

Supplemental data for the reviewer 2

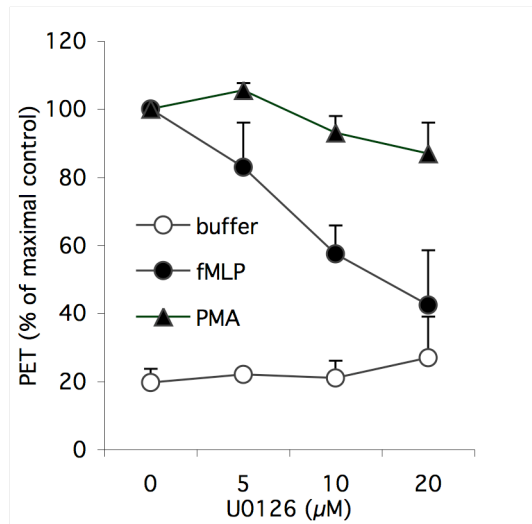


Figure 1S: The MEK antagonist U0126 inhibits fMLP-induced production of phosphatidylethanol (PET) by PLD in human neutrophils. Cells labelled with tritiated lysophosphatidylcholine as described (Djerdjouri et al, *Biochem. Biophys. Res Commun*, 264, 371-375, 1999) were pretreated with various concentrations of U0126 before stimulation with $1 \mu\text{M}$ fMLP or PMA for 3 min. Results are expressed as percentage of maximal control values. Data are the mean of 4 experiments.

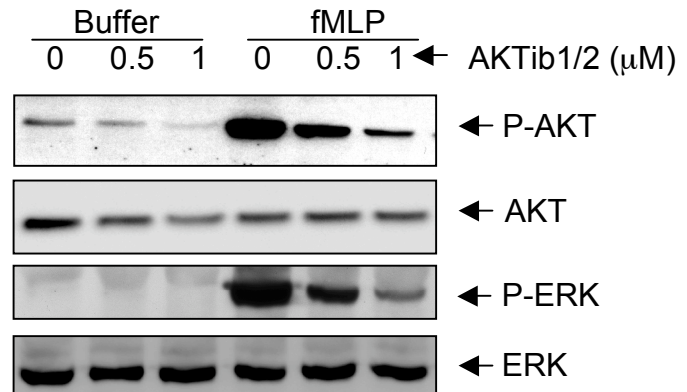


Figure 2S: fMLP-mediated activation of ERK in human neutrophils is dependent on the AKT signaling pathway. Human neutrophils were treated in the absence or presence of the AKT antagonist AKTib1/2 for 15 min before stimulation with fMLP ($1 \mu\text{M}$) for 2 min. Representative western blots show the phosphorylated form of AKT (S473, P-AKT) and ERK1/2 (P-ERK) detected with a specific antibody.

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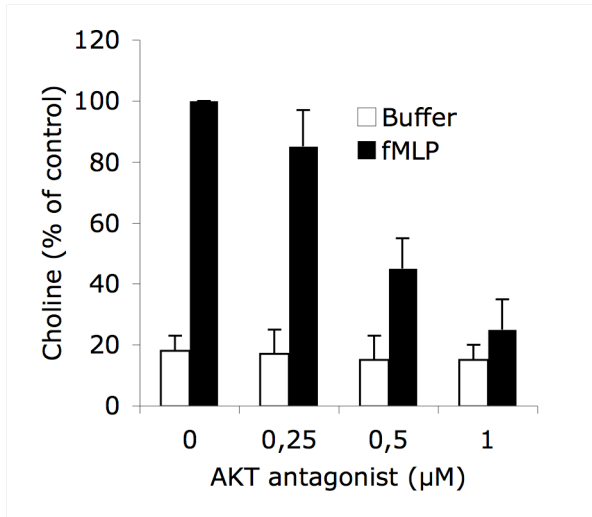


Figure 3S: fMLP-mediated PLD activity in human neutrophils is dependent on the AKT signaling pathway. Human neutrophils were treated in the absence or presence of the AKT antagonist AKTib-8 for 15 min before stimulation with fMLP (1µM) for 3 min. PLD activity was quantified by measuring the production of choline (Paruch S, Faseb J. 20, 142- 2006). Results are expressed as percentage of maximal control values (mean of 5 experiments).

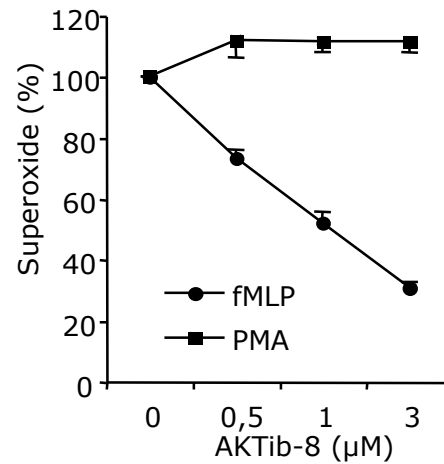


Figure 4S: fMLP-mediated production of superoxide in human neutrophils is dependent on the AKT signaling pathway. Human neutrophils were treated in the absence or presence of the AKT antagonist AKTib-8 for 15 min before stimulation with 1 µM fMLP or PMA for 5 min. Results represent the production of superoxide and are expressed as percentage of maximal control values (mean of 5 experiments).

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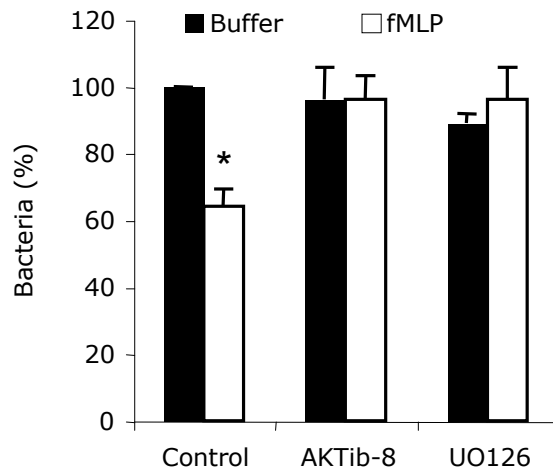


Figure 5S : fMLP-mediated human neutrophil bactericidal activity is inhibited by the AKT antagonist AKTib1/2 and by the MEK antagonist UO126. Human neutrophils (0.5×10^6 cells/100 μ L) were treated at 37°C in the absence (Control) or presence of AKTib1/2 (1 μ M) or UO126 (5 μ M) for 10 min. Cells were incubated with opsonized bacteria (50×10^3 *Staphylococcus Aureus*) in the absence (buffer) or presence of 1 μ M fMLP for 15 min. Cells were lysed with saponine and diluted, and the number of bacterial colonies was determined after overnight culture on LB plate of the bacteria in the cell homogenates. Results represent the mean \pm SEM of 5 experiments. Significant differences between control and treated cells are designated by * ($p < 0.05$).