

## **Preparation of BEC28 VLPs**

Total RNA was extracted with QIAamp Viral RNA Mini Kit (Qiagen) from 170  $\mu$ L of intestinal content sample from an Italian calf infected with BEC strain BEC/28IT identified in a survey performed in Northern Italy. The poly(A)-containing RNA was subjected to reverse transcription using the oligo(dT)<sub>20</sub> and SuperScript® III First-Strand Synthesis System (Invitrogen), following the manufacturer's instructions. Amplification of the entire capsid region by PCR was performed by a first PCR with primers J11U (5'-CCATCAACCATTGGATTTTGAC (1) and FwVP (5'- ataagaatg cgg ccg cgt aaa tga aga tga ctg ac-3'), followed by a second PCR, semi-nested, with primers FwVP and RORF2 (5'- Cggggt accgaattcagaagccatcaag-3') containing *NotI* and *KpnI*, respectively. The DNA fragment obtained, flanked by restriction enzyme sites *NotI* and *KpnI*, was ligated into the transfer vector, pFastBac™ 1 (Invitrogen) digested with the same enzymes. The bacmid with the entire BEC ORF2 (called BACBEC28, Acc # GQ397857) was transfected into the Sf9 insect cell line and the high titre BACBEC28 virus stock solution obtained was used to purify assembled VLPs, as confirmed by EM visualization. Sf9 cells were infected with high titre BACBEC28 and incubated at 27°C. Seven days postinfection, when a diffuse cytopathic effect was observable, cells and supernatant were collected. The VLPs were purified from cell culture supernatant and cell lysate by ultracentrifugation through a 30% sucrose cushion (wt/vol), followed by CsCl density gradient (1.362 g/cm<sup>3</sup>) at 30.000 rpm for 24 hours at 12°C.

1. **Smiley, J. R., A. E. Hoet, M. Traven, H. Tsunemitsu, and L. J. Saif.** 2003. Reverse transcription-PCR assays for detection of bovine enteric caliciviruses (BEC) and analysis of the genetic relationship among BEC and human caliciviruses. *J. Clin. Microbiol.* 42:5214-5224.

**Figure A1:** Dendrogram, drawn using Bionumerics software packages (Applied Maths, Belgium) method UPGMA, was based on the entire ORF2. GenBank accession no., origin, and genotype are reported for all strains. Strains involved in this study are indicated in boldface type.

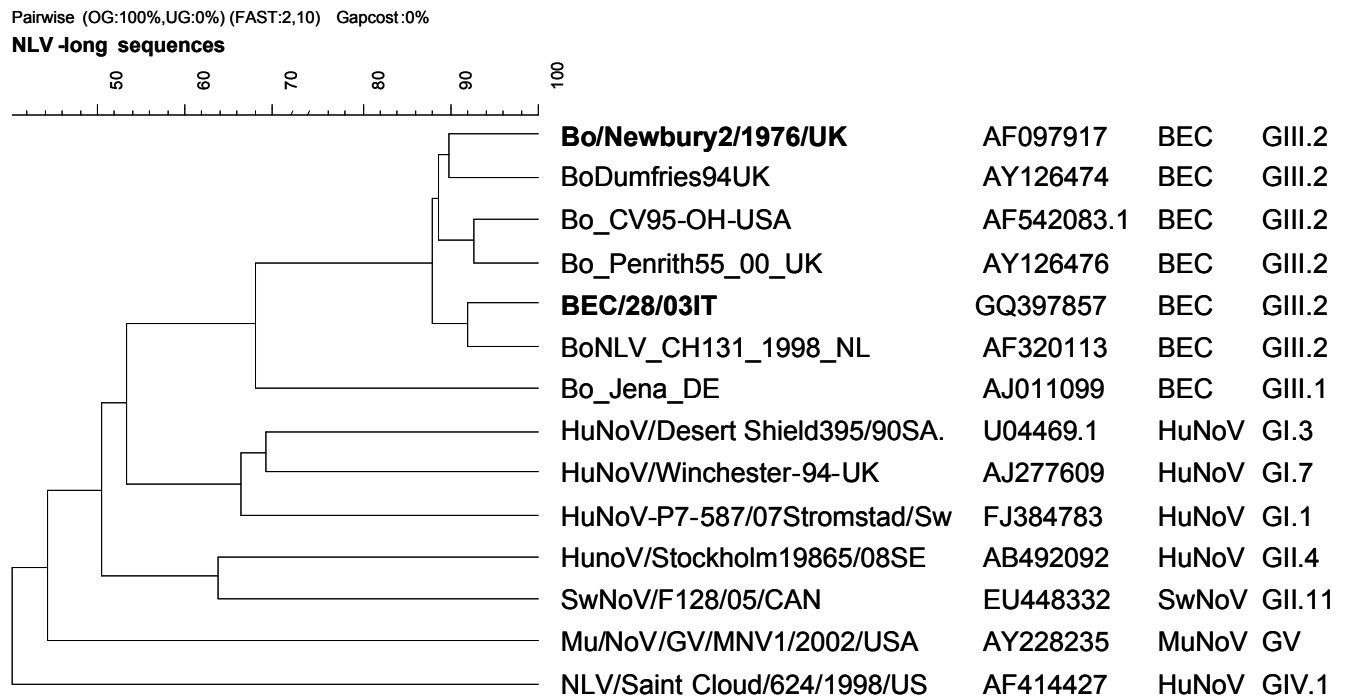


Table A1. Neoglycoconjugates used to determine the carbohydrate specificity of BEC28

VLPs

Trivial name	OLIGOSACCHARIDE STRUCTURE <sup>A</sup>
Tn	GalNAc $\alpha$ -R1
$\alpha$ -galactose monosaccharide	Gal $\alpha$ -R1
$\alpha$ -fucose monosaccharide	Fuc $\alpha$ -R1
A disaccharide	GalNAc $\alpha$ 3Gal $\beta$ -R1
B disaccharide	Gal $\alpha$ 3Gal $\beta$ -R1
	Gal $\alpha$ 2Gal $\beta$ -R1
	Gal $\alpha$ 6Glc $\beta$ -R1
Core 5	GalNAc $\alpha$ 3GalNAc $\alpha$ -R1
H disaccharide	Fuc $\alpha$ 2Gal $\beta$ -R1
Forsmann disaccharide	GalNAc $\alpha$ 3GalNAc $\beta$ -R1
Core 8	Gal $\alpha$ 3GalNAc $\alpha$ -R1
Type 2 precursor	Gal $\beta$ 4GlcNAc $\beta$ -R1
T $\alpha\beta$	Gal $\alpha$ 3GalNAc $\beta$ -R1
A trisaccharide	GalNAc $\alpha$ 3(Fuc $\alpha$ 2)Gal $\beta$ -R1, R2
B trisaccharide	Gal $\alpha$ 3(Fuc $\alpha$ 2)Gal $\beta$ -R1, R2
H type 1	Fuc $\alpha$ 2Gal $\beta$ 3GlcNAc $\beta$ -R1
H type 2	Fuc $\alpha$ 2Gal $\beta$ 4GlcNAc $\beta$ -R1, R2
H type 3	Fuc $\alpha$ 2Gal $\beta$ 3GalNAc $\alpha$ -R1
$\alpha$ Gal trisaccharide	Gal $\alpha$ 3Gal $\beta$ 4GlcNAc $\beta$ -R1
Gb3 (Pk)	Gal $\alpha$ 4Gal $\beta$ 4Glc $\beta$ -R1
iGb3	Gal $\alpha$ 3Gal $\beta$ 4Glc $\beta$ -R1

P1 trisaccharide	Gal $\alpha$ 4Gal $\beta$ 4GlcNAc $\beta$ -R1
Lewis a	Gal $\beta$ 3(Fuc $\alpha$ 4)GlcNAc $\beta$ -R1
Lewis x	Gal $\beta$ 4(Fuc $\alpha$ 3)GlcNAc $\beta$ -R1
3'-Sulfo-Lewis a	Su-O-3Gal $\beta$ 3(Fuc $\alpha$ 4)GlcNAc $\beta$ -R1
3'-Sulfo-Lewis x	Su-O-3Gal $\beta$ 4(Fuc $\alpha$ 3)GlcNAc $\beta$ -R1
A type 2	GalNAc $\alpha$ 3(Fuc $\alpha$ 2)Gal $\beta$ 4GlcNAc $\beta$ -R1
B type 2	Gal $\alpha$ 3(Fuc $\alpha$ 2)Gal $\beta$ 4GlcNAc $\beta$ -R1
Lewis b	Fuc $\alpha$ 2Gal $\beta$ 3(Fuc $\alpha$ 4)GlcNAc $\beta$ -R1, R3
Lewis y	Fuc $\alpha$ 2Gal $\beta$ 4(Fuc $\alpha$ 3)GlcNAc $\beta$ -R1, R2
Sialyl-Lewis a	NeuAc $\alpha$ 2,3Gal $\beta$ 3(Fuc $\alpha$ 4)GlcNAc $\beta$ -R1, R3
Sialyl-Lewis x	NeuAc $\alpha$ 2,3Gal $\beta$ 3(Fuc $\alpha$ 4)GlcNAc $\beta$ -R1, R3
6-sulfo Sialyl-Lewis x	NeuAc $\alpha$ 2,3Gal $\beta$ 3(Fuc $\alpha$ 4)(Su-O-6)GlcNAc $\beta$ -R1
Tk	GlcNAc $\beta$ 3(GlcNAc $\beta$ 6)GlcNAc $\beta$ 3Gal $\beta$ -R1
Lacto-N-tetraose (LNT)	Gal $\beta$ 3GlcNAc $\beta$ 3Gal $\beta$ 4Glc $\beta$ -R3
Lacto-N-neotetraose (LNnT)	Gal $\beta$ 4GlcNAc $\beta$ 3Gal $\beta$ 4Glc $\beta$ -R3
$\alpha$ -Gal-Lewis x	Gal $\alpha$ 3Gal $\beta$ 4(Fuc $\alpha$ 3)GlcNAc $\beta$ -R1
$\alpha$ -Gal pentasaccharide	Gal $\alpha$ 3Gal $\beta$ 4GlcNAc $\beta$ Gal $\beta$ 4Glc $\beta$ -R1
Sialyl-Lewis x pentasaccharide	NeuAc $\alpha$ 2,3Gal $\beta$ 3(Fuc $\alpha$ 4)GlcNAc $\beta$ 3Gal $\beta$ -R1
Sialyl-lacto-N-neotetraose (Sia-LNnT)	NeuAc $\alpha$ 2,3Gal $\beta$ 4GlcNAc $\beta$ 3Gal $\beta$ 4Glc $\beta$ -R1
Lactoneofucopentaose I (LNF I)	Fuc $\alpha$ 2Gal $\beta$ 3GlcNAc $\beta$ 3Gal $\beta$ 4Glc $\beta$ -R1, R3
Lacto-N-fucopentaose II (LNF II)	Gal $\beta$ 3(Fuc $\alpha$ 4)GlcNAc $\beta$ 3Gal $\beta$ 4Glc $\beta$ -R3
Lacto-N-fucopentaose III (LNF III)	Gal $\beta$ 4(Fuc $\alpha$ 3)GlcNAc $\beta$ 3Gal $\beta$ 4Glc $\beta$ -R3
A hexasaccharide	GalNAc $\alpha$ 3(Fuc $\alpha$ 2)Gal $\beta$ 3GlcNAc $\beta$ 3Gal $\beta$ 4Glc $\beta$ -R3
A heptasaccharide	GalNAc $\alpha$ 3(Fuc $\alpha$ 2)Gal $\beta$ 3(Fuc $\alpha$ 4)GlcNAc $\beta$ 3Gal $\beta$ 4Glc $\beta$ -R3

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<sup>a</sup>Oligosaccharides were used coupled to either polyacrylamide via an 3 carbon spacer (R1), or to human serum albumin via either a p-aminophenylethyl spacer (R2) or an acetyl phenylenediamine spacer (R3).