Supporting Information

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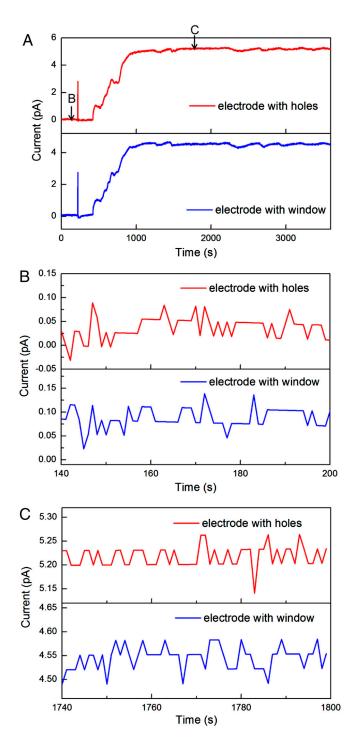


Fig. S1. Noise analysis for short-circuit current measurement in Fig. 2A. (A) Short-circuit current recording on two adjacent electrodes separated by 25 μm, with nanoholes (red) and window opening (blue). Data are the same as shown in Fig. 2A, with black arrows here marking the regions where the current noise was analyzed. (B and C) Comparison of the current noise from adjacent nanohole and window electrodes before (B) and after (C) cell addition. No correlation in noise patterns was observed for these two electrodes, with or without MR-1 cells in solution. These data combined with the voltammetry results in Fig. 1F, which showed that an adjacent but fully passivated electrode exhibited negligible current signal, demonstrate that there is no current cross-talk (signal or noise) between measured electrodes in our experiments.

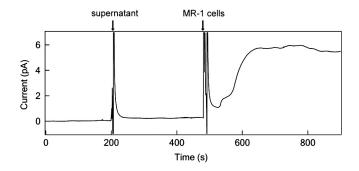
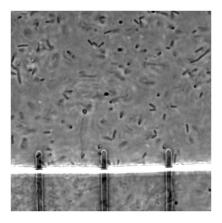
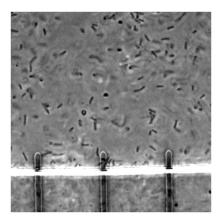


Fig. S2. Short-circuit current measurement on nanoelectrode when supernatant and Shewanella oneidensis MR-1 (MR-1) cells were added sequentially. The measured electrode ($2 \mu m$ wide) has a 6 $\mu m \times 10 \mu m$ opening to yield a total exposed electrode area of $\sim 12 \mu m^2$. The baseline was recorded with minimal media (MM) only. The 18-h MR-1 culture in MM was centrifuged at 3,000 rpm for 5 min, and 0.5 mL supernatant was added to the measurement chamber at ca. 200 s (after recording the stable baseline). An immediate increase of current followed by a rapid drop to background level was observed within 30 s. Subsequently, when 0.5 mL of culture containing MR-1 cells was injected (at ca. 480 s) the current increased steadily after the initial spike, and saturated at \sim 5 pA. These results indicate that supernatant or mediators alone cannot produce sustained currents, and moreover, that living MR-1 cells are required for the persistent current output observed in Fig. 2A.



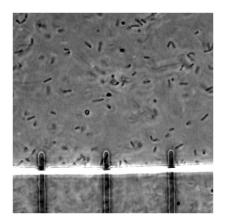
Movie S1. Real-time phase-contrast microscopy videos of *Shewanella* MR-1 cells recorded at position of the electrode used to record data in Fig. 5A. The videos were recorded before (Movie S1), during (Movie S2), and after (Movie S3) the sharp current dip occurring at ca. 22 min.

Movie S1 (WMV)



Movie S2. Real-time phase-contrast microscopy videos of *Shewanella* MR-1 cells recorded at position of the electrode used to record data in Fig. 5A. The videos were recorded before (Movie S1), during (Movie S2), and after (Movie S3) the sharp current dip occurring at ca. 22 min.

Movie S2 (WMV)



Movie S3. Real-time phase-contrast microscopy videos of *Shewanella* MR-1 cells recorded at position of the electrode used to record data in Fig. 5A. The videos were recorded before (Movie S1), during (Movie S2), and after (Movie S3) the sharp current dip occurring at ca. 22 min.

Movie S3 (WMV)