU.871	<i>pfkB</i>	putative fructose-1-phosphate kinase	-2.70	-0.64
SMU.872	<i>fpxC</i>	putative fructose-PTS II ABC	-2.75	-0.51
SMU.877	<i>agaL</i>	alpha-galactosidase	3.60	0.96
SMU.878	<i>msmE</i>	multiple sugar-binding ABC transporter, sugar-binding protein	3.60	0.65
SMU.879	<i>msmF</i>	multiple sugar-binding ABC transporter, permease	3.25	0.36
SMU.880	<i>msmG</i>	multiple sugar-binding ABC transporter, permease	3.41	0.58
SMU.881	<i>gtfA</i>	sucrose phosphorylase	3.04	0.22
SMU.882	<i>msmK</i>	multiple sugar-binding ABC transporter, ATP-binding protein	3.00	0.21
SMU.883	<i>dexB</i>	dextran glucosidase DexB	3.10	0.23
SMU.885	<i>galR</i>	galactose operon repressor GalR	2.00	0.64
SMU.886	<i>galK</i>	galactokinase	3.86	0.37
SMU.887	<i>galT</i>	galactose-1-phosphate uridylyltransferase	3.76	0.48

SMU.888	<i>galE</i>	UDP-galactose 4-epimerase, GalE	1.72	0.88
SMU.113	<i>pfk</i>	putative fructose-1-phosphate kinase	0.59	-4.25
SMU.114	<i>fruC</i>	putative fructose-PTS IIBC	0.63	-4.17
SMU.115	<i>fruD</i>	putative fructose-PTS IIA	0.43	-3.05
SMU.116	<i>lacD2</i>	tagatose 1,6-diphosphate aldolase	0.79	-3.47
SMU.1956c	<i>levX</i>	hypothetical protein	1.49	2.91
SMU.1957	<i>levG</i>	putative fructose/mannose-PTS IID	1.34	2.71
SMU.1958c	<i>levF</i>	putative fructose/mannose-PTS IIC	1.18	2.55
SMU.1960c	<i>levE</i>	putative fructose/mannose-PTS IIB	1.58	3.13
SMU.1961c	<i>levD</i>	putative fructose/mannose-PTS IIA	1.50	3.17
SMU.78	<i>fruA</i>	fructan hydrolase	3.14	2.64
SMU.79	<i>fruB</i>	fructan hydrolase	2.46	2.56
SMU.1421	<i>pdhC</i>	branched-chain alpha-keto acid	1.18	-0.99
		dehydrogenase subunit E2		
SMU.1422	<i>pdhB</i>	putative pyruvate dehydrogenase E1	1.75	-1.29
		component beta subunit		
SMU.1423	<i>pdhA</i>	putative pyruvate dehydrogenase	1.78	-2.04
SMU.1424	<i>pdhD</i>	putative dihydrolipoamide dehydrogenase	1.70	-2.15
SMU.148	<i>adhE</i>	bifunctional acetaldehyde-CoA/alcohol	0.98	-1.35
		dehydrogenase		

#### Amino acid transport

SMU.1062	<i>opuAb</i>	putative proline/glycine betaine ABC transporter, permease protein	-1.55	-1.84
SMU.1063	<i>opuAa</i>	putative proline/glycine betaine ABC transporter, ATP-binding	-1.54	-1.93

SMU.2116	<i>opuCa</i>	putative osmoprotectant amino acid ABC transporter, ATP-binding	-1.14	-0.95
SMU.2117	<i>opuCb</i>	putative osmoprotectant ABC transporter; permease	-1.19	-1.02
SMU.2118	<i>opuCc</i>	putative osmoprotectant-binding protein	-1.17	-1.04
SMU.2119	<i>opuCd</i>	putative osmoprotectant ABC transporter; permease protein	-1.2	-1.09
<b>Iron transport</b>				
SMU.995	-	ferrichrome ABC transporter, permease	0.89	0.88
SMU.996	-	ferrichrome ABC transporter, permease	0.95	1.12
SMU.997	-	ferrichrome ABC transporter, ATP-binding	1.09	1.35
SMU.998	-	ferrichrome ABC transporter, periplasmic ferrichrome-binding	1.06	1.27

---

Table T4. Confirmation of differential gene expression by quantitative PCR in comparison to the microarray results.

Microarray Fold change ( $\Delta rpoE$ / wild type)						Real-time PCR Fold change ( $\Delta rpoE$ / wild type)					
	M0/W0	M1/W1	M2/W2	M3/W3	M4/W4	ajusted	M0/W0	M1/W1	M2/W2	M3/W3	M4/W4
						p value					
<i>levD</i>	2.83	8.99	0.95	0.33	0.65	1.25	$6.88 \pm 0.06$	$19.76 \pm 0.08$	$0.38 \pm 0.48$	$0.25 \pm 0.04$	$0.17 \pm 0.52$
						E-08					
<i>msmE</i>	12.15	1.57	0.44	0.99	1.44	1.35	$23.17 \pm 0.22$	$1.74 \pm 0.42$	$0.31 \pm 0.13$	$0.72 \pm 0.01$	$0.80 \pm 0.06$
						E-12					
<i>hisC</i>	0.25	0.21	0.31	0.48	0.21	6.07	$0.24 \pm 0.02$	$0.21 \pm 0.07$	$0.29 \pm 0.04$	$0.39 \pm 0.01$	$0.20 \pm 0.02$
						E-15					
<i>fruC</i>	1.55	0.06	1.47	1.28	0.99	1.92	$2.48 \pm 0.01$	$0.40 \pm 0.01$	$1.50 \pm 0.01$	$3.54 \pm 0.01$	$1.59 \pm 0.00$
						E-10					