

Supplementary figures:

Figure S1

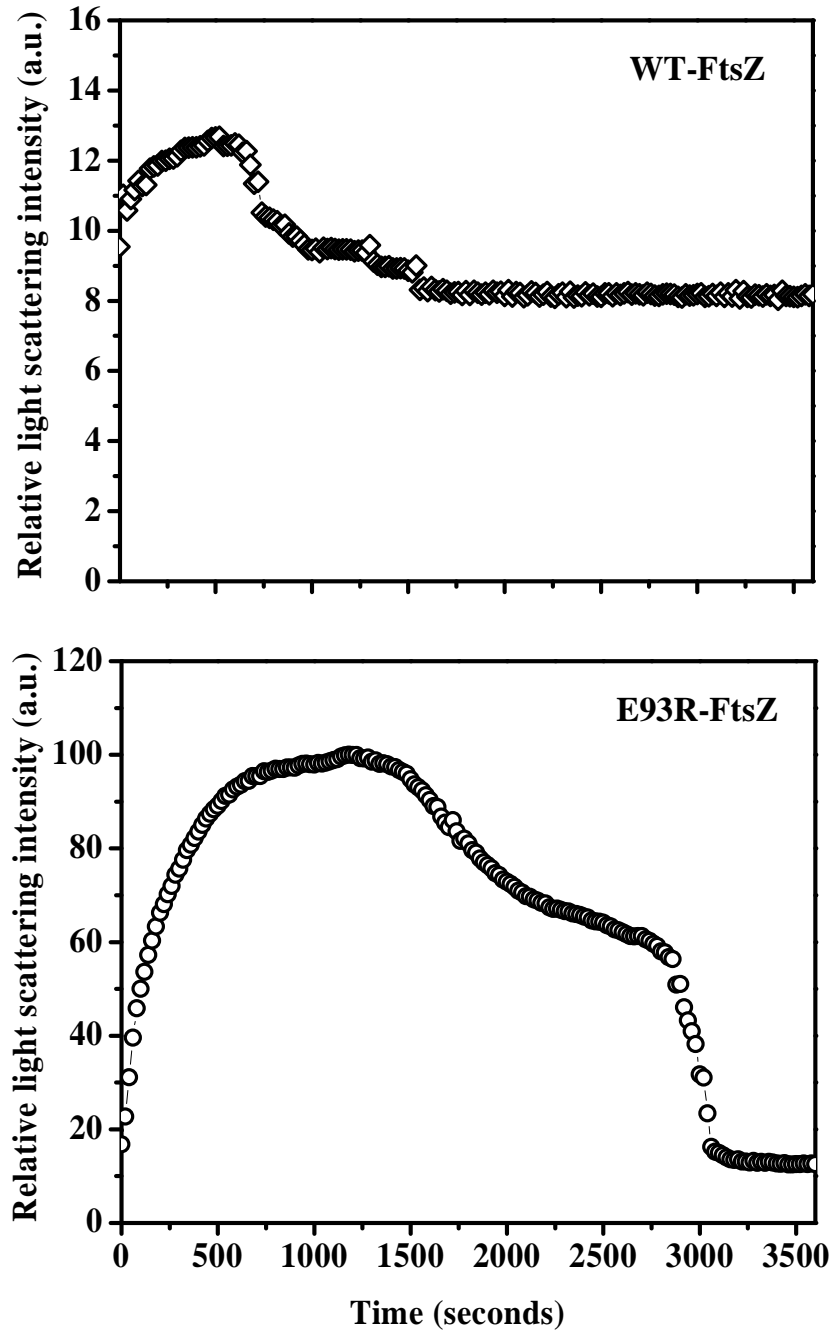


Figure S1: E93R mutation stabilized FtsZ polymers. WT and E93R FtsZ (14.4 μ M) were polymerized at 37 $^{\circ}$ C and the assembly kinetics was monitored through 90 $^{\circ}$ light scattering at 500 nm.

Figure S2

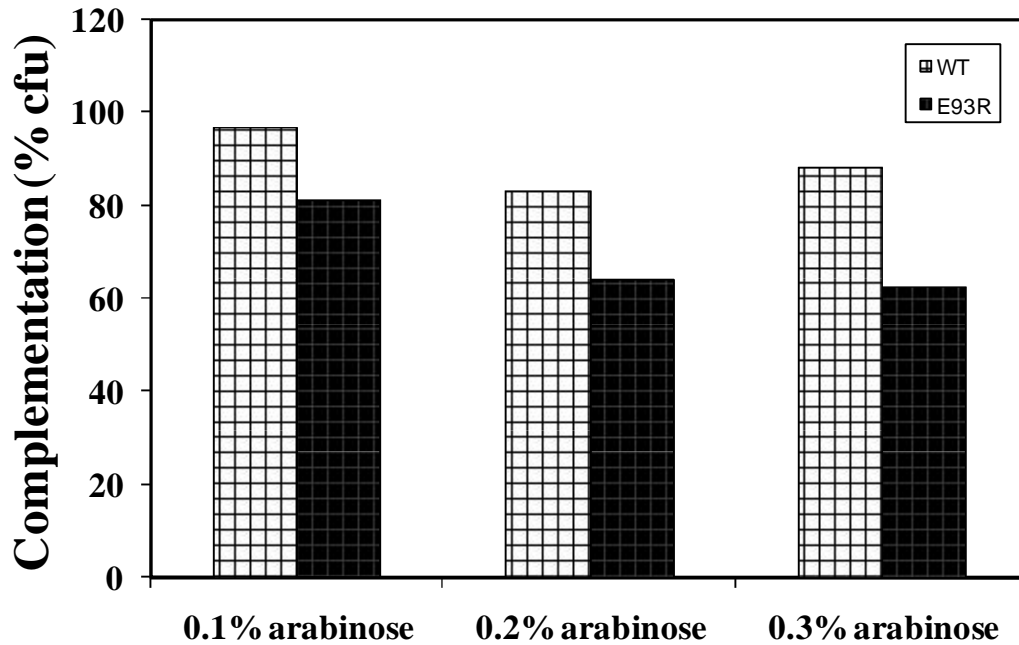


Figure S2: E93R mutation complements WT-FtsZ in JKD7-1/pKD3 cells. WT-FtsZ and E93R-FtsZ genes were inserted in complementation vector pJSB2. JKD7-1/pKD3 cells were transformed with the recombinant plasmids. Graph is showing the percent colonies formed in induction media (with 0.1%, 0.2% and 0.3% arabinose at 42 °C) calculated against the number of colonies formed in repression plates (with 0.5% glucose at 30 °C). The experiment was done twice.

Figure S3

Figure S3A

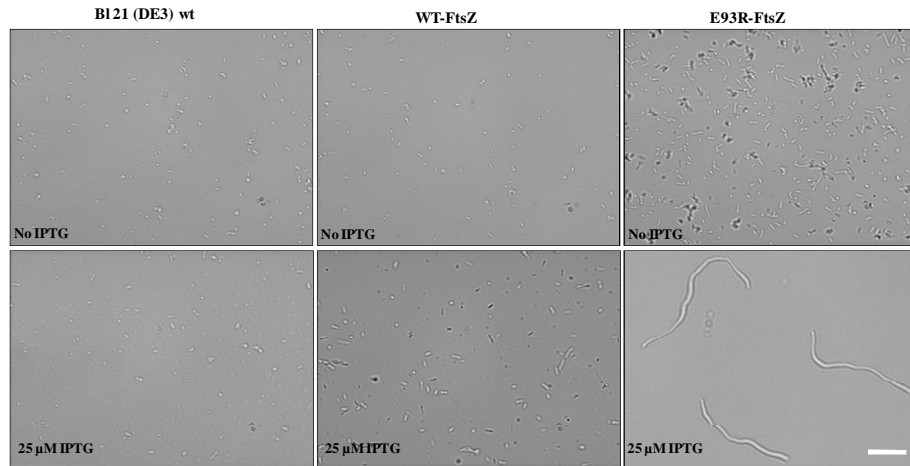


Figure S3B

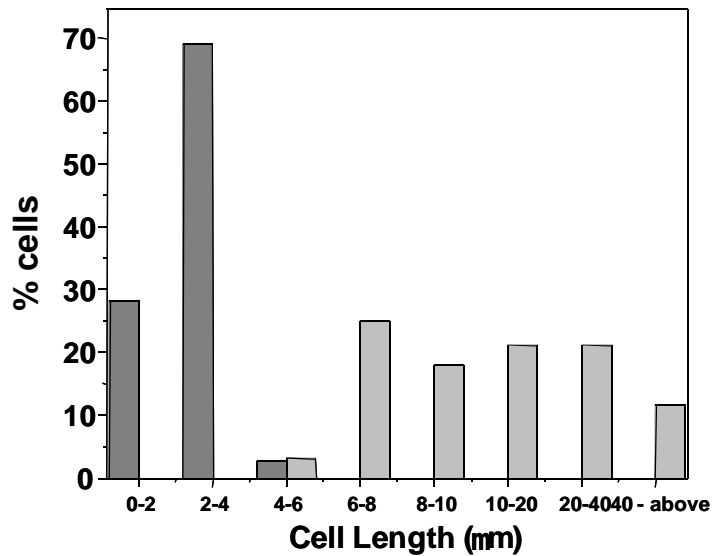


Figure S3: (A) E93R-FtsZ overexpression induced filamentation in *E. coli* cells. Cells harboring WT FtsZ or E93R mutant plasmid were grown for 3 hours in the absence and presence of 25 μ M IPTG and observed under differential interference contrast microscope. Shown are the wild type *E. coli* BL21 (DE3) cells containing no vector, *E. coli* BL21 (DE3) cells containing plasmid for WT-FtsZ and *E. coli* BL21 (DE3) cells containing plasmid for E93R-FtsZ in the absence and presence of IPTG (upper and lower panels), respectively. Scale bar is 10 μ m. **(B) Distribution of cell lengths in cells containing WT-FtsZ and E93R-FtsZ plasmids.** Percent of cells in total population was calculated as a function of cell length. Dark grey bars denote cells containing WT-FtsZ and light grey bars show cells containing E93R-FtsZ grown in the presence of 25 μ M IPTG.

Figure S4

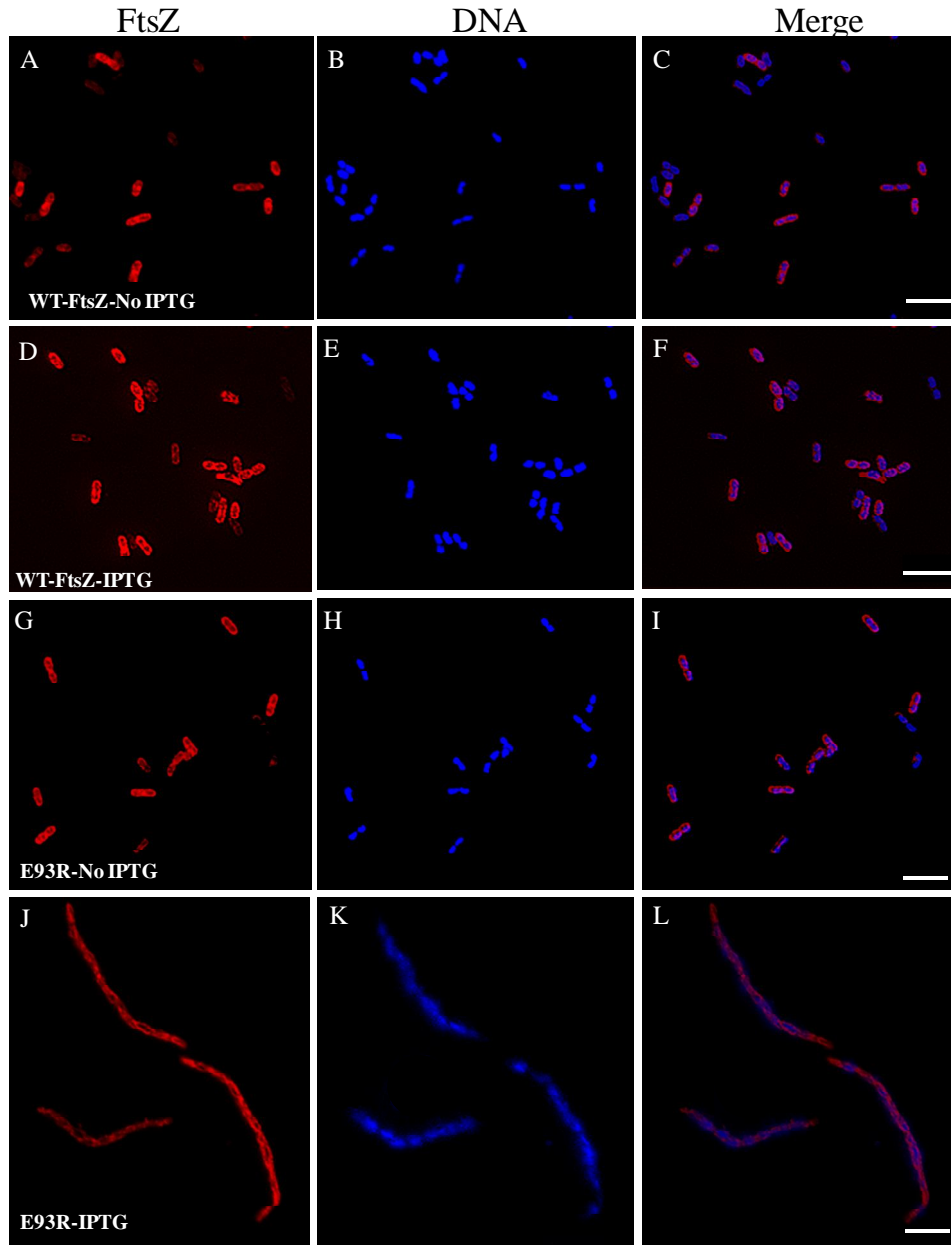


Figure S4: Effect of E93R mutation on the Z-ring assembly and nucleoid segregation. Z-ring and DNA of bacterial cells containing WT-FtsZ and E93R-FtsZ mutant plasmids were grown in the absence or presence of 25 μ M IPTG. Z-rings (red) were immunostained with anti FtsZ antibody and DNA (blue) was stained with DAPI. Shown are the cells containing WT-FtsZ in the absence (A, B, and C) and presence of IPTG (D, E, and F) and E93R mutant cells in the absence (G, H, and I) and presence of IPTG (J, K, and L). Experiment was performed three times. Scale bar is 10 μ m.

Figure S5

Figure S5A

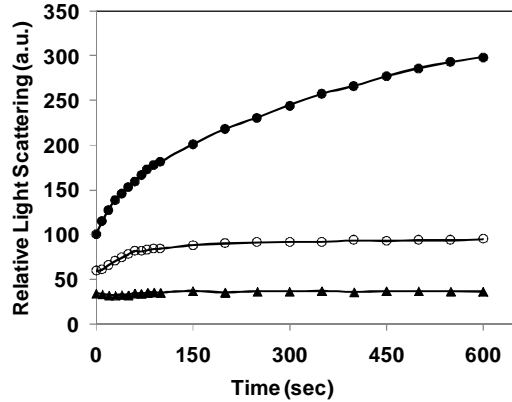


Figure S5B

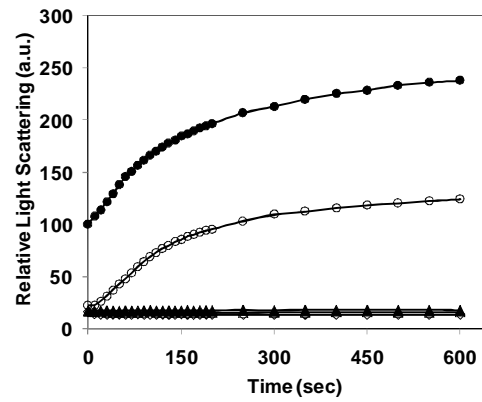


Figure S5C

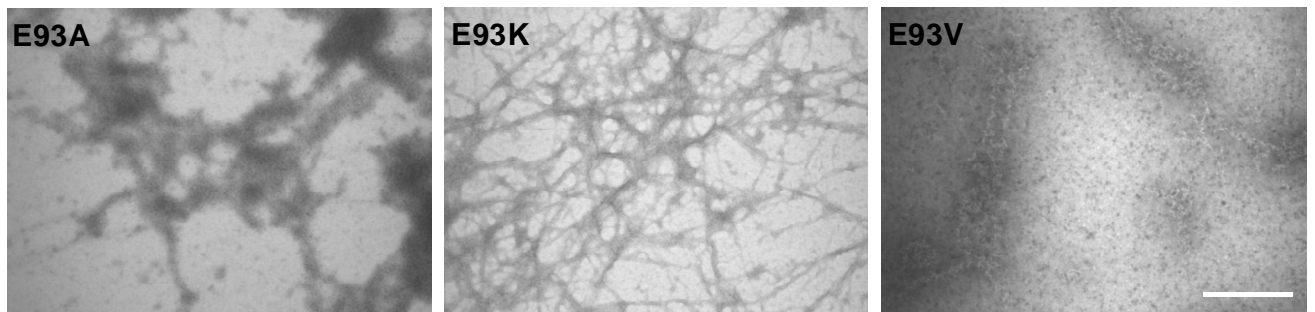


Figure S5: Effect of substitution of E93 to alanine, lysine, or valine on the assembly of FtsZ. The assembly kinetics of native and mutant FtsZ proteins were analyzed by light scattering at 500 nm (Panel A and B). WT, E93R, E93A, E93K, E93V (14.4 μ M) were polymerized at 37 $^{\circ}$ C in the assembly buffer containing 25 mM PIPES pH 6.8, 50 mM KCl, 10 mM $MgCl_2$ and 1mM GTP. Shown are the (A) Light scattering traces of WT (▲), E93R (●) and E93K (○) FtsZ. (B) Light scattering traces of WT (▲), E93R (●), E93A (○), and E93V (△) FtsZ. (C) Electron micrographs of E93A, E93K and E93V FtsZ polymers are shown. 7.2 μ M of E93A, E93K and E93V FtsZ proteins were polymerized as described in the experimental procedures. Scale bar is 1000 nm.

Figure S6

Figure S6A

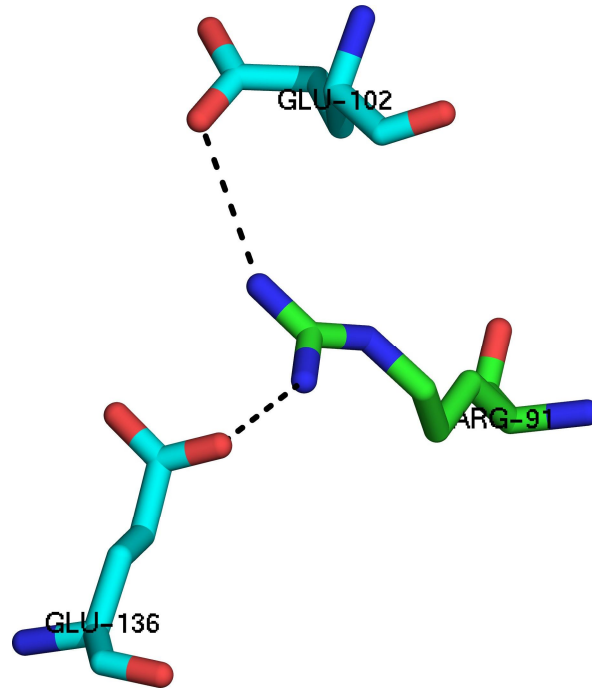


Figure S6B

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FTSZ_MYCTU 1  MTPPHNYL--AVIKVVCITGGGGVNAVNRMIQGLKGVFEFIATNTDAQALLMSDADVKLDVGRDSTRGLGA
FTSZ_ECOLI 1  MFEPMELTNDAVIKVLCVGGGGGNAVEHMRERIEGVFFAVNTDAQALRKTAVGQTIQIGSGITKGLGA

FTSZ_MYCTU 69  GADPEVGRKAAEDAKDEIEELIRGADMVVFVTAEGGGGTGTGAPVVASIARKLGALTVGVVTRPFSFEGK
FTSZ_ECOLI 71  GANPEVGRNAADEDRLRALRAALEGADMVFVLAAGMGGGTGTGAAPVVAEVAKDLGILTVAVVTRKPFNFEGK

FTSZ_MYCTU 139 RRSNQAEANGIAADRESCDTLIVIPNDRLLQMGDAAVSLMDAERSADEVLLNGVQGITDLITTPGLINVDF
FTSZ_ECOLI 141 KRMAFAEQGITELSKHVDSLITIPNDKLLKVLGRGITSLLDAFGAANDVLKGAVQGI AELITRPGLMNVDF

FTSZ_MYCTU 209 ADVKGIMSGACTALMGTGSARGEGRSLKAAETAINSPILLEAS-MEQAQGVLMSTAGGSDLGLFPEINEAAS
FTSZ_ECOLI 211 ADVRTVMSEMGYAMMGSGVASGEDRAEEAAEMAISPLLEDIDLSGARGVLVNITACFDLRLDEFETVGN

FTSZ_MYCTU 278 LVQDAAHPDANIIFGTVIDDSLGDEVVRTVLAAGFDVSGPGRKPVMGETGGAHRIESAKAGKLTSTLTFEP
FTSZ_ECOLI 281 TIRAFASDNATVVIGTSLDPDMNDELRTVVVATGIGMDKRPEITLVTNKQVQPVMDRYQQHGMAPLITQE

FTSZ_MYCTU 348 VDAVSVPLHTNGATLSIGGDDDDVDVPPFMR---
FTSZ_ECOLI 351 QKPVAKVVNDNAPQAK--EPDYLDIAPFLRQAD
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Figure S6: (A) Salt bridges at the interface in the crystal structure of *MtbFtsZ* (1RQ7). Arg91 is from chain A and Glu102 and Glu136 are from chain B. The snapshot was created using PyMol (36). (B) Alignment of *EcFtsZ* (FTSZ_ECOLI; Uniprot id: P0A9A6) and *MtbFtsZ* (FTSZ_MYCTU; Uniprot id: P64170) was obtained using ClustalW (S1). Identical and similar residues are displayed in black and grey backgrounds, respectively, using Bioedit (S2). The residues forming salt bridge in crystal structure of *MtbFtsZ* are shown with asterisk (*) below the alignment. The numbering on the left of sequence shows the residue numbering.

References:

- S1. Thompson, J. D., Higgins, D. G., and Gibson, T. J. (1994) *Nucleic Acids Res.* 22, 4673-4680
- S2. Hall, T. A. (1999) *Nucleic Acids Symp. Series* 41, 95-98