

# Alternative Splicing and Polyadenylation Contribute to the Generation of hERG1 C-Terminal Isoforms

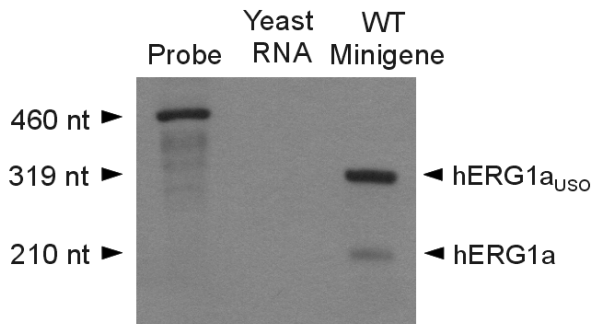
Qiuming Gong, Matthew R. Stump, A. Russell Dunn, Vivianne Deng and Zhengfeng Zhou

## SUPPLEMENTAL FIGURES:

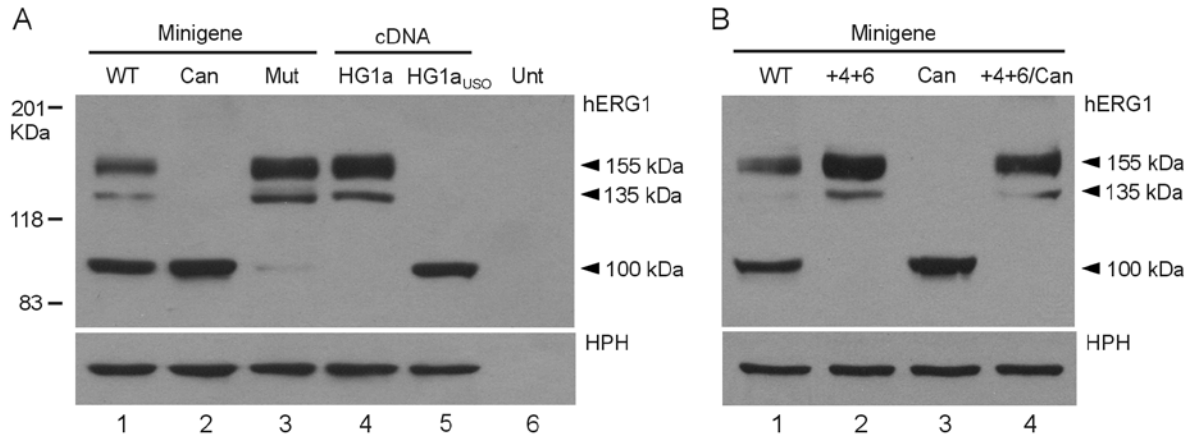
```

-130
CACAAATACACACCCCCACAAAACCTAAAATCAAAGTTTTCACTACATAACACTGGGCCTTACTGCATGTGGT
TCATTCTAGCATTCTCTGTTCTGTGCTGTGCTAAGCTATACTACTGTATGTTCTTTAGTAAAAAAAAAAAAA
AAAAAAAAAAAAATGCTGGTTTTGATTCACTACTGTGTCTGATCTTTGGTTTGAAGAACATTGCTTATAA
GGGTGCAGTGATTGGCTAAGAGGGTGTGGGACCTGGGGTTGGGGTAGTCCTGCTGGCTGGGTCTCATT
TGCTGGGAGACCCTGGGCTCC
+172
  
```

**Figure S1. Sequence surrounding hERG1 intron 9 poly(A) signal.** The hERG1 intron 9 poly(A) signal and flanking sequences (-130/+172 bp) are shown. The noncanonical poly(A) signal (AGTAAA) is shown in bold. Putative downstream U/GU rich elements are underlined.



**Figure S2. RPA analysis of hERG1 mRNA alternative processing.** Analysis of the wild-type minigenes by RPA with a probe containing hERG1<sub>a\_USO</sub> sequences from exon 9 to intron 9. This probe generated a protected fragment of 319 nt for hERG1<sub>a\_USO</sub> transcript and a 210 nt fragment for hERG1a transcript. The length of the probe was 460 nt and contained sequences from the pCRII vector at both ends.



**Figure S3. Immunoblot analysis of hERG1a and hERG1a<sub>USO</sub> proteins.** A, Flp-In HEK293 cells were stably transfected with minigenes carrying the wild-type (WT), canonical (Can) and mutant (Mut) intron 9 poly(A) signals, and hERG1a (HG1a) and hERG1a<sub>USO</sub> (HG1a<sub>USO</sub>) cDNAs. Untransfected control (Unt). Cell lysates were subjected to SDS-PAGE and immunoblotted with an anti-hERG1 antibody against the N-terminus of hERG1 protein. The expression level of hygromycin B phosphotransferase (HPH) encoded by hygromycin B resistant gene in pcDNA5 vector served as a loading control. Results shown are representative of four independent experiments. B, Flp-In HEK293 cells were stably transfected with minigenes carrying the wild-type (WT), 5' splice site mutation +4U>A/+6G>U (+4+6), canonical (Can) intron 9 poly(A) signal and +4U>A/+6G>U plus canonical poly(A) signal mutation (+4+6/Can). Results shown are representative of three independent experiments.