

Supplementary Discussion

Structural determination of CusA

Data collection and refinement statistics are summarized in Tables S1 and S2. The resulting experimental electron density maps (Fig. S1) based on MIRAS reveal that the asymmetric unit consists of one protomer. The native crystal structure of CusA has been determined to a resolution of 3.52 Å (Table S1). Currently, 97.9% of the residues (residues 5-504 and 516-1040) are included in our final model. The final structure is refined to R_{work} and R_{free} of 23.7% and 27.9%, respectively. Superimposition of the structure of CusA with the structure of AcrB (pdb code:1IWG)⁵ results in a high RMSD of 11.4 Å for 1,003 C^α atoms, suggesting highly significant differences between these two transporters.

Reconstitution and stop-flow transport assay

Although all of our experimental results strongly suggest that these five methionine pairs/clusters (Fig. 4) are important for metal transport, we cannot definitively rule out the possibility that these mutations may affect the structure of the pump and in that way impair metal binding. Nonetheless, our collective experiments provide direct evidence that CusA is capable of taking up Ag⁺ from the cytoplasm. It should be noted that no decrease in fluorescence signal was detected in the absence of a pH gradient (i.e. no differential pH) between the inside and outside of the proteoliposomes, or if the intravesicular pH is higher than the extravesicular pH (i.e. reverse pH gradient) (Fig. S13).

Superimposition of the structures of CusA and AcrB

We superimposed a protomer of apo-CusA onto each protomer of the “asymmetric” AcrB (pdb code: 2DHH).⁷ Likewise, we also superimposed a protomer of CusA-Cu(I) onto those of the AcrB pump. These superimpositions gave high overall RMSDs, each RMSD exceeding 11 Å. However, when the portion of the apo-CusA protomer that contains only sub-domains PC1, PC2, PN1 and PN2 was superimposed onto those of the “extrusion”, “binding” and “access” conformers of AcrB (2DHH),⁷ we obtained much smaller RMSDs of 4.7 Å, 5.6 Å and 5.5 Å. Similarly, superimpositions of the periplasmic domain (only containing PC1, PC2, PN1 and PN2) of a protomer of CusA-Cu(I) onto those of the “extrusion”, “binding” and “access” conformers of AcrB (2DHH)⁷ gave RMSDs of 6.2 Å, 5.0 Å and 5.3 Å, respectively. These superimpositions suggest that the conformation of the apo-CusA protomer may correspond to the “extrusion” state, and that the CusA-Cu(I) protomer structure may represent the “binding” conformation of the pump. If this is the case, CusA may go through a cyclic conformational change, from the “access”, through to the “binding” and finally to the “extrusion” conformer, as suggested for the AcrB pump,⁷⁻⁹ to export copper/silver.

We also superimposed the Cu(I)-binding site of CusA onto the doxorubicin binding site of AcrB⁷. The superimposition suggests that the locations of the bound copper and drug molecule are within 3 Å apart (Fig. S14).

Supplemental Figures

Fig. S1. Stereo view of the experimental electron density map at a resolution of 3.8 Å.

(a) The electron density map contoured at 1.2 σ is in blue. Each subunit of CusA consists of 34 methionines. 33 selenium sites are found with a single SeMet crystal. The anomalous maps of the selenium sites contoured at 4 σ are in green. The C α traces of CusA are in red. (b) Anomalous maps of the 33 selenium sites (contoured at 4 σ). The selenium sites corresponding to the 11 methionines forming the methionine-residue relay network are in green. The rest of the 22 selenium sites are in purple. The C α traces of CusA are in red. (c) Representative section of the electron density in the second domain of CusA. The electron density (colored blue) is contoured at the 1.2 σ level and superimposed with the final refined model (yellow, carbon; red, oxygen; blue, nitrogen).

Fig. S2. Sequence and topology of CusA and AcrB. Alignment of the amino acid sequences of CusA and AcrB were done using CLUSTAL W. *, identical residues; :, >60% homologous residues. Secondary structural elements are indicated: TM, transmembrane helix; N α and N β , helix and strand, respectively, in the N-terminal half; C α and C β , helix and strand, respectively, in the C-terminal half. The CusC or TolC docking domain is divided into two sub-domains, DN and DC; whereas the pore domain is divided into four sub-domains, PN1, PN2, PC1 and PC2. The sequence and topology of CusA are shown at the top, and those for the AcrB pump are shown at the bottom.

Fig. S3. Experimental electron density of the periplasmic domain of CusA. (a) The electron density map of apo-CusA (3.8 Å-resolution) contoured at 1.2 σ is in blue. The

C α traces of the periplasmic domain of apo-CusA are in red. (b) The electron density map of CusA-Cu(I) (4.1 Å-resolution) contoured at 1.2 σ is in purple. The C α traces of the periplasmic domain of CusA-Cu(I) are in green. The bound copper is shown as a brown sphere.

Fig. S4. Ion pairs in the transmembrane domain viewed from the cytoplasmic side. Residues D405 of TM4, E939 of TM10 and K984 of TM11 that form ion pairs, which may play an important role in proton translocation, are in yellow sticks. The six methionines, M391, M403, M410, M486, M501 and M1009, are shown as pink sticks.

Fig. S5. Docking of CusB to CusA. (a) Side view of the docked complex of CusBA. The three CusB protomers are shown in green ribbons. The trimeric CusA is in gray surfaces. Sub-domains PN2 and PC1 of CusA are in red and blue, respectively. Specific interaction is found to occur between Domain 2 of CusB and the groove formed between DN and DC sub-domains of CusA to further stabilize the complex. (b) Top view of the docked complex of CusBA.

Fig. S6. Stereo view of the methionine-residue relay network. (a) The transmembrane domain of a subunit of CusA viewed from the cytoplasmic side. The six methionines that form three pairs in the transmembrane region are in green sticks. (b) The periplasmic domain of a subunit of CusA viewed from the periplasmic side. The five methionines that form a triad and a pair in the periplasmic domain are in green sticks.

Fig. S7. Alignment of amino acid sequences of the HME-RND-type Cu(I) and/or Ag(I) efflux pumps using CLUSTAL W. *, identical residues; :, >60% homologous residues. The alignment suggests that the methionine residues forming the relay network are conserved among these 30 different pumps. Six of these methionines located at the transmembrane region are highlighted with gray bars.

Fig. S8. Channel in the CusA pump. (a) Stereo view of the channel formed by the front protomer of apo-CusA (red) leading through the transmembrane and periplasmic domains is in gray color. This channel was calculated using the position of the sulfur atom of residue M672 as a starting point. The diameter of this channel ranges between 4.4 and 8.9 Å. The 11 methionines forming the relay network are in spheres (green, carbon; red, oxygen; blue, nitrogen; orange, sulfur). The distances between consecutive methionine pairs, from the cytoplasm to the periplasm, are 10.1, 12.1, 17.5 and 14.0 Å, respectively. Two other CusA protomers behind the front protomer are shown as blue wires. (b) Stereo view of the channel formed by the front protomer of Cu(I) bound-CusA (green) leading through the transmembrane and periplasmic domains is in gray color. This channel was calculated using the position of the sulfur atom of residue M672 as a starting point. The diameter of this channel ranges between 5.6 and 9.1 Å. The 11 methionines forming the relay network are in spheres (pink, carbon; red, oxygen; blue, nitrogen; orange, sulfur). The distances between consecutive methionine pairs, from the cytoplasm to the periplasm, are 9.4, 13.0, 20.0 and 14.7 Å, respectively. The bound Cu(I) is completely buried inside the channel. Two other CusA protomers behind the front protomer are shown as blue wires. The funnel formed by sub-domains DN and DC is

indicated with a dotted curve. For clarity, the channels formed by the other two protomers at the back are omitted. The calculations were done using the program CAVER (<http://loschmidt.chemi.muni.cz/caver>).

Fig. S9. A cartoon of a proteoliposome containing the CusA trimers. The CusA trimers are in green. The intravesicular space is loaded with the fluorescence indicator PGSK (blue star). Ag^+ (red sphere) is then added into the extravesicular medium for metal transport.

Fig. S10. The neighbor dependent opening closing motions. We show the effects of the strong internal motions on the CusA trimer. In the center is a diagram of the periplasmic domain looking downward, towards the inner membrane. We build an elastic network model using the formalism of Atilgan et al.³⁴ to predict the natural motions of CusA. On each of three sides we show the effect of two of the structure's natural motions. These motions describe coupled opening and closing of adjacent periplasmic metal entry sites. Black wedges are shown in open and closed form, indicating the approximate angle between the edges of PC1 and PC2 in each motion. From these the alternating open/close motion is evident.

Fig. S11. The swinging motion of PC2 coupled to TM8, as computed with the same elastic network model used for Fig. S10. This motion is similar to the difference between the apo and bound crystal forms displayed in Fig. 2. An arrow indicates the motion of

swinging from the red conformation to the green. For clarity PC2 and TM8 are shown in thicker lines than the remainder of the monomer.

Fig. 12. Expression level of the CusA pumps. An immunoblot against CusA of crude extracts from 50 µg dry cells of strain BL21(DE3) Δ *cueO* Δ *cusA* expressing the CusA pumps (M755I, lane 1; M391I, lane 2; M410I, lane 3; M486I, lane 4; wild-type, lane 5; marker, lane 6; M573I, lane 7; M623I, lane 8; M672I, lane 9) is shown.

Fig. S13. Stopped-flow transport assay of reconstituted wild-type CusA with extravesicular Ag⁺ ion at different intravesicular and extravesicular pHs. The stopped-flow traces are the cumulative average of four successive recordings (intravesicular pH = 6.6 and extravesicular pH = 7.0, black curve; intravesicular pH = extravesicular pH = 6.6, blue curve; intravesicular pH = extravesicular pH = 7.0, green curve; intravesicular pH = 7.0 and extravesicular pH = 6.6, pink curve; control liposome, red curve).

Fig. S14. Comparison of the CusA and AcrB binding sites. Superimposition of a subunit of Cu(I) bound-CusA (green) onto the “binding” protomer of AcrB (orange) with doxorubicin (pdb code: 2DH6).⁷ In this superimposition, the bound copper and doxorubicin are 3 Å apart. The bound Cu(I) and doxorubicin are colored red and blue, respectively.

Table S1. Data collection and refinement statistics of apo-CusA.

	Native CusA	Au(III)	Ta ₆ Br ₁₂ ²⁺	Se (peak)
Data collection				
Space group	<i>R</i> 32	<i>R</i> 32	<i>R</i> 32	<i>R</i> 32
Cell dimensions				
<i>a</i> , <i>b</i> , <i>c</i> (Å)	178.42,	179.31,	178.34,	178.21,
	178.42,	179.31,	178.34,	178.21,
	285.75	287.10	284.23	285.57
α , β , γ (°)	90, 90, 120	90, 90, 120	90, 90, 120	90, 90, 120
Wavelength (Å)	0.9791	1.0398	1.0089	0.9791
Resolution (Å)	50-3.52	50-4.31	50-3.52	50-3.68
	(3.62-3.52)	(4.50-4.31)	(3.66-5.52)	(3.81-3.68)
<i>R</i> _{sym} or <i>R</i> _{merge}	5.7 (43.5)	10.2 (35.9)	6.2 (37.7)	7.5 (46.9)
<i>I</i> / σ <i>I</i>	32.5 (4.0)	17.3 (5.2)	21.7 (3.4)	19.7 (2.6)
Completeness (%)	94.2 (95.8)	100 (100)	99.9 (100)	100 (100)
Redundancy	6.9 (6.9)	3.2 (2.8)	3.8 (3.9)	3.9 (3.9)
Refinement				
Resolution (Å)	50-3.52			
No. reflections	432,346			
<i>R</i> _{work} / <i>R</i> _{free}	0.238/0.275			
No. atoms				
Protein	7,884			
Ligand/ion	0			
Water	0			
R.m.s deviations				
Bond lengths (Å)	0.004			
Bond angles (°)	0.874			

*Highest resolution shell is shown in parenthesis.

Table S2. Data collection and refinement statistics of the CusA-Cu(I) and CusA-Ag(I) complexes.

	Cu(I) (peak)	Cu(I)	Ag(I)
Data collection			
Space group	<i>R</i> 32	<i>R</i> 32	<i>R</i> 32
Cell dimensions			
<i>a</i> , <i>b</i> , <i>c</i> (Å)	178.21, 178.21, 285.57	179.14, 179.14, 286.12	179.99, 179.99, 287.98
α , β , γ (°)	90, 90, 120	90, 90, 120	90, 90, 120
Wavelength (Å)	1.3779	1.0333	1.3779
Resolution (Å)	50-4.10 (4.25-4.10)	50-3.88 (4.03-3.88)	50-4.35 (4.51-4.35)
<i>R</i> _{sym} or <i>R</i> _{merge}	8.9 (42.4)	9.0 (42.6)	9.2 (39.6)
<i>I</i> / σ <i>I</i>	11.8 (1.8)	10.5 (2.1)	14.9 (2.0)
Completeness (%)	96.6 (98.7)	99.5 (98.8)	99.7 (99.9)
Redundancy	2.5 (2.5)	3.2 (3.1)	3.3 (3.4)
Refinement			
Resolution (Å)		50-3.88	50-4.35
No. reflections		346,057	258,908
<i>R</i> _{work} / <i>R</i> _{free}		0.261/0.296	0.271/0.312
No. atoms			
Protein		7,802	7,782
Ligand/ion		1	1
Water		0	0
R.m.s deviations			
Bond lengths (Å)		0.005	0.006
Bond angles (°)		0.995	1.252

*Highest resolution shell is shown in parenthesis.

Table S3. MICs of copper for different CusA mutants expressed in *E. coli* BL21(DE3) Δ *cueO* Δ *cusA*

Gene in BL21(DE3) Δ <i>cueO</i> Δ <i>cusA</i>	MIC (mM) of CuSO ₄
Empty vector	0.50
<i>cusA</i> (wild-type)	2.25
<i>cusA</i> (M573I)	0.50
<i>cusA</i> (M623I)	0.50
<i>cusA</i> (M672I)	0.50
<i>cusA</i> (M391I)	1.25
<i>cusA</i> (M410I)	1.75
<i>cusA</i> (M486I)	1.75
<i>cusA</i> (M755I)	1.75
<i>cusA</i> (D405A)	0.50
<i>cusA</i> (E939A)	0.50
<i>cusA</i> (K984A)	0.50
<i>cusA</i> (E412A)	1.00 ^a
<i>cusA</i> (S453A)	1.00 ^a
<i>cusA</i> (C353A)	2.25 ^b
<i>cusA</i> (C375A)	2.25 ^b

^aResidues E412 and S453, located in the vicinity of the triad formed by D405, E939 and K984, may also involve in the proton-relay network.

^bThe CusA pump contains only two cysteine residues, C353 and C375, which may not involve in copper resistance.

Table S4. MICs of silver for different CusA mutants expressed in *E. coli* BL21(DE3) Δ *cueO* Δ *cusA*

Gene in BL21(DE3) Δ <i>cueO</i> Δ <i>cusA</i>	MIC (μ M) of AgNO ₃
Empty vector	10.0
<i>cusA</i> (wild-type)	30.0
<i>cusA</i> (M573I)	12.5
<i>cusA</i> (M623I)	12.5
<i>cusA</i> (M672I)	12.5
<i>cusA</i> (M391I)	10.0
<i>cusA</i> (M410I)	17.5
<i>cusA</i> (M486I)	17.5
<i>cusA</i> (M755I)	12.5
<i>cusA</i> (D405A)	12.5
<i>cusA</i> (E939A)	12.5
<i>cusA</i> (K984A)	12.5
<i>cusA</i> (E412A)	12.5 ^a
<i>cusA</i> (S453A)	15.0 ^a

^aResidues E412 and S453, located in the vicinity of the triad formed by D405, E939 and K984, may also involve in the proton-relay network.

Fig. S1a

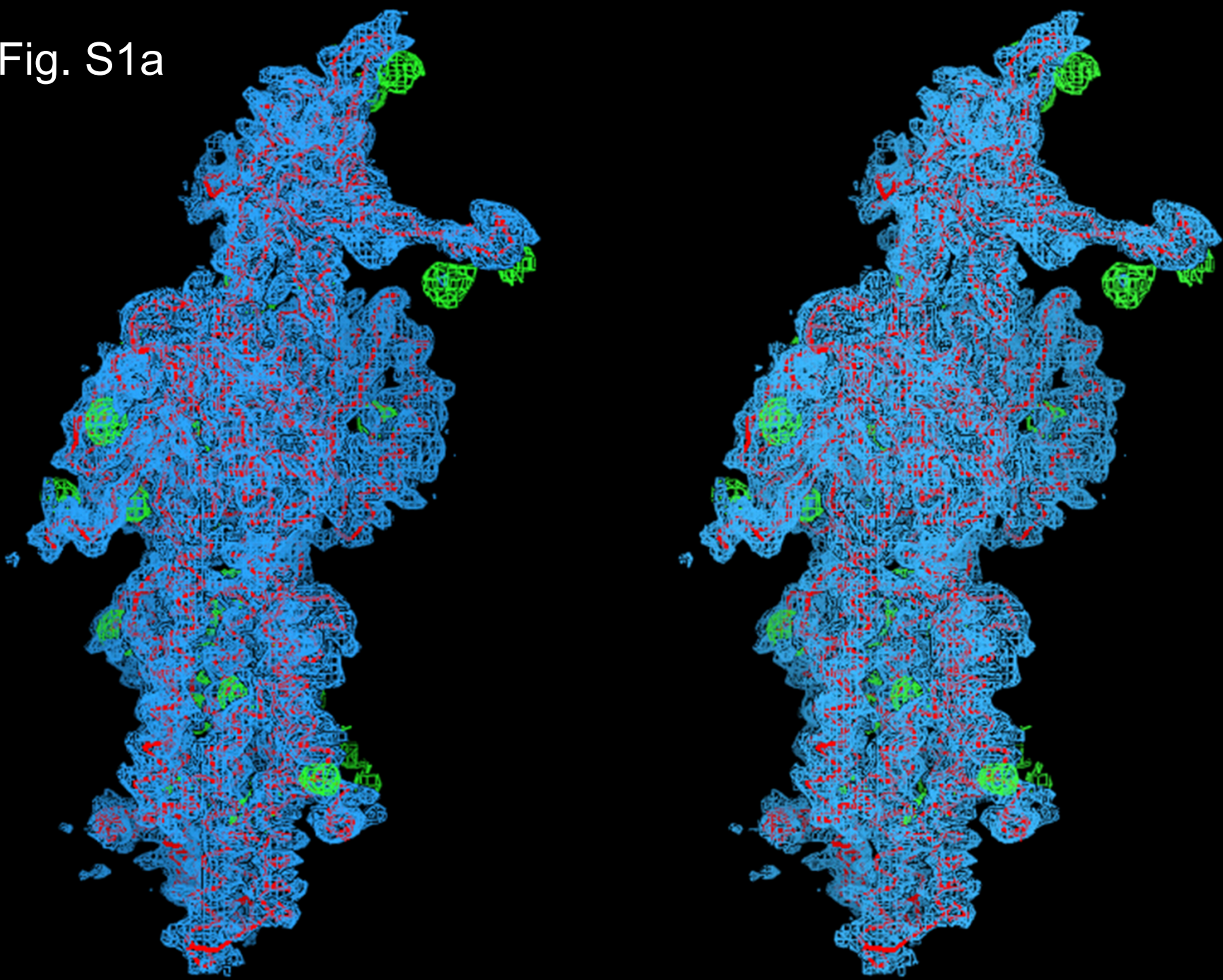


Fig. S1b

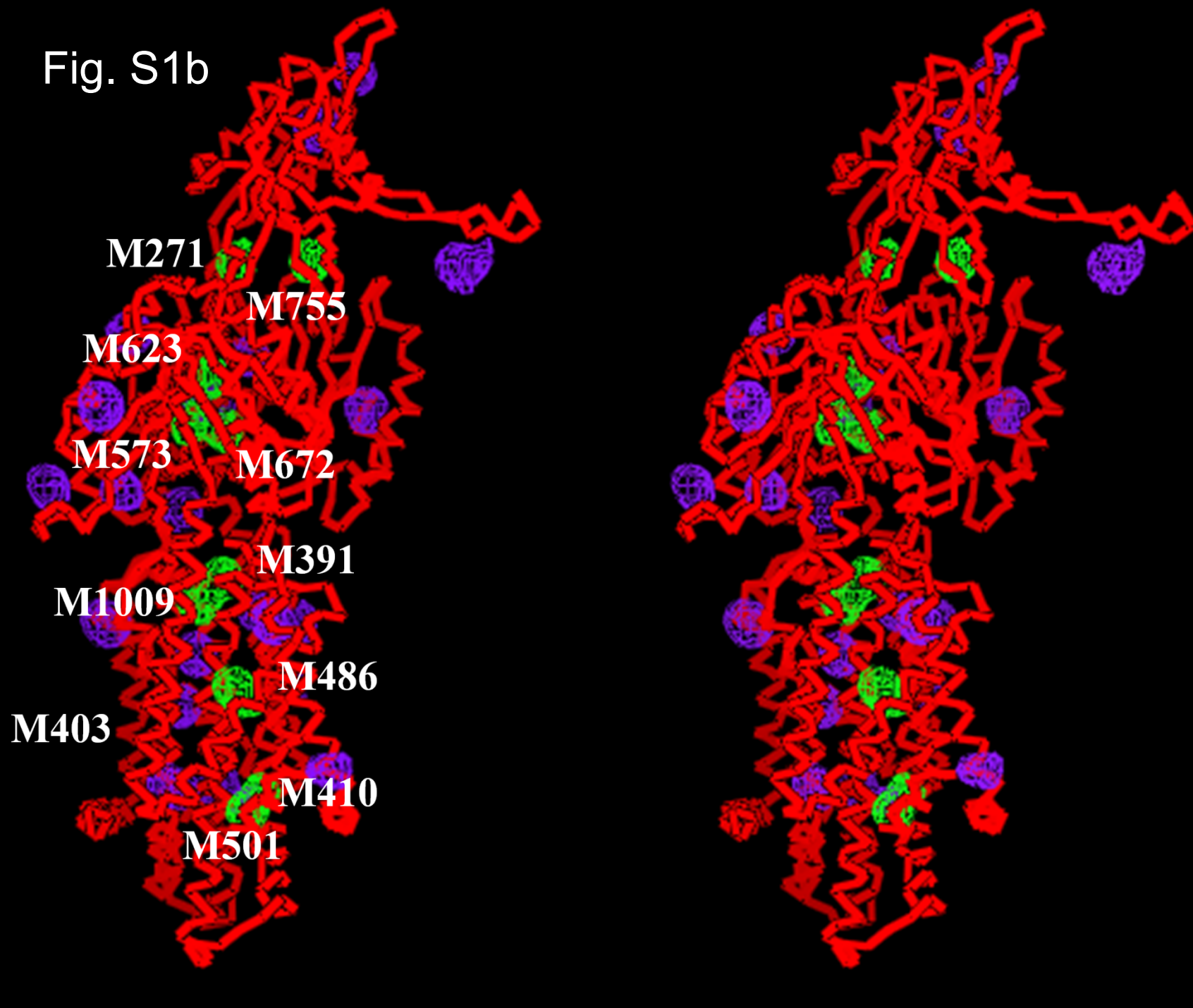


Fig. S1c

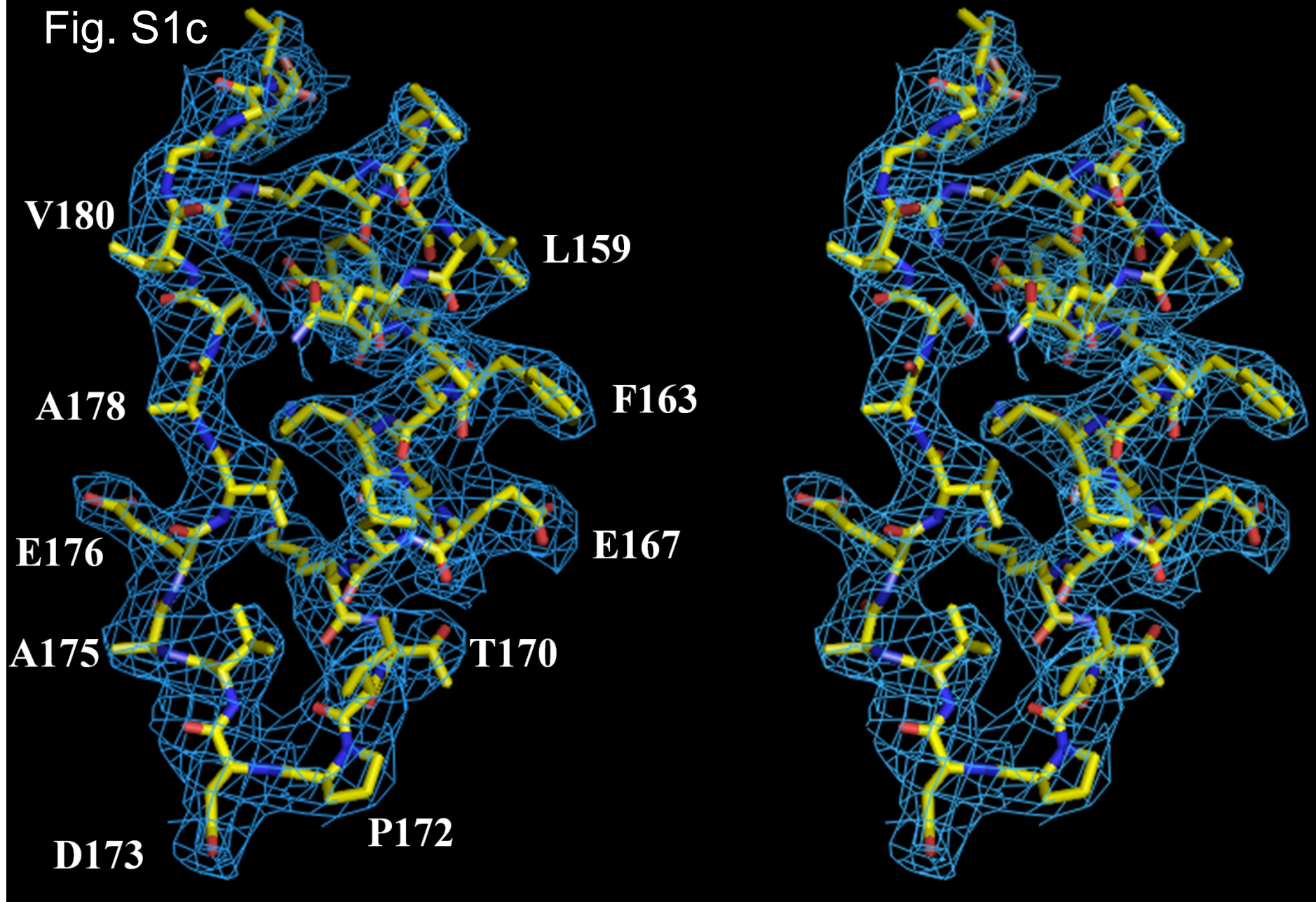


Fig. S2

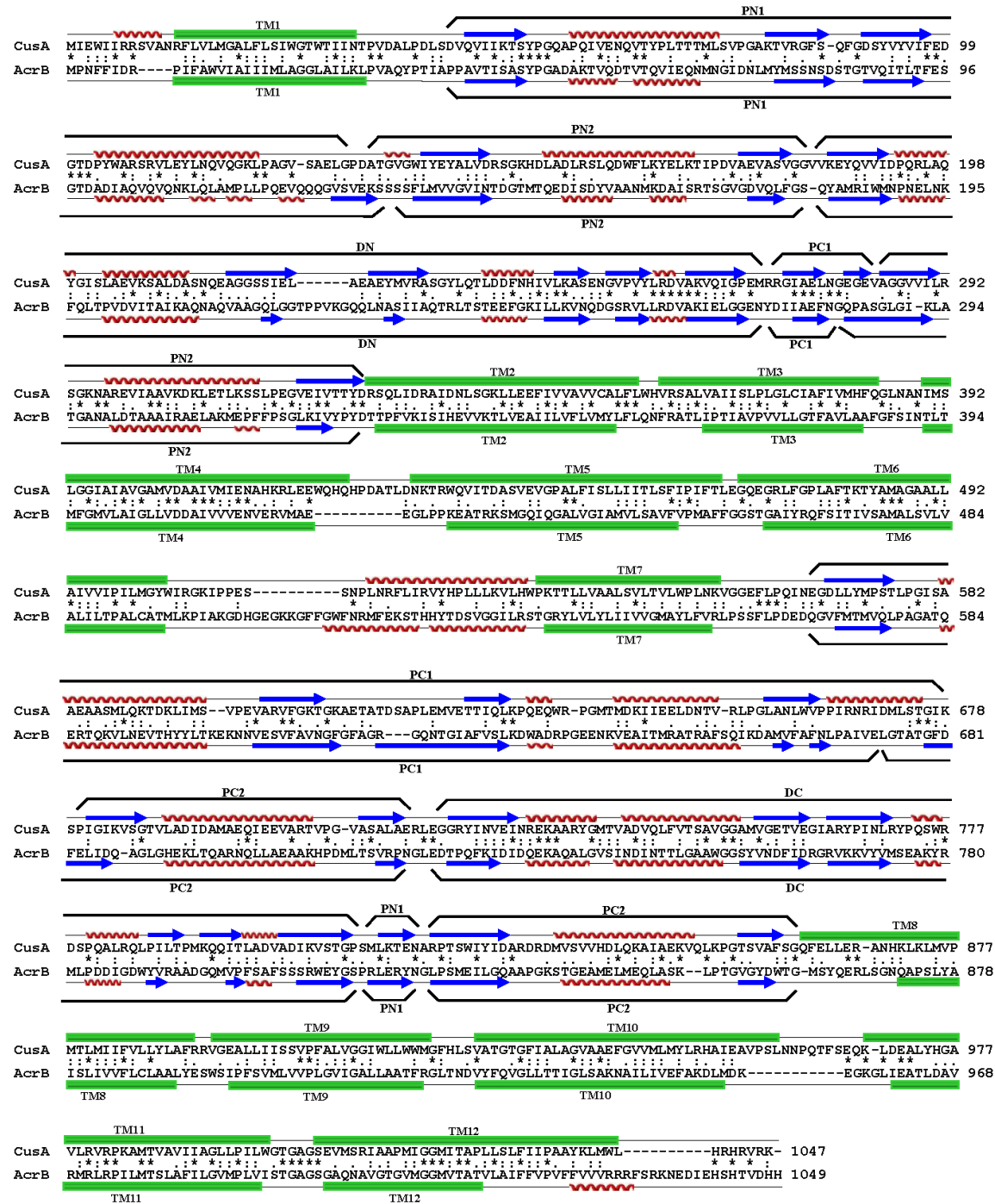


Fig. S3

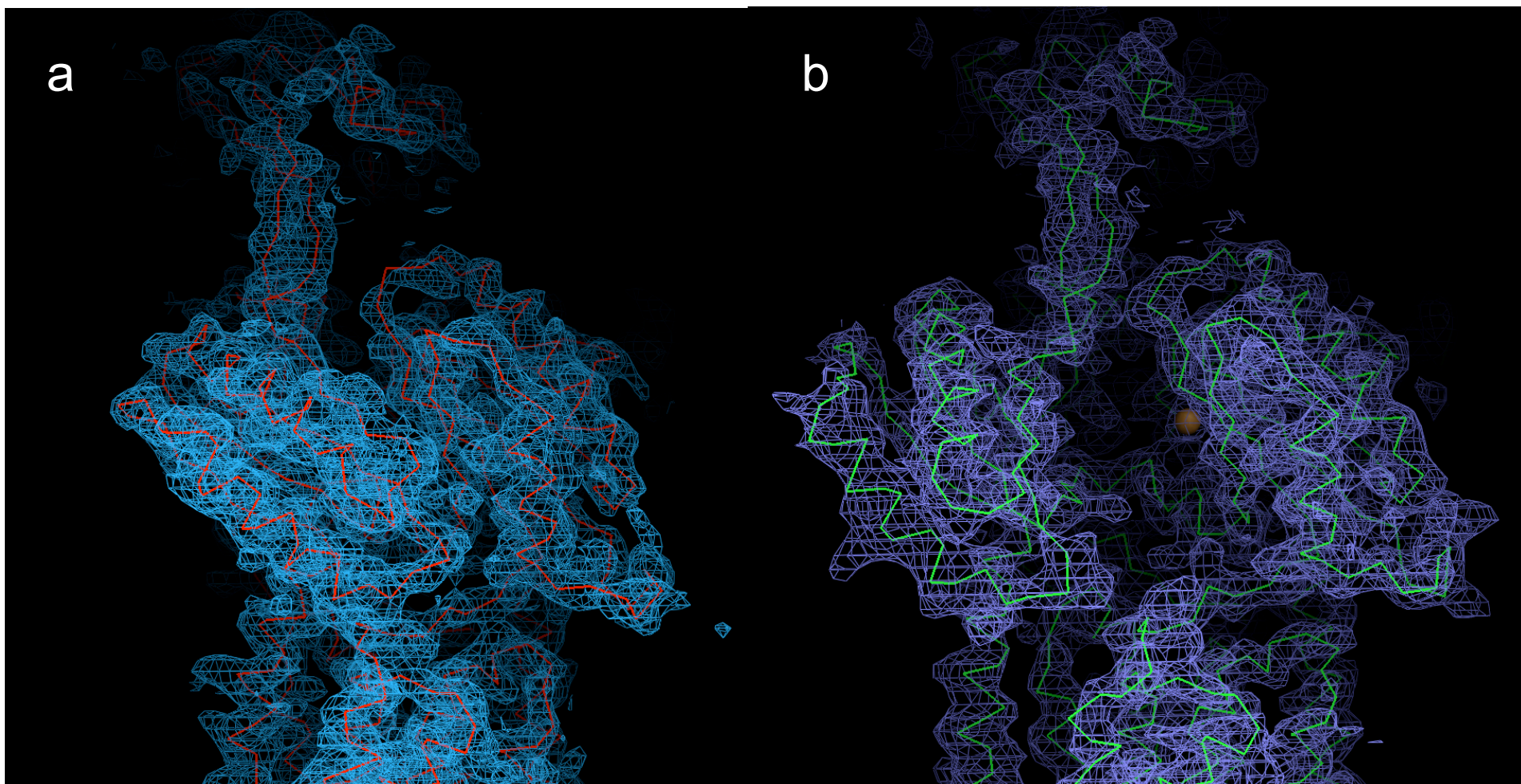


Fig. S4

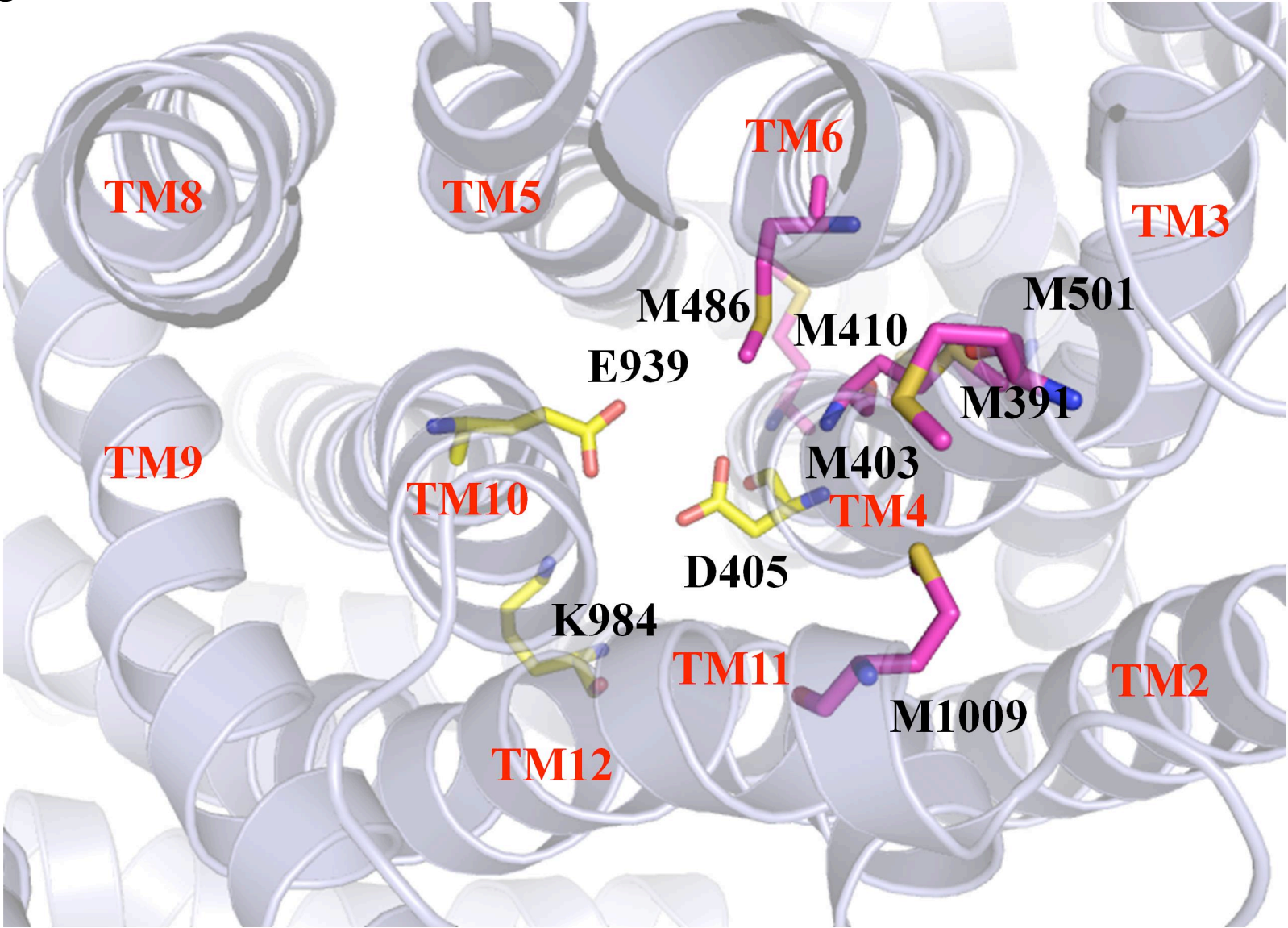


Fig. S5

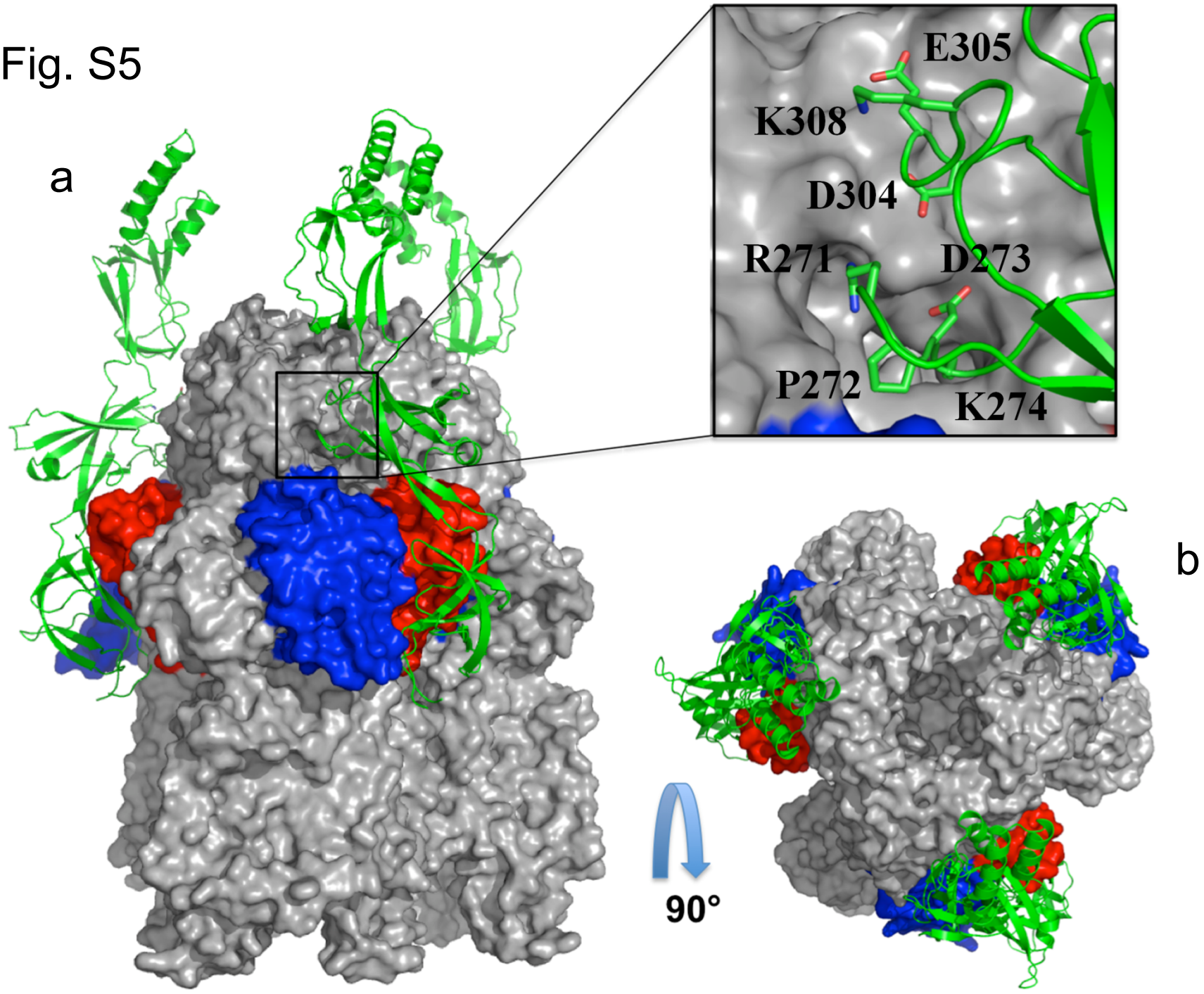


Fig. S6a

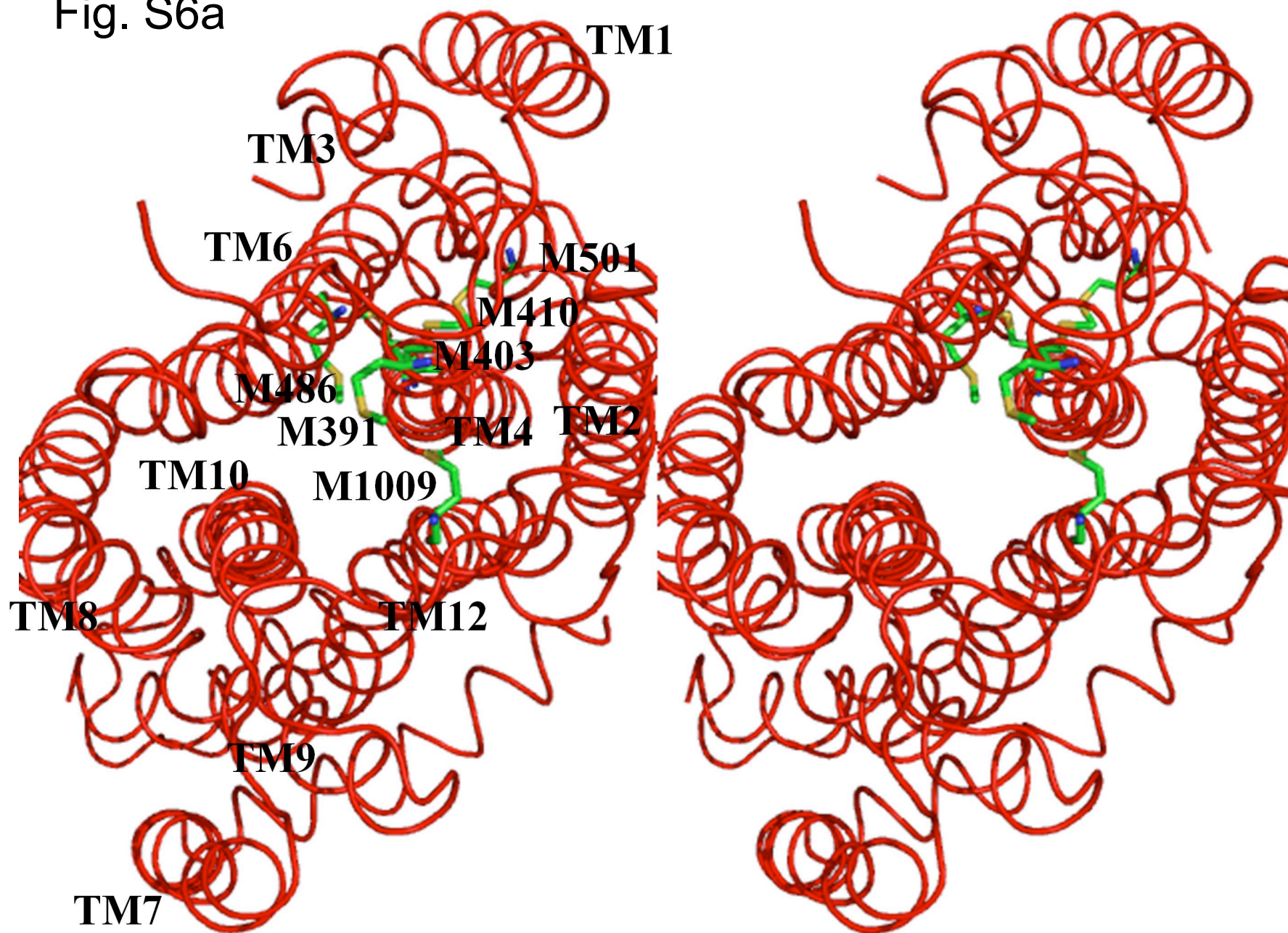
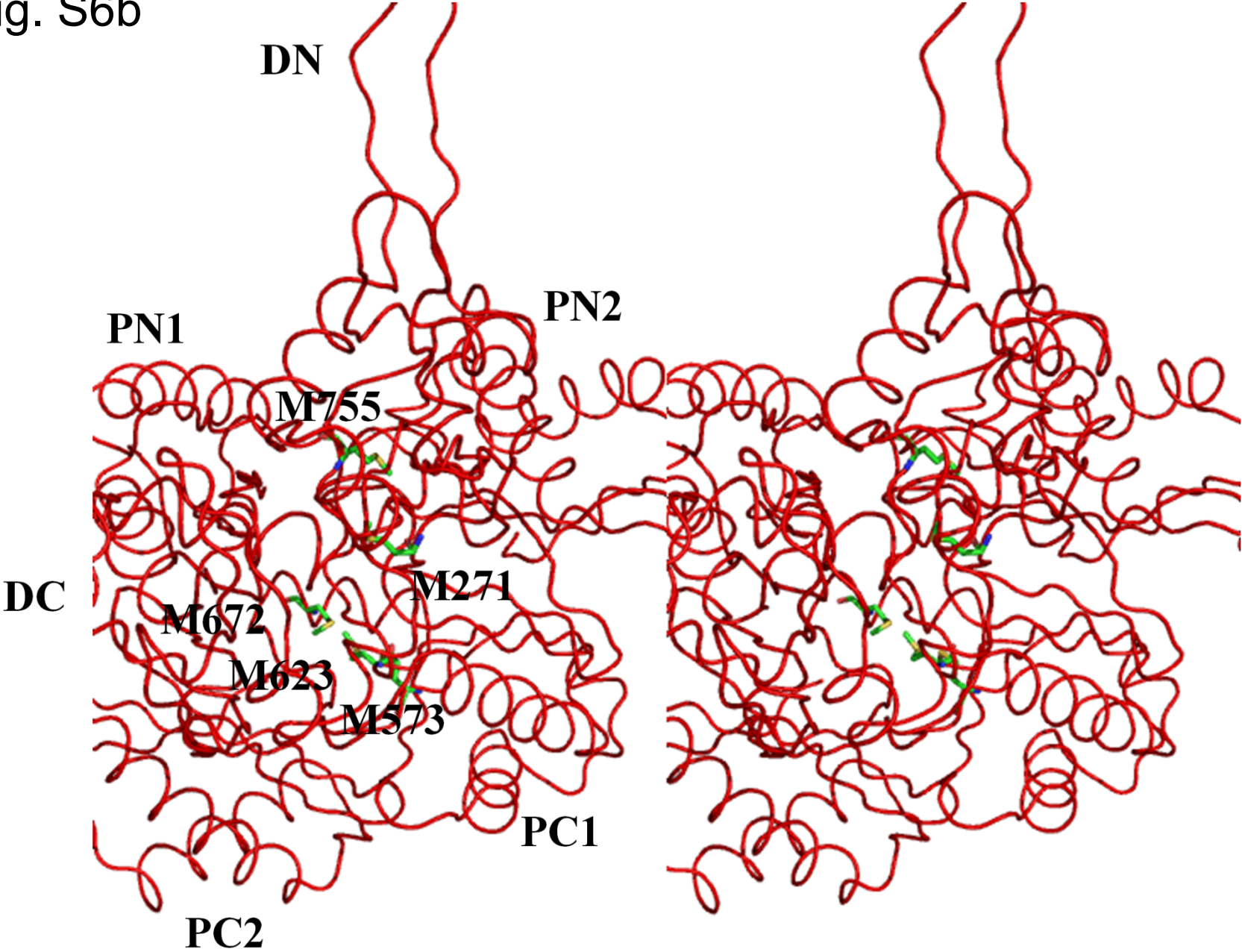


Fig. S6b



CusA_Escherichia coli K12
Putative Shigella sonnei
CzCA_Shevanella sp. ANA-3
SilA_Klebsiella pneumoniae
Putative Serratia marcescens
SilA_Salmonella Typhimurium
CzCA_Erythrobacter sp. NAP1
CzCA_Sphingopyxis alaskensis RB2256
CzCA_Caulobacter sp. K31
CzCA_Phenylobacterium zucineum HLK1
CzCA_Asticcacaulis excentricus CB 48
CzCA_Pseudomonas fluorescens Pf0-1
CzCA_Thiobacillus denitrificans
CzCA_Stenotrophomonas sp. SKA14
CzCA_Acidovorax sp. JS42
CzCA_Leptothrix cholodnii SP-6
CusA_curvibacter
CzCA_Ralstonia pickettii 12J
SilA_Ralstonia metallidurans CH34
Putative Methylibium petroleiphilum PM1
CzCA_Delftia acidovorans SPH-1
CzCA_Yersinia kristensenii
CusA_Klebsiella pneumoniae
CzCA_Polaromonas naphthalenivorans CJ2
CusA_Ewardsiella ictaluri
CusA_Enhydrobacter aerosaccus SK60
CusA_Citrobacter sp. 30-2
CzCA_Acinetobacter junii SH205
Putative Pectobacterium carotovorum
CusA_Providencia rettgeri DSM1131

CusA_Escherichia coli K12
Putative Shigella sonnei
CzCA_Shevanella sp. ANA-3
SilA_Klebsiella pneumoniae
Putative Serratia marcescens
SilA_Salmonella Typhimurium
CzCA_Erythrobacter sp. NAP1
CzCA_Sphingopyxis alaskensis RB2256
CzCA_Caulobacter sp. K31
CzCA_Phenylobacterium zucineum HLK1
CzCA_Asticcacaulis excentricus CB 48
CzCA_Pseudomonas fluorescens Pf0-1
CzCA_Thiobacillus denitrificans
CzCA_Stenotrophomonas sp. SKA14
CzCA_Acidovorax sp. JS42
CzCA_Leptothrix cholodnii SP-6
CusA_curvibacter
CzCA_Ralstonia pickettii 12J
SilA_Ralstonia metallidurans CH34
Putative Methylibium petroleiphilum PM1
CzCA_Delftia acidovorans SPH-1
CzCA_Yersinia kristensenii
CusA_Klebsiella pneumoniae
CzCA_Polaromonas naphthalenivorans CJ2
CusA_Ewardsiella ictaluri
CusA_Enhydrobacter aerosaccus SK60
CusA_Citrobacter sp. 30-2
CzCA_Acinetobacter junii SH205
Putative Pectobacterium carotovorum
CusA_Providencia rettgeri DSM1131

CusA_Escherichia coli K12
Putative Shigella sonnei
CzCA_Shevanella sp. ANA-3
SilA_Klebsiella pneumoniae
Putative Serratia marcescens
SilA_Salmonella Typhimurium
CzCA_Erythrobacter sp. NAP1
CzCA_Sphingopyxis alaskensis RB2256
CzCA_Caulobacter sp. K31
CzCA_Phenylobacterium zucineum HLK1
CzCA_Asticcacaulis excentricus CB 48
CzCA_Pseudomonas fluorescens Pf0-1
CzCA_Thiobacillus denitrificans
CzCA_Stenotrophomonas sp. SKA14

CzcA_Stenotrophomonas sp. SKA14 AGVAEEFGVVMILYKQALAEPCDR----REPTREELLDAREGAVLRVRPKAMTVAVILAGLVPIVWSSGTGSEVMSR
CzcA_Acidovorax sp. JS42 AGVAEEFGVVMILYKQALAEPCDG----REPTREELLDAREGAVLRVRPKAMTVAVILAGLVPIVWSSGTGSEVMSR
CzcA_Leptothrix cholodnii SP-6 AGVAEEFGVVMILYKHALQDRLAAG----ASPSAALVDDAREGAVLRVRPKAMTVAVILAGLVPIVWSSGTGSEVMSR
Cusa_curvibacter AGVAEEFGVVMILYKQALADRESQG----AAPGVAMVLDAREGAVLRVRPKAMTVAVILAGLVPIVWSSGTGSEVMSR
CzcA_Ralstonia pickettii 12J AGVAEEFGVIMLLYLKHAHWTERQENG----QTSTQALLEAIQEGAVLRVRPKAMTVAVILAGLPIVWSSGTGSEVMQR
Sila_Ralstonia metallidurans CH34 AGVAEEFGVIMLLYLKHAHWDRLRG----EDTLDALLDAREGAVLRVRPKAMTVAVILAGLPIVWSSGTGSEVMQR
Putative Methylibium petroleiphilum PM1 AGVAEEFGVIMLLYLKQAWEARLARG----LSSDADLMDAREGAVLRVRPKAMTVAVILAGLVPIVWSSGTGSEVMSR
CzcA_Delftia acidovorans SPH-1 AGVSAEFGVIMLLYLKNAWQERADQG----KTSEADLLDAREGAVLRVRPKAMTVAVILAGLPIVWSSGTGSEVMQR
CzcA_Yersinia kristensenii AGVAEEFGVIMVLYLNQAVKHKHRPQ----IAMTASEMSAAIHGAVLRVRPKAMTVAVILAGLPIVWSSGTGSEVMQR
Cusa_Klebsiella pneumoniae AGVAEEFGVVMMLYLKHAIEAEPSLKN--PQTFSDVKLDEALYQGAVLRVRPKAMTVAVILAGLPIVWSSGTGSEVMSR
CzcA_Polaromonas naphthalenivorans CJ2 AGVSAEFGVIMLLYLRLHAWD--ERLAQGGKTN---AEDLLDAREGAVLRVRPKAMTVAVILAGLPIVWSSGTGSEVMQR
Cusa_Edwardsiella ictaluri AGVAEEFGVVMMLYLKQAIIE--QDPALSHTRGFTPAALDEALYHGAVLRVRPKAMTVAVILAGLPIVWSSGTGSEVMSR
Cusa_Enhydrobacter aerosaccus SK60 AGVAEEFGVVMFLYLRKQAIIEHAQQSLSSSTASLTEQQLNEAHTGAVLRVRPKAMTVAVILAGLPIVWSSGTGSEVMSR
CzcA_Citrobacter sp. 30-2 AGVAEEFGVVMMLYLKHAIE--AEPALEN--PQFTADKLEALYHGAVLRVRPKAMTVAVILAGLPIVWSSGTGSEVMSR
CzcA_Acinetobacter junii SH205 AGVAEEFGVVMFLYLRKQAVEHAQQSLSSSTASLTEQQLNDAIRTGAVLRVRPKAMTVAVILAGLPIVWSSGTGSEVMSR
Putative Pectobacterium carotovorum AGVSAEFGVIMLLYLRLHAWKRR--V--AGQALSQQQLMDAREGAVLRVRPKAMTVAVILAGLPIVWSSGTGSEVMSR
Cusa_Providencia rettgeri DSM1131 AGVAEEFGVVMMLYLKHAIEDKKEKQGG--ISKLSNQLDRALYEGAVLRVRPKAMTVAVILAGLPIVWSSGTGSEVMSR
:*:*.**: : : * : * : **** * : * : * : * : *

Cusa_Escherichia coli K12 IAAPMIGGMITAPLLSLFIIIPAAAYKLMWLHR----HRVRK----- 1047
Putative Shigella sonnei IAAPMIGGMITAPLLSLFIIIPAAAYKLMWLHR----HRVRK----- 1047
CzcA_Shevanella sp. ANA-3 IAAPMIGGMITAPLLSLFIIIPAAAYKLIWLRR---HKKSVS----- 1048
Sila_Klebsiella pneumoniae IAAPMIGGMITAPLLSLFIIIPAAAYKLIWLRR---HKKSVS----- 1048
Putative Serratia marcescens IAAPMIGGMITAPLLSLFIIIPAAAYKLIWLRR---HKKSVS----- 1048
Sila_Salmonella Typhimurium IAAPMIGGMITAPLLSLFIIIPAAAYKLIWLRR---HKKSVS----- 1048
CzcA_Erythrobacter sp. NAP1 IAAPMIGGMITAPLLSLFIIIPAAAYLMLRRR---PERPQPQGEEECVQPS--- 1055
CzcA_Sphingopyxis alaskensis RB2256 IAAPMVGGMITAPLLSLFVIPAAYLLMRQRK---AKPNPQPD-----QP--- 1047
CzcA_Caulobacter sp. K31 IAAPMIGGMLTAPLLSLMFVVPAAAYLLIERR---LSRPRGPT-----IAHG--- 1049
CzcA_Phenylobacterium sucineum HLK1 IAAPMIGGMLTAPLLSLMFVVPAGYLLLR---PSRTSILK-----GDQP--- 1049
CzcA_Asticcacaulis excentricus CB 48 IAAPMIGGMLTAPLLSLMVLVPAAYLLIERR---RSNPPQSN-----SQQ--- 1048
CzcA_Pseudomonas fluorescens Pf0-1 IAAPMVGGMITAPLLSLFVIPAAYRLMR---LPAENKSPQSDDASTIQP--- 1056
CzcA_Thiobacillus denitrificans IAAPMVGGMITAPLLSLMFVIPAAYLMLRRR---PQPHAPHPHHSSEGHP--- 1060
CzcA_Stenotrophomonas sp. SKA14 IAAPMIGGMITAPLLSLFVIPAAYLLMRKFR---PQPHAPHPHHSSEGHP--- 1039
CzcA_Acidovorax sp. JS42 IAAPMIGGMITAPLLSLFVIPAAYLLMRKFR---PQPHAPHPHHSSEGHP--- 1039
CzcA_Leptothrix cholodnii SP-6 IAAPMIGGMITAPLLSLFVIPAAYLLMRKFR---PQPHAPHPHHSSEGHP--- 1066
Cusa_curvibacter IAAPMIGGMITAPLLSLFIIIPAAAYRMMRVRT---SRRLN----- 1044
CzcA_Ralstonia pickettii 12J IAAPMIGGMITAPLLSLFVIPAAYLLMRKFR---PQPHAPHPHHSSEGHP--- 1055
Sila_Ralstonia metallidurans CH34 IAAPMVGGMITAPLLSLMFVVPAAAYLLLR---LPSSTTAKQAELEAV--- 1056
Putative Methylibium petroleiphilum PM1 IAAPMVGGMITAPLLSLMFVIPAAYRLLRRRRASAGEAAAAPGKSSIPTPAPSAA 1063
CzcA_Delftia acidovorans SPH-1 IAAPMVGGMITAPLLSLMFVIPAAYLLMRKFR---ERKASPMWFRHRAAAA--- 1056
CzcA_Yersinia kristensenii IAAPMIGGMITAPLLSLFIIIPAAAYLLMRKFR---ERKASPMWFRHRAAAA--- 1034
Cusa_Klebsiella pneumoniae IAAPMIGGMITAPLLSLFIIIPAAAYKLMWLSR---HRGKRSE----- 1049
CzcA_Polaromonas naphthalenivorans CJ2 IAAPMVGGMITAPLLSLMFVIPAAYLLMRKFR---ERKASPMWFRHRAAAA--- 1055
Cusa_Edwardsiella ictaluri IAAPMIGGMITAPLLSLFIIIPAAAYKLIWLHRQRATR---RTDSSA----- 1054
Cusa_Enhydrobacter aerosaccus SK60 IAAPMVGGMITAPLLSLMFVIPAAYQLLIKRLKSKN----- 1048
Cusa_Citrobacter sp. 30-2 IAAPMIGGMITAPLLSLFIIIPAAAYKLMWLRHRGQK----- 1047
CzcA_Acinetobacter junii SH205 IAAPMVGGMITAPLLSLMFVIPAAYQLLIKRLKSKN----- 1048
Putative Pectobacterium carotovorum IAAPMIGGMITAPLLSLMFVIPAAYLLMRKFR---ERKASPMWFRHRAAAA--- 1045
Cusa_Providencia rettgeri DSM1131 IAAPMIGGMITAPLLSLFIIIPAAAYKLIWLHRQRATR---RTDSSA----- 1045
** ** : * : * : * : * : * : * : * : * : * : * : * : * : *

Fig. S8a

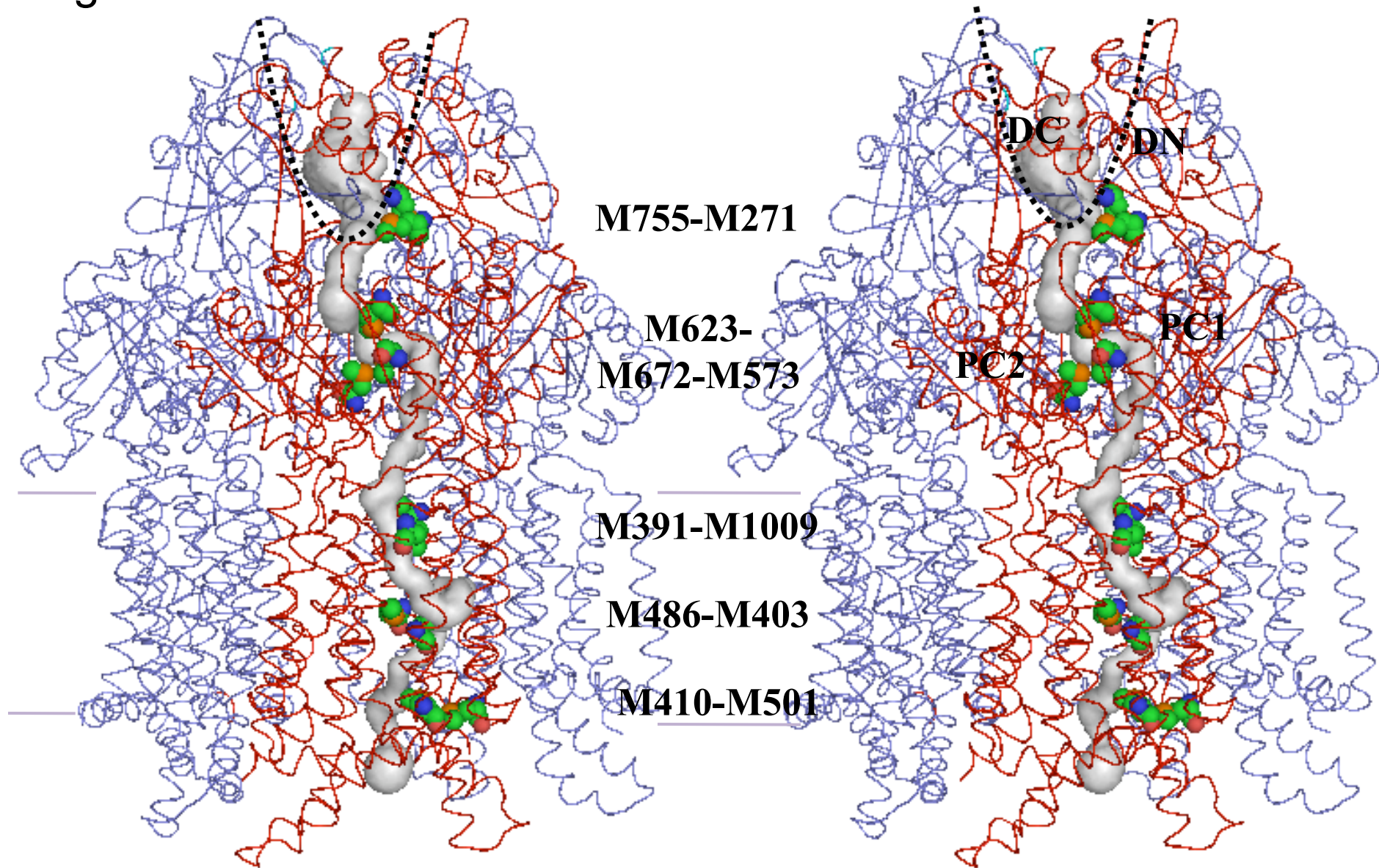


Fig. S8b

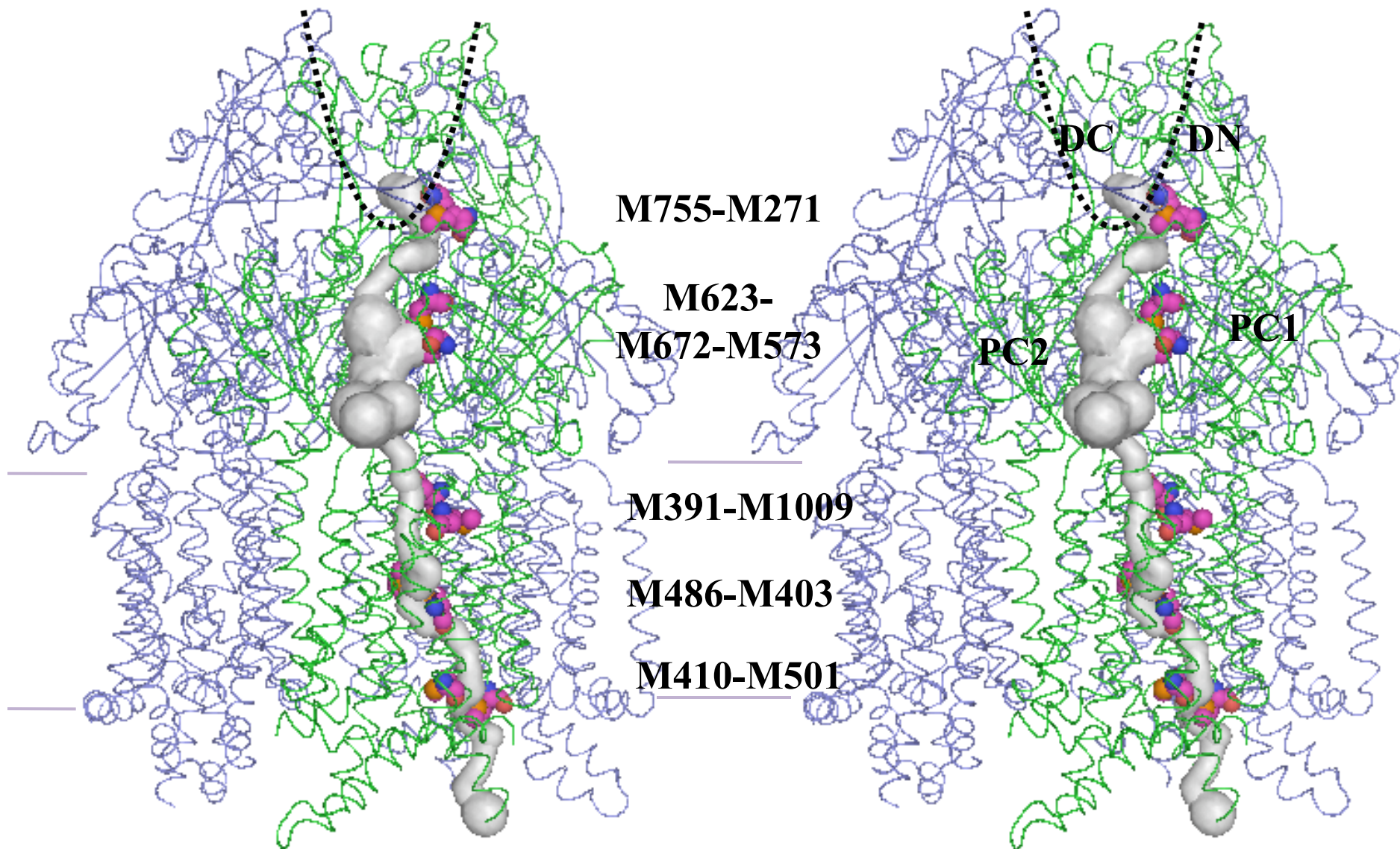


Fig. S9

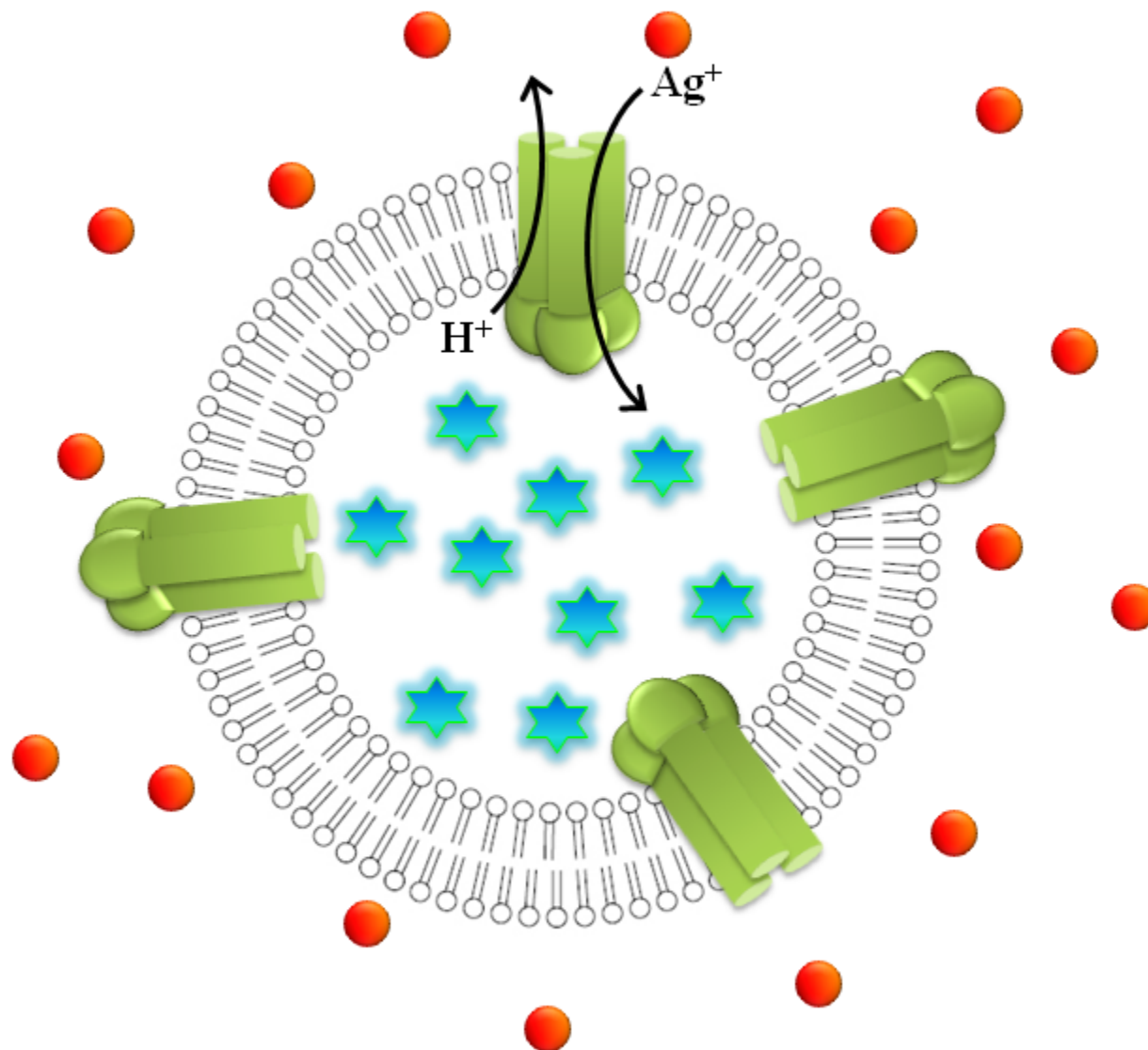


Fig. S10

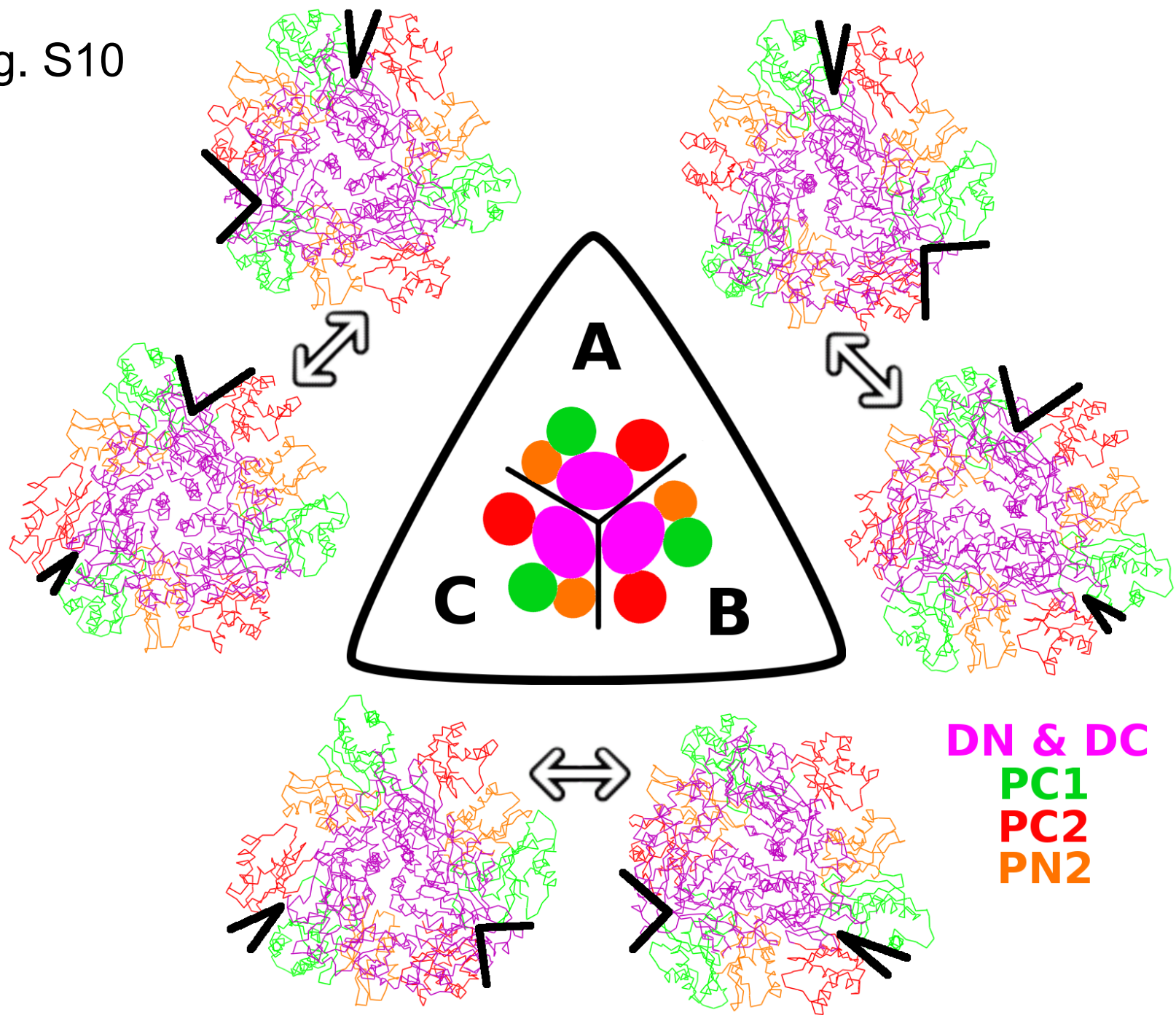


Fig. S11

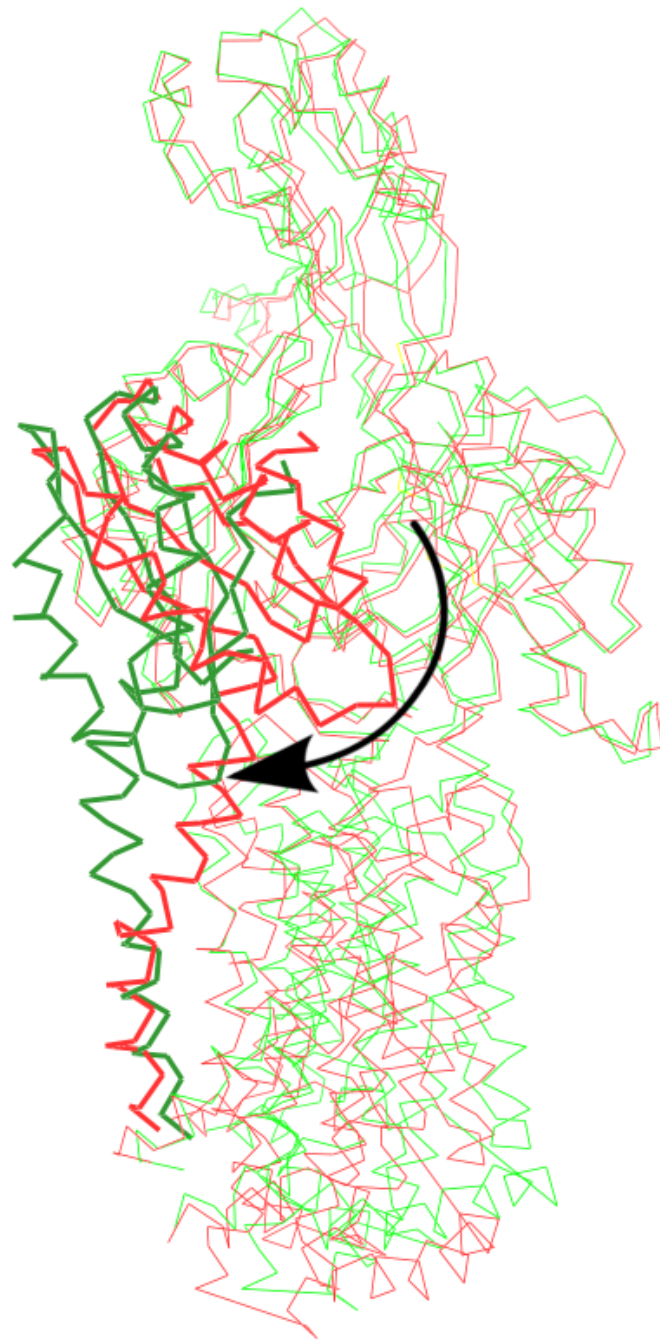


Fig. S12

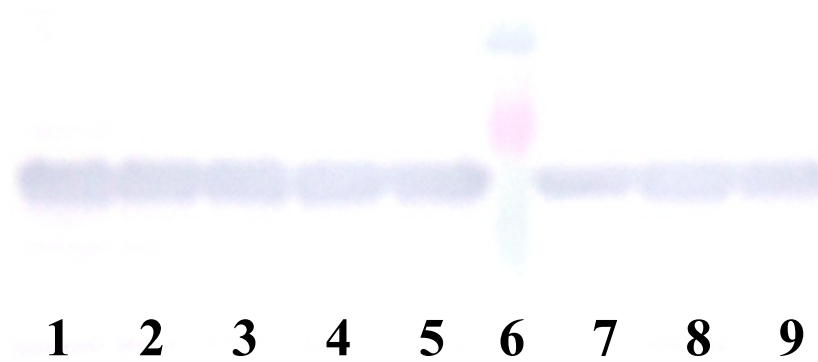


Fig. S13

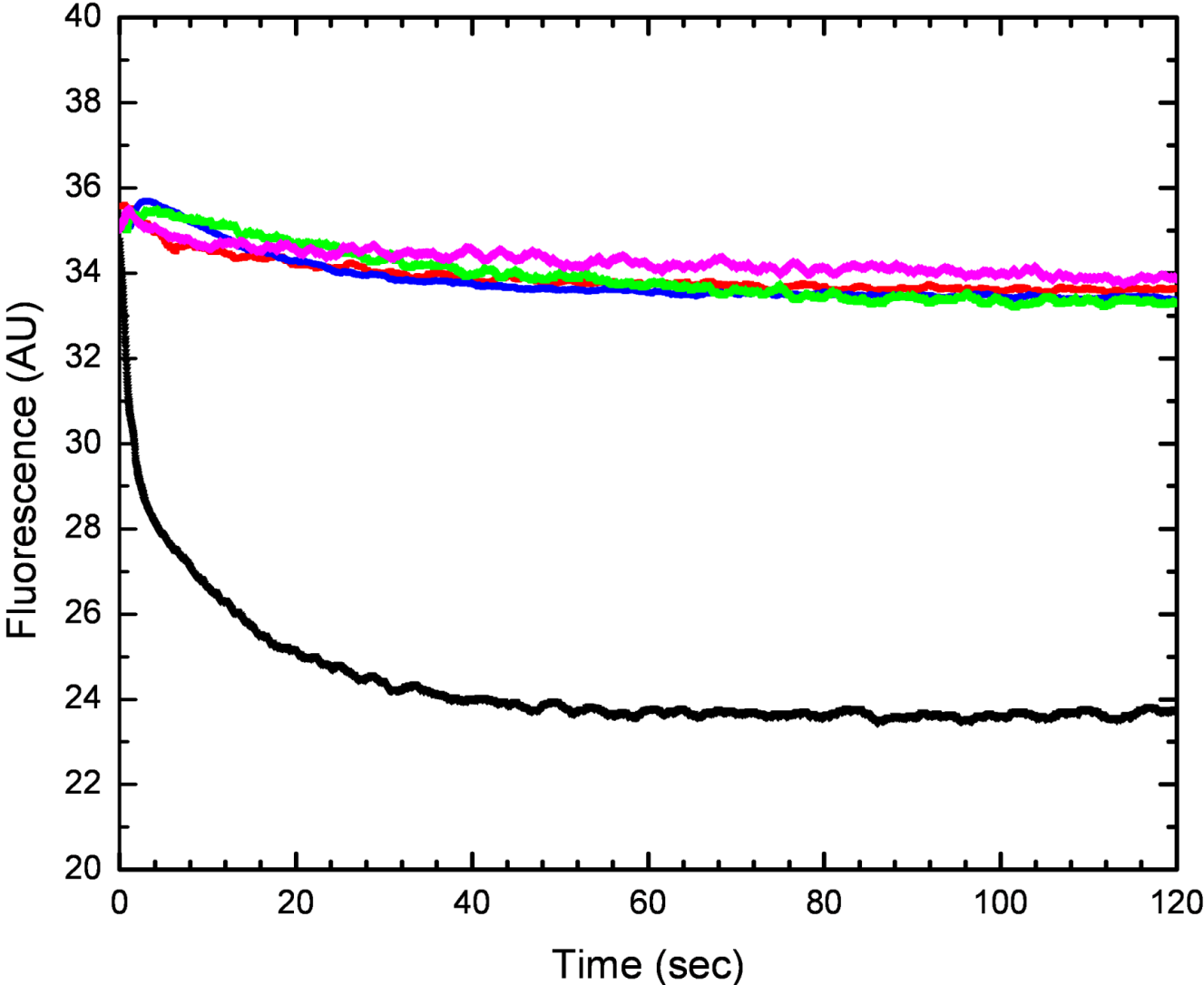
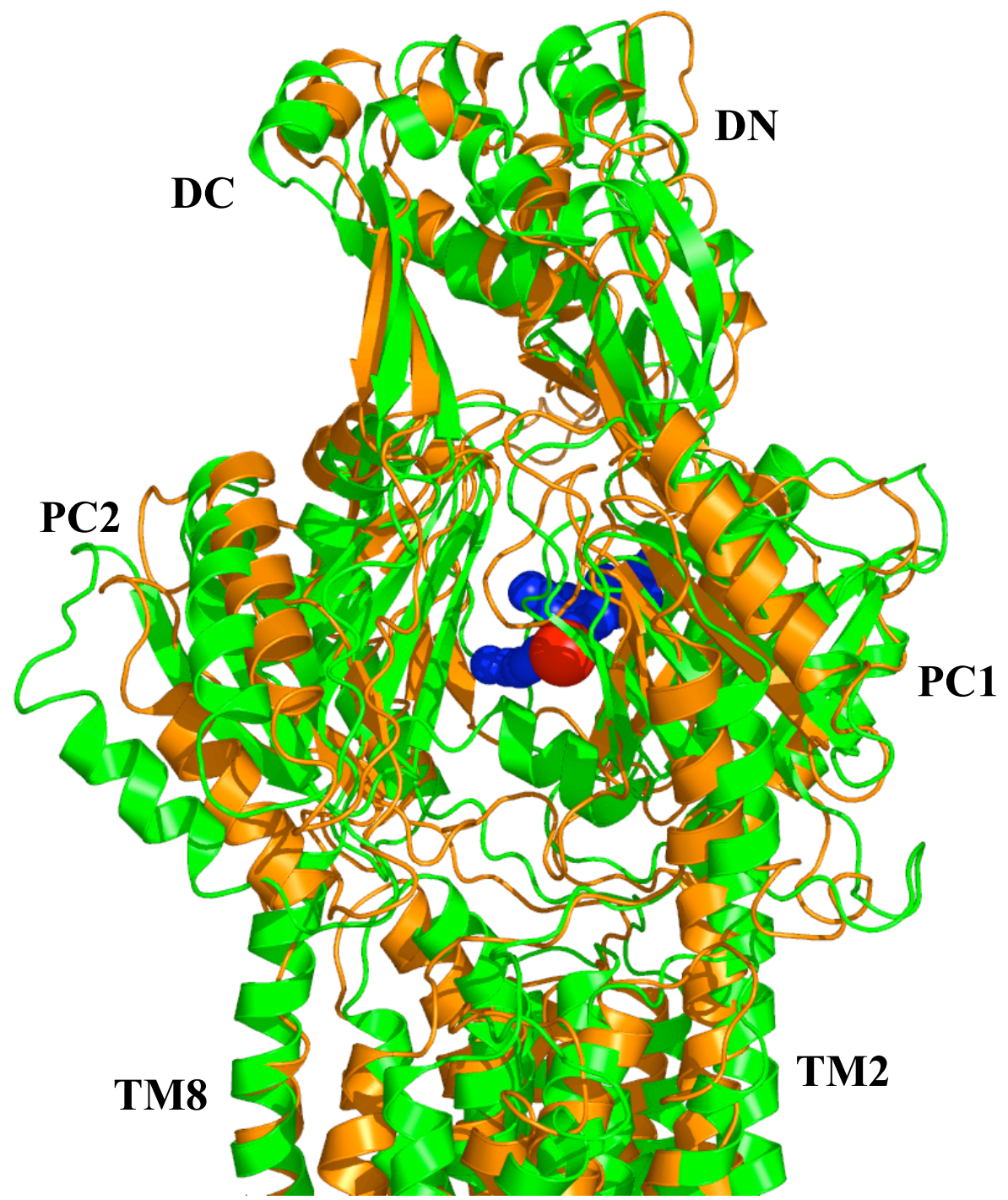


Fig. S14



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