

Supporting Information

Stevenson et al. 10.1073/pnas.1010390107

SI Materials and Methods

Hepatocyte Culture and TPO Measurement. Livers from WT or *Urah^{plt2/plt2}* mice aged 5 to 6 wk were perfused retrogradely via the inferior vena cava to isolate individual hepatocytes, as previously described (1). Then, 6×10^6 hepatocytes were cultured in RPMI containing 10% vol/vol bovine calf serum (HyClone) and 5×10^{-5} M 2-mercaptoethanol for 60 h at 37 °C in 10% (vol/vol) CO₂ in air before culture supernatant collection and concentration with a centrifugal filter unit (Amicon Ultra-15 10000 MWCO; Millipore). The number of hepatocytes in individual culture flasks was determined by measurement of the total genomic DNA content of each flask using a Quant-iT PicoGreen dsDNA Assay Kit (Invitrogen). TPO content of individual flasks was then corrected for the number of cultured hepatocytes as measured by the total dsDNA content of the cell ly-

sate. TPO concentration in serum, liver lysates, and cell culture supernatant was determined by a Quantikine murine TPO ELISA (R&D Systems).

Production of *Urah* Transgenic Mice. Transgenic mice were generated via injection of *C57BL/6* zygotes with a DNA construct in which the *Urah^{long}* cDNA was linked to the ubiquitin C promoter (2). DNA extracted from tail tips of founder mice was screened for the presence of the transgene by Southern blot analysis. Colonies were maintained by mating transgene-positive mice with *C57BL/6* partners. Transgenic rescue was performed by mating *Urah* transgenic mice with *Urah^{plt2/plt2}* mice and then backcrossing to additional *Urah^{plt2/plt2}* mice.

1. Brysha M, et al. (2001) Suppressor of cytokine signaling-1 attenuates the duration of interferon gamma signal transduction in vitro and in vivo. *J Biol Chem* 276: 22086–22089.

2. Schorpp M, et al. (1996) The human ubiquitin C promoter directs high ubiquitous expression of transgenes in mice. *Nucleic Acids Res* 24:1787–1788.

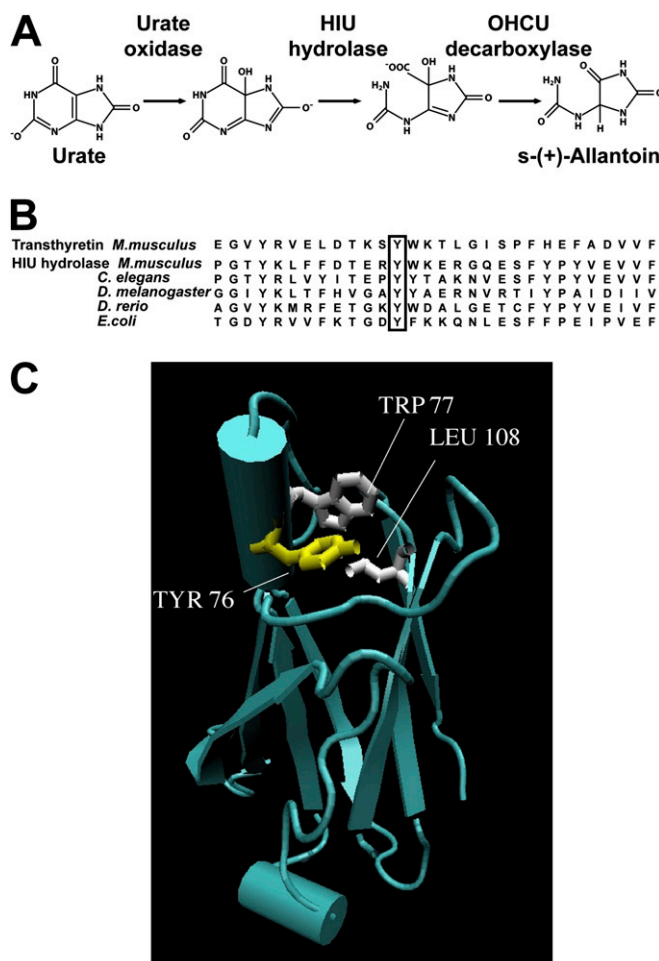


Fig. S1. HIU hydrolase structure and function. (A) HIU hydrolase catalyzes the second step of the three-step conversion of urate to allantoin in the mouse and most other nonhuman mammals. (B) Protein homology of HIU hydrolase in the region of the *plt2* mutation with predicted genes from other species (*Mus musculus*, *Caenorhabditis elegans*, *Drosophila melanogaster*, *Danio rerio*, and *Escherichia coli*) demonstrating that the Tyr substituted in *Urah^{plt2/plt2}* mice (boxed) has been conserved throughout evolution. (C) Structure of zebrafish HIU hydrolase (1) showing the tyrosine (TYR76 according to numbering as in ref. 1) substituted in *Urah^{plt2/plt2}* mice making strong hydrophobic contacts with conserved neighboring residues.

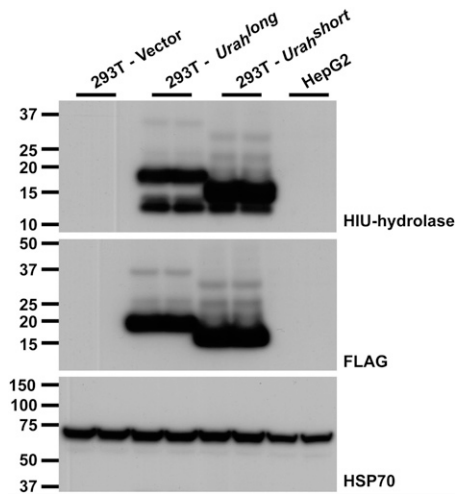


Fig. S2. Anti-HIU-hydrolase antiserum detects protein in cells transfected with murine *Urah* cDNA but not in human Hep-G2 cells. 293T cells were transiently transfected with vectors containing the short or long form of the murine *Urah* cDNA, including an N-terminal FLAG epitope tag and lysates examined by Western blot analysis with a rabbit anti-mouse HIU hydrolase polyclonal antiserum, anti-FLAG antibody, or control antibodies to HSP70. Sizes of molecular weight markers are shown in kilodaltons.

Table S1. Hematological and serum biochemical analysis in *Urah*^{plt2/plt2} mice

	WT	<i>Urah</i> ^{plt2/plt2}
Hematology		
Red cell count, 10 ¹² /L	10.3 ± 0.4	10.1 ± 0.4
White cell count, 10 ⁹ /L	9.0 ± 1.7	9.1 ± 2.1
Neutrophils	0.9 ± 0.3	0.9 ± 0.8
Lymphocytes	7.7 ± 1.6	7.6 ± 1.3
Monocytes	0.1 ± 0.1	0.1 ± 0.1
Eosinophils	0.2 ± 0.1	0.2 ± 0.1
Platelets, 10 ⁹ /L	1,424 ± 139	2,234 ± 838*
Serum biochemistry		
Total protein, g/L	46 ± 7	46 ± 6
Albumin, g/L	30 ± 2	30 ± 2
Bilirubin, mmol/L	3.1 ± 0.6	2.6 ± 0.5
Alanine aminotransferase, IU/L	28 ± 10	35 ± 9
Aspartate aminotransferase, IU/L	69 ± 34	67 ± 15
γ-Glutamyl transferase, IU/L	0.5 ± 0.8	0.3 ± 0.6

n = 8–11 mice for biochemical analyses and *n* = 18–24 mice for hematology.

**P* < 0.01.