

SUPPORTING INFORMATION

A New Pathway for α -Ribazole-Phosphate Synthesis in *Listeria innocua*.

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TABLES

Table S1. Strains and plasmids used in this study

Strain or plasmid	Marker(s) ^a	Relevant genotype	Reference or source ^b
<u><i>L. innocua</i> strain:</u>			
DD680			K. Boor ^c
<u><i>S. enterica</i> strains:</u>			
TR6583		<i>metE205 ara-9</i> derivative of strain LT2	K. Sanderson ^d via J.R. Roth ^e
JE1244	Tc ^R	<i>metE205 ara-9 cobT10::Tn10d(tet⁺)</i>	lab collection
JE2701	Tc ^R	<i>metE205 ara-9 DELcob291 cobB1176::Tn10d(tet⁺)</i>	lab collection
JE8511	Tc ^R Ap ^R	<i>metE205 ara-9 cobT10::Tn10d(tet⁺)</i> / pBAD24	
JE8612	Tc ^R Ap ^R	<i>metE205 ara-9 cobT10::Tn10d(tet⁺)</i> / pCOBT48	
JE12331	Tc ^R Ap ^R	<i>metE205 ara-9 DELcob291 cobB1176::Tn10d(tet⁺)</i> / pBAD24	
JE12332	Tc ^R Ap ^R	<i>metE205 ara-9 DELcob291 cobB1176::Tn10d(tet⁺)</i> / pCOBT48	
JE12334	Tc ^R Ap ^R	<i>metE205 ara-9 DELcob291 cobB1176::Tn10d(tet⁺)</i> / pCBLTS1	
JE12550	Tc ^R Ap ^R	<i>metE205 ara-9 cobT10::Tn10d(tet⁺)</i> / pCBLT1	
JE12552	Tc ^R Ap ^R	<i>metE205 ara-9 cobT10::Tn10d(tet⁺)</i> / pCBLTS1	
JE12588	Tc ^R Ap ^R	<i>metE205 ara-9 cobT10::Tn10d(tet⁺)</i> / pCBLS4	
<u><i>E. coli</i> strains:</u>			
DH5 α /F'		F' / <i>endA1 hsdR17</i> (r _k ⁻ m _k ⁺) <i>glnV44 thi-1 recA1 gyrA</i> (Nal ^R) <i>relA1</i> Δ (<i>lacIZYA-argF</i>)U169	(Woodcock et al., 1989, Raleigh et al., 1989)

		<i>deoR</i> (ϕ 80 <i>dlac</i> Δ (<i>lacZ</i>)M15)	
BL21(DE3)		F ⁻ <i>ompT gal dcm lon hsdSB</i> (<i>r_B⁻ m_B⁻</i>) λ (DE3 [<i>lacI lacUV5-T7 gene 1 ind1 sam7 nin5</i>])	Novagen
Plasmids:			
pGEM [®] -T Easy	Ap ^R	TA cloning vector	Promega
pBAD24	Ap ^R	cloning vector with P _{BAD} arabinose-inducible promoter	(Guzman <i>et al.</i> , 1995)
pTEV5	Ap ^R	TEV protease-cleavable His ₆ tag overexpression vector	(Rocco <i>et al.</i> , 2008)
pKLD116	Ap ^R	TEV protease-cleavable His ₆ / maltose-binding protein tag overexpression vector	(Rocco <i>et al.</i> , 2008)
pCOBT48	Ap ^R	<i>S. enterica cobT</i> ⁺	lab collection
pCBLT1	Ap ^R	<i>L. innocua cbIT</i> ⁺	
pCBLS3	Ap ^R	<i>L. innocua cbIS</i> ⁺ translational fusion to His ₆ / maltose-binding protein tags for protein purification	
pCBLS4	Ap ^R	<i>L. innocua cbIS</i> ⁺	
pCBLS5	Ap ^R	<i>L. innocua cbIS</i> ⁺ translational fusion to His ₆ tag for protein purification	
pCBLTS1	Ap ^R	<i>L. innocua cbIT</i> ⁺ <i>cbIS</i> ⁺	

^a Tc^R, tetracycline resistance; Ap^R, ampicillin resistance.

^b Unless otherwise indicated, all strains and plasmids were constructed during the course of these studies.

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Table S2. Composition of MLM (minimal *Listeria* medium)

Component	Concentration (mM)
fructose	100
4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES, pH 7.0)	20
MgSO ₄	2
adenine HCl	0.5
CaCl ₂	0.1
	Concentration (g l⁻¹)
Na ₂ HPO ₄ •7H ₂ O	6.8
KH ₂ PO ₄	3
NaCl	0.5
	Concentration (mg l⁻¹)
cysteine	10
leucine	10
isoleucine	10
valine	10
riboflavin	5
thiamine	1
nicotinamide	1
pyridoxal	1
<i>p</i> -aminobenzoic acid	1
biotin	0.5
FeCl ₂	0.135
ZnCl ₂	0.017
MnCl ₂ •4H ₂ O	0.01
CoCl ₂ •6H ₂ O	0.006
NaMoO ₄ •2H ₂ O	0.006
α -lipoic acid	0.005
CuCl ₂ •2H ₂ O	0.005
Na ₂ SeO ₄	0.005

Table S3. Comparison of the presence of *cobT*, *bluB*, and *cbiZ* homologs in bacterial genomes encoding *cbIS* or *cbIT* homologs

Species ^a	Locus tag of predicted <i>cbIT</i> homolog (% identity, % similarity to CbiT from <i>L. innocua</i>) ^b	Locus tag of predicted <i>cbIS</i> homolog (% identity, % similarity to CbiS from <i>L. innocua</i>) ^c	Locus tag of predicted <i>cobT</i> homolog(s) (% identity, % similarity to CobT from <i>S. enterica</i>) ^d	Locus tag of predicted <i>bluB</i> homolog (% identity, % similarity to BluB from <i>R. rubrum</i>) ^g	Locus tag of predicted <i>cbiZ</i> homolog (% identity, % similarity to CbiZ from <i>M. mazei</i>) ^h
<i>Alkaliphilus metalliredigens</i> QYMF	Amet_0066 (41%, 58%)	Amet_0067 (34%, 54%)	Amet_0464 (45%, 62%)		
<i>Alkaliphilus oremlandii</i> OhILAs	Clos_2207 (33%, 56%)	Clos_2206 (36%, 58%)			
<i>Anaerotruncus colihominis</i> DSM 17241		ANACOL_03809 (35%, 55%)	ANACOL_02043 (41%, 61%)		
<i>Anoxybacillus flavithermus</i> WK1	Aflv_1051 (33%, 53%)	Aflv_1052 (28%, 48%)	Aflv_2169 (42%, 59%)		Aflv_1043 ⁱ (28%, 50%)
<i>Bacillus coahuilensis</i> m4-4	Bcoam_010100007990 (30%, 47%)				Bcoam_010100007955 ⁱ (25%, 46%)
<i>Bacillus halodurans</i> C-125		BH0853 (33%, 54%)	BH0284 (40%, 60%)		BH1587 ⁱ (34%, 50%)
<i>Carboxydotherrmus hydrogenoformans</i> Z-2901	CHY_0775 (37%, 53%)	CHY_0776 (32%, 51%)	CHY_0480 (38%, 61%)		
<i>Clostridium botulinum</i> A Hall	CLC_1007 (43%, 71%)	CLC_0877 (32%, 53%)			
<i>Clostridium perfringens</i> 13	CPE1308 (42%, 69%)	CPE1307 (26%, 50%)	CPE1034 (38%, 60%)		
<i>Clostridium sporogenes</i> ATCC 15579	CLOSPO_01660 (42%, 71%)	CLOSPO_01525 (31%, 52%)			
<i>Clostridium tetani</i> E88	CTC00741 (42%, 64%)	CTC00742 (36%, 58%)	CTC02290 (39%, 60%)		
<i>Desulfitobacterium hafniense</i> Y51		DSY2725 (34%, 50%)	DSY2114 (42%, 61%)		
<i>Geobacillus kaustophilus</i> HTA426	GK2256 (33%, 51%)	GK2255 (30%, 45%)	GK1793 (40%, 57%)		GK2264 ⁱ (29%, 48%)

<i>Geobacillus thermodenitrificans</i> NG80-2	GTNG_2189 (32%, 50%)	GTNG_2188 (30%, 46%)	GTNG_1683 (40%, 57%)		GTNG_2197 ⁱ (27%, 47%)
<i>Heliobacterium modesticaldum</i> Ice1	HM1_1580 (33%, 51%)	HM1_1579 (34%, 54%)	HM1_2401 (41%, 62%)		HM1_1855 (27%, 43%)
<i>Lactobacillus brevis gravesensis</i> ATCC 27305	HMPREF0496_0 555 (47%, 68%)	HMPREF0496_0 556 (42%, 59%)			
<i>Lactobacillus buchneri</i> ATCC 11577	HMPREF0497_0 511 (47%, 67%)	HMPREF0497_0 512 (43%, 60%)			
<i>Lactobacillus hilgardii</i> ATCC 8290	HMPREF0519_1 876 (54%, 70%)	HMPREF0519_1 306 (42%, 60%)			
<i>Listeria innocua</i> Clip11262	lin1153 (100%, 100%)	lin1110 (100%, 100%)			
<i>Listeria monocytogenes</i> EGD-e	lmo1190 (97%, 98%)	lmo1146 (81%, 87%)			
<i>Listeria welshimeri</i> sv 6b SLCC5334	lwe1147 (94%, 98%)	lwe1104 (78%, 88%)			
<i>Lysinibacillus sphaericus</i> C3-41	Bsph_2450 (31%, 50%)	Bsph_2451 (26%, 47%)	Bsph_1664 (48%, 64%)	Bsph_2918 (33%, 49%)	Bsph_0967 ⁱ (25%, 45%)
<i>Moorella thermoacetica</i> ATCC 39073	Moth_0191 (48%, 69%)	Moth_0190 (35%, 51%)	Moth_1101 (41%, 63%)		
			Moth_1721 (41%, 59%)		
<i>Natranaerobius thermophilus</i> JW/NM-WN-LF	Nther_0938 (45%, 68%)	Nther_0939 (33%, 54%)			
<i>Propionibacterium acnes</i> KPA171202		PPA0108 (27%, 46%)	PPA0441 ^e (40%, 57%)	PPA0953 ^f (38%, 53%)	
			PPA0953 ^f (33%, 50%)		
<i>Symbiobacterium thermophilum</i> IAM 14863	STH1931 (41%, 54%)	STH1930 (35%, 55%)			
<i>Thermoanaerobacter brockii</i> subsp. <i>finnii</i> Ako-1	ThebrDRAFT_14 29 (34%, 57%)	ThebrDRAFT_14 30 (33%, 53%)			
<i>Thermoanaerobacter ethanolicus</i> CCSD1	TeCCSD1DRAF T_0616 (33%, 56%)	TeCCSD1DRAF T_0615 (35%, 54%)			
<i>Thermoanaerobacter italicus</i> Ab9	ThitDRAFT_083	ThitDRAFT_083			

	7 (33%, 55%)	6 (33%, 54%)			
<i>Thermoanaerobacter mathranii</i> subsp. <i>mathranii</i> str. A3	TmathDRAFT_1 629 (34%, 55%)	TmathDRAFT_1 628 (33%, 54%)			
<i>Thermoanaerobacter pseudethanolicus</i> ATCC 33223	Teth39_1898 (34%, 57%)	Teth39_1897 (33%, 53%)			
<i>Thermoanaerobacter tengcongensis</i> MB4	TTE0378 (37%, 57%)	TTE0379 (36%, 53%)			TTE2426 (44%, 65%)
<i>Thermoanaerobacterium thermosaccharolyticum</i> DSM 571	TtheDRAFT_223 9 (35%, 55%)	TtheDRAFT_224 0 (32%, 50%)			

^a For species with multiple sequenced strains, a single representative example has been selected.

^b Identified by BLAST search of the Integrated Microbial Genomes database (<http://img.jgi.doe.gov>), accessed March 23, 2010 (Markowitz *et al.*, 2006).

^c Identified by BLAST search of the Integrated Microbial Genomes database (<http://img.jgi.doe.gov>), accessed March 23, 2010 (Markowitz *et al.*, 2006).

^d Identified by BLAST search of the Integrated Microbial Genomes database (<http://img.jgi.doe.gov>), using the *Salmonella enterica* CobT sequence (Trzebiatowski *et al.*, 1994) as the search term, accessed June 18, 2010 (Markowitz *et al.*, 2006). Percent identity and percent similarity were calculated with the BLAST 2 Sequences program (McGinnis & Madden, 2004).

^e This gene encodes a fusion protein between CobT and a CobU (adenosylcobinamide kinase / adenosylcobinamide-phosphate guanylyltransferase) ortholog (Markowitz *et al.*, 2006).

^f This gene encodes a fusion protein between CobT and a BluB (aerobic DMB synthase) ortholog (Markowitz *et al.*, 2006).

^g Identified by BLAST search of the Integrated Microbial Genomes database (<http://img.jgi.doe.gov>), using the *Rhodospirillum rubrum* BluB sequence (Gray & Escalante-Semerena, 2007) as the search term, accessed April 8, 2010 (Markowitz *et al.*, 2006).

^h Identified by BLAST search of the Integrated Microbial Genomes database (<http://img.jgi.doe.gov>), using the *Methanosarcina mazei* CbiZ sequence (Gray *et al.*, 2008) as a search term, accessed April 8, 2010 (Markowitz *et al.*, 2006).

ⁱ These genes encode fusion proteins between CbiZ and homologs of BtuD, the ATPase component of the Btu corrinoid-specific ABC transport system (Gray *et al.*, 2008; Markowitz *et al.*, 2006).

Table S4: Primers used in this study

Primer	Sequence ^a
[1]	CTG TTT CTC CAT ACC CGT T
[2]	GGC TGA AAA TCT TCT CTC AT
[3]	ACT GAA <u>GAA TTC</u> ATG AA ATT CAA AAA TTA GTA TTA TGT GCG
[4]	ACT GAA <u>TCT AGA TTA</u> GTA CCC GGC AAT TCG TCT T
[5]	GAC GTC <u>CCG GGC TAG CAT</u> GCC TCA AGT GAG GGA TTT
[6]	GTG CGG CCG <u>CAA GCT TTC</u> AGC CAT GGT AAT TCC TCA A
[7]	CGA GCG GAA CCG CCT CG
[8]	CCA TTC GCC AAT CCG GAT
[9]	TCG TAC TAC CAT CAC CAT CAC
[10]	CTA GCA GGA <u>GGA ATT CCA</u> CCA TGC CTC AAG TGA GGG ATT T
[11]	CAA AAC AGC <u>CAA GCT TTC</u> AGC CAT GGT AAT TCC TCA A
[12]	CTA GCA GGA <u>GGA ATT CTT</u> GAA GAT TCA AAA ATT AGT ATT ATG TGC G
[13]	CTT GAG GCA TTC TAG ATT AGT ACC CGG CAA TTC GTC TT
[14]	CGG GTA CTA ATC TAG AAT GCC TCA AGT GAG GGA TTT
[15]	CCA AGC TTG CAT <u>GCC TGC AGT</u> CAG CCA TGG TAA TTC CTC AA

^a Restriction sites introduced for cloning are underlined.

FIGURES

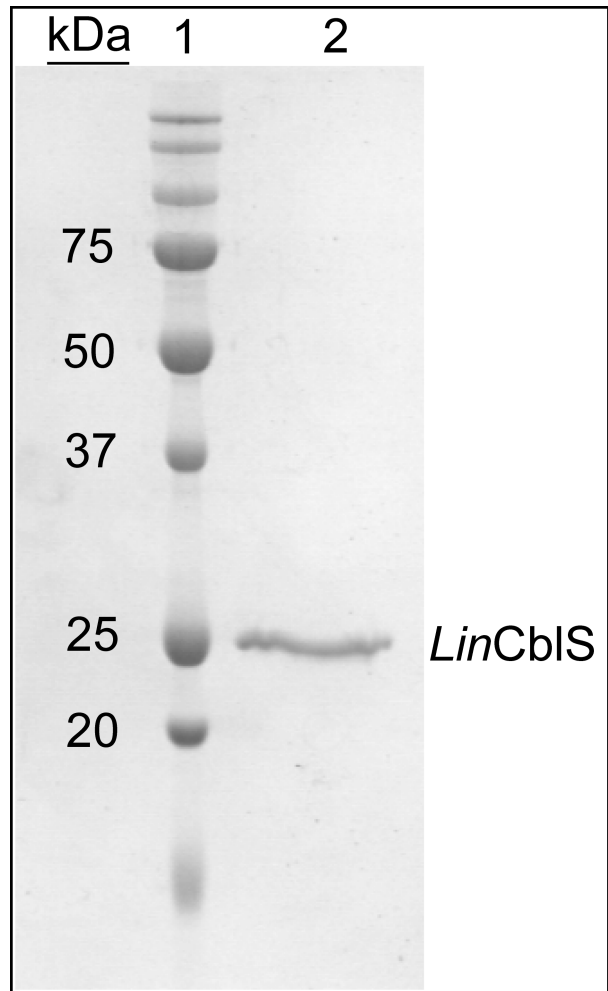


Figure S1. Purification of *LinCbIS* protein. Purified *LinCbIS* (2 μ g) was resolved by 12% SDS-PAGE (Laemmli, 1970) and stained with Coomassie Blue (Sasse, 1991). Lane 1. Electrophoretic mobility of molecular mass markers (Precision Plus Protein™ Standards, Bio-Rad Laboratories). Molecular mass in kilodaltons (kDa) is indicated. Lane 2. *LinCbIS* protein, > 98% homogenous.

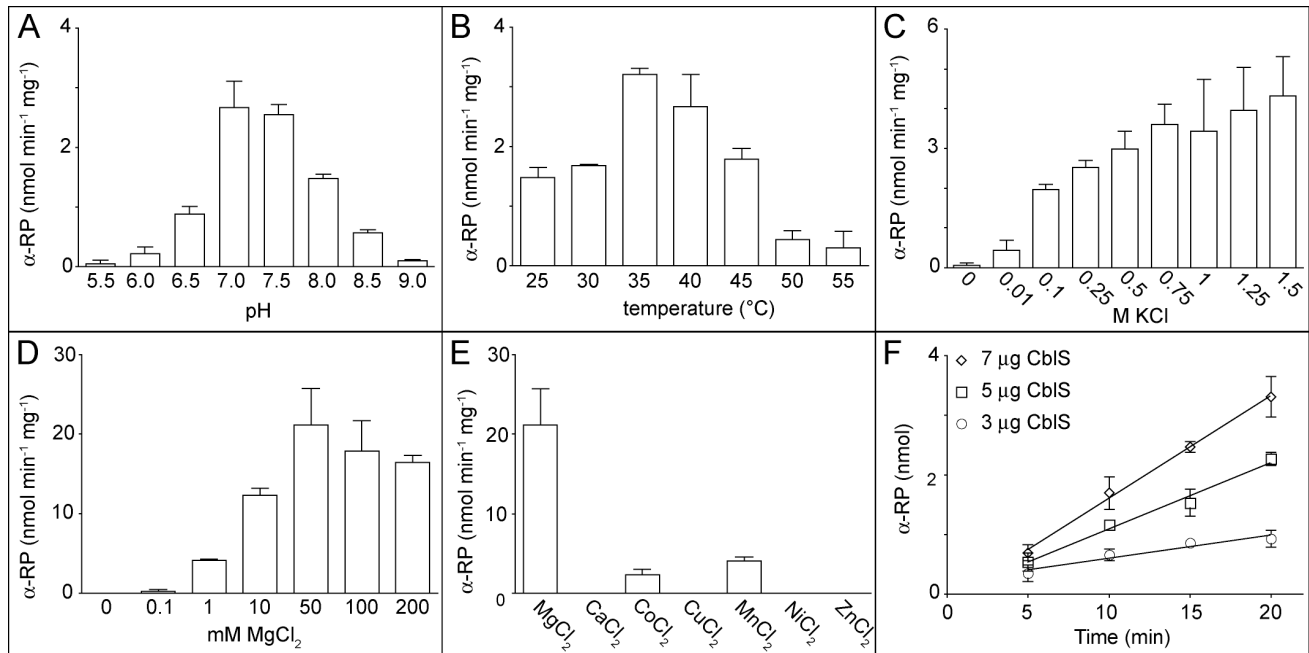


Figure S2. Optimization of *LinCbIS* reaction conditions. α -R kinase activity of *LinCbIS* (expressed as nanomoles of α -RP min⁻¹ mg⁻¹ protein) is shown, with error bars of one standard deviation. Unless indicated, reactions contained Tris-HCl buffer (100 mM, pH 7.0), TCEP (5 mM), KCl (0.75 M), MgCl₂ (50 mM), α -R (30 μ M), *LinCbIS* (3 - 10 μ g) and ATP (1 mM) and were incubated at 35°C. **A.** *LinCbIS* activity as a function of pH at 37°C, 0.5 M KCl, 1 mM MgCl₂. **B.** *LinCbIS* activity as a function of temperature at 0.5 M KCl, 1 mM MgCl₂. **C.** *LinCbIS* activity as a function of KCl concentration at 1 mM MgCl₂. **D.** *LinCbIS* activity as a function of MgCl₂ concentration. **E.** *LinCbIS* activity in the presence of different divalent cations (50 mM). **F.** Product formation as a function of time and *LinCbIS* concentration at 60 μ M α -R.

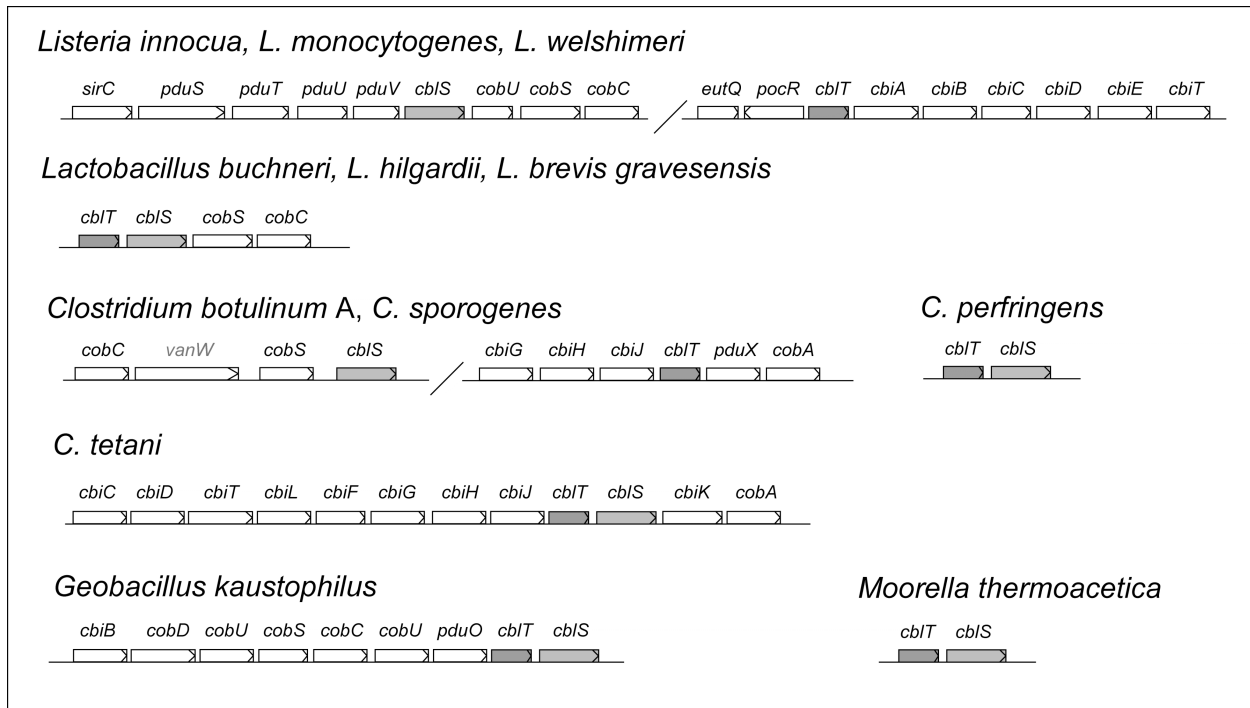


Figure S3. Genomic context of *cbiT* and *cbIS* homologs in selected bacteria. The organization of genes encoding products predicted to be involved in coenzyme B₁₂ biosynthesis or utilization near predicted *cbiT* (dark grey) and *cbIS* (light grey) homologs in representative bacterial genomes is indicated. Arrows within boxes indicate the direction of transcription of the indicated gene. Diagonal slashes indicate gaps between *cbiT*- and *cbIS*-associated loci in a single genome. The products of the indicated genes are as follows (the locus tags of the *Listeria innocua* Clip11262 homolog of each gene is provided in parentheses): *sirC* (*lin1105*), precorrin-2 dehydrogenase; *pduS* (*lin1106*) and *pduV* (*lin1107*), predicted 1,2-propanediol utilization proteins; *pduT* (*lin1108*) and *pduU* (*lin1109*), metabolosome shell proteins involved in 1,2-propanediol utilization; *cbIS* (*lin1110*), α -R kinase; *cobU* (*lin1111*), bifunctional AdoCbi kinase / GTP:AdoCbi-P guanylyltransferase; *cobS* (*lin1112*), AdoCbi-P synthase; *cobC* (*lin1113*), AdoCbi-5'-P phosphatase; *eutQ* (*lin1151*), predicted ethanolamine utilization protein; *pocR* (*lin1152*), putative 1,2-propanediol-sensing transcription factor; *cbiT* (*lin1153*), α -R transporter; *cbiA* (*lin1154*), cobyrinic acid *a,c*-diamide synthase; *cbiB* (*lin1155*), AdoCbi-P synthetase; *cbiC* (*lin1156*), precorrin-8X methylmutase; *cbiD* (*lin1157*), cobalt-precorrin-6A synthase; *cbiE* (*lin1158*), precorrin-6B methylase; *cbiT* (*lin1159*), precorrin-8W decarboxylase; *vanW*, vancomycin B-type resistance protein (*C. botulinum* locus tag CLC_0875, no *L. innocua* homolog and no known role in coenzyme B₁₂ metabolism); *cbiG* (*lin1161*), cobalt-precorrin-5A hydrolase; *cbiH* (*lin1162*), precorrin-3B C17-methyltransferase; *cbiJ* (*lin1163*), precorrin-6X reductase; *pduX* (*lin1134*), L-threonine kinase; *cobA* (*C. tetani* locus tag CTC00744, no *L. innocua* homolog), ATP:corrinoid adenosyltransferase; *cbiL* (*lin1166*), precorrin-2 methyltransferase; *cbiF* (*lin1160*), precorrin-4 C-11 methyltransferase; *cbiK* (*lin1165*), cobalt chelatase; *pduO* (*lin1128* and *lin1172*), ATP:corrinoid adenosyltransferase.

<i>A. metaliredigens</i>	-----	
<i>A. oremlandii</i>	-----	
<i>A. flavithermus</i>	-----	
<i>B. coahuilensis</i>	-----	
<i>C. hydrogenoformans</i>	-----	
<i>C. botulinum</i>	-----	
<i>C. perfringens</i>	-----	
<i>C. sporogenes</i>	-----	
<i>C. tetani</i>	-----	
<i>G. kaustophilus</i>	-----	
<i>G. thermodenitrificans</i>	-----	
<i>H. modesticaldum</i>	-----MSNMP	5
<i>L. gravesensis</i>	-----	
<i>L. buchneri</i>	-----	
<i>L. hilgardii</i>	-----	
<i>L. innocua</i>	-----	
<i>L. monocytogenes</i>	-----	
<i>L. welshimeri</i>	-----	
<i>L. sphaericus</i>	-----	
<i>M. thermoacetica</i>	-----MAKPE	5
<i>N. thermophilus</i>	-----MLS--	3
<i>S. thermophilum</i>	MPLEHLTTTSGIGAPKHRCTVACRTGTGESRAQRGAAYGHNGVPTKRLL	50
<i>T. brockii</i>	-----	
<i>T. ethanolicus</i>	-----	
<i>T. italicus</i>	-----	
<i>T. mathranii</i>	-----	
<i>T. pseudethanolicus</i>	-----	
<i>T. tengcongensis</i>	-----	
<i>T. thermosaccharolyticum</i>	-----	
<i>A. metaliredigens</i>	-MISRNRKQMDTKMLVKMAMLIASGIGAMIKIQG---SIALD	46
<i>A. oremlandii</i>	-MNNTKNVMSMSNIQVITRSGLLIGLSAIGAMIKIQG---TIAFDS	46
<i>A. flavithermus</i>	-----MRRFTLLVMFISLSVIGSLIKIPFTIGSIALDS	38
<i>B. coahuilensis</i>	-----MKTSRVNLIAAFIALSVIGGFIKIPSPISSIALDS	40
<i>C. hydrogenoformans</i>	-----MNIRTLTLTALIALIALSAVGAFIKIPSPGTVALD	40
<i>C. botulinum</i>	-----MKTKRLVIMGLFIALSFVGANIKIMG---SIAFDS	37
<i>C. perfringens</i>	-MNFTEKPKRDLVKSLCFSAILIAISVILANFPIFS---SIALDS	46
<i>C. sporogenes</i>	-----MKTKRLVIVGLFIALSFVGANIKIMG---SIAFDS	37
<i>C. tetani</i>	-----MKTNKMVITALFIALSFVGANIKIMG---TVAFDA	37
<i>G. kaustophilus</i>	-----MNRRLAWTAVCLIALSAIGSFIKLPTFVGSIALDS	39
<i>G. thermodenitrificans</i>	-----MNRRFVWLAVCMALSVIGSFIKLPTFVGSIALDS	39
<i>H. modesticaldum</i>	TQYNEVGPWSKSVRSIALMGLLIALSAVGAMVKLPSPVGTIGLDS	55
<i>L. gravesensis</i>	-----MRNDYLKKLMLTTIFLALCVGANVKILG---SIALDS	40
<i>L. buchneri</i>	-----MRNDYLKKLMLTTIFLALCVGANVKILG---SIALDS	40
<i>L. hilgardii</i>	-----MTQSDRLNRLTLAAMLLALCVGANVKIMG---SVAFDA	41
<i>L. innocua</i>	-----MKIQKLVLCAMLIAMCVIGANIKIMG---SVAFDA	37
<i>L. monocytogenes</i>	-----MKIQKLVLCAMLIAMCVIGANIKIMG---SVAFDA	37
<i>L. welshimeri</i>	-----MKIQKLVLCAMLIAMCVIGANIKIMG---SVAFDA	37
<i>L. sphaericus</i>	-----MDRQTLMTWILTAMVAAICAVGAAIKVPAFISTAALDS	43
<i>M. thermoacetica</i>	LSSPAVKASRWTARRLATLAMLIALSTVGANLKIPTSITGTPAFDS	55
<i>N. thermophilus</i>	-----LDHRELVKIALIALSMIGSQIKIPSLTGTPALDS	43
<i>S. thermophilum</i>	LGRGGLLVLLARTNTLVRGLIALSVLGAYIKLGP--SSIAFDA	98
<i>T. brockii</i>	-MK-EKANALKNVKTLTLVAMLIALSavgALIKIFN---TVAFDA	45
<i>T. ethanolicus</i>	-MK-EKANALKNVKTLTLVAMLIALSavgALIKIFN---TVAFDA	45
<i>T. italicus</i>	-MK-EKANALKNVKTLTLVAMLIALSavgALIKIFN---TVAFDA	45
<i>T. mathranii</i>	-MK-EKANTLKNVKTTLTLVAMLIALSavgALIKVFN---TVAFDA	45
<i>T. pseudethanolicus</i>	-MK-EKANALKNVKTLTLVAMLIALSavgALIKIFN---TVAFDA	45
<i>T. tengcongensis</i>	-MK-EKEVSLGNVKTTLTLVAMLIALSavgALIKIFN---TVAFDA	45
<i>T. thermosaccharolyticum</i>	-MN-TKTTTIKNVKTLTLVAMLIAMSAVGAMIKIYN---TVAFDS	45
<i>A. metaliredigens</i>	AALLGPMAGGIVAFAGELISALTAGEPMTVPMHLVVAVEMFIIVALFSV	96
<i>A. oremlandii</i>	AALFISPMAGGAVASLGEHLLTAFTSCFPLTLPMHMLTVVMGIAYFFGV	96
<i>A. flavithermus</i>	AAVLLGPTAAAVVASVGEHVVSALFGGEPILG-PFHILVAIEMAALLYIVGL	87
<i>B. coahuilensis</i>	AAMFSGMIGGFISIGHELLSSLSGGCFPLG-PFHFIIMIEMFICILFIYAL	89
<i>C. hydrogenoformans</i>	AALYLSPLGALVAALGHELLSAATAGFPPLTLPLHLLVALEMATFAAVFGV	90
<i>C. botulinum</i>	GTLLIGPIMGAIIGAVAFHLSALTSCEPPLSLPVMIVMVDMAVTMILFGI	87
<i>C. perfringens</i>	GGIILSPVGGIIGLLAHFLVALRTGFPPLSLPVHILVALEMVVVYITSI	96
<i>C. sporogenes</i>	GTLLIGPIMGAIIGAVAFHLSALTSCEPPLSLPVMIVMVDMAVTMLLFGV	87
<i>C. tetani</i>	GALLLGPVYGGIIGGVGHEFLTALTSCEPPLSLPVHLVITVIMAAATMAIFGL	87
<i>G. kaustophilus</i>	AAGVLGPRAGAAVAGLGHLSVALIGGCFPLG-PVHWFVALEMAGLGALFAV	88
<i>G. thermodenitrificans</i>	AAAVLGPQAGATVAGLGHFISAYIGGCFPLG-PHEHLVACEMAVLGAFAA	88
<i>H. modesticaldum</i>	AAALGATGGMIVIALGHELLTAMVVGFPPLTLPVHLFIALQAMWAYVFRR	105
<i>L. gravesensis</i>	GAILLGAIGAFILGFFGHLLSAMLAGEPPLTLPVHLIIGVMMACMFVFGV	90
<i>L. buchneri</i>	GAILLGAIGAFILGFFGHLLSAMLAGEPPLTLPVHLIIGVMMACMFVFGV	90

<i>L. hilgardii</i>	GAVLLGPFWFGATLGLFGELVSAALAGFPLTLPIHLIIGVAMGICMLIFGL	91
<i>L. innocua</i>	GTLLLGPMYGAVALGIFGELTSAALLAGFPLTLPIHLIVAGMMGVTMIAYGF	87
<i>L. monocytogenes</i>	GTLLLGPMYGAVALGIFGELTSAALLAGFPLTLPIHLIVAGMMGVTMIAYGF	87
<i>L. welshimeri</i>	GTLLLGPMYGAVALGIFGELTSAALLAGFPLTLPIHLIVAGMMGVTMIAYGF	87
<i>L. sphaericus</i>	GVVFLSPLLAGIGGFIFGELTSAALLAGFPLG-PLHVIIVAVEMFIVVWIFGI	92
<i>M. thermoacetica</i>	GALLLGPADGALIAALGELTSAALLAGFPLTPPLHLVIAAGMAAVVALFAI	105
<i>N. thermophilus</i>	AAFVWVGKYAAAIIGFFGELTSAALLAGFPLTLPIHLIVAGMMGVTMIAYGF	93
<i>S. thermophilum</i>	AALLMGPAAAGALICGLGELTSAALLAGFPLTLPIHLIVAGMMGVTMIAYGF	148
<i>T. brockii</i>	AALYLGSWYGALVISLGEMLTAITSGFPLGLTNHIYIAVQMGLYAYLKF	95
<i>T. ethanolicus</i>	AALYLGSWYGALVISLGEMLTAITSGFPLGLTNHIYIAVQMGLYAYLKF	95
<i>T. italicus</i>	AALYLGSWYGALVISLGEMLTAITSGFPLGLTNHIYIAVQMGLYAYLFRF	95
<i>T. mathranii</i>	AALYLGSWYGALVISLGEMLTAITSGFPLGLTNHIYIAVQMGLYAYLFRF	95
<i>T. pseudethanolicus</i>	AALYLGSWYGALVISLGEMLTAITSGFPLGLTNHIYIAVQMGLYAYLKF	95
<i>T. tengcongensis</i>	AALYLGSLYGAIIVIALGELTSAALLAGFPLTLPIHLIVAGMMGVTMIAYGF	95
<i>T. thermosaccharolyticum</i>	ASLYFGSYGAIIVISLGEMLTAITSGFPLGLTNHIYIAVQMGLYAYLFRF	95
<i>A. metaliredigens</i>	VW-----QKINPVAIVGILLNGVAGLLVVPMSILLGLPLNGWALF	139
<i>A. oremlandii</i>	IE-----RKVNGVLACVIAILLNGPVATFIAGITASLLGLPLSGSAMF	139
<i>A. flavithermus</i>	LIR-----RGWFIFSYVCFFIG-NALLAPLPFV-----WIMS-PAFV	122
<i>B. coahuilensis</i>	FYK-----TNKWV--AFLFFIGANTFLAPLPFL-----FFHGPIFI	124
<i>C. hydrogenoformans</i>	LG-----KRN-VIVGIVVATLLNGVIAPLSFAI-----MPKFGMAFF	126
<i>C. botulinum</i>	VYNKFKNKNNILAAIVATVVAVIINGPVSFVAI-----IPIAGKG-V	128
<i>C. perfringens</i>	IF-----NRGKVLIVGIVGTLNGIVFTLITGVFMYFVLGGMNPIDFL	139
<i>C. sporogenes</i>	VYNKLNKNKNNILAAIVATIVAVIINGPVSFVAI-----IPITGKG-V	128
<i>C. tetani</i>	VYKVLVKKNKVLAAMILAIVGIVVNGPINLLVLTPLL---MPIMGKAGI	133
<i>G. kaustophilus</i>	LHR-----RGWKIGSAVVFFIG-NAFLAPLPLA-----VSFG-WPFV	123
<i>G. thermodenitrificans</i>	MYG-----RGWRFGGAIAFFVG-NVFLAPLPFV-----IAFG-WPLV	123
<i>H. modesticaldum</i>	VN-----QFGLTAAFAAAVCLNGVSSSLTM-----LLGGWGAV	140
<i>L. gravesensis</i>	LRQRLN-LSRGLVIVISDAIGYLVNPLELTL-----YPIKQSV	130
<i>L. buchneri</i>	LRQRLT-LSRGLVIVISDAIGYLVNPLELTL-----YPIKQSV	130
<i>L. hilgardii</i>	VRKWLK-KDTLKGVLTSVLYGAINVPIELVLL-----YPLMKQA-V	131
<i>L. innocua</i>	TRHKLAEKNQLVAISASSVAVFVFNCPPLSLLAL-----YPLMQA-V	128
<i>L. monocytogenes</i>	TRQKLAENQLVAISVSSIVAFVFNCPPLSLLAL-----YPLMQA-V	128
<i>L. welshimeri</i>	TRQKLADKNQLIAVSVSSMVAFVFNCPPLSLLAL-----YPLMQA-V	128
<i>L. sphaericus</i>	MHK-----KGMHFWKVVALVL-NGIVAPLPFY-----FIIS-PAFF	127
<i>M. thermoacetica</i>	FY-----RFS-PWLGIAAGIALNGLLPLALFIP-----LPGFGKAF	141
<i>N. thermophilus</i>	VN-----NYSGRAAGVLAGMVLNGLLPLGIFVL-----IPGFGIFF	130
<i>S. thermophilum</i>	AA-----RRFGLVAGAAVLVAVANGILAPALLALLP-----NPLGLGLF	186
<i>T. brockii</i>	FY-----RKFNIYIAVIAATILNGPVATLLF-----VPIFGWGF	130
<i>T. ethanolicus</i>	FY-----QKFNIIYIAVIAATILNGPVATLLF-----VPIFGWGF	130
<i>T. italicus</i>	FY-----RKFNIYIAVIVATILNGPVATLLF-----VPIFGWGF	130
<i>T. mathranii</i>	FY-----RKFNIYIAVIVATILNGPVATLLF-----VPIFGWGF	130
<i>T. pseudethanolicus</i>	FY-----RKFNIYIAVIAATILNGPVATLLF-----VPIFGWGF	130
<i>T. tengcongensis</i>	FY-----KKFNIVYVAIVLTTLLNGPLATLLF-----VPMFGWGF	130
<i>T. thermosaccharolyticum</i>	AY-----KKFNISYVAVAVGTILNGPVATLIF-----VPQYGWGF	130
<i>A. metaliredigens</i>	AVIWMPLLIGSTVNIILIAASLYKIMGKGSVNGN-----	173
<i>A. oremlandii</i>	TALVIPLTVVAAVNVILAYIIFKVLPRK-----	167
<i>A. flavithermus</i>	LSIIPPLALATGVNLAIAVAVVSKALQRAWGKQ-----HA----	156
<i>B. coahuilensis</i>	FSLPLSLGIATVNVNIIILVILLPKIQWIYSHRFSGTFO-----	162
<i>C. hydrogenoformans</i>	TAMVVPLLVASFVNILLAGLTARALRGKTER-----	157
<i>C. botulinum</i>	LALLPILSLAALANVITAIIIYRFIPEKYFQR----DK-----	162
<i>C. perfringens</i>	KLLGLPLTLASLVNIVVAFIVSKGLKNANI-----	169
<i>C. sporogenes</i>	LAILPILSLAALANVITAIIIYRFIPEKYFER---NK-----	162
<i>C. tetani</i>	FALVPVLSGVAAINAVVAVLIYKFLPRSLKKY----ENK----	168
<i>G. kaustophilus</i>	FAVIPPLSAAAANVLIALAVMPVVVRLAAKAGVEAPHA---	162
<i>G. thermodenitrificans</i>	IAVIPPLSVATAMNIIIVAMVMPVSVVRLAAKVGKAPHA---	162
<i>H. modesticaldum</i>	MAVMPFLVAASAVNVGLAAAAYRGMKGIL-----	169
<i>L. gravesensis</i>	VALFLPLTIATVNLNIIICELVSAALPKRVRR--FKALKH----	168
<i>L. buchneri</i>	VALLPLTIATVNLNIIICELVSAALPKRVRR--FKALKH----	168
<i>L. hilgardii</i>	WAFVPLTIATVNLNIIICELVSAALPKRVRR--FKALKH----	170
<i>L. innocua</i>	FVLFVPLAIGSICNIFVAEVVYQVLPERWKRR--IAGY-----	164
<i>L. monocytogenes</i>	FVLFVPLAIGSICNIFVAEVVYQVLPERWKRR--IAGY-----	164
<i>L. welshimeri</i>	FVLFVPLAIGSICNIFVAEVVYQVLPERWKRR--IAGY-----	164
<i>L. sphaericus</i>	WGALGSIFATINLIIIVAVVMPILSKVFVRKAGRV-----	163
<i>M. thermoacetica</i>	LAMVPLLIIASALNIVLAATAFTSLRRVFPASYAAGRKGEGK	184
<i>N. thermophilus</i>	MGMVVPVTVASAVNIGLAVTAAPVCSQFLNRKPV-----	164
<i>S. thermophilum</i>	AAALPLTVAAAGANAVALLVVGLRRRAGVEG-----	218
<i>T. brockii</i>	AAWVPLTIASFANVFLAALVYKAIPKRSRE-----	161
<i>T. ethanolicus</i>	AAWVPLTIASFANVFLAALVYKAIPKRSRE-----	161
<i>T. italicus</i>	AAWVPLTIASFANVFLAALVYKAIPKRSRE-----	161
<i>T. mathranii</i>	AAWVPLTIASFANVFLAALVYKAIPKRSRE-----	161
<i>T. pseudethanolicus</i>	AAWVPLTIASFANVFLAALVYKAIPKRSRE-----	161
<i>T. tengcongensis</i>	ITWVPLTIASFANVFLAALVYKAIKILSKREQ-----	160

T. thermosaccharolyticum VQMVFPLTTIASFANVFLAAVIFKAISTKIKR----- 161

Figure S4. Alignment of CbIT homologs. ClustalW2 (Larkin *et al.*, 2007) alignment of predicted CbIT proteins from 29 bacterial species (Table S4). Conserved residues are shaded. Putative transmembrane helices are underlined and were predicted from the *L. innocua* Clip11262 CbIT protein sequence with the toppred algorithm (<http://mobyli.pasteur.fr/cgi-bin/portal.py?form=toppred>, accessed March 24, 2010) (von Heijne, 1992; Claros & von Heijne, 1994).

CblS	-----MPQVRDLSDVDVPG--GCVLTSCDISAGFGEKVH	32
PurM	MTDKTSLSYK DAGVDIDAGNALVGRIGVVKTRRPEVMGGLGGFGALCALPQKYREPV	60
ThiL	-----MACGEFSLIARYFDRVRSRRLDVELG--IGDDCALLNIPEKQTLAI	44
HypE	-----MQQLINSLFMEAFANFWLAEQED--QARLDAQLVAEGDRLAF	41
SelD	-MSENSIRLTQYSHGAGCGCKISPKVLETILHSEQAKFVD--PNLLVGNETRDDAAVYDL	57
CblS	DGLRVAPEVTARLTTLRVALLEMLASGAVVVAVS	92
PurM	VSGTDGVTGKLRRLAMDLKRHDTIGIDLVMCVNDL	118
ThiL	STDTLVAGN--HFLPDID-PADLAYKALAVNLS	99
HypE	STDSYVIDP--LEFFPGGN-IGKLAICGTAN--D	94
SelD	GNGTSVISTTDFEMPIVDNPFDFGRIAAATNAIS	117
CblS	GHIELNGSTEENMNVTQTSVG-----VLVTGFATKAALKLINVHEA	133
PurM	ASAVISGIAEGCLQSGCSLVGGETAEMPGMYHGEDYDVAGFCVGVVEKSEIIDGSKVSDG	178
ThiL	LESFSDSLFDLLNYYDMQLIGGDTTRGP-----LSMTLGIHGFPVPMGRALTRSGAKPG	152
HypE	LKAVVTSMAETARAAGIAIVTGDTKVVQRGAV-DKLFINTAGMGAI PANIHWAQTLTAG	153
SelD	AREVTEGGRYACRQAGIALAGHSIDAP-----EPIFGLAVTGIVPTERVKKNSTAQAG	171
CblS	AVLFAFG---EPIVGAEVLQRMEMPDPYPLVKQLVSD-----	167
PurM	DVLIALGSSGPHSNGYSLVRKILEVSGCDPQTELDGKPLA--DHLLAPTRIYVKS	235
ThiL	DWLYVTGTGPGDSAAGLAILQNRLLQVADAKDADYLIKRRHLP--SPRILQG-QALRDLANS	209
HypE	DVLLVSGTLGDHGATILNLRQLGLDGLVSDCAVLTPLIQ--TLRDI PGVKALRDATR-	210
SelD	CKLFLTKPLGIGVLTAEKKSLLKPEHQGLATEVVMCRMNIAGASFANIEGVKAMTDVTG-	230
CblS	-----SRVLEVVPVSGKGMAYEANTLARLNDCVFEASG-----	200
PurM	-----LIEKVDVHAIAHLTGGGFWEENIPRVLPDNTQAVID-----ESSWQWPEVFNWL	283
ThiL	AIDLSDGLISDLGHIVKASDCGARIDLALLPFSDALSRHVEP---EQALRWALSGGEDYE	266
HypE	-----GGVNAVVEFAAACCGGIELSEAAPVPAVRGVCEL---LGLDALNFANEGKLV	262
SelD	-----FGLLGHLSSEMCQAGVQARVDYEAIPKLPGVVEEYIKLGAVPGGTERNFASYGHLM	285
CblS	-----VFNEATMNKTAG-----PASVILVAVKASEVKAFEQNFPAKCLGELRNYHG--	247
PurM	QTAGNVEHHEMYRTFNCGVGMIIALPAPEVDKALALLNANGENAWKIGI IKASDSEQRVV	343
ThiL	LCFTVPELNRGALDVALG---HLGVPTFCIGQMTADIEGLCFIRDGEFVTLDWKGYDHFA	323
HypE	IAVERNAAEQVLAALHSH---PLGKDAALIGE VVERKGVRLAGLYGVKRTLDLPHAEPLP	319
SelD	GEMPREVRDLLCDPQTSGG-LLAVMPEAENEVKATAAEFGIELTAIGELVPARGGRAMV	344
CblS	---	
PurM	IE- 345	
ThiL	TP- 325	
HypE	RIC 322	
SelD	EIR 347	

Figure S5. Alignment of *LinCblS* with PurM ATP-binding superfamily proteins. ClustalW2 (Larkin *et al.*, 2007) alignment of *L. innocua* CblS with PurM, ThiL, HypE, and SelD from *E. coli* MG1655. Conserved residues are shaded.

<i>A. metaliredigens</i>	-----M	1
<i>A. oremlandii</i>	-----	
<i>A. colihominis</i>	MCYDNAIEQAAQPPKIDSKEGVPMTMPNQESLWTDYQELLEMTROCGFSS	50
<i>A. flavithermus</i>	-----	
<i>B. halodurans</i>	-----	
<i>C. hydrogenoformans</i>	-----M	1
<i>C. botulinum</i>	-----M	1
<i>C. perfringens</i>	-----M	1
<i>C. sporogenes</i>	-----MYTRYKRKEVSFM	13
<i>C. tetani</i>	-----M	1
<i>D. hafniense</i>	-----	
<i>G. kaustophilus</i>	-----	
<i>G. thermodenitrificans</i>	-----	
<i>H. modesticaldum</i>	-----	
<i>L. gravesensis</i>	-----	
<i>L. buchneri</i>	-----	
<i>L. hilgardii</i>	-----	
<i>L. innocua</i>	-----	
<i>L. monocytogenes</i>	-----	
<i>L. welshimeri</i>	-----	
<i>L. sphaericus</i>	-----	
<i>M. thermoacetica</i>	-----MTPATL	6
<i>N. thermophilus</i>	-----	
<i>P. acnes</i>	-----MTAPNRVPSRCL	12
<i>S. thermophilum</i>	-----	
<i>T. brockii</i>	-----	
<i>T. ethanolicus</i>	-----	
<i>T. italicus</i>	-----	
<i>T. mathranii</i>	-----	
<i>T. pseudethanolicus</i>	-----	
<i>T. tengcongensis</i>	-----	
<i>T. thermosaccharolyticum</i>	-----	
<i>A. metaliredigens</i>	EIKRCRDLTLIERAGQPNLVIACDSSGGIGEKPDQIKVPAE VVGYFTAR	51
<i>A. oremlandii</i>	-MFKFRDLTVIDIPPNHRMIIACDSSGGIGNKKHDVVQAEPE TLGYFTA H	49
<i>A. colihominis</i>	RIRKYRDL SILRLGDISLVVACDSNASNGEKPNDTHQNSYEETA VSA LK	100
<i>A. flavithermus</i>	---MRDVLVLPDEENELVLA TDCSSGGI GLKQDDVVNVVPYDVVAYYGAR	46
<i>B. halodurans</i>	-----MATDASGGVGEKAKD HVHVPYEVVSYAAR	30
<i>C. hydrogenoformans</i>	AGFKYRDLTIVPLSRE-KMVI AADSIGGVGPKAGDIVKTSGEIVGRYGAR	50
<i>C. botulinum</i>	NVKKVRDLT LISLDKDKTLVVACDSSSGISGSKNDILKIPAFYTGKFTIR	51
<i>C. perfringens</i>	QIYKFRDLTVLENEKN-KLVIACDSSGGIGENEGDFVKASNEIVSYF SAR	50
<i>C. sporogenes</i>	NVKKVRDLT LISLDKDKTLVVACDSSSGISGPKNDILQIPAFYTGKFAIR	63
<i>C. tetani</i>	RISKVRDLTLIKLTEDKNLVVACDSSGGIGSKPEDALKVPAIVGKLTAR	51
<i>D. hafniense</i>	MGYQGRDVEVVALNDAQYLVAACDSCGAI GEKELDAVKVPWRVTGRMTAR	50
<i>G. kaustophilus</i>	---MRDVLFLFPADGME LAVAADGSAAVGDKPGD VVSVPDVVAYF SAR	46
<i>G. thermodenitrificans</i>	---MRDVLFLPLTDGVELAIAADGSAAVGEKQGD AVFVPAETTAYFAAR	46
<i>H. modesticaldum</i>	MGYRGRDVEVVALSPEQCLVVA CDACGAI GAKELDAVQVSPYIVGIF TTR	50
<i>L. gravesensis</i>	--MKFRDLTIKPI SDKTALVIACDVSAGI GEKPD DLVHVTADVTA AFALR	48
<i>L. buchneri</i>	--MKFRDLTIKPI SDKTALVIACDVSAGI GEKPD DLVHVTADVTA AFALR	48
<i>L. hilgardii</i>	--MKFRDLTIKPI SDKTALVIACDVSAGI GEKPD DLVHVTADVTA AFALR	48
<i>L. innocua</i>	-MPQVRDL SVIDVPGG-CVLTS CDISAGFGEKVH DGLRVAPEVTARLTLR	48
<i>L. monocytogenes</i>	-MPQVRDL SVIDVPGG-CVLTS CDISAGFGEKTH DGLMVAPEVTARLTLR	48
<i>L. welshimeri</i>	-MPQVRDL SVIDVPGG-CILTS CDISAGFGEKAH DGLVVTPEVTARLTLR	48
<i>L. sphaericus</i>	-----MKVGAFIATMDNAAAI GEKPD IVPASDQLTAYMTAR	37
<i>M. thermoacetica</i>	SPRRYRDLTILDLD AQRSLVIACD SAGAI GPKEADVVRVPGYV LGRFTAR	56
<i>N. thermophilus</i>	MIFQLDDVKMIKMSKNEYLAVACD SLGGI GEKSLDKVKVPCEWVGRV LTR	50
<i>P. acnes</i>	SITRLRDL LLLADIPAS-RLVIACDT IGGIGPRPDDSYPADPVWCAHLGAR	61
<i>S. thermophilum</i>	--MRWRDLTLVDLP CGGRLVIACDAAGGI GPKERD VIRVAGYVIGRFTAR	48
<i>T. brockii</i>	MIERYRDLVVIYEN-DVAYVIS CDISLGAIGNKEHDV LKVDEEIVGRTTVK	49
<i>T. ethanolicus</i>	MIERYRDLVVIYEN-DVAYVIS CDISLGAIGNKEHDV LKVDEEIVGRTTVK	49
<i>T. italicus</i>	MIERYRDLVVIYEN-DVAYVIS CDISLGAIGNKENDV LKVDEEIVGRTTVK	49
<i>T. mathranii</i>	MIERYRDLVVIYEN-DVAYVIS CDISLGAIGNKENDV LKVDEEIVGRTTVK	49
<i>T. pseudethanolicus</i>	MIERYRDLVVIYEN-DVAYVIS CDISLGAIGNKEHDV LKVDEEIVGRTTVK	49
<i>T. tengcongensis</i>	MIERYRDLVVIYEK-DVAFVVS CDISVGAIGSKENDI LKVDEEIVGRTTVK	49
<i>T. thermosaccharolyticum</i>	MVERYRDLVVIYES-DTVYAIACD SIGAIGNKEGD I LKVDEEIVGRTTVK	49
<i>A. metaliredigens</i>	VSLMEVMSV GARVMTVINTLSVEREPTGEKMIKGIQKMIEEVKLP----	96
<i>A. oremlandii</i>	VALMELLATGATPLTVVNTLGVEMEDSGVRIIEGIQKALEPLNLK---ED	96
<i>A. colihominis</i>	VPLMEVLATGAAPIVIADNLCVEMEPGRRIIAAMQEELRGCGLYD----	146
<i>A. flavithermus</i>	VAWMELMSIGATPKAFVLQNFVN-DDAWHALVAGVQQTMEELQLSLP---	92
<i>B. halodurans</i>	VALLECMSVGATPFTFLVQNFSG-DEPYQKMDGINRALGEANQAQAT--	77
<i>C. hydrogenoformans</i>	VVLMELLAVKARQVIVSCTVSCWEPTGKEIYQGVLAEVKTL PFA-----	95
<i>C. botulinum</i>	VGILEVMCTGAEIVTNTALCCEMNP TGREIIDGKIGELKRAGID----	96
<i>C. perfringens</i>	VCLFELLAFRAKPLVIVNNLGM SMNNGEKKIIQGINRAIKEYNAENFFEE	100

<i>C. sporogenes</i>	VGILEVMCTGAEIVTVTNAVCCMNPTGKEIIDGKIGELKCRAGID-----	108
<i>C. tetani</i>	VALMEVLCTGAEIVTITDAVCNEMPTGKEIIRGIKEELKEAKIN-----	96
<i>D. hafniense</i>	VALLEVLAVGAVPQMLSAIANEFLPAGEEIMKGVREELKAMNLL-----	95
<i>G. kaustophilus</i>	VALMELLVSGAEARVVVLQNFIA-DDRWEALCRGVRRRCRELGDLP----	92
<i>G. thermodenitrificans</i>	VALMELVSVGAEKAVVLQNFIA-DERWEALCRGIRQAGSELGLDLP---	92
<i>H. modesticaldum</i>	VALMEILATGAEFPVALTVTVANEPFPTGGEVIAGVRDELATIGKR-----	95
<i>L. gravesensis</i>	VPLMELLCFGATPISVVDTVGNEMPTPTGENMIAGLKQELKRAGLS-----	93
<i>L. buchneri</i>	VPLMELLCFGATPISVVDTVGNEMPTPTGENMIAGLKQELKRAGLS-----	93
<i>L. hilgardii</i>	VPLMELLCFGATPISVVDTVGNEMPTPTGENMIAGLKQELRRGGLS-----	93
<i>L. innocua</i>	VALLEMLASGAVVAVSDVIGAEMEPTGKRVIAGLKDELFKADLG-----	93
<i>L. monocytogenes</i>	VALIEMIASGAEVAVSDVVGEMEPTGRRVIVGLKDELTKANLS-----	93
<i>L. welshimeri</i>	VALLEMIASGADVVSVDVGAEMEPTGRRIIIGLKEELSKANLN-----	93
<i>L. sphaericus</i>	VTFLQALAAQALPIQILLANFSG-DAAWSRYERGIQQVFEETGLPCPV--	84
<i>M. thermoacetica</i>	VALMEVLALGAWPVCVNTLCVEPEPAGAAIREGVADEMRVLGIDPEK--	104
<i>N. thermophilus</i>	VAVMELLALRIDPFLVINTLANEMVPTGEQIIISGIKEELDTVEIDP----	96
<i>P. acnes</i>	VPLLEVLFCAGARPLVLDLTCQDSASAQPMIAE-FRRCALDAGIDPD----	107
<i>S. thermophilum</i>	VALMDLLAAGAQLPHLVNNTCEVPDPTGREILQIGICDEAALAGLSAD---	95
<i>T. brockii</i>	VALSEVLCVGAKPLVISDTLSVEMNPTGQKILRGIKSELEENGLS-----	94
<i>T. ethanolicus</i>	VALSEVLCVGAKPLVISDTLSVEMNPTGQKILRGIKSELEENGLS-----	94
<i>T. italicus</i>	VALSEVLCVGAKPLVISDTLSVEMNPTGKILKGIKNELEDNGLL-----	94
<i>T. mathranii</i>	VALSEVLCVGAKPLVISDTLSVEMNPTGKILKGIKNELEDNGLL-----	94
<i>T. pseudethanolicus</i>	VALSEVLCVGAKPLVISDTLSVEMNPTGQKILRGIKSELEENGLS-----	94
<i>T. tengcongensis</i>	VALSELLSVGATPLVSDTLSEMYPTGKILKGIKKELEENGL-----	93
<i>T. thermosaccharolyticum</i>	VAVSELLCIGAWPIIISDTLSNEMNPTGIKIIGGIKKELENDNEIY-----	94
<i>A. metaliredigens</i>	----ITALNGSTEEENVVTCQTAMGITVIGEVERESIKIGCSK-----	134
<i>A. oremlandii</i>	----IVVTGSTEENIPVSQTSMGITIIIGMMEKSRWRAQKVE-----	133
<i>A. colihominis</i>	----AVSFTGSTEDNMRTLQGTIGVTVIGLVSGASLRRLGRTO-----	184
<i>A. flavithermus</i>	-----ITGSSSENMPMQSAVGFVAVGTVRKTEKRINVTP-----	127
<i>B. halodurans</i>	-----IAGSTSENMVLEQSCLGVTVIGRVHQSALKIGITP-----	112
<i>C. hydrogenoformans</i>	-----IDITGSSSEKNFTTNTTGAGFTAVGLGDELLVNRICET-----	132
<i>C. botulinum</i>	----EVLVTGSTEENFPFSFSTGLGITVIGVIVDNSVIKVNNSVNNFGNNEK	142
<i>C. perfringens</i>	KSNLSECVTGSTEDDNFKTLQTFGLGTTIIGEKEET--LKVNF-----	140
<i>C. sporogenes</i>	----EVLVTGSTEENFTTFTSTGVGIVTGLMVDNSVIKVNNSVNNFGNNEK	154
<i>C. tetani</i>	----EVLVTGSTEENFPFKATGLGVTVIGVIVDNNMKNVNNIT-----	134
<i>D. hafniense</i>	----SLPAAVSTENKMPTEQTLGITVIGLQDQDKLRIGRAR-----	133
<i>G. kaustophilus</i>	-----ITGSTEENFPVQSAVGVAIGTVANERKRIGITP-----	127
<i>G. thermodenitrificans</i>	-----ITGSTEENFATVQSAVGVAIGTVAHGQKRIGITP-----	127
<i>H. modesticaldum</i>	----DLPLAISTEENKPIPTRQTGVTITLVGAVEKERLRVGCSSR-----	133
<i>L. gravesensis</i>	----DISLNGSTEDNMPTRTTSIGVTVIGIATRVDLDFQ-K-----	130
<i>L. buchneri</i>	----DISLNGSTEDNMPQTTSIGVTVIGIATRVDLDFQ-K-----	130
<i>L. hilgardii</i>	----DISLNGSTEDNMPQTTSIGVTVIGIATRVDLDFQ-K-----	130
<i>L. innocua</i>	----HIELNGSTEEENMNVTQTSVGVVLTGFATKAALKLINVH-----	131
<i>L. monocytogenes</i>	----HIELNGSTEEENMKVTQTSVGVVLTGFATKAALKLINVH-----	131
<i>L. welshimeri</i>	----HIELNGSTEEENMKVTQTSVGVVLTGFATKAALKLINVH-----	131
<i>L. sphaericus</i>	-----IAGSSSENMPQLQSLAITMLGEIQQ--RKAWDL-----	116
<i>M. thermoacetica</i>	-----TLTGSSSEKNIPTTQSGIGITVIGLATTAEELMGRLA-----	140
<i>N. thermophilus</i>	----EISLTGSCETNIVTDQGTAGVTVLQGS-PEFPGWGYSY-----	133
<i>P. acnes</i>	----AVTGSTEDNVATQGTGVGVTIIGVMHHEDEVPK--SL-----	141
<i>S. thermophilum</i>	-----QINGSTSEKNIPTVQTLGVTVAIGYLAPGRSLR-TAR-----	130
<i>T. brockii</i>	----DVVFTGSTEENFPSTITGIGITVIAKANVCDLKIKKVK-----	132
<i>T. ethanolicus</i>	----DVVFTGSTEENFPSTITGIGITVIAKANVCDLKIKKVK-----	132
<i>T. italicus</i>	----DVVFTGSTEENFPSTITGIGITVIAKAYVCDLKIKKVK-----	132
<i>T. mathranii</i>	----DVVFTGSTEENFPSTITGIGITVIAKANVCDLKIKKVK-----	132
<i>T. pseudethanolicus</i>	----DVVFTGSTEENFPSTITGIGITVIAKANVCDLKIKKVK-----	132
<i>T. tengcongensis</i>	----EVLVTGSTEENFPSTITGIGITAVGRARKEDLKIKKAR-----	131
<i>T. thermosaccharolyticum</i>	----DVALTGSTEENFPSTMTGVTVAIGKAEKALKVRKAM-----	132
<i>A. metaliredigens</i>	-----PGDLIVALGTEPKVGNIEK--LPI-DDEICSIKDFQALVKMKNVK	175
<i>A. oremlandii</i>	-----KGDVALVVGIPKVGQEVLEDEGR--EIMSIPLLLELLKKSIIH	174
<i>A. colihominis</i>	-----RGDAVYCAQVPSQSVGLERYSEHDP--SVAKISTVTRLCALDYIH	226
<i>A. flavithermus</i>	-----TDACWAVIGEPVGEAVIQKKDR---IIPLSLFRLLQLDGIY	167
<i>B. halodurans</i>	-----KDAGIALLGKPLVGEVVSQEKD---VFPLKQFDELLEQEEGVY	152
<i>C. hydrogenoformans</i>	-----SDLYIVGVPVGEVVKFPPELQ---ATPHLVYELSQNNDVL	170
<i>C. botulinum</i>	HNK---DDILLISVGIKPVGEEIN--IYD-DKEIVDYEDIKILLENPKVY	186
<i>C. perfringens</i>	-----KGDSICLLGIPKVGQEVLEDINNNLGEIVTFKDFKILMDNKDVK	184
<i>C. sporogenes</i>	HNKFDLDDVLLISVGIKPLGKEIN--IYD-DKEIVDYVDIKILLENPRVY	201
<i>C. tetani</i>	-----KDCLLISIGIPKVGKEID--IFGYDEEIASYNNISTILNNQEAY	176
<i>D. hafniense</i>	-----PGNSLFCGLGPKVGTAEADPEDP--DILQGIHLIQLLNIPGVY	174
<i>G. kaustophilus</i>	-----EEAKFAVIGRPLVGSVAVLSHPPEW---VAPLPLFAALLRTPYIY	167
<i>G. thermodenitrificans</i>	-----ETAKFAVIGRPLVGSVAVLVHSDW---IAPLSLVAELLASPVYVY	167
<i>H. modesticaldum</i>	-----PGDCVYSLGWPVGAEVVVDDGK---AIARADHVQALLAHPGVH	174
<i>L. gravesensis</i>	-----QPMVIYQLGRPLVG--DAVKQHF--DLFSYNLINELRSDSAVI	170
<i>L. buchneri</i>	-----QPMVIYQLGRPLVG--YAVKQHF--DLFSYNLINELRSDSAVI	170

<i>L. hilgardii</i>	-----QPMVIYQLGRPLVGV--YAVKQHFN--DLFSYNLINELRSDSAVI	170
<i>L. innocua</i>	-----EAAVLFAFGEPIVGV--AEVLQRM--EMPDYPLVKQLVSDSRVL	171
<i>L. monocytogenes</i>	-----EAAVLFAFGEPIVGV--AEVLQKLR--DMPDYRLVKKLVEHNDVL	171
<i>L. welshimeri</i>	-----EAAVLFAFGEPIVGV--EEVLQKMT--EMPDYHLVKKLVADSSVL	171
<i>L. sphaericus</i>	-----EQLSWYTYGLPLVGEELLAQPDD---VAQLQPIYQAWLAEIVQ	156
<i>M. thermoacetica</i>	-----AGDTLALFGRPKVGV--TEVFLDD--PEIVDLKTVRLLDQPGIR	180
<i>N. thermophilus</i>	-----PGIALYMTGTPKVGAEVDLND-P---EIMDLQTMKELSEINGVL	173
<i>P. acnes</i>	-----DGDVLCVCGAPISAPNDDVALGR--REIVGVTEVKALMASGVKH	183
<i>S. thermophilum</i>	-----PGDLVVAIGRPKVGV--AEVRLDD--PELPDLPLVRRLAGDPLVH	170
<i>T. brockii</i>	-----AGMHVSLGYPKRVGSEVLGAK-----DVLNLSYDIKISNSKEIV	171
<i>T. ethanolicus</i>	-----TGMHVSILGYPKRVGNEVLSSN-----DVLTLKDYVKISNSKEIV	171
<i>T. italicus</i>	-----AGMHVSLGYPKRVGSEVLSSK-----DVLTLKDYIKISNSKEII	171
<i>T. mathranii</i>	-----AGMHVSLGYPKRVGSEVLNSK-----DVLTLKDYIKISNSKEII	171
<i>T. pseudethanolicus</i>	-----AGMHVSLGYPKRVGSEVLGAK-----DVLNLSYDIKISNSKEIV	171
<i>T. tengcongensis</i>	-----KGMHVVLIGYPKRVGSEVLGAE-----DVMTLKDYIKISQTKEVV	170
<i>T. thermosaccharolyticum</i>	-----AGMEVGLFYGYPKRVGQEVLCCH-----DILSLKDYVKIFRCGEIV	171
<i>A. metaliredigens</i>	DIHPIGSKGMYEBAQLLASLNHCEFKSREA-TGVD--LKKSAGPATAVIF	222
<i>A. oremlandii</i>	DIIIPVGSKGIAYEVGQMAESNGIGYNLY-EHVGID--LNKSAGPATCVIV	221
<i>A. colihominis</i>	EIIPVGSKGAAYEAGQLAECVCGTFAAD-AQPPID--LATSAGSCTAVLV	273
<i>A. flavithermus</i>	EIIPVGSKGISHEBWKLMYGE-----RPLRCS--LP--PHASAGPATCVLV	208
<i>B. halodurans</i>	EIIPVGSKGVKYLEVLAATNEWNTLQVPVNGNVD--LTKSSGPTSLLI	200
<i>C. hydrogenoformans</i>	EIIPVGSKGVGABLMWLNKQG---FEIENVIAPPFS--LEKSAGPGTSLLV	216
<i>C. botulinum</i>	EIVPVGSKGIILYEGEILARNNNLKLKLEEN-ISID--IKKSNCPSTCLIF	233
<i>C. perfringens</i>	DIIPIGSKGIHIEISELENSDKIRINL--SYEGDY--LRKSAGPATALLF	230
<i>C. sporogenes</i>	EIVPVGSKGIILYEGEVLAKNNLKLKLEEN-IPID--IKRSNCPATTVIA	248
<i>C. tetani</i>	EIVPVGSKGIILFAEELAKNNRCEFYLNKDKIDVD--IRRSAGPATVIA	224
<i>D. hafniense</i>	DIIIPVGSQGIIRGEAEALAQAVGARFGE-DSQCRLD--INKSAGSCTAVLV	221
<i>G. kaustophilus</i>	ELIPIGSKGIYYEWTQLLAANGRQGRACACP--LP--LFSSGCPATSVLV	213
<i>G. thermodenitrificans</i>	ELIPIGSKGIYYEWTQLLAANGRQWCACACP--LP--LFASGCPATSLLI	213
<i>H. modesticaldum</i>	EIVPVGSKGIIRABAEKLAADVHCRICEWQSVAGLD--LDKSAGPSTCLVF	222
<i>L. gravesensis</i>	DMLPVGSKGIAFIEINQLAKTHQLKVVDGSSVMETEE--MVKSAGPATVALI	218
<i>L. buchneri</i>	DMLPVGSKGIAFIEINQLAKTHQLEIVDSSVMETEE--MAKSAGPATVALI	218
<i>L. hilgardii</i>	DMLPVGSKGIAFIEINQLAKTHQLEIVDSSVMETEE--MAKSAGPATVALI	218
<i>L. innocua</i>	EIVPVGSKGMAYEANTLARLNDVCFEASGVFNAT--MNKTAGPASVILV	219
<i>L. monocytogenes</i>	EIVPVGSKGMAYEANTLARLNGGIFVPSDTFSEAV--MNKTAGPASVILA	219
<i>L. welshimeri</i>	EIVPVGSKGIILYEGEINLAKLNNRVFKETSSLDEIS--LNKTAGPASVIL	219
<i>L. sphaericus</i>	QVWVPGSKGLQGEICARLFGQ-----QLVECS--HD--MTKSAGPSTVILL	197
<i>M. thermoacetica</i>	EIVPAGSRGILABARDLAALYGLQINWRPGLTGPA--LLKSAGPATCILA	228
<i>N. thermophilus</i>	EIIPVGSQGLLKESENTLANRSELRLQLNNAIKTDEGLLTKSAGPSTCVIF	223
<i>P. acnes</i>	DCVPVGSKGVAVAEALQASTAGLRVAFQP--TDVD--LTRSGCPSTCLVM	229
<i>S. thermophilum</i>	DLLPVGSRGIRABAEEDLAASAGLEVEWAPAEAGFP--LGKSAGPVTCLLV	218
<i>T. brockii</i>	EAIIPVGSKGVKYLEGILEKISGLKVEA-NFPQHLD--VLKSGCPSTCCLV	218
<i>T. ethanolicus</i>	EAIIPVGSKGVKYLEGILEKISGLKVEA-NFPQHLD--VLKSGCPSTCCLV	218
<i>T. italicus</i>	EAIIPVGSKGIKYEGILEKISGLKVEA-NFPQYLD--VLKSGCPSTCCLV	218
<i>T. mathranii</i>	EAIIPVGSKGIKYEGILEKISGLKVEA-NFPQYLD--VLKSGCPSTCCLV	218
<i>T. pseudethanolicus</i>	EAIIPVGSKGVKYLEGILEKISGLKVEA-NFPQHLD--VLKSGCPSTCCLV	218
<i>T. tengcongensis</i>	EAIIPVGSKGIAYELGVLEDLYGFRIKE-DDGLNID--LFKSAGPATCCLV	217
<i>T. thermosaccharolyticum</i>	EAIIPVGSKGIKHELDVLLKSSGLEFLK-EYKSLD--DTKSGCPSTCCLV	218
<i>A. metaliredigens</i>	SVSKE-QLPT--VESQLQQQVKVIGSLEN-----	248
<i>A. oremlandii</i>	AVNQE-NYEA--LKESMPIPVHVLVGSFT-----	246
<i>A. colihominis</i>	SLPLQ---AGGRLAADMDVPCFLIGRIQ---	298
<i>A. flavithermus</i>	SYHQK---QEPLQLSLAGTLFYPIVNEL---	233
<i>B. halodurans</i>	TFDGT---MEALKAKYSSWTTTLASKIST-----	227
<i>C. hydrogenoformans</i>	AVKEG-----IELKAAIPVHYLGKAIKVKKT-----	242
<i>C. botulinum</i>	AIHKE-EYEN--IRAKINN-VNIIGKLER-----	258
<i>C. perfringens</i>	ILRNN-SLED--IKNKIKTPIMEIGNII-----	255
<i>C. sporogenes</i>	AIHKE-EYEN--IRAKINN-VNIIGELESV-----	274
<i>C. tetani</i>	AVSKK-AYEN--LKD-IQN-INLLGEIKKGT-----	250
<i>D. hafniense</i>	TAED-----IELGDFGKLPiHKIGRLEEDGGGGLLTENED	257
<i>G. kaustophilus</i>	SYDPD---GEREIKKQAGSLFFPLHVEW-----	238
<i>G. thermodenitrificans</i>	SYDPA---GEQVLRKQAGRLFFSLFAEL-----	238
<i>H. modesticaldum</i>	TAGP-----KPPEIAGAPIHYLGRLESAG-----	247
<i>L. gravesensis</i>	GVKPE---KRRVFERHFQVSYLSMLT-----	242
<i>L. buchneri</i>	GVKPE---KRRVFERHFQAHYLLSLN-----	242
<i>L. hilgardii</i>	GVKPE---KRRVFERHFQAHYLLSLN-----	242
<i>L. innocua</i>	AVKAS---EVKAFEQNPPAAKCLGELRNYHG-----	247
<i>L. monocytogenes</i>	AVKED---NVTAFEREFGLGAKRIGLLRGNEEW-----	249
<i>L. welshimeri</i>	AVKKR---DCSIFEKHFSGAQYLGELCGNEEQI-----	249
<i>L. sphaericus</i>	GMDPE---KEQLAHQFFQRNFEKLRISAE-----	223
<i>M. thermoacetica</i>	AGDRA-ALETTGRR--EKPFCLLGTLGPAASR-----	258
<i>N. thermophilus</i>	TADHDGADQAQKLAQVSGKMKMTQLGRLY-----	251
<i>P. acnes</i>	ACAKT-NLEQLHALVNPERPVAWIGHLRS-----	257

<i>S. thermophilum</i>	AAAPS-ALQGLALTL--TQPWAVVAQLR-----	243
<i>T. brockii</i>	VHNEE---DTVSIKGLTDKPLTYVGVLI-----	243
<i>T. ethanolicus</i>	VHSEE---DTASIKGLTDKPLTYVGVLI-----	243
<i>T. italicus</i>	VHNEE---DTVSIKGLTDKPLTYIGVLI-----	243
<i>T. mathranii</i>	VHSEE---DTVSIKGLTDKPLTYIGVLI-----	243
<i>T. pseudethanolicus</i>	VHNEE---DTVSIKGLTDKPLTYVGVLI-----	243
<i>T. tengcongensis</i>	VYREE---DTDFIKAITDKPFTHVGIIE-----	242
<i>T. thermosaccharolyticum</i>	VYKEG---DRRKIENLLDKPFVRLGRLIDGR-----	246

Figure S6. Alignment of CbIS homologs. ClustalW2 (Larkin *et al.*, 2007) alignment of predicted CbIS proteins from 32 bacterial species (Table S5). Conserved residues are shaded.

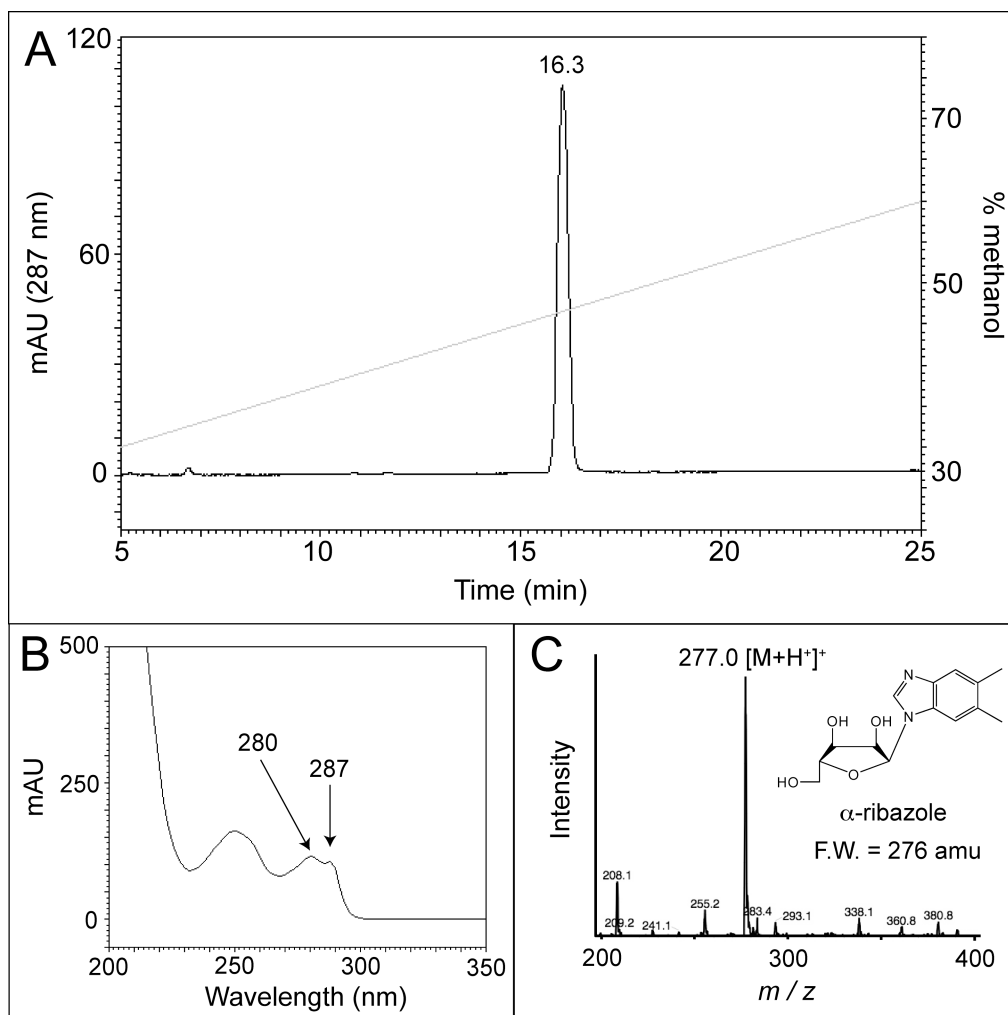


Figure S7. Purification of α -R. Purified α -R was resolved by RP-HPLC using a Beckman Coulter System Gold® 126 HPLC system equipped with a Phenomenex 150 x 4.6 mm Synergi 4 μ Hydro-RP column. Products were detected by their absorbance at 287 nm using a photodiode array detector. The column was equilibrated at 1 ml min⁻¹ with 30% methanol. 5 minutes after injection, the column was developed for 20 min with a linear gradient to 60% methanol, then developed for 5 min with a linear gradient to 100% methanol. Panel A shows elution of a product at 16 min. The grey line (right axis) indicates the methanol gradient used to elute α -R. Panels B and C show the UV-visible absorbance and mass spectra of α -R, respectively. The structure of α -R is indicated in the inset to panel C.

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