

Supporting Information

One-Pot Etherification of Purine Nucleosides and Pyrimidines

Hari Prasad Kokatla and Mahesh K. Lakshman*

Department of Chemistry, The City College and The City University of New York,
160 Convent Avenue, New York, New York 10031-9198

Table of Contents

Information	Page
General Experimental Considerations	S-4
General Procedure for One-Pot Etherification Reactions	S-4
2',3',5'-Tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -methylinosine (2)	S-4
<i>O</i> ⁶ -(Allyl)-2',3',5'-tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)inosine (4)	S-5
2',3',5'-Tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -propargylinosine (5)	S-5
2',3',5'-Tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -isopropylinosine (6)	S-5
2',3',5'-Tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -(2-hydroxyethyl)inosine (7)	S-6
Ethylene glycol bis- <i>O</i> ⁶ -[9-(2',3',5'-tri- <i>O</i> - <i>tert</i> -butyldimethylsilyl-β-D-ribofuranosyl)]purinyl ether	S-6
2',3',5'-Tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -(4-nitrophenyl)inosine (8)	S-7
2',3',5'-Tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -methylguanosine (9)	S-7
2',3',5'-Tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -(4-methoxyphenyl)guanosine (10)	S-7
3',5'-Di- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -methyl-2'-deoxyguanosine (11)	S-8
3',5'-Di- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -phenyl-2'-deoxyguanosine (12)	S-8
4-Methoxyquinazoline (16a)	S-8
4-Ethoxyquinazoline (16b)	S-9
4-Propoxyquinazoline (16c)	S-9
4-Isopropoxyquinazoline (16d)	S-9
2-(4-Quinazolinyl)ethanol (16e)	S-9
4-(Allyloxy)quinazoline (16f)	S-10
4-(2-Propynyloxy)quinazoline (16g)	S-10
4-(Cyclopentyloxy)quinazoline (16h)	S-10
4-(Cyclohexyloxy)quinazoline (16i)	S-10
4-(Benzyl)quinazoline (16j)	S-10
4-(4-Methoxyphenoxy)quinazoline (16k)	S-11
2-(Methoxy)-4-methylpyrimidine (17a)	S-11

2-(Isopropoxy)-4-methylpyrimidine (17b)	S-11
2-(Allyloxy)-4-methylpyrimidine (17c)	S-11
2-[(4-Methylpyrimidin-2-yl)oxy]ethanol (17d)	S-12
5-Bromo-2-methoxypyrimidine (18a)	S-12
5-Bromo-2-isopropoxypyrimidine (18b)	S-12
2-(Allyloxy)-5-bromopyrimidine (18c)	S-12
2-(5-Bromopyrimidin-2-yloxy)ethanol (18d)	S-12
5-Bromo-2-(4-methoxyphenoxy)pyrimidine (18e)	S-13
500 MHz ^1H NMR spectrum of 2',3',5'-tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -methylinosine (2) in CDCl ₃	S-14
500 MHz ^1H NMR spectrum of <i>O</i> ⁶ -(allyl)-2',3',5'-tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)inosine (4) in CDCl ₃	S-14
500 MHz ^1H NMR spectrum of 2',3',5'-tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -propargylinosine (5) in CDCl ₃	S-15
500 MHz ^1H NMR spectrum of 2',3',5'-tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -isopropylinosine (6) in CDCl ₃	S-15
500 MHz ^1H NMR spectrum of 2',3',5'-tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -(2-hydroxyethyl)inosine (7) in CDCl ₃	S-16
500 MHz ^1H NMR spectrum of ethylene glycol bis- <i>O</i> ⁶ -[9-(2',3',5'-tri- <i>O</i> - <i>tert</i> -butyldimethylsilyl)- β -D-ribofuranosyl] <i>p</i> urinyl ether in CDCl ₃	S-16
500 MHz ^1H NMR spectrum of 2',3',5'-tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -(4-nitrophenyl)inosine (8) in CDCl ₃	S-17
500 MHz ^1H NMR spectrum of 2',3',5'-tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -methylguanosine (9) in CDCl ₃	S-17
500 MHz ^1H NMR spectrum of 2',3',5'-tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -(4-methoxyphenyl)guanosine (10) in CDCl ₃	S-18
500 MHz ^1H NMR spectrum of 3',5'-di- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -methyl-2'-deoxyguanosine (11) in CDCl ₃	S-18
500 MHz ^1H NMR spectrum of 3',5'-di- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -phenyl-2'-deoxyguanosine (12) in CDCl ₃	S-19
500 MHz ^1H NMR spectrum of 4-methoxyquinazoline (16a) in CDCl ₃	S-19
500 MHz ^1H NMR spectrum of 4-ethoxyquinazoline (16b) in CDCl ₃	S-20
500 MHz ^1H NMR spectrum of 4-propoxyquinazoline (16c) in CDCl ₃	S-20
500 MHz ^1H NMR spectrum of 4-isopropoxyquinazoline (16d) in CDCl ₃	S-21
500 MHz ^1H NMR spectrum of 2-(4-quinazolinylloxy)ethanol (16e) in CDCl ₃	S-21
500 MHz ^1H NMR spectrum of 4-(allyloxy)quinazoline (16f) in CDCl ₃	S-22
500 MHz ^1H NMR spectrum of 4-(2-propynylloxy)quinazoline (16g) in CDCl ₃	S-22
500 MHz ^1H NMR spectrum of 4-(cyclopentyloxy)quinazoline (16h) in CDCl ₃	S-23

500 MHz ^1H NMR spectrum of 4-(cyclohexyloxy)quinazoline (16i) in CDCl_3	S-23
500 MHz ^1H NMR spectrum of 4-(benzyloxy)quinazoline (16j) in CDCl_3	S-24
500 MHz ^1H NMR spectrum of 4-(4-methoxyphenoxy)quinazoline (16k) in CDCl_3	S-24
500 MHz ^1H NMR spectrum of 2-(methoxy)-4-methylpyrimidine (17a) in CDCl_3	S-25
500 MHz ^1H NMR spectrum of 2-(isopropoxy)-4-methylpyrimidine (17b) in CDCl_3	S-25
500 MHz ^1H NMR spectrum of 2-(allyloxy)-4-methylpyrimidine (17c) in CDCl_3	S-26
500 MHz ^1H NMR spectrum of 2-[(4-methylpyrimidin-2-yl)oxy]ethanol (17d) in CDCl_3	S-26
500 MHz ^1H NMR spectrum of 5-bromo-2-methoxypyrimidine (18a) in CDCl_3	S-27
500 MHz ^1H NMR spectrum of 5-bromo-2-isopropoxypyrimidine (18b) in CDCl_3	S-27
500 MHz ^1H NMR spectrum of 2-(allyloxy)-5-bromopyrimidine (18c) in CDCl_3	S-28
500 MHz ^1H NMR spectrum of 2-(5-bromopyrimidin-2-yloxy)ethanol (18d) in CDCl_3	S-28
500 MHz ^1H NMR spectrum of 5-bromo-2-(4-methoxyphenoxy)pyrimidine (18e) in CDCl_3	S-29
500 MHz ^1H - ^1H COSY spectrum of 2',3',5'-tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -(4-nitrophenyl)-inosine (8) in CDCl_3	S-30
500 MHz ^1H - ^1H COSY spectrum of 2',3',5'-tri- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -methyl-guanosine (9) in CDCl_3	S-31
500 MHz ^1H - ^1H COSY spectrum of 3',5'-di- <i>O</i> -(<i>tert</i> -butyldimethylsilyl)- <i>O</i> ⁶ -methyl-2'-deoxy-guanosine (11) in CDCl_3	S-32
References	S-33

General Experimental Considerations

Thin layer chromatography was performed on 250 μm silica plates and column chromatographic purifications were performed on 200–300 mesh silica gel. THF was distilled over LiAlH₄ and then over Na, MeCN was distilled over CaH₂. All other reagents were obtained from commercial sources and were used without further purification. ¹H NMR spectra were recorded at 500 MHz in CDCl₃ and are referenced to the residual protonated solvent resonance. ¹³C NMR spectra were recorded at 125 MHz in CDCl₃ and are referenced to the solvent resonance. Chemical shifts (δ) are reported in parts per million (ppm) and coupling constants (J) are in hertz (Hz).

General Procedure for the One-Pot Etherification Reactions

To a solution of the substrate (0.16 mmol of protected inosine or 79 μmol of protected guanosine or 0.10 mmol of protected 2'-deoxyguanosine or 0.68 mmol of either quinazolin-4(3*H*)-one or 4-methylpyrimidin-2(1*H*)-one•HCl or 0.57 mmol of 5-bromopyrimidin-2(1*H*)-one) in dry THF (2 mL) were added 2 molar equiv each of BOP and Cs₂CO₃ under a nitrogen atmosphere. The mixture was stirred at room temperature (10 min for inosine, 1 h for guanosine and 2'-deoxyguanosine, 50 min for pyrimidinones).

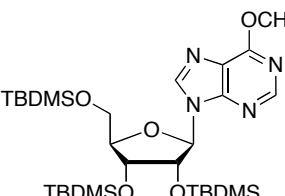
For reactions with alcohols: the resulting mixture was evaporated under reduced pressure, 2 molar equiv of Cs₂CO₃ and 20 molar equiv of alcohol were added.

For reactions with phenols: 2 molar equiv of Cs₂CO₃, and 2 molar equiv of the phenol were added without evaporation of the mixture.

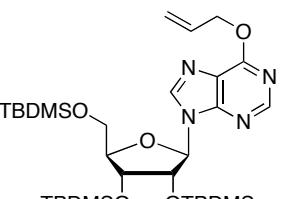
In each case, the mixture was allowed to stir at the appropriate temperature (indicated in Tables 1 and 2), until TLC indicated complete reaction. The reaction mixture was diluted with water (10 mL) and extracted with EtOAc (3 x 10 mL). The organic layer was dried over Na₂SO₄ and concentrated under reduced pressure. The crude products were purified by column chromatography on silica gel using the solvent indicated under each compound heading. Any deviation from the general procedure is described under the specific compound heading.

2',3',5'-Tri-*O*-(*tert*-butyldimethylsilyl)-*O*⁶-methylinosine (2).

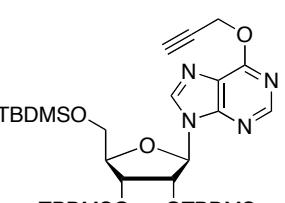
Purification using 20% EtOAc in hexanes gave 96 mg (94%) of **2** as a yellowish solid. R_f (SiO₂/20% EtOAc in hexanes) = 0.4. ¹H NMR: δ 8.52 (s, 1H, Ar-H), 8.32 (s, 1H, Ar-H), 6.08 (d, 1H, H-1', J = 4.8 Hz), 4.62 (t, 1H, H-2', J = 4.6 Hz), 4.31 (t, 1H, H-3', J = 4.1 Hz), 4.18 (s, 3H,


 OCH₃), 4.13 (q, 1H, H-4', *J* = 3.1 Hz), 4.03 (dd, 1H, H-5', *J* = 3.9, 11.2 Hz), 3.79 (dd, 1H, H-5', *J* = 2.6, 11.2 Hz), 0.95, 0.93, and 0.75 (3s, 27H, *tert*-Bu), 0.14, 0.13, 0.10, 0.09, -0.04, and -0.22 (6s, 18H, SiCH₃). ¹³C NMR: δ 161.2, 152.2, 151.9, 141.3, 122.2, 88.6, 85.6, 76.3, 72.1, 62.7, 54.3, 26.3, 26.0, 25.8, 18.7, 18.3, 18.0, -4.1, -4.4, -4.8, -5.1. HRMS (ESI) calcd for C₂₉H₅₇N₄O₅Si₃ [M + H]⁺ 625.3631, found 625.3636.

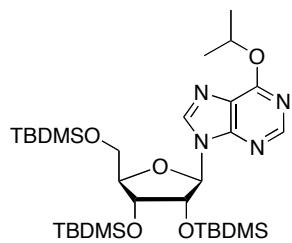
O⁶-(Allyl)-2',3',5'-tri-O-(tert-butyldimethylsilyl)inosine (4).


 Purification using 20% EtOAc in hexanes gave 96 mg (90%) of **4** as a white solid. *R*_f (SiO₂/30% EtOAc in hexanes) = 0.7. ¹H NMR: δ 8.50 (s, 1H, Ar-H), 8.30 (s, 1H, Ar-H), 6.17 (m, 1H, -CH=), 6.08 (d, 1H, H-1', *J* = 4.8 Hz) 5.45 (d, 1H, =CH_{trans}, *J* = 17.0 Hz) 5.29 (d, 1H, =CH_{cis}, *J* = 10.2 Hz), 5.15 (br d, 2H, OCH₂, *J* = 5.8 Hz), 4.63 (t, 1H, H-2', *J* = 8.3 Hz), 4.31 (t, 1H, H-3', *J* = 3.4 Hz), 4.13 (br d, 1H, H-4', *J* = 2.4 Hz), 4.02 (dd, 1H, H-5', *J* = 3.4, 11.2 Hz), 3.79 (d, 1H, H-5', *J* = 11.2 Hz), 0.95, 0.93, and 0.78 (3s, 27H, *tert*-Bu), 0.14, 0.13, 0.10, 0.09, -0.04, and -0.23 (6s, 18H, SiCH₃). ¹³C NMR: δ 160.5, 152.1, 141.3, 132.6, 122.1, 118.7, 88.5, 85.7, 76.3, 72.1, 67.7, 62.7, 26.3, 26.0, 25.8, 18.7, 18.3, 18.0, -4.1, -4.4, -4.8, -5.1. HRMS (ESI) calcd for C₃₁H₅₉N₄O₅Si₃ [M + H]⁺ 651.3788, found 651.3787.

2',3',5'-Tri-O-(tert-butyldimethylsilyl)-O⁶-propargylinosine (5).

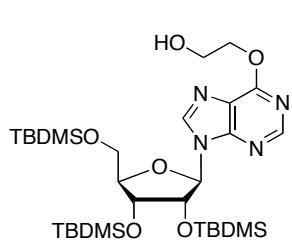

 Purification using 20% EtOAc in hexanes gave 91 (86%) mg of **5** as a white solid. *R*_f (SiO₂/30% EtOAc in hexanes) = 0.7. ¹H NMR: δ 8.54 (s, 1H, Ar-H), 8.36 (s, 1H, Ar-H), 6.08 (d, 1H, H-1', *J* = 4.8 Hz), 5.23 (dd, 2H, OCH₂, *J* = 1.4, 2.4 Hz), 4.59 (t, 1H, H-2', *J* = 4.4 Hz), 4.32 (t, 1H, H-3', *J* = 4.1 Hz), 4.13 (q, 1H, H-4', *J* = 3.4 Hz), 4.00 (dd, 1H, H-5', *J* = 3.6, 11.4 Hz), 3.79 (dd, 1H, H-5', *J* = 2.4, 11.7 Hz), 2.49 (t, 1H, =C-H, *J* = 2.4 Hz), 0.95, 0.92, and 0.79 (3s, 27H, *tert*-Bu), 0.14, 0.13, 0.10, 0.09, -0.04, and -0.21 (6s, 18H, SiCH₃). ¹³C NMR: δ 159.5, 152.3, 151.9, 141.7, 122.1, 88.7, 85.6, 78.3, 76.4, 75.3, 71.9, 62.6, 54.3, 26.3, 26.0, 25.8, 18.7, 18.2, 18.0, -4.1, -4.5, -4.7, -5.1. HRMS (ESI) calcd for C₃₁H₅₇N₄O₅Si₃ [M + H]⁺ 649.3631, found 649.3634.

2',3',5'-Tri-O-(tert-butyldimethylsilyl)-O⁶-isopropylinosine (6).¹



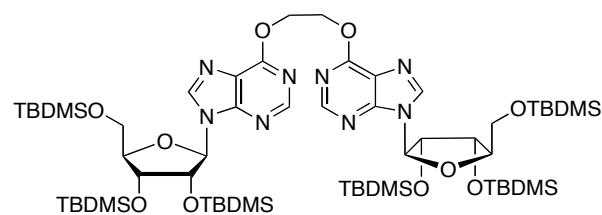
Purification 15% EtOAc in hexanes gave 80 mg (75%) of **6** as a clear gummy material. R_f (SiO_2 /30% EtOAc in hexanes) = 0.7. ^1H NMR: δ 8.48 (s, 1H, Ar-H), 8.26 (s, 1H, Ar-H), 6.08 (d, 1H, H-1', J = 4.8 Hz), 5.66 (septet, 1H, OCH, J = 6.3 Hz), 4.63 (t, 1H, H-2', J = 4.6 Hz), 4.31 (t, 1H, H-3', J = 3.6 Hz), 4.12 (br d, 1H, H-4', J = 2.4 Hz), 4.01 (dd, 1H, H-5', J = 3.6, 11.4 Hz), 3.79 (dd, 1H, H-5', J = 1.9, 11.2 Hz), 1.47 (d, 6H, $(\text{CH}_3)_2$, J = 6.3 Hz), 0.94, 0.92, and 0.78 (3s, 27H, *tert*-Bu), 0.13, 0.12, 0.10, 0.09, -0.04, -0.22 (6s, 18H, SiCH_3). ^{13}C NMR: δ 160.7, 152.2, 152.1, 141.0, 122.3, 88.5, 85.6, 76.2, 72.1, 70.4, 62.7, 26.3, 26.0, 25.8, 22.1, 18.7, 18.3, 18.0, -4.1, -4.4, -4.8, -5.1.

2',3',5'-Tri-*O*-(*tert*-butyldimethylsilyl)-*O*⁶-(2-hydroxyethyl)inosine (7).



Purification using 70% EtOAc in hexanes gave 68 mg (63%) of **7** as a white solid. R_f (SiO_2 /EtOAc) = 0.3. ^1H NMR: δ 8.49 (s, 1H, Ar-H), 8.35 (s, 1H, Ar-H), 6.87 (d, 1H, H-1', J = 4.4 Hz), 4.72 (m, 2H, OCH₂), 4.61 (t, 1H, H-2', J = 4.4 Hz), 4.31 (m, 1H, H-3'), 4.13 (br s, 1H, H-4'), 4.02 (m, 3H, H-5' and OCH₂), 3.79 (d, 1H, H-5', J = 11.2 Hz), 3.52 (s, 1H, OH), 0.95, 0.92 and, 0.79 (3s, 27H, *tert*-Bu), 0.14, 0.13, 0.09, 0.08, -0.04, -0.22 (6s, 18H, SiCH_3). ^{13}C NMR: δ 160.8, 152.2, 152.0, 141.6, 122.0, 88.6, 85.7, 76.4, 72.0, 69.8, 62.6, 61.9, 26.3, 26.0, 25.8, 18.7, 18.2, 18.0, -4.1, -4.5, -4.8, -5.1. HRMS (ESI) calcd for $\text{C}_{30}\text{H}_{59}\text{N}_4\text{O}_6\text{Si}_3$ [M + H]⁺ 655.3737, found 655.3739.

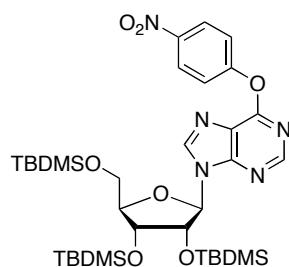
Ethylene glycol bis-*O*⁶-[9-(2',3',5'-tri-*O*-*tert*-butyldimethylsilyl- β -D-ribofuranosyl)]purinyl ether.



Minor product obtained as a white solid (32 mg, 15%) in the reaction of **3** with ethylene glycol. R_f (SiO_2 /20% EtOAc in hexanes) = 0.4. ^1H NMR: δ 8.49 (s, 2H, Ar-H), 8.29 (s, 2H, Ar-H), 6.09 (d, 2H, H-1', J = 4.8 Hz), 5.06 (m, 4H, OCH₂), 4.61 (t, 2H, H-2', J = 4.4 Hz), 4.31 (t, 2H, H-3', J = 2.9 Hz), 4.13 (br d, 2H, H-4', J = 2.4 Hz), 4.02 (dd, 2H, H-5', J = 3.4, 11.2), 3.79 (d, 2H, H-5', J = 11.7 Hz), 0.94, 0.92, and 0.79 (3s, 54H, *tert*-Bu), 0.13, 0.12, 0.10, 0.09, -0.04, -0.22 (6s, 36H, SiCH_3). ^{13}C NMR: δ 160.5, 152.2,

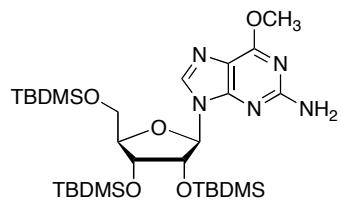
152.0, 141.3, 122.0, 88.6, 85.7, 76.3, 72.1, 64.9, 62.7, 26.3, 26.0, 25.9, 18.7, 18.3, 18.0, -4.1, -4.5, -4.7, -5.1. HRMS (ESI) calcd for $C_{58}H_{111}N_8O_{10}Si_6$ [M + H]⁺ 1247.7033, found 1247.7032.

2',3',5'-Tri-O-(*tert*-butyldimethylsilyl)-O⁶-(4-nitrophenyl)inosine (8).¹



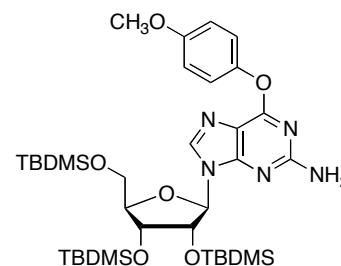
Purification using 20% EtOAc in hexanes gave 102 mg (85%) of **8** as a white solid. R_f (SiO₂/20% EtOAc in hexanes) = 0.5. ¹H NMR: δ 8.52 (s, 1H, Ar-H), 8.50 (s, 1H, Ar-H), 8.33 (d, 2H, Ar-H, J = 8.7 Hz), 7.47 (d, 2H, Ar-H, J = 8.7 Hz), 6.14 (d, 1H, H-1', J = 4.7 Hz), 4.60 (t, 1H, H-2', J = 4.4 Hz), 4.33 (t, 1H, H-3', J = 3.9 Hz), 4.17 (m, 1H, H-4'), 4.04 (dd, 1H, H-5', J = 3.3, 11.4), 3.82 (d, 1H, H-5', J = 11.3 Hz), 0.97, 0.93, and 0.81 (3s, 27H, *tert*-Bu), 0.16, 0.15, 0.11, 0.10, -0.01, and -0.18 (6s, 18H, SiCH₃). ¹³C NMR: δ 159.0, 157.5, 153.5, 151.9, 145.3, 143.0, 125.6, 122.6, 122.3, 88.8, 85.7, 76.6, 71.9, 62.5, 26.3, 26.0, 25.8, 18.7, 18.2, 18.0, -4.1, -4.4, -4.7, -5.1. (The structure of this compound shown on the ¹H NMR spectrum in the Supporting Information to ref 1 contains a typographical error. It should have an O atom in place of the NH).

2',3',5'-Tri-O-(*tert*-butyldimethylsilyl)-O⁶-methylguanosine (9).²



Purification using 20% EtOAc in hexanes gave 37 mg (73%) of **9** as an orange solid. R_f (SiO₂/20% EtOAc in hexanes) = 0.3. ¹H NMR: δ 7.97 (s, 1H, Ar-H), 5.91 (d, 1H, H-1', J = 5.3 Hz), 5.12 (s, 2H, NH₂), 4.46 (t, 1H, H-2', J = 4.9 Hz), 4.26 (t, 1H, H-3', J = 4.3 Hz), 4.08 (m, 1H, H-4'), 4.04 (s, 3H, OCH₃), 3.96 (dd, 1H, H-5', J = 3.9, 11.2 Hz), 3.76 (dd, 1H, H-5', J = 2.4, 11.2 Hz), 0.94, 0.92, and 0.79 (s, 27H, *tert*-Bu), 0.13, 0.12, 0.11, 0.09, -0.04, and -0.18 (6s, 18H, SiCH₃). ¹³C NMR: δ 161.6, 159.5, 153.8, 137.9, 116.0, 87.7, 85.4, 76.4, 72.2, 62.8, 53.9, 26.2, 26.0, 25.9, 18.7, 18.2, 18.1, -4.1, -4.5, -4.8, -5.2.

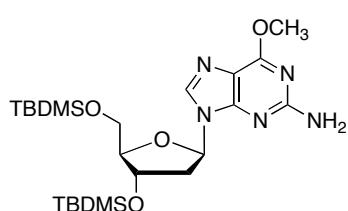
2',3',5'-Tri-O-(*tert*-butyldimethylsilyl)-O⁶-(4-methoxyphenyl)guanosine (10).



Purification using 20% EtOAc in hexanes gave 44 mg (75%) of **10** as a white solid. R_f (SiO₂/20% EtOAc in hexanes) = 0.4. ¹H NMR: δ 8.08 (s, 1H, Ar-H), 7.16 (d, 2H, Ar-H, J = 8.8 Hz), 6.91 (d, 2H, Ar-H, J = 8.8 Hz), 5.92 (d, 1H, H-1', J = 4.5 Hz), 4.79 (s, 2H, NH₂), 4.51 (t, 1H, H-2', J = 4.4 Hz), 4.31 (t, 1H, H-3', J = 4.0 Hz), 4.16 (m, 1H, H-4'), 3.99 (dd, 1H, H-5', J = 3.5, 11.4 Hz), 3.82 (s, 3H, OCH₃), 3.79 (dd, 1H, H-

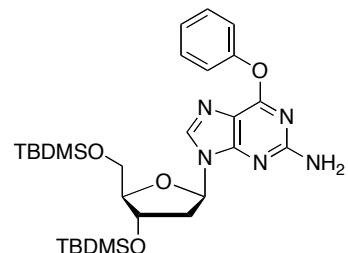
$J = 1.9, 11.4$ Hz), 0.95, 0.92, and 0.83 (3s, 27H, *tert*-Bu), 0.14, 0.13, 0.10, 0.09, 0.00, and –0.10 (6s, 18H, SiCH₃). ¹³C NMR: δ 160.9, 159.3, 157.0, 154.7, 146.2, 138.9, 122.9, 116.1, 114.4, 88.2, 85.1, 76.3, 71.9, 62.6, 55.7, 26.3, 26.0, 25.9, 18.7, 18.7, 18.2, 18.1, –4.1, –4.4, –4.6, –5.1. HRMS (ESI) calcd for C₃₅H₆₂N₅O₆Si₃ [M + H]⁺ 732.4002, found 732.4007.

3',5'-Di-*O*-(*tert*-butyldimethylsilyl)-O⁶-methyl-2'-deoxyguanosine (**11**).²



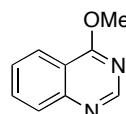
Purification using 20% EtOAc in hexanes gave 38 mg (75%) of **11** as a yellowish solid (from 0.1 mmol of 3',5'-di-*O*-TBDMS 2'-deoxyguanosine). R_f (SiO₂/40% EtOAc in hexanes) = 0.3. ¹H NMR: δ 7.70 (s, 1H, Ar-H), 6.31 (t, 1H, H-1', $J = 6.3$ Hz), 4.89 (s, 2H, NH₂), 4.57 (m, 1H, H-3'), 4.05 (s, 3H, OCH₃), 3.89 (m, 1H, H-4'), 3.80 (dd, 1H, H-5', $J = 4.4, 11.2$ Hz), 3.74 (dd, 1H, H-5', $J = 2.9, 11.2$ Hz), 2.55 (app quint, 1H, H-2', $J_{app} \sim 6.3$ Hz), 2.34 (ddd, 1H, H-2', $J = 2.2, 6.0, 13.1$ Hz), 0.90 (br s, 18H, *tert*-Bu), 0.08, 0.07, and 0.06 (3s, 12H, SiCH₃). ¹³C NMR: δ 161.7, 159.5, 153.6, 137.7, 116.1, 87.8, 83.7, 72.0, 63.0, 53.9, 41.1, 26.1, 25.9, 18.6, 18.1, –4.4, –4.5, –5.2, –5.3.

3',5'-Di-*O*-(*tert*-butyldimethylsilyl)-O⁶-phenyl-2'-deoxyguanosine (**12**).²



Purification using 20% EtOAc in hexanes gave 42 mg (73%) of **12** as a yellowish solid (from 0.1 mmol of 3',5'-di-*O*-TBDMS 2'-deoxyguanosine). R_f (SiO₂/40% EtOAc in hexanes) = 0.3. ¹H NMR: δ 8.00 (s, 1H, Ar-H), 7.40 (t, 2H, Ar-H, $J = 7.7$ Hz), 7.25 (m, 3H, Ar-H), 6.34 (t, 1H, H-1', $J = 6.4$ Hz), 4.77 (s, 2H, NH₂), 4.60 (m, 1H, H-3'), 3.99 (m, 1H, H-4'), 3.83 (dd, 1H, H-5', $J = 4.1, 11.1$ Hz), 3.77 (dd, 1H, H-5', $J = 2.6, 11.1$ Hz), 2.59 (app quint, 1H, H-2', $J_{app} \sim 6.4$ Hz), 2.37 (ddd, 1H, H-2', $J = 1.4, 5.8, 12.9$ Hz), 0.923 and 0.920 (2s, 18H, *tert*-Bu), 0.10 and 0.09 (2s, 12H, SiCH₃). ¹³C NMR: δ 160.6, 159.3, 154.7, 152.7, 138.7, 129.4, 125.4, 122.1, 116.2, 87.9, 83.9, 72.1, 63.0, 41.2, 26.1, 25.9, 18.6, 18.2, –4.4, –4.5, –5.1, –5.2.

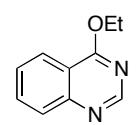
4-Methoxyquinazoline (**16a**).^{3,4}



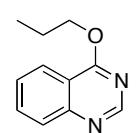
Purification using 15% EtOAc in hexanes gave 89 mg (81%) of **16a** as a colorless oil (solidifies at 0 °C). R_f (SiO₂/20% EtOAc in hexanes) = 0.4. ¹H NMR: δ 8.77 (s, 1H, Ar-H), 8.10 (d, 1H, Ar-H, $J = 8.3$ Hz), 7.89 (d, 1H, Ar-H, $J = 8.5$ Hz), 7.77 (dt, 1H,

Ar-H, $J = 1.3, 8.3$ Hz), 7.50 (t, 1H, Ar-H, $J = 7.8$ Hz), 4.13 (s, 3H, OCH₃). ¹³C NMR: δ 167.2, 154.5, 151.0, 133.6, 127.8, 127.1, 123.6, 116.7, 54.4.

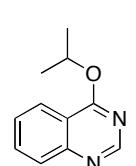
4-Ethoxyquinazoline (**16b**).⁴

 Purification using 15% EtOAc in hexanes gave 92 mg (77%) of **16b** as a colorless oil (solidifies at 0 °C). R_f (SiO₂/20% EtOAc in hexanes) = 0.4. ¹H NMR: δ 8.79 (s, 1H, Ar-H), 8.18 (d, 1H, Ar-H, $J = 8.3$ Hz), 7.92 (d, 1H, Ar-H, $J = 8.3$ Hz), 7.82 (t, 1H, Ar-H, $J = 7.3$ Hz), 7.55 (t, 1H, Ar-H, $J = 7.5$ Hz), 4.64 (q, 2H, OCH₂, $J = 7.3$ Hz), 1.52 (t, 3H, CH₃, $J = 7.3$ Hz). ¹³C NMR: δ 166.9, 154.6, 151.1, 133.6, 127.8, 127.0, 123.7, 116.9, 63.2, 14.5.

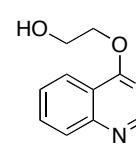
4-Propoxyquinazoline (**16c**).⁴

 Purification using 15% EtOAc in hexanes gave 101 mg (78%) of **16c** as a colorless oil. R_f (SiO₂/30% EtOAc in hexanes) = 0.5. ¹H NMR: δ 8.78 (s, 1H, Ar-H), 8.17 (d, 1H, Ar-H, $J = 8.3$ Hz), 7.91 (d, 1H, Ar-H, $J = 8.3$ Hz), 7.81 (dt, 1H, Ar-H, $J = 1.4, 7.6$ Hz), 7.54 (t, 1H, Ar-H, $J = 7.5$ Hz), 4.52 (t, 2H, OCH₂, $J = 6.8$ Hz), 1.92 (sextet, 2H, CH₂, $J = 7.5$), 1.09 (t, 3H, CH₃, $J = 7.5$ Hz). ¹³C NMR: δ 166.9, 154.6, 151.0, 133.5, 127.8, 127.0, 123.6, 116.8, 68.8, 22.2, 10.6.

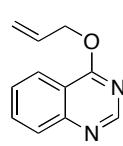
4-Isopropoxyquinazoline (**16d**).⁵

 Purification using 15% EtOAc in hexanes gave 90 mg (70%) of **16d** as a yellowish oil. R_f (SiO₂/30% EtOAc in hexanes) = 0.6. ¹H NMR: δ 8.77 (s, 1H, Ar-H), 8.15 (d, 1H, Ar-H, $J = 8.3$ Hz), 7.89 (d, 1H, Ar-H, $J = 8.3$ Hz), 7.79 (t, 1H, Ar-H, $J = 7.5$ Hz), 7.52 (t, 1H, Ar-H, $J = 7.5$ Hz), 5.62 (septet, 1H, OCH, $J = 6.3$ Hz), 1.45 (d, 6H, (CH₃)₂, $J = 6.3$ Hz). ¹³C NMR: δ 166.4, 154.6, 151.1, 133.4, 127.7, 126.8, 123.8, 117.2, 70.3, 22.0.

2-(4-Quinazolinyl)ethanol (**16e**).

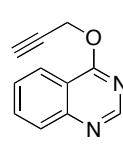
 Purification using 70% EtOAc in hexanes gave 100 mg (77%) of **16e** as a white solid. R_f (SiO₂/100% EtOAc) = 0.2. ¹H NMR: δ 8.76 (s, 1H, Ar-H), 8.16 (d, 1H, Ar-H, $J = 8.3$ Hz), 7.93 (d, 1H, Ar-H, $J = 8.3$ Hz), 7.81 (td, 1H, Ar-H, $J = 1.4, 8.3$ Hz), 7.55 (t, 1H, Ar-H, $J = 7.5$ Hz), 4.75 (t, 2H, OCH₂, $J = 4.4$ Hz), 4.08 (br s, 2H, CH₂OH), 3.36 (br s, 1H, OH). ¹³C NMR: δ 167.1, 154.0, 151.1, 134.0, 127.9, 127.4, 123.7, 116.6, 69.7, 61.9. HRMS (ESI) calcd for C₁₀H₁₁N₂O₂ [M + H]⁺ 191.0815, found 191.0815.

4-(Allyloxy)quinazoline (16f).⁶



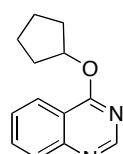
Purification using 20% EtOAc in hexanes gave 96 mg (75%) of **16f** as a yellowish liquid. R_f (SiO₂/30% EtOAc in hexanes) = 0.6. ¹H NMR: δ 8.79 (s, 1H, Ar-H), 8.20 (d, 1H, Ar-H, J = 8.3 Hz), 7.93 (d, 1H, Ar-H, J = 8.3 Hz), 7.82 (td, 1H, Ar-H, J = 1.4, 8.3 Hz), 7.56 (t, 1H, Ar-H, J = 7.5 Hz), 6.16 (m, 1H, -CH=), 5.48 (dd, 1H, =CH_{trans}, J = 1.4, 17.0 Hz), 5.33 (dd, 1H, =CH_{cis}, J = 1.4, 10.7 Hz), 5.10 (d, 2H, OCH₂, J = 5.3 Hz). ¹³C NMR: δ 166.5, 154.5, 151.2, 133.6, 132.5, 127.9, 127.2, 123.7, 118.6, 116.8, 67.6.

4-(2-Propynyoxy)quinazoline (16g).⁶



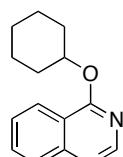
Purification using 20% EtOAc in hexanes gave 94 mg (75%) of **16g** as a white solid. R_f (SiO₂/20% EtOAc in hexanes) = 0.5. ¹H NMR: δ 8.81 (s, 1H, Ar-H), 8.19 (d, 1H, Ar-H, J = 7.8 Hz), 7.88 (d, 1H, Ar-H, J = 8.3 Hz), 7.83 (t, 1H, Ar-H, J = 8.3 Hz), 7.56 (t, 1H, Ar-H, J = 7.5 Hz), 5.21 (s, 2H, OCH₂), 2.55 (s, 1H, ≡CH). ¹³C NMR: δ 165.7, 154.1, 151.2, 133.9, 127.9, 127.4, 123.6, 116.5, 78.1, 75.4, 54.5.

4-(Cyclopentyloxy)quinazoline (16h).



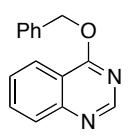
Purification using 20% EtOAc in hexanes gave 115 mg (78%) of **16h** as a colorless oil. R_f (SiO₂/30% EtOAc in hexanes) = 0.5. ¹H NMR: δ 8.78 (s, 1H, Ar-H), 8.12 (d, 1H, Ar-H, J = 8.3 Hz), 7.89 (d, 1H, Ar-H, J = 8.3 Hz), 7.79 (t, 1H, Ar-H, J = 8.3 Hz), 7.51 (t, 1H, Ar-H, J = 7.5 Hz), 5.72 (m, 1H, OCH), 2.08–2.01 (m, 2H, CH₂), 1.95–1.89 (m, 2H, CH₂), 1.88–1.81 (m, 2H, CH₂), 1.73–1.65 (m, 2H, CH₂). ¹³C NMR: δ 166.7, 154.7, 151.1, 133.5, 127.8, 126.9, 123.8, 117.3, 79.9, 33.0, 24.1. HRMS (ESI) calcd for C₁₃H₁₅N₂O [M + H]⁺ 215.1179, found 215.1176.

4-(Cyclohexyloxy)quinazoline (16i).⁷

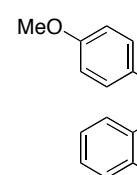


Purification using 20% EtOAc in hexanes gave 120 mg (77%) of **16i** as a colorless oil. R_f (SiO₂/30% EtOAc in hexanes) = 0.5. ¹H NMR: δ 8.77 (s, 1H, Ar-H), 8.17 (d, 1H, Ar-H, J = 7.8 Hz), 7.90 (d, 1H, Ar-H, J = 8.3 Hz), 7.80 (t, 1H, Ar-H, J = 8.0 Hz), 7.53 (t, 1H, Ar-H, J = 7.5 Hz), 5.41 (m, 1H, OCH), 2.08–2.05 (m, 2H, CH₂), 1.85–1.82 (m, 2H, CH₂), 1.73–1.66 (m, 2H, CH₂), 1.62–1.58 (m, 1H, CH₂), 1.54–1.46 (m, 2H, CH₂), 1.42–1.37 (m, 1H, CH₂). ¹³C NMR: δ 166.5, 154.7, 151.2, 133.5, 127.8, 126.9, 123.8, 117.3, 75.0, 31.6, 25.7, 23.8.

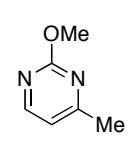
4-(Benzylxy)quinazoline (16j).⁸


 Purification using 5% MeOH in hexanes gave 155 mg (96%) of **16j** as a colorless oil. R_f (SiO_2 /40% EtOAc in hexanes) = 0.4. ^1H NMR: δ 8.83 (s, 1H, Ar-H), 8.20 (dd, 1H, Ar-H, J = 0.7, 8.2 Hz), 7.94 (d, 1H, Ar-H, J = 8.4 Hz), 7.82 (td, 1H, Ar-H, J = 1.4, 7.0 Hz), 7.54 (m, 3H, Ar-H), 7.41 (t, 2H, Ar-H, J = 7.2 Hz), 7.36 (t, 1H, Ar-H, J = 7.2 Hz), 5.64 (br s, 2H, OCH_2). ^{13}C NMR: δ 166.7, 154.5, 151.2, 136.3, 133.8, 128.8, 128.5, 128.3, 127.9, 127.2, 123.7, 116.8, 68.7.

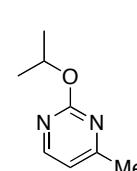
4-(4-Methoxyphenoxy)quinazoline (16k).⁹


 Purification using 30% EtOAc in hexanes gave 165 mg (96%) of **16k** as a white solid. R_f (SiO_2 /30% EtOAc in hexanes) = 0.5. ^1H NMR: δ 8.77 (s, 1H, Ar-H), 8.36 (d, 1H, Ar-H, J = 8.1 Hz), 7.99 (d, 1H, Ar-H, J = 8.4), 7.89 (t, 1H, Ar-H, J = 7.1 Hz), 7.63 (t, 1H, Ar-H, J = 7.5 Hz), 7.18 (br d, 2H, Ar-H, J = 8.9 Hz), 6.98 (br d, 2H, Ar-H, J = 8.9 Hz), 3.83 (s, 3H, OCH_3). ^{13}C NMR: δ 167.4, 157.5, 154.5, 151.7, 145.8, 134.1, 128.0, 127.6, 123.7, 122.8, 116.6, 114.9, 55.7.

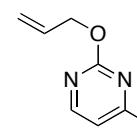
2-(Methoxy)-4-methylpyrimidine (17a).¹⁰


 Purification using 15% EtOAc in hexanes gave 65 mg (77%) of **17a** as a clear oil. R_f (SiO_2 /20% EtOAc in hexanes) = 0.2. ^1H NMR: δ 8.33 (d, 1H, Ar-H, J = 4.8 Hz), 6.78 (d, 1H, Ar-H, J = 4.8 Hz), 3.98 (s, 3H, OCH_3), 2.44 (s, 3H, CH_3). ^{13}C NMR: δ 170.2, 165.7, 158.7, 114.6, 54.8, 24.2.

2-(Isopropoxy)-4-methylpyrimidine (17b).

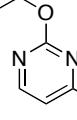

 Purification using 15% EtOAc in hexanes gave 76 mg (73%) of **17b** as a yellowish oil. R_f (SiO_2 /20% EtOAc in hexanes) = 0.5. ^1H NMR: δ 8.31 (d, 1H, Ar-H, J = 4.8 Hz), 6.74 (d, 1H, Ar-H, J = 4.8 Hz), 5.27 (septet, 1H, OCH , J = 6.3 Hz), 2.43 (s, 3H, CH_3), 1.38 (d, 6H, $(\text{CH}_3)_2$, J = 6.3 Hz). ^{13}C NMR: δ 170.1, 164.9, 158.6, 114.2, 70.0, 24.3, 22.1. HRMS (ESI) calcd for $\text{C}_8\text{H}_{13}\text{N}_2\text{O}$ [$\text{M} + \text{H}$]⁺ 153.1022, found 153.1033.

2-(Allyloxy)-4-methylpyrimidine (17c).

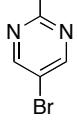

 Purification using 20% EtOAc in hexanes gave 77 mg (75%) of **17c** as a yellowish oil. R_f (SiO_2 /40% EtOAc in hexanes) = 0.5. ^1H NMR: δ 8.31 (d, 1H, Ar-H, J = 4.8 Hz), 6.76 (d, 1H, Ar-H, J = 4.8 Hz), 6.06 (m, 1H, -CH=), 5.39 (d,

1H, =CH_{trans}, *J* = 17.5 Hz), 5.22 (d, 1H, =CH_{cis}, *J* = 10.2 Hz), 4.86 (d, 2H, OCH₂, *J* = 5.3 Hz), 2.43 (s, 3H, Ar-CH₃). ¹³C NMR: δ 170.2, 165.0, 158.6, 133.0, 117.8, 114.7, 68.0, 24.2. HRMS (ESI) calcd for C₈H₁₁N₂O [M + H]⁺ 151.0866, found 151.0861.

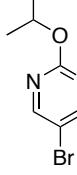
2-[(4-Methylpyrimidin-2-yl)oxy]ethanol (17d).

 Purification using 60% EtOAc in hexanes gave 74 mg (70%) of **17d** as a pale yellow liquid. *R*_f (SiO₂/EtOAc) = 0.3. ¹H NMR: δ 8.34 (d, 1H, Ar-H, *J* = 4.9 Hz), 6.82 (d, 1H, Ar-H, *J* = 4.9 Hz), 4.49 (t, 2H, OCH₂, *J* = 4.3 Hz), 3.97 (t, 2H, OCH₂, *J* = 4.3 Hz), 2.97 (br s, 1H, OH), 2.46 (s, 3H, CH₃). ¹³C NMR: δ 170.4, 165.2, 158.7, 115.0, 69.6, 62.0, 24.2. HRMS (ESI) calcd for C₇H₁₁N₂O₂ [M + H]⁺ 155.0815, found 155.0814.

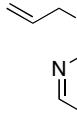
5-Bromo-2-methoxypyrimidine (18a).¹¹

 Purification using 30% EtOAc in hexanes gave 85 mg (78%) of **18a** as a colorless liquid. *R*_f (SiO₂/40% EtOAc in hexanes) = 0.5. ¹H NMR: δ 8.52 (s, 2H, Ar-H), 3.98 (s, 3H, OCH₃). ¹³C NMR: δ 164.4, 159.7, 112.0, 55.6.

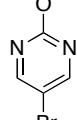
5-Bromo-2-isopropoxypyrimidine (18b).¹²

 Purification using 30% EtOAc in hexanes gave 100 mg (80%) of **18b** as a clear oil. *R*_f (SiO₂/40% EtOAc in hexanes) = 0.6. ¹H NMR: δ 8.48 (s, 2H, Ar-H), 5.19 (septet, 1H, OCH, *J* = 6.3 Hz), 1.37 (d, 6H, (CH₃)₂, *J* = 6.3 Hz). ¹³C NMR: δ 163.6, 159.7, 111.3, 71.3, 21.8.

2-(Allyloxy)-5-bromopyrimidine (18c).¹³

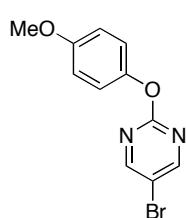
 Purification using 30% EtOAc in hexanes gave 97 mg (79%) of **18c** as a yellowish solid. *R*_f (SiO₂/40% EtOAc in hexanes) = 0.5. ¹H NMR: δ 8.50 (s, 2H, Ar-H), 6.06 (m, 1H, -CH=), 5.41 (dq, 1H, =CH_{trans}, *J* = 1.5, 17.2 Hz), 5.39 (dd, 1H, =CH_{cis}, *J* = 1.2, 10.5 Hz), 4.72 (dt, 2H, OCH₂, *J* = 1.4, 6.8 Hz). ¹³C NMR: δ 163.7, 159.7, 132.3, 118.4, 112.0, 68.8.

2-(5-Bromopyrimidin-2-yloxy)ethanol (18d).

 Purification using 40% EtOAc in hexanes gave 94 mg (75%) of **18d** as a white solid. *R*_f (SiO₂/40% EtOAc in hexanes) = 0.4. ¹H NMR: δ 8.51 (s, 2H, Ar-H), 4.45 (t, 2H, OCH₂, *J* = 4.6 Hz), 3.94–3.97 (m, 2H, OCH₂), 2.98 (t, 1H, OH, *J* = 6.3 Hz).

¹³C NMR: δ 164.0, 159.9, 112.3, 70.1, 61.4. HRMS (ESI) calcd for C₆H₈BrN₂O₂ [M + H]⁺ 218.9764, found 218.9763.

5-Bromo-2-(4-methoxyphenoxy)pyrimidine (18e**).¹⁴**



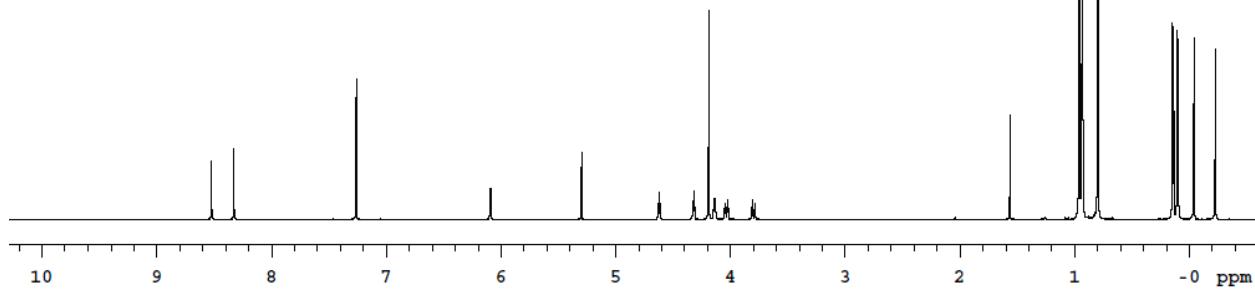
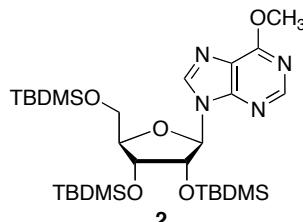
Purification using 20% EtOAc in hexanes gave 114 mg (71%) of **18e** as a white solid. *R*_f (SiO₂/20% EtOAc in hexanes) = 0.4. ¹H NMR: δ 8.55 (s, 2H, Ar-H), 7.09 (d, 2H, Ar-H, *J* = 8.9 Hz), 6.93 (d, 2H, Ar-H, *J* = 8.9 Hz), 3.81 (s, 3H, OCH₃). ¹³C NMR: δ 164.4, 160.1, 157.3, 146.3, 122.4, 114.8, 113.1, 55.7.

1203-hp-01-050

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-hp-01-050
INOVA-500 "riga"

Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
10 repetitions
OBSERVE H1, 499.7707217 MHz
DATA PROCESSING
FT size 32768
Total time 0 min, 34 sec

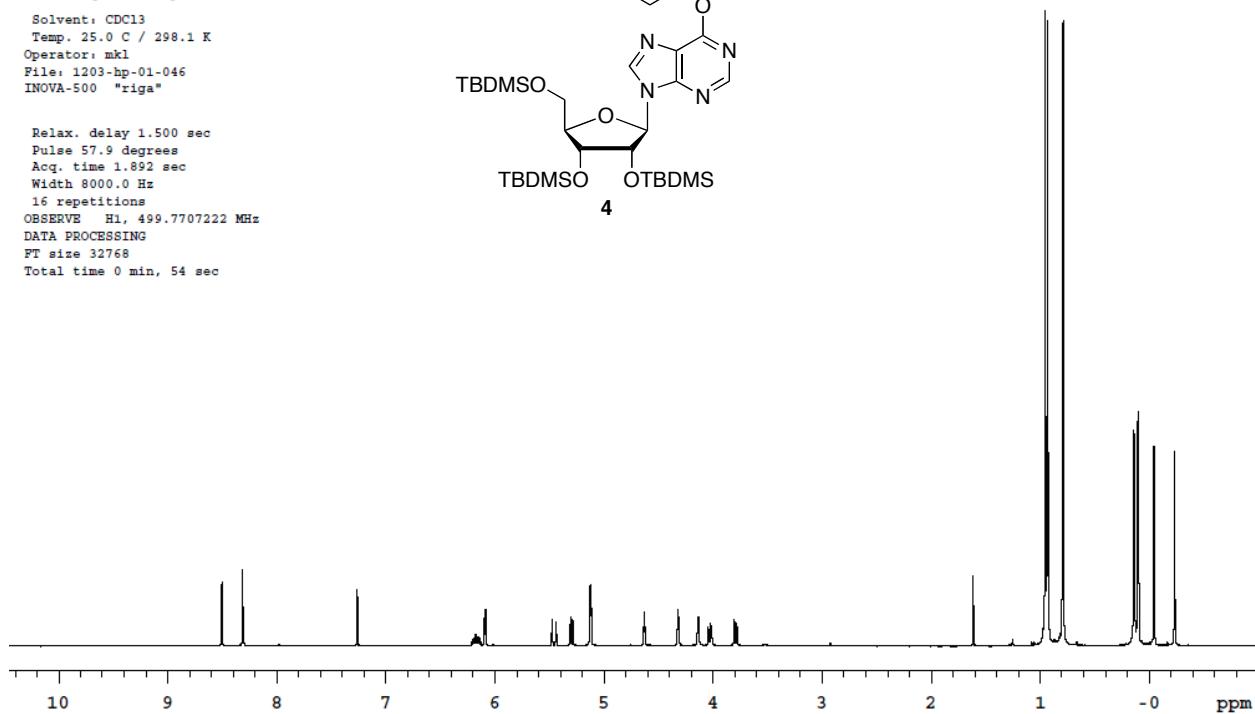
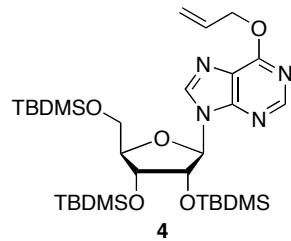


1203-hp-01-046

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-hp-01-046
INOVA-500 "riga"

Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
16 repetitions
OBSERVE H1, 499.7707222 MHz
DATA PROCESSING
FT size 32768
Total time 0 min, 54 sec

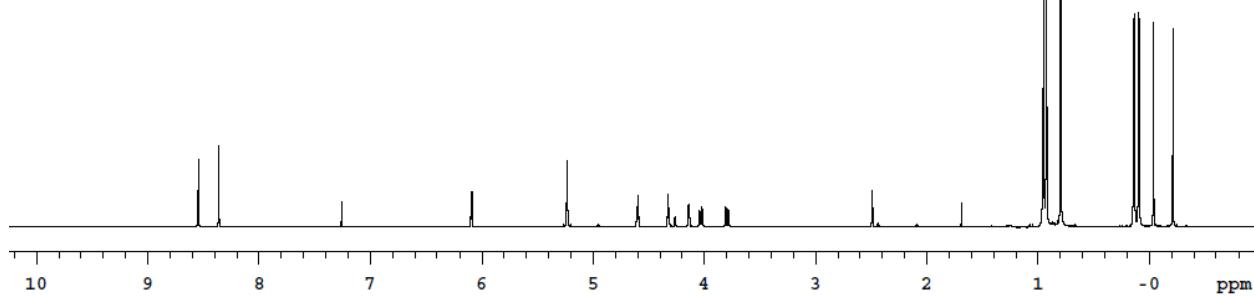
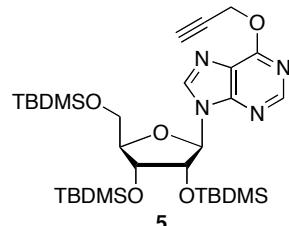


1203-hp-01-048

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-hp-01-048
INOVA-500 "riga"

Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
10 repetitions
OBSERVE H1, 499.7707217 MHz
DATA PROCESSING
FT size 32768
Total time 0 min, 34 sec

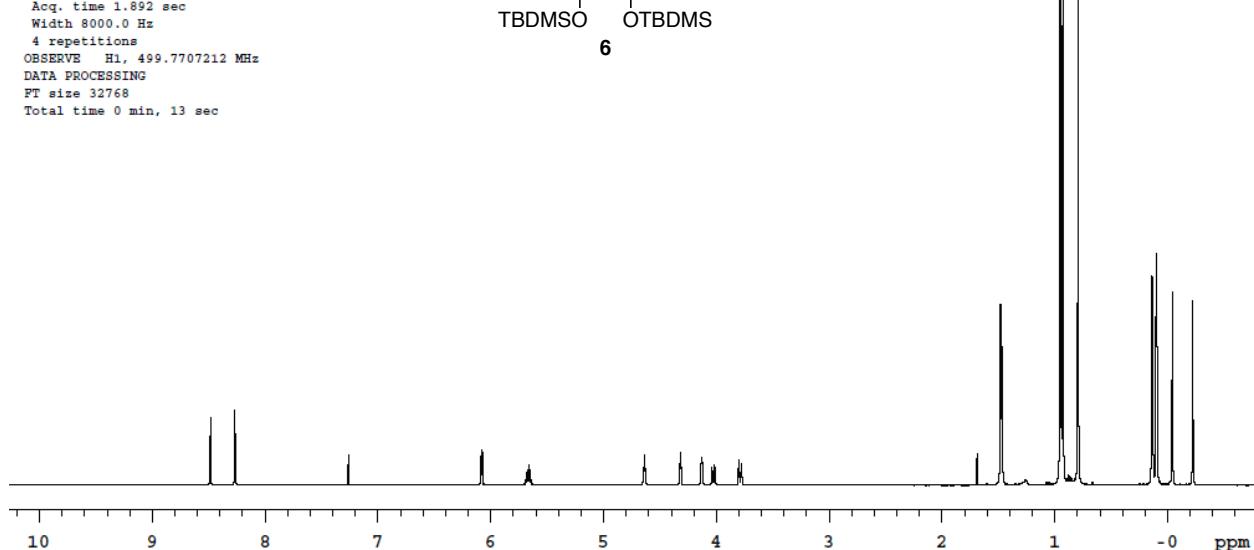
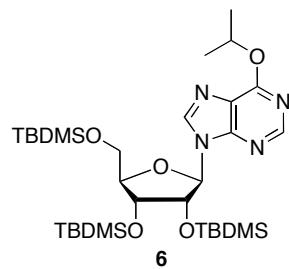


1203-hp-01-047

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-hp-01-047
INOVA-500 "riga"

Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
4 repetitions
OBSERVE H1, 499.7707212 MHz
DATA PROCESSING
FT size 32768
Total time 0 min, 13 sec

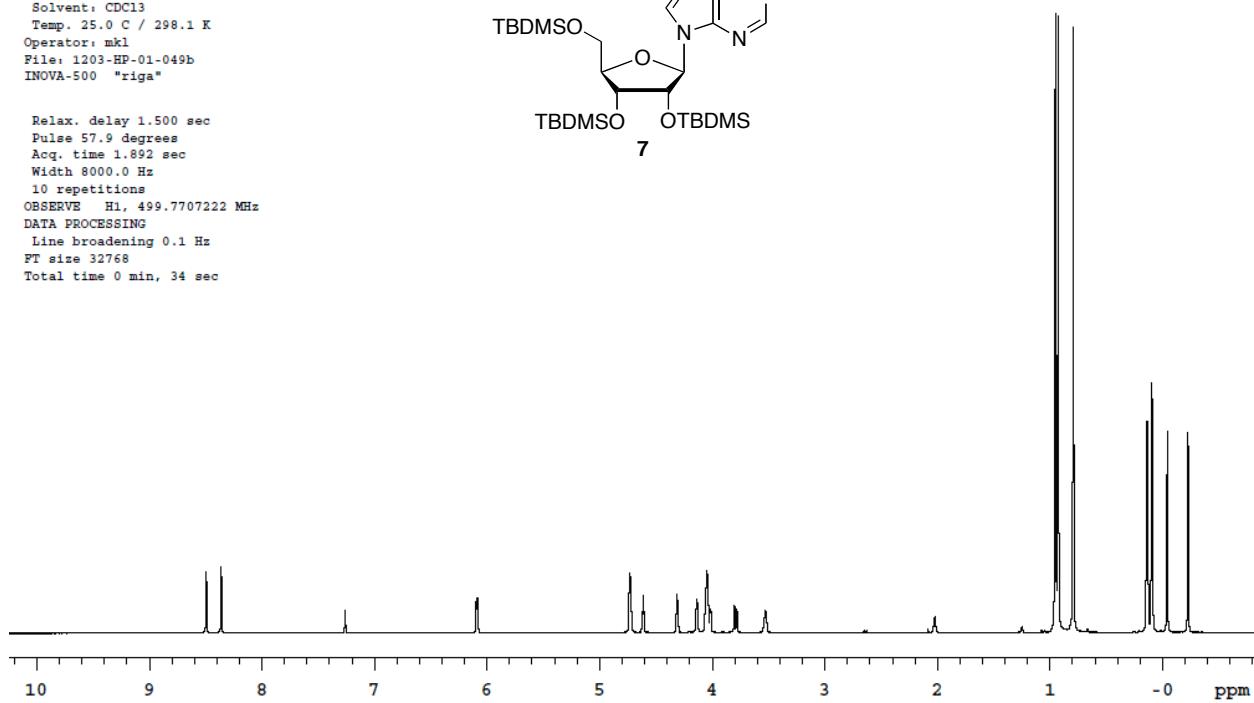
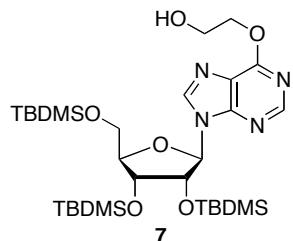


1203-HP-01-049b

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01-049b
INOVA-500 "riga"

Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
10 repetitions
OBSERVE H1, 499.7707222 MHz
DATA PROCESSING
Line broadening 0.1 Hz
FT size 32768
Total time 0 min, 34 sec

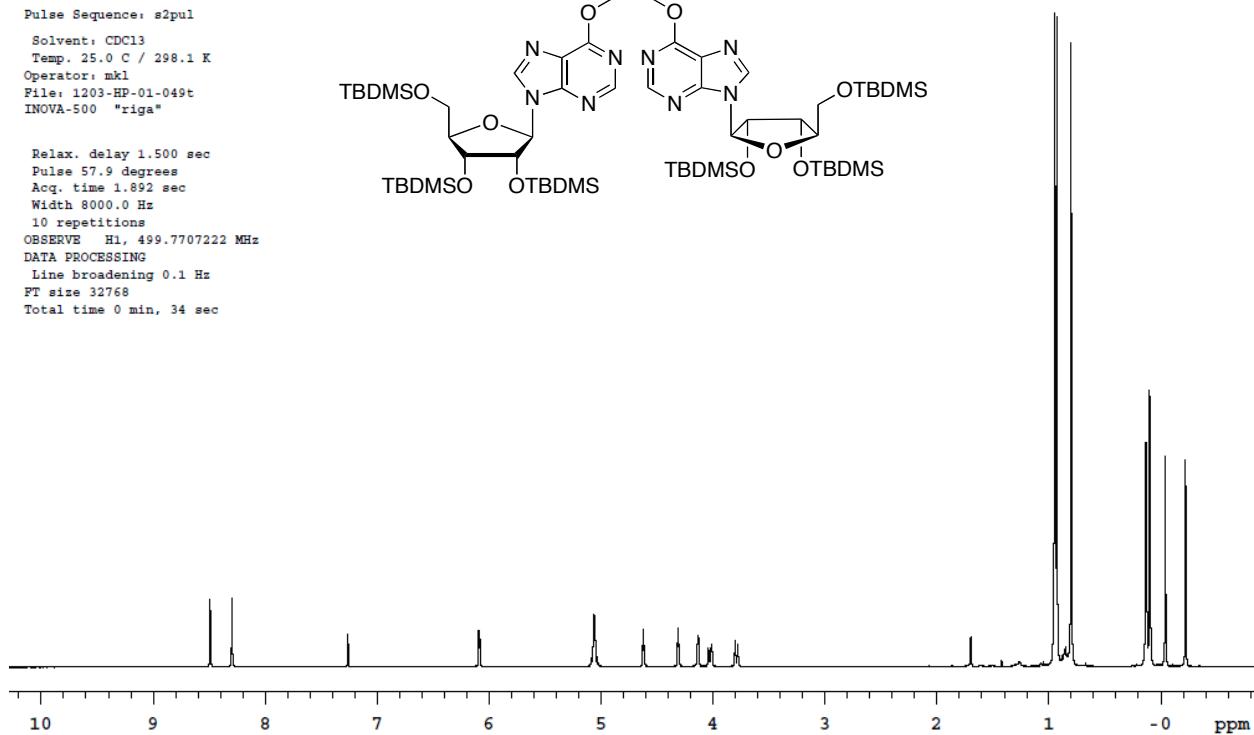
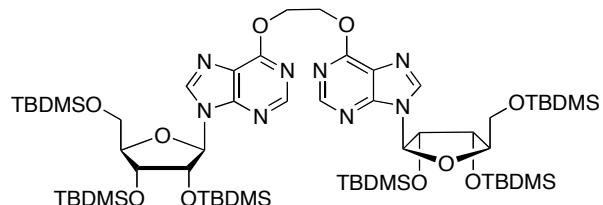


1203-HP-01-049t

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01-049t
INOVA-500 "riga"

Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
10 repetitions
OBSERVE H1, 499.7707222 MHz
DATA PROCESSING
Line broadening 0.1 Hz
FT size 32768
Total time 0 min, 34 sec

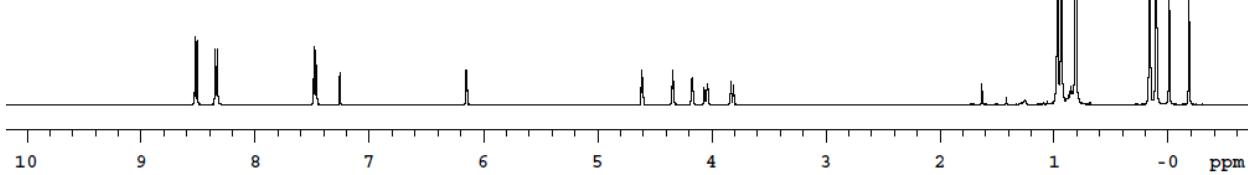
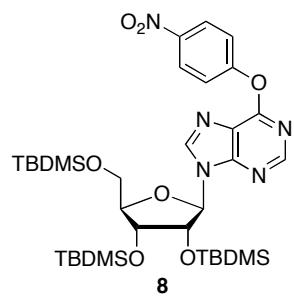


1203-HP-01-064-1H

Pulse Sequence: s2pul

Solvent: *cdcl*3
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01-064-1H
INNOVA-500 "riga"

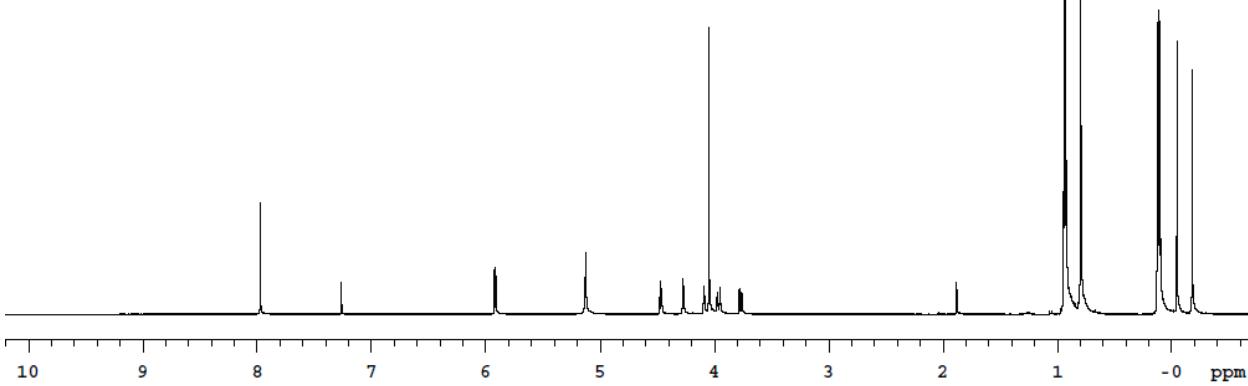
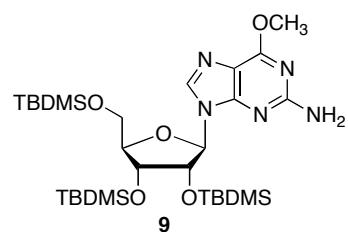
Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
82 repetitions
OBSERVE H1, 499.7707217 MHz
DATA PROCESSING
FT size 32768
Total time 5 min, 43 sec



1203-HP-01-066repeat-1H

Pulse Sequence: s2pul
Solvent: *cdcl*3
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01-066repeat-1H
INNOVA-500 "riga"

Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
79 repetitions
OBSERVE H1, 499.7707217 MHz
DATA PROCESSING
FT size 32768
Total time 5 min, 43 sec

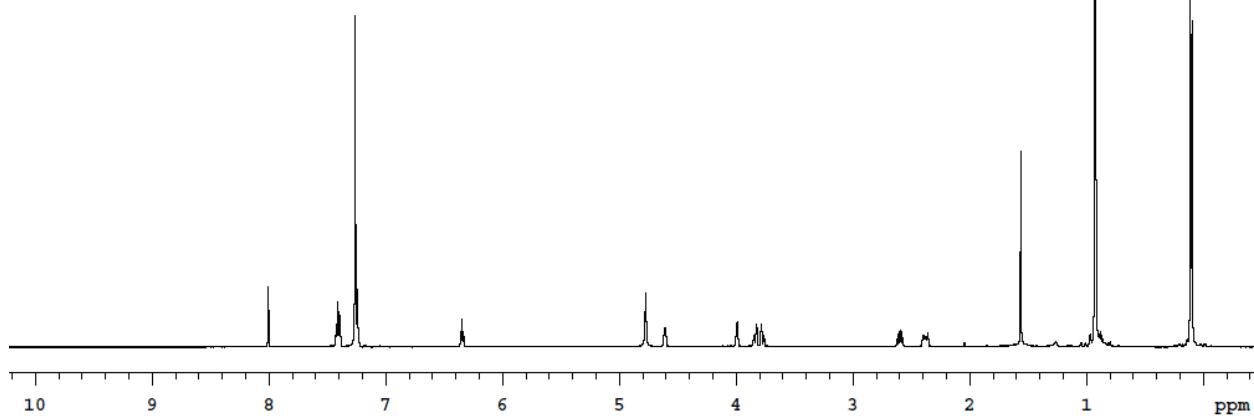
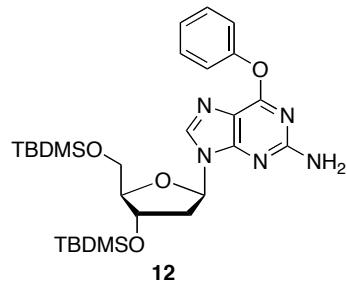


1203-HP-01-067-1H

Pulse Sequence: s2pul

Solvent: cdcl3
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01-067-1H
INOVA-500 "riga"

Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
67 repetitions
OBSERVE H1, 499.7707222 MHz
DATA PROCESSING
FT size 32768
Total time 5 min, 43 sec

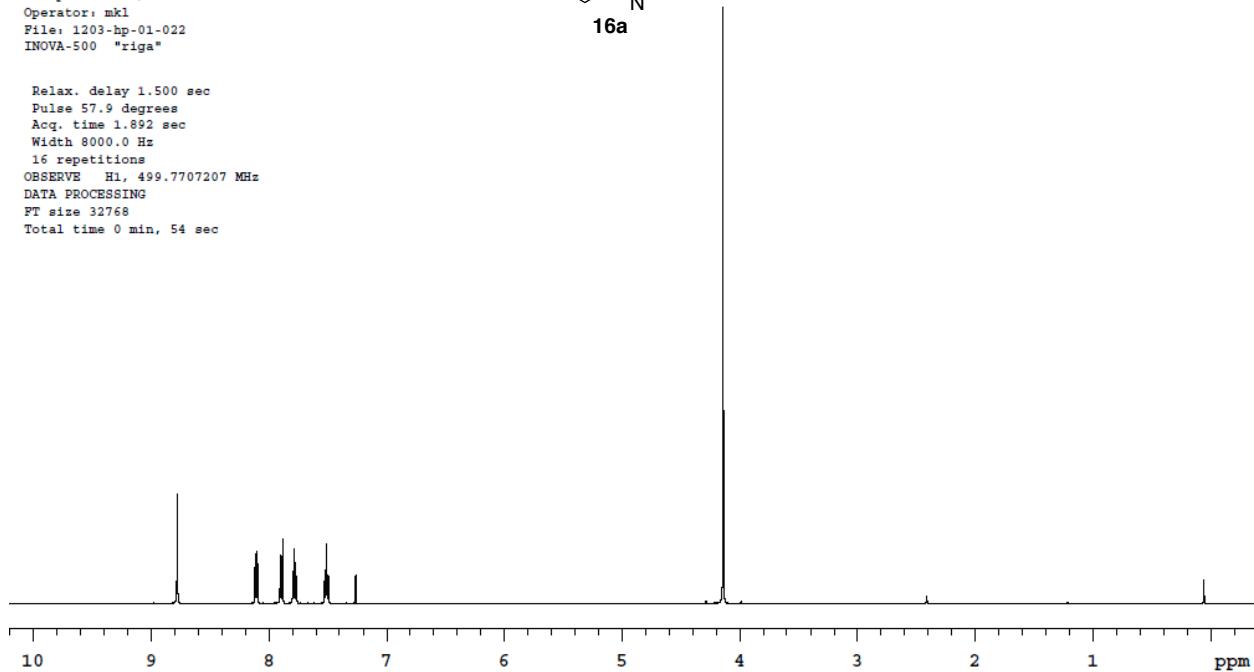
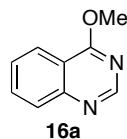


1203-hp-01-022

Pulse Sequence: s2pul

Solvent: CDCl3
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-hp-01-022
INOVA-500 "riga"

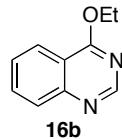
Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
16 repetitions
OBSERVE H1, 499.7707207 MHz
DATA PROCESSING
FT size 32768
Total time 0 min, 54 sec



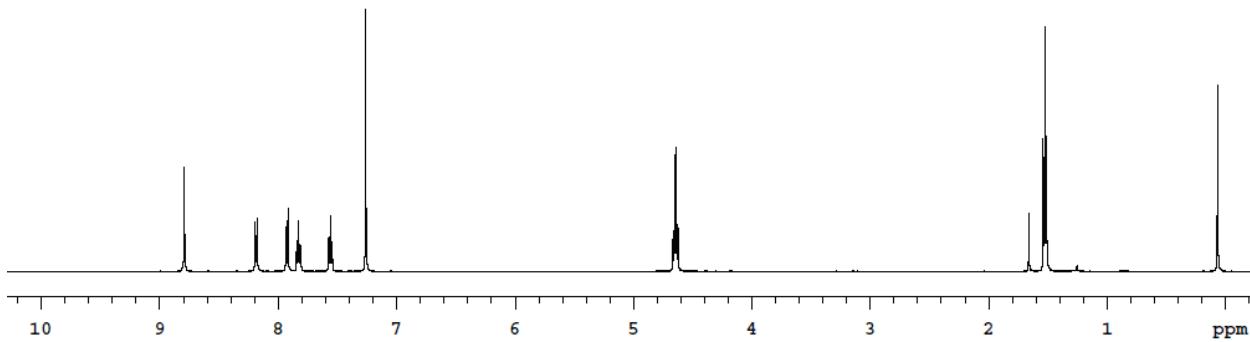
1203-HP-01-023

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01-023
INOVA-500 "riga"



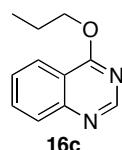
Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
12 repetitions
OBSERVE H1, 499.7707212 MHz
DATA PROCESSING
Line broadening 0.1 Hz
FT size 32768
Total time 11 min, 20 sec



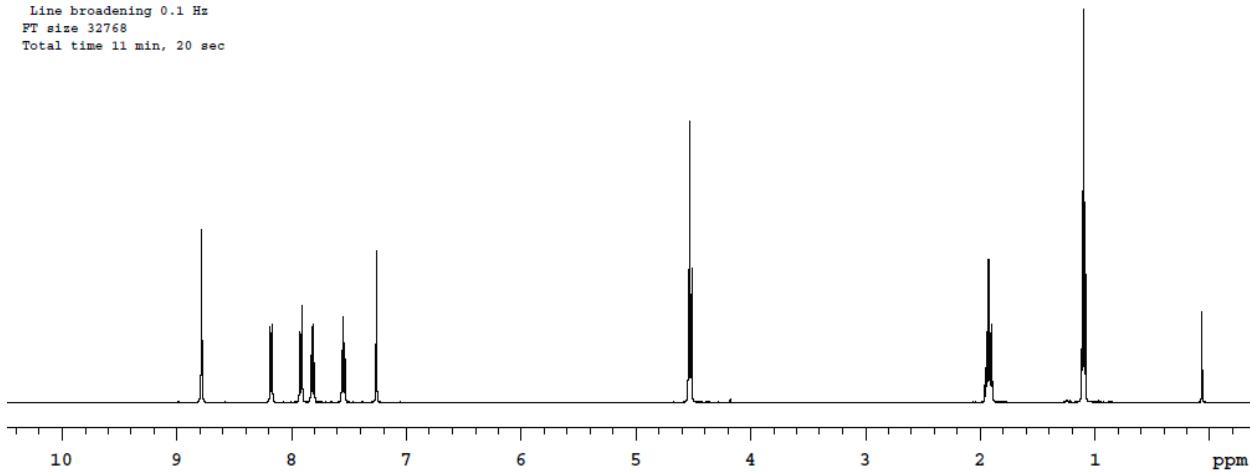
1203-HP-01-024

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01-024
INOVA-500 "riga"



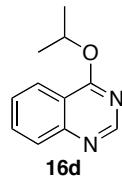
Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
32 repetitions
OBSERVE H1, 499.7707212 MHz
DATA PROCESSING
Line broadening 0.1 Hz
FT size 32768
Total time 11 min, 20 sec



1203-hp-01-025

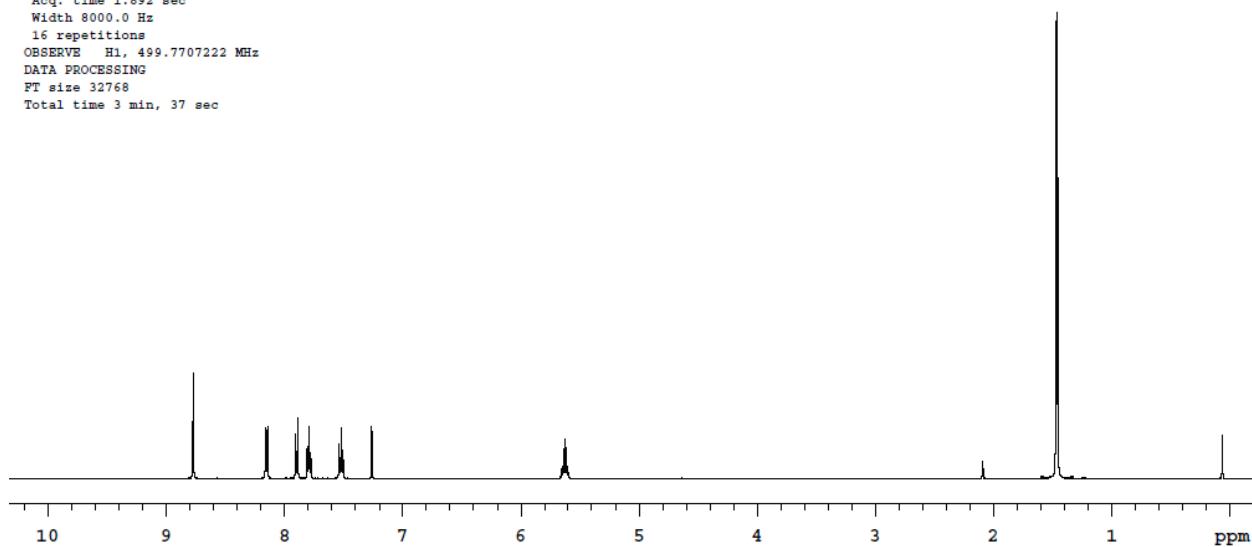
Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-hp-01-025
INOVA-500 "riga"



16d

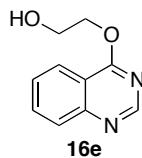
Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
16 repetitions
OBSERVE H1, 499.7707222 MHz
DATA PROCESSING
FT size 32768
Total time 3 min, 37 sec



1203-HP-01-028m

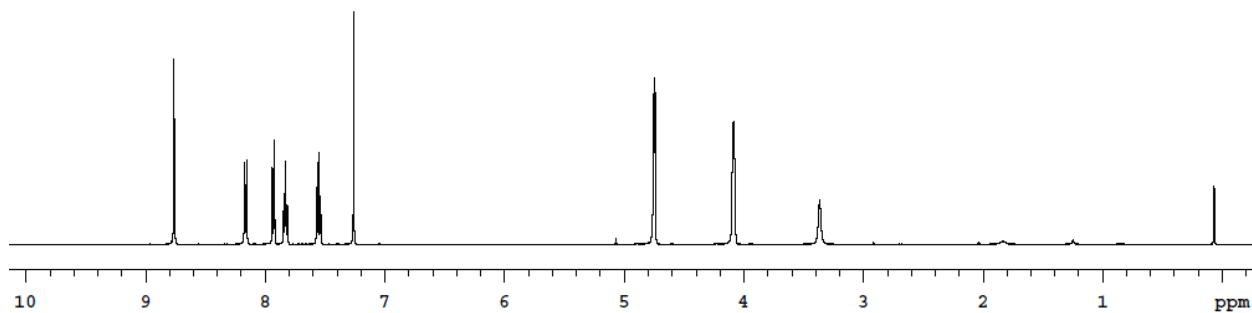
Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01-028m
INOVA-500 "riga"



16e

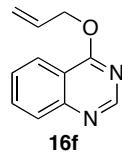
Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
64 repetitions
OBSERVE H1, 499.7707212 MHz
DATA PROCESSING
FT size 32768
Total time 3 min, 37 sec



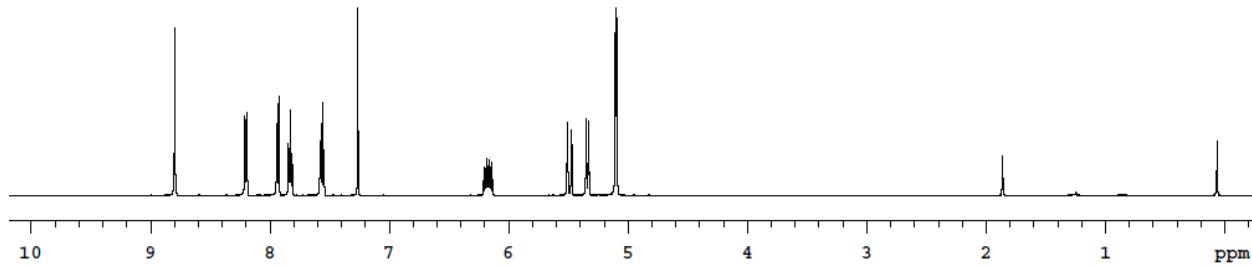
1203-HP-01-029

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01-029
INOVA-500 "riga"



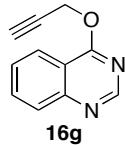
Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
164 repetitions
OBSERVE H1, 499.7707212 MHz
DATA PROCESSING
Line broadening 0.1 Hz
FT size 32768
Total time 11 min, 20 sec



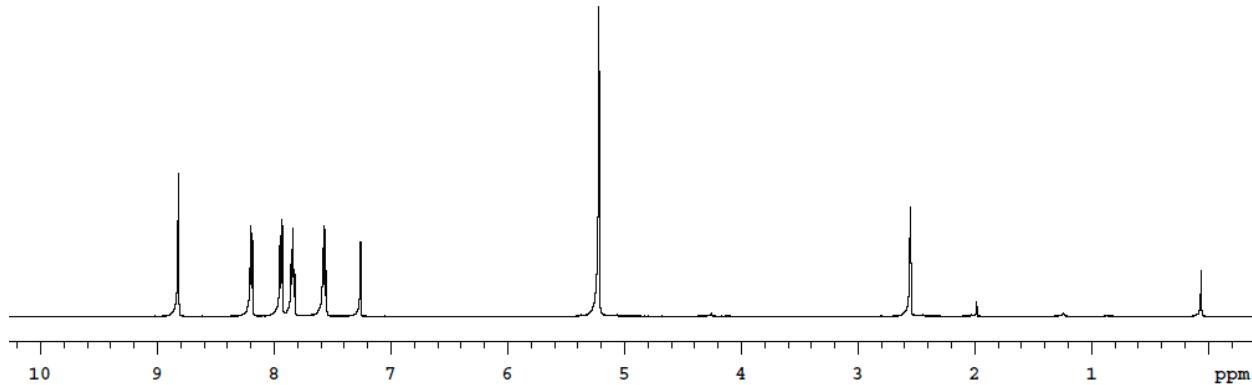
1203-HP-01-030

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01-030
INOVA-500 "riga"



Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
20 repetitions
OBSERVE H1, 499.7707212 MHz
DATA PROCESSING
Line broadening 0.1 Hz
FT size 32768
Total time 11 min, 20 sec



1203-hp-01-032p

Pulse Sequence: s2pul

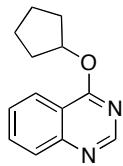
Solvent: CDCl₃

Temp. 25.0 C / 298.1 K

Operator: mkl

File: 1203-hp-01-032p

INOVA-500 "riga"



16h

Relax. delay 1.500 sec

Pulse 57.9 degrees

Acq. time 1.892 sec

Width 8000.0 Hz

16 repetitions

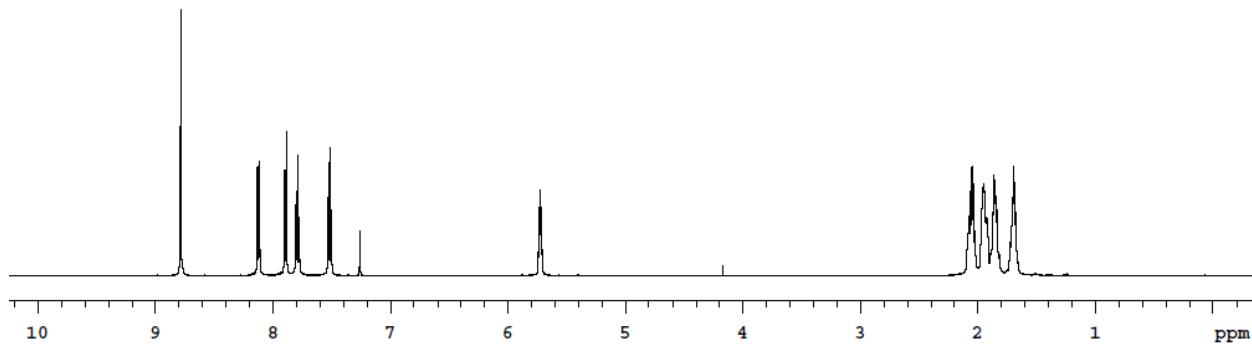
OBSERVE H1, 499.7707217 MHz

DATA PROCESSING

Line broadening 0.1 Hz

FT size 32768

Total time 0 min, 54 sec



1203-hp-01-031p

Pulse Sequence: s2pul

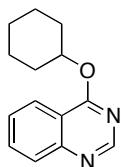
Solvent: CDCl₃

Temp. 25.0 C / 298.1 K

Operator: mkl

File: 1203-hp-01-031p

INOVA-500 "riga"



16i

Relax. delay 1.500 sec

Pulse 57.9 degrees

Acq. time 1.892 sec

Width 8000.0 Hz

16 repetitions

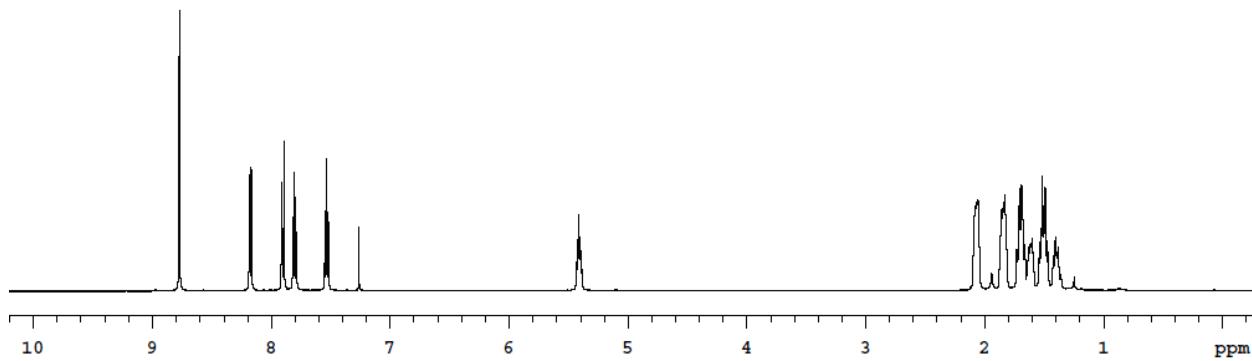
OBSERVE H1, 499.7707212 MHz

DATA PROCESSING

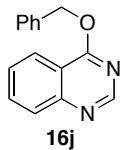
Line broadening 0.1 Hz

FT size 32768

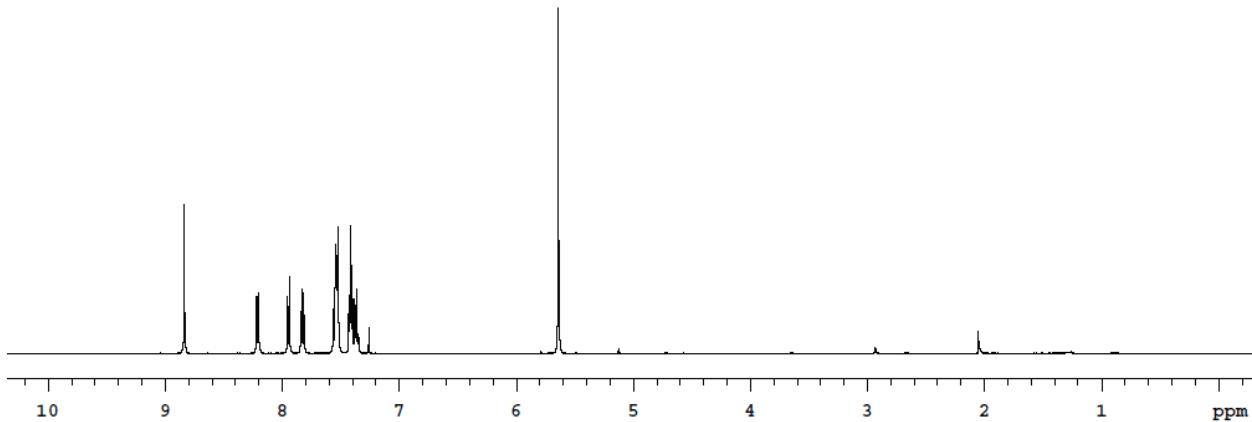
Total time 0 min, 54 sec



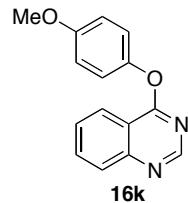
1203-HP-01-033repeat-puri1-1H
 Pulse Sequence: s2pul
 Solvent: cdcl3
 Temp. 25.0 C / 298.1 K
 Operator: mkl
 File: 1203-HP-01-033repeat-puri1-1H
 INOVA-500 "riga"



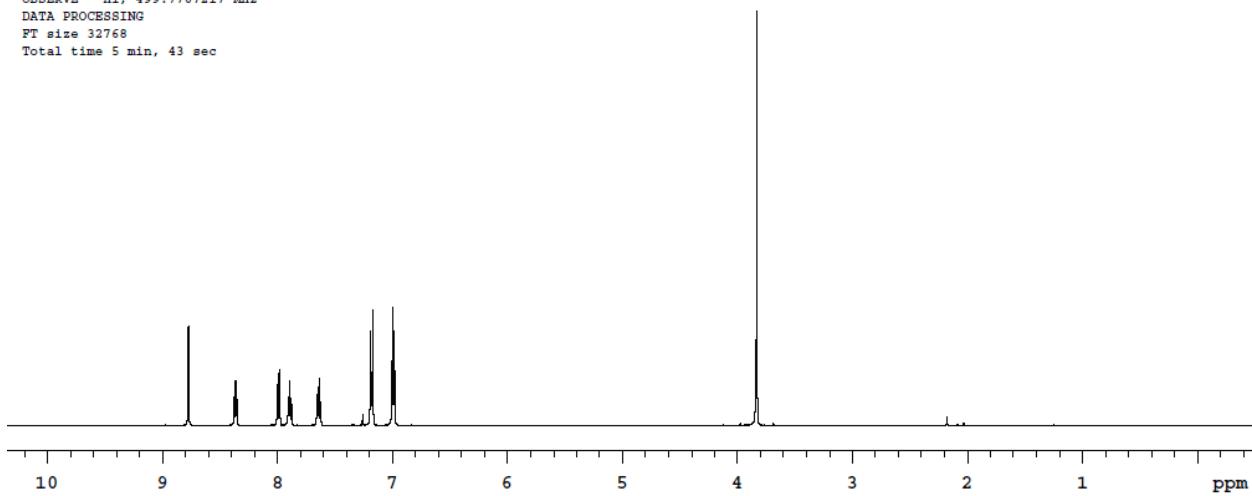
Relax. delay 2.000 sec
 Pulse 45.0 degrees
 Acq. time 1.892 sec
 Width 8000.0 Hz
 99 repetitions
 OBSERVE H1, 499.7707217 MHz
 DATA PROCESSING
 FT size 32768
 Total time 13 min, 0 sec



1203-HP-01-062-1h
 Pulse Sequence: s2pul
 Solvent: cdcl3
 Temp. 25.0 C / 298.1 K
 Operator: mkl
 File: 1203-HP-01-062-1h
 INOVA-500 "riga"



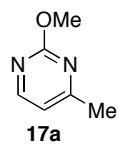
Relax. delay 2.000 sec
 Pulse 45.0 degrees
 Acq. time 1.892 sec
 Width 8000.0 Hz
 58 repetitions
 OBSERVE H1, 499.7707217 MHz
 DATA PROCESSING
 FT size 32768
 Total time 5 min, 43 sec



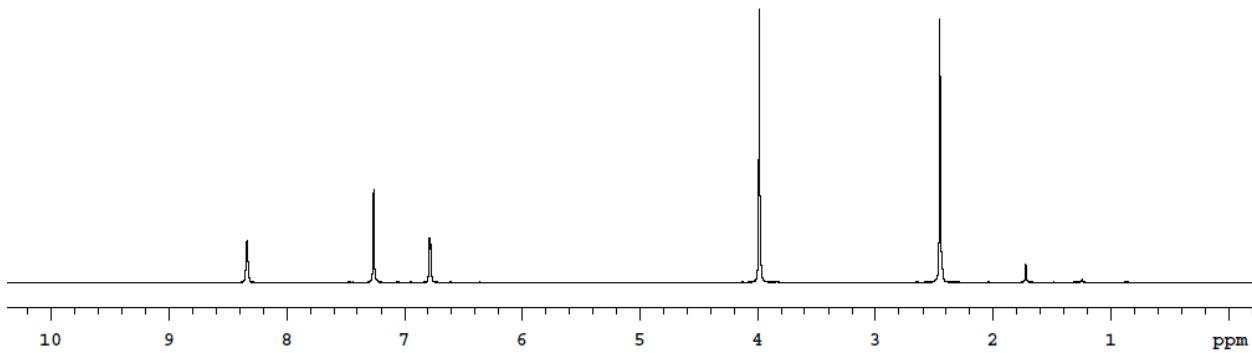
1203-hp-01-037

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-hp-01-037
INOVA-500 "riga"



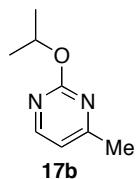
Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
64 repetitions
OBSERVE H1, 499.7707197 MHz
DATA PROCESSING
FT size 32768
Total time 3 min, 37 sec



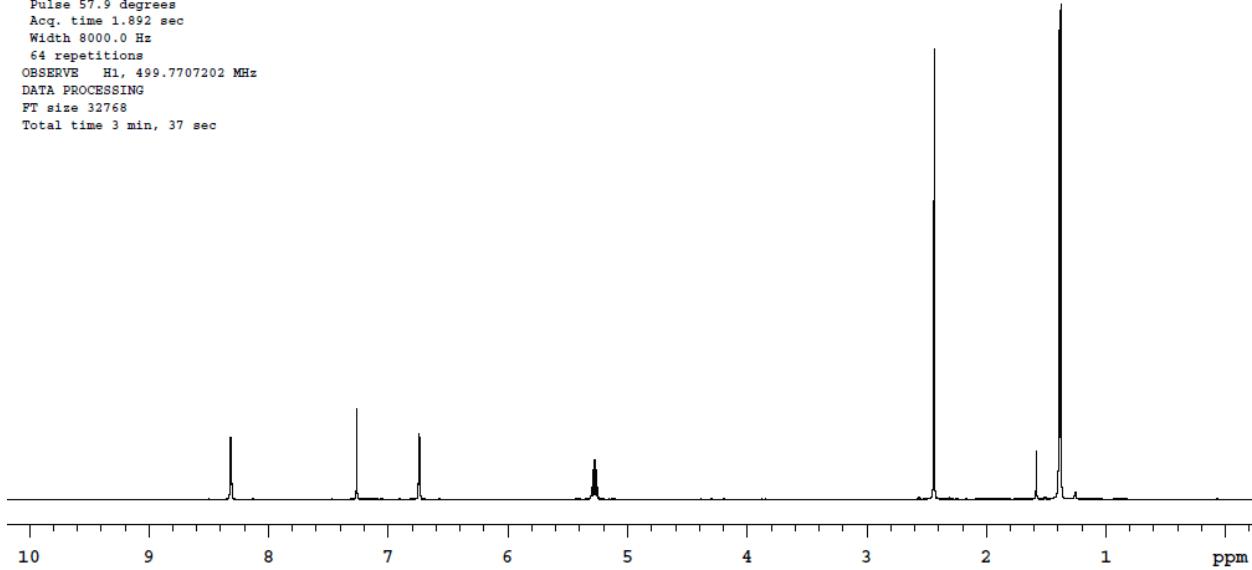
1203-hp-01-038

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-hp-01-038
INOVA-500 "riga"



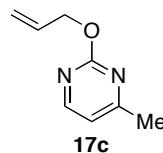
Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
64 repetitions
OBSERVE H1, 499.7707202 MHz
DATA PROCESSING
FT size 32768
Total time 3 min, 37 sec



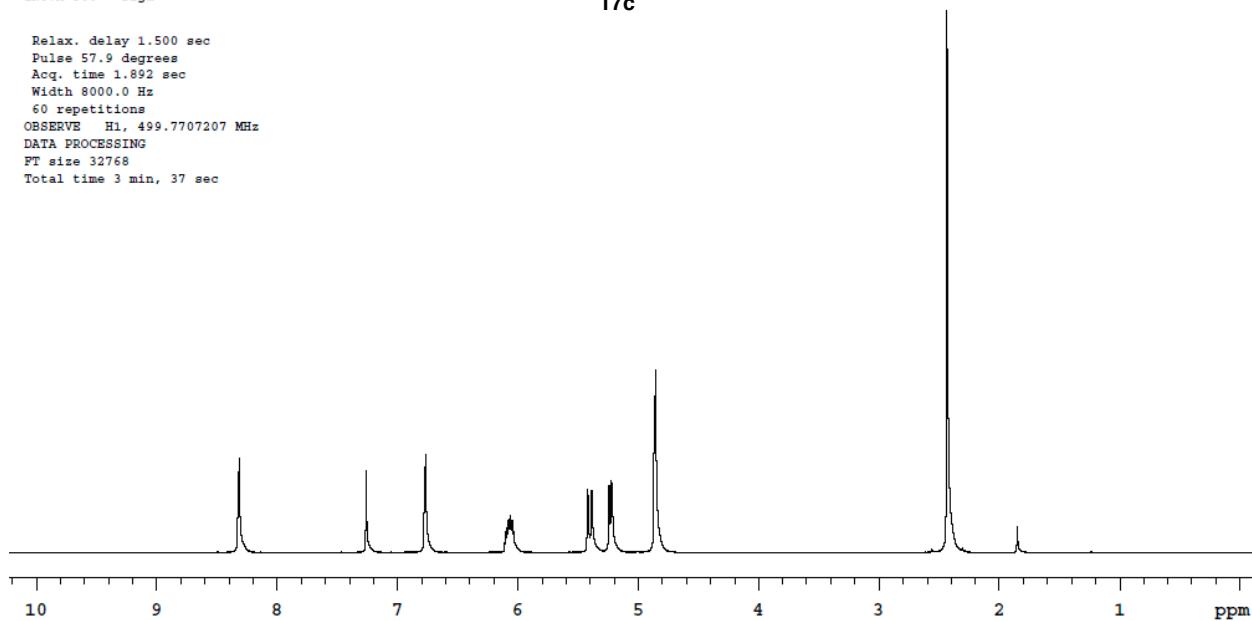
1203-hp-01-039

Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-hp-01-039
INOVA-500 "riga"



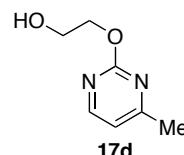
Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
60 repetitions
OBSERVE H1, 499.7707207 MHz
DATA PROCESSING
FT size 32768
Total time 3 min, 37 sec



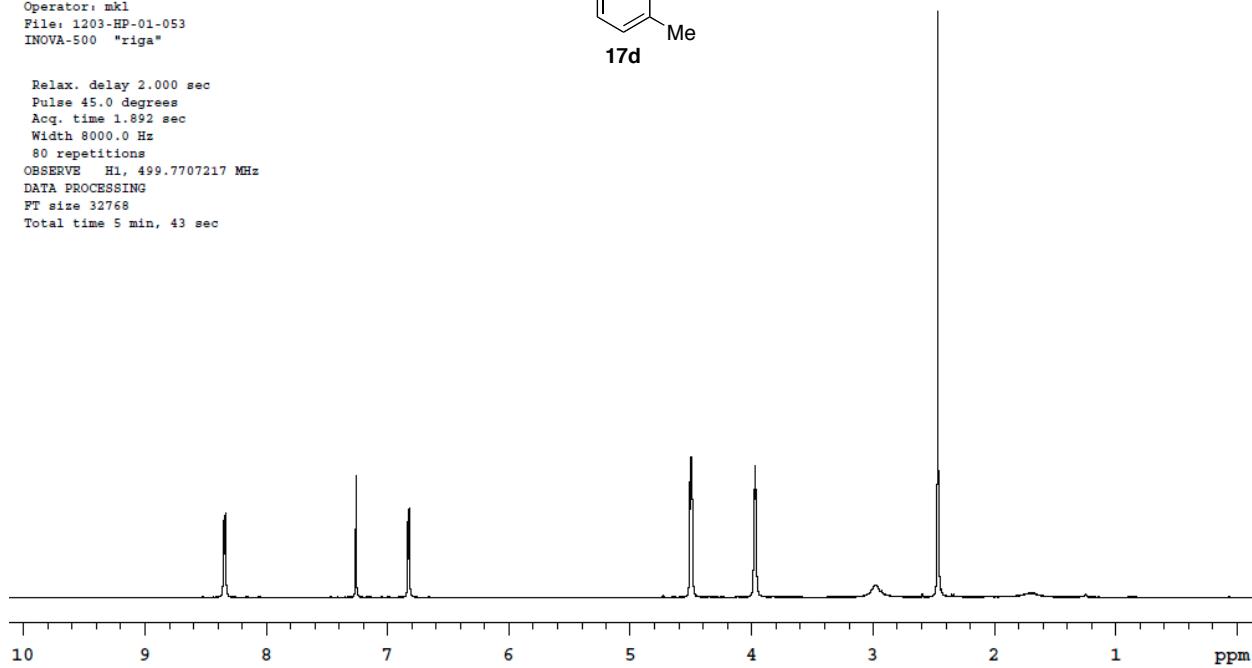
1203-HP-01-053

Pulse Sequence: s2pul

Solvent: cdcl3
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01-053
INOVA-500 "riga"



Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
80 repetitions
OBSERVE H1, 499.7707217 MHz
DATA PROCESSING
FT size 32768
Total time 5 min, 43 sec



1203-hp-01-041b

Pulse Sequence: s2pul

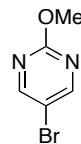
Solvent: CDCl₃

Temp. 25.0 C / 298.1 K

Operator: mkl

File: 1203-hp-01-041b

INOVA-500 "riga"



Relax. delay 2.000 sec

Pulse 57.9 degrees

Acq. time 1.892 sec

Width 8000.0 Hz

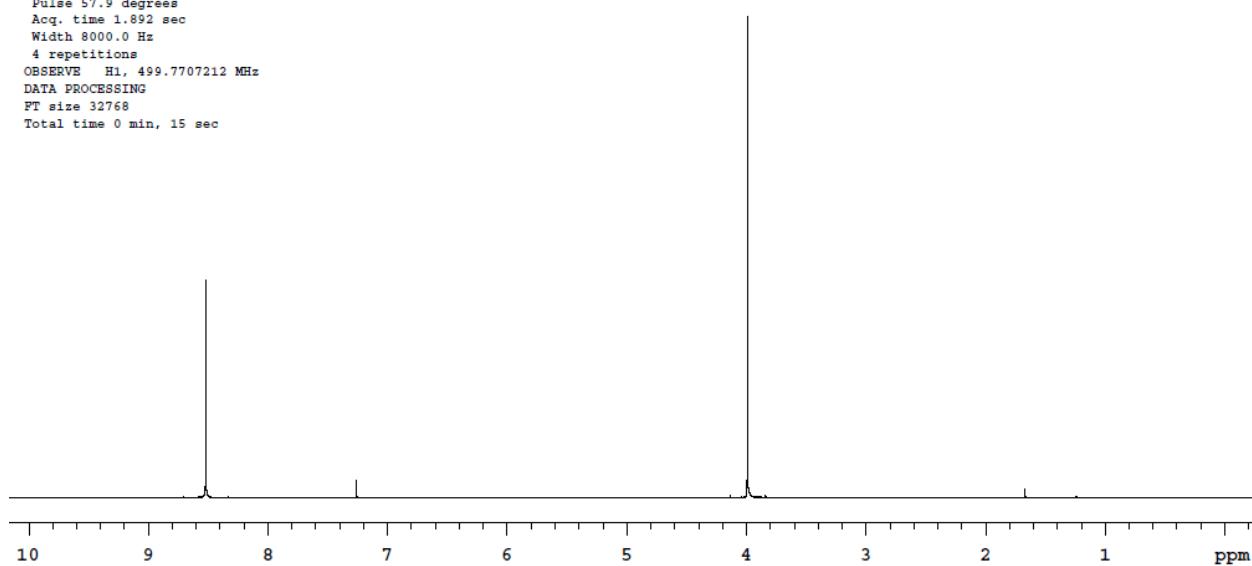
4 repetitions

OBSERVE H1, 499.7707212 MHz

DATA PROCESSING

FT size 32768

Total time 0 min, 15 sec



1203-hp-01-042

Pulse Sequence: s2pul

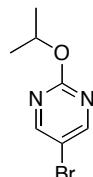
Solvent: CDCl₃

Temp. 25.0 C / 298.1 K

Operator: mkl

File: 1203-hp-01-042

INOVA-500 "riga"



Relax. delay 2.000 sec

Pulse 57.9 degrees

Acq. time 1.892 sec

Width 8000.0 Hz

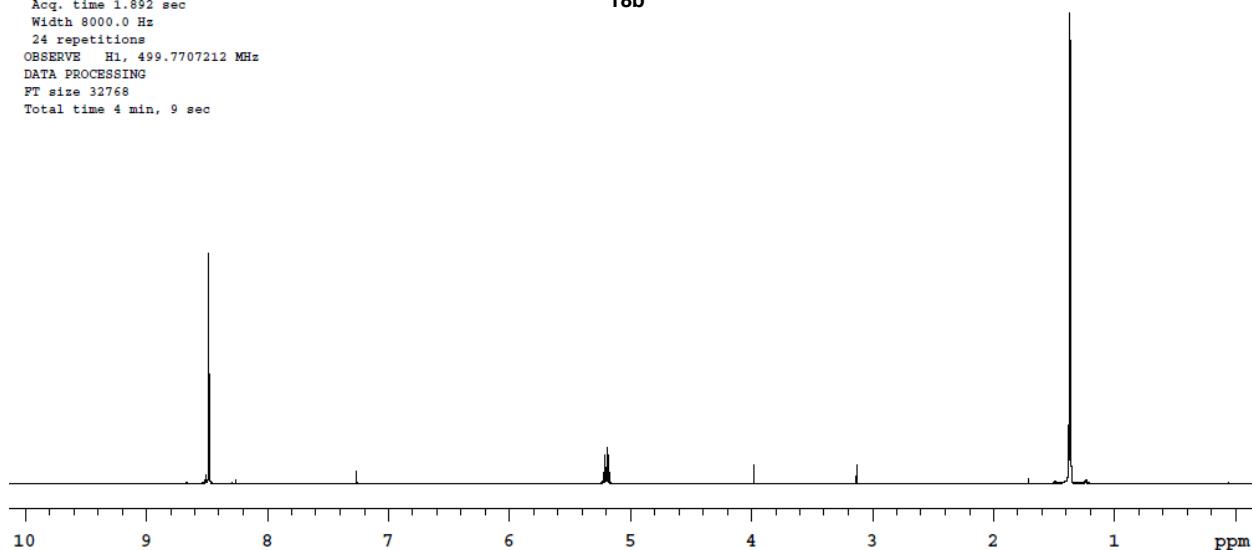
24 repetitions

OBSERVE H1, 499.7707212 MHz

DATA PROCESSING

FT size 32768

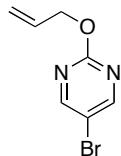
Total time 4 min, 9 sec



1203-hp-01-043

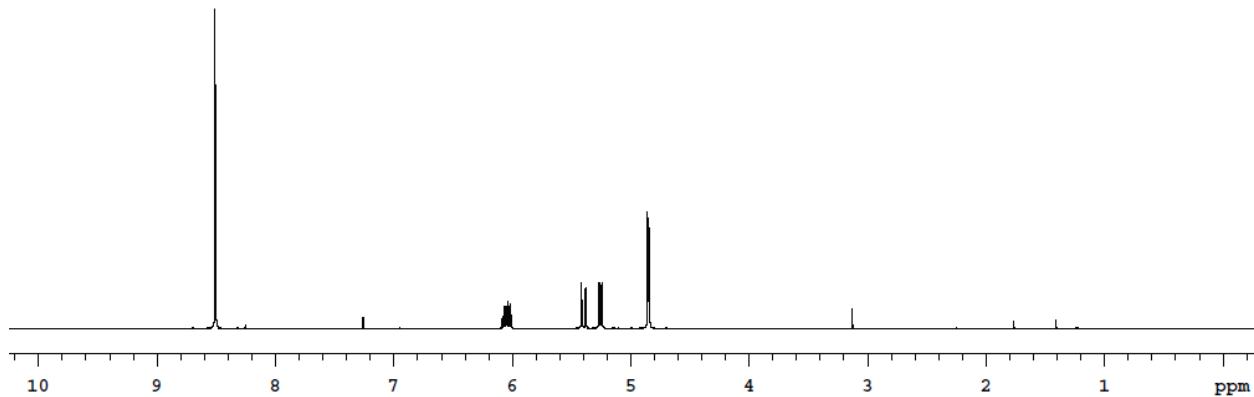
Pulse Sequence: s2pul

Solvent: CDCl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-hp-01-043a
INOVA-500 "riga"



18c

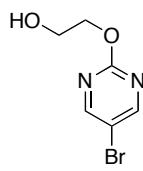
Relax. delay 1.500 sec
Pulse 57.9 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
32 repetitions
OBSERVE H1, 499.7707217 MHz
DATA PROCESSING
FT size 32768
Total time 3 min, 37 sec



1203-HP-01044b1

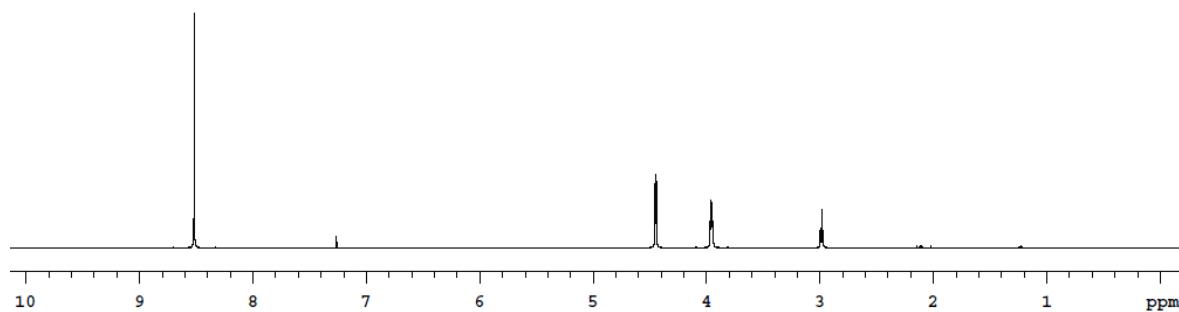
Pulse Sequence: s2pul

Solvent: cdcl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01044b1
INOVA-500 "riga"



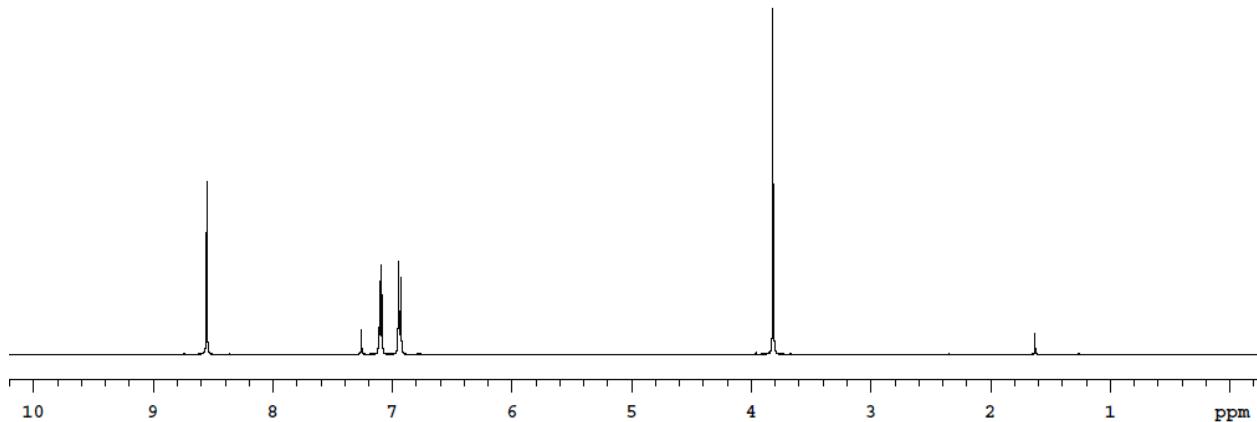
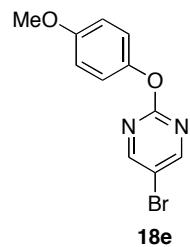
18d

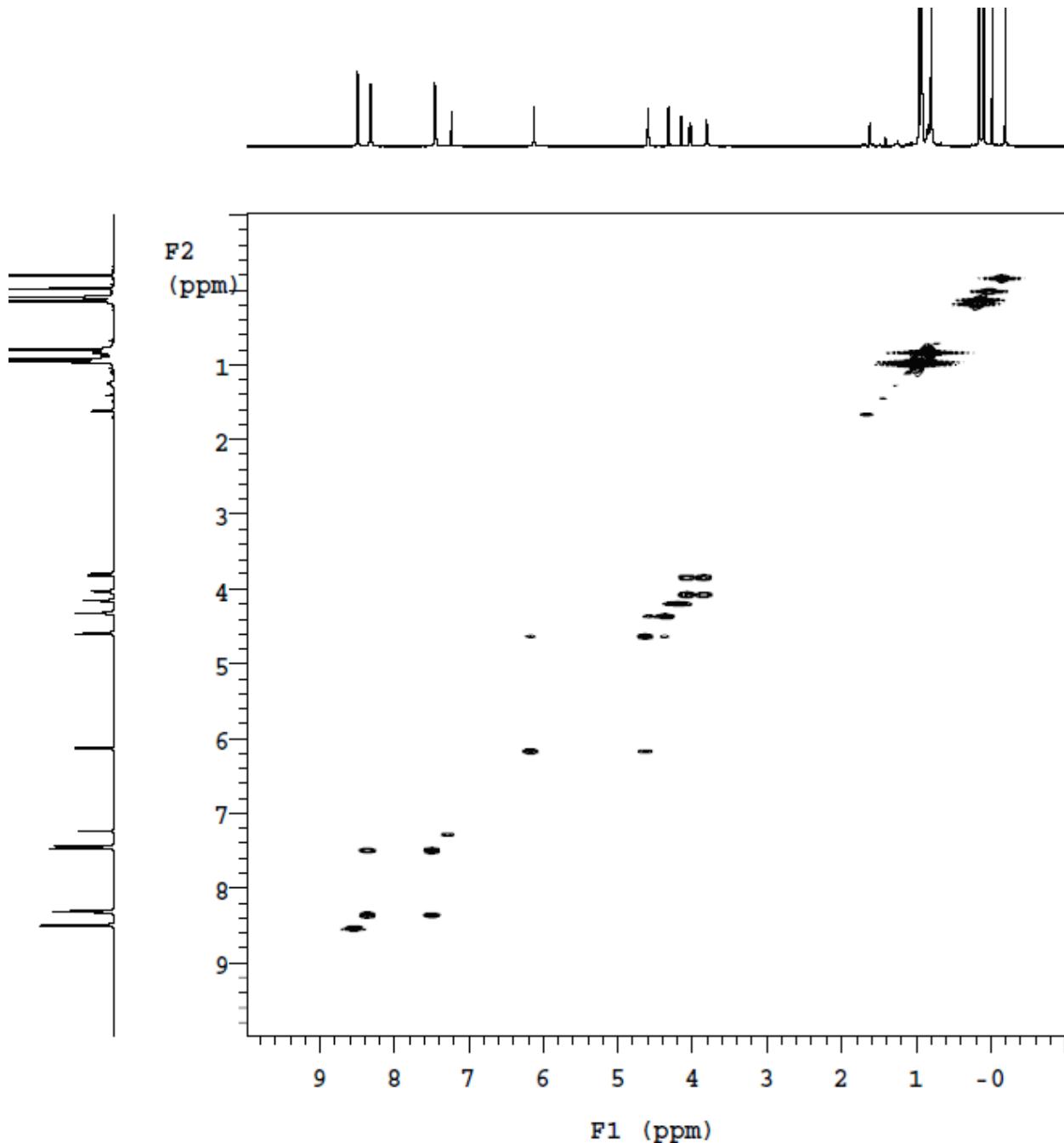
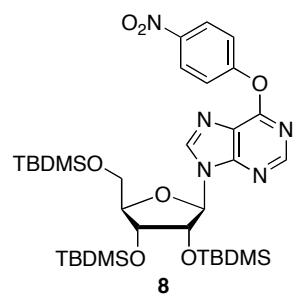
Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
32 repetitions
OBSERVE H1, 499.7707217 MHz
DATA PROCESSING
FT size 32768
Total time 2 min, 4 sec

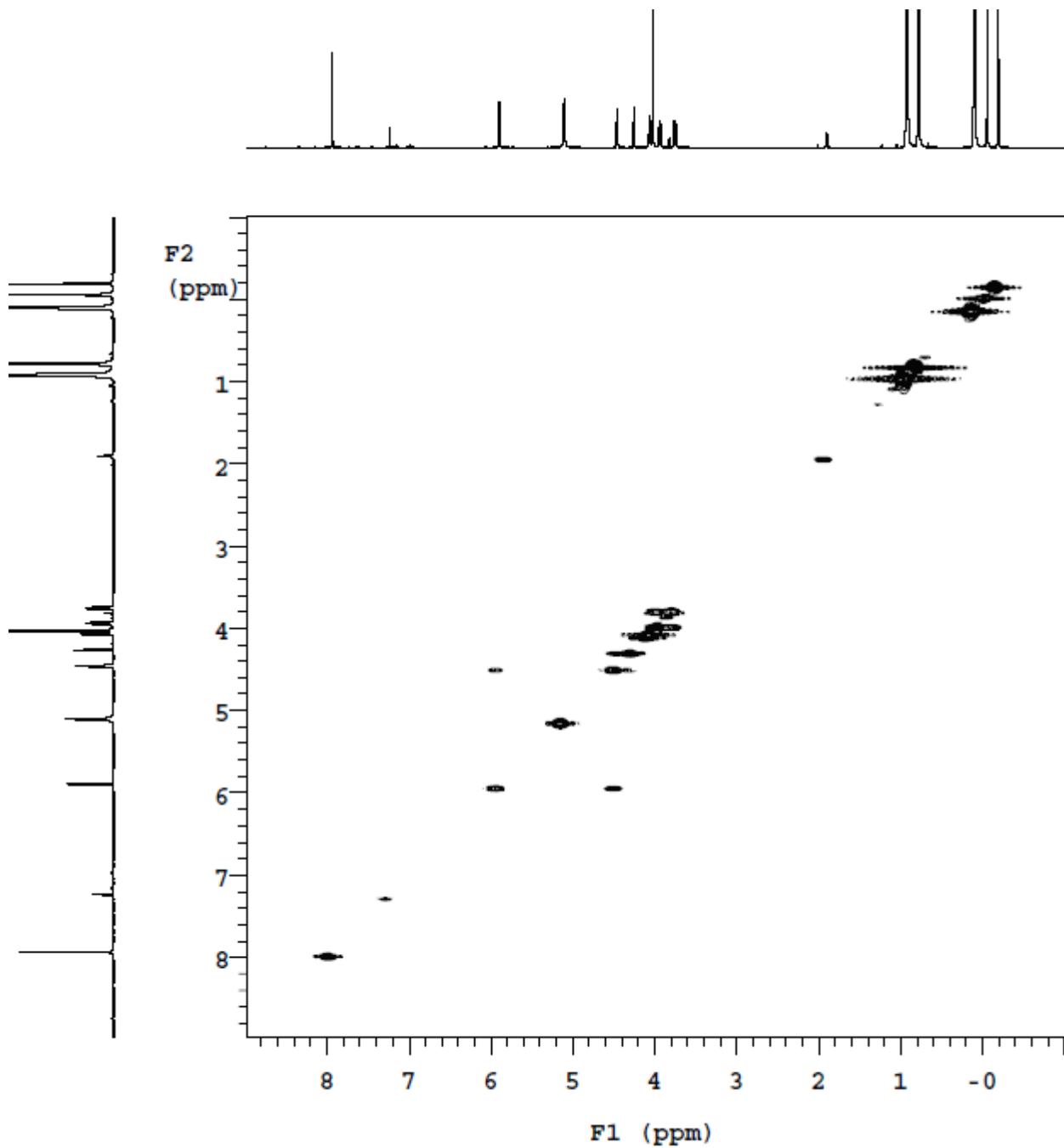
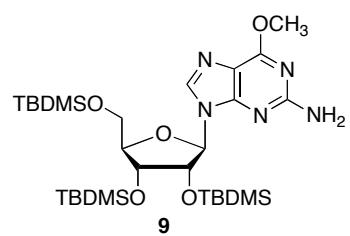


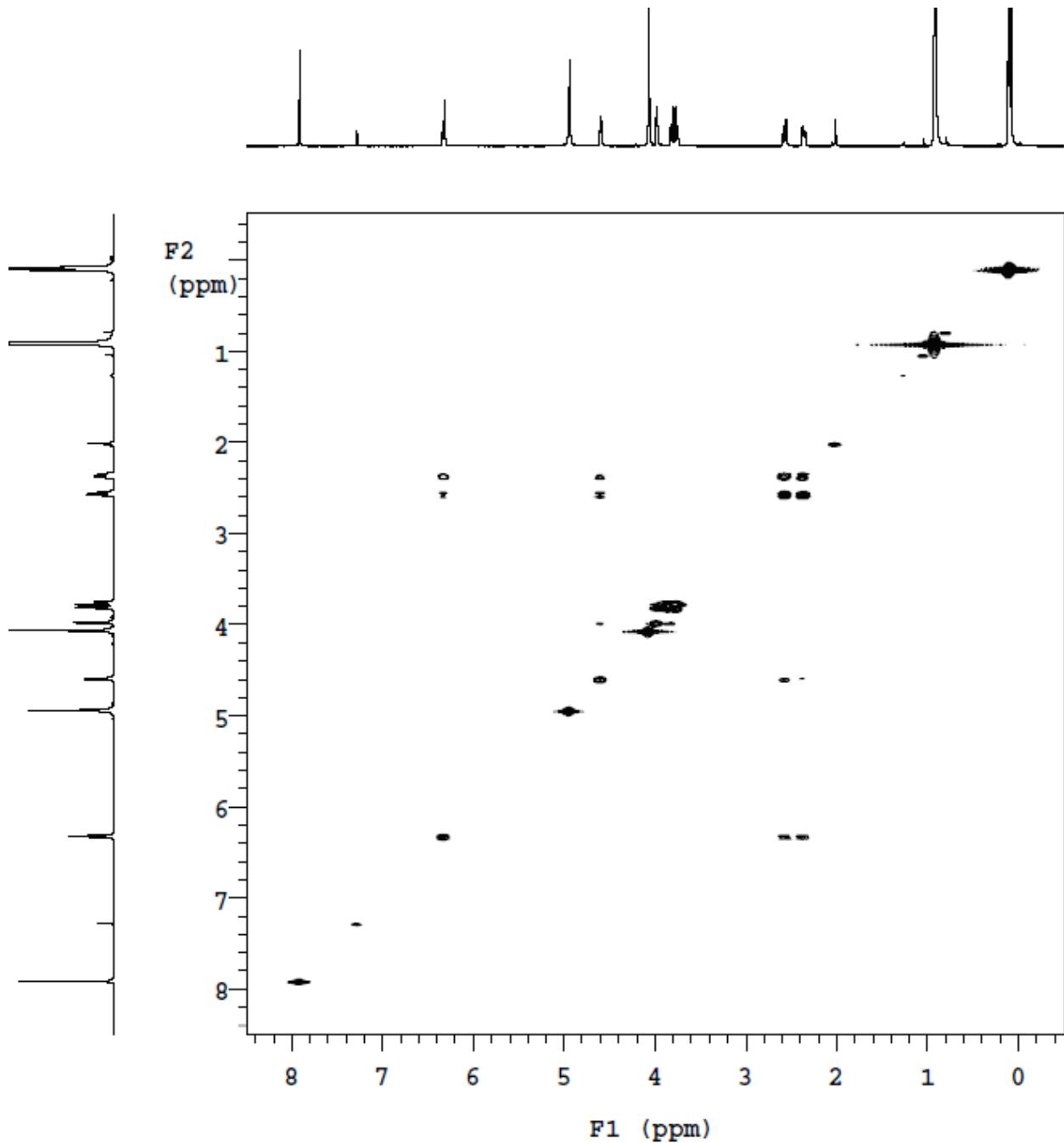
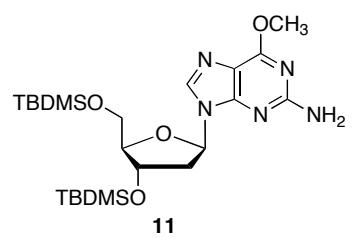
1203-HP-01-061-1h
Pulse Sequence: s2pul
Solvent: cdcl₃
Temp. 25.0 C / 298.1 K
Operator: mkl
File: 1203-HP-01-061-1h
INOVA-500 "riga"

Relax. delay 2.000 sec
Pulse 45.0 degrees
Acq. time 1.892 sec
Width 8000.0 Hz
48 repetitions
OBSERVE H1, 499.7707217 MHz
DATA PROCESSING
FT size 32768
Total time 5 min, 43 sec









References

- (1) Bae, S.; Lakshman, M. K. *J. Am. Chem. Soc.* **2007**, *129*, 782–789.
- (2) Lakshman, M. K.; Frank, J. *Org. Biomol. Chem.* **2009**, *7*, 2933–2940.
- (3) Štefane, B.; Polanc, S. *Synlett* **2008**, 1279–1282.
- (4) Bogert, M. T.; May, C. E. *J. Am. Chem. Soc.* **1909**, *31*, 507–513.
- (5) Adachi, K. *Yakugaku Zasshi* **1955**, *75*, 1426–1429.
- (6) Ranganathan, D.; Rathi, R.; Keshavan, K.; Singh, W. P. *Tetrahedron* **1986**, *42*, 4873–4878.
- (7) Mantell, S. J.; Gibson, K. R.; Osborne, S. A.; Maw, G. N.; Rees, H.; Dodd, P. G.; Greener, B.; Harbottle, G. W.; Million, W. A.; Poinsard, C.; England, S.; Carnell, P.; Betts, A. M.; Monhemuis, R.; Prime, R. L. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 2190–2194.
- (8) Miyashita, A.; Matsuda, H.; Lijima, C.; Higashino, T. *Chem. Pharm. Bull.* **1992**, *40*, 43–48.
- (9) Wan, Z.-K.; Wacharasindhu, S.; Levins, C. G.; Lin, M.; Tabei, K.; Mansour, T. S. *J. Org. Chem.* **2007**, *72*, 10194–10210.
- (10) Marshall, J. R.; Walker, J. *J. Chem. Soc.* **1951**, 1004–1017.
- (11) Nara, S. J.; Jha, M.; Brinkhorst, J.; Zemanek, T. J.; Derek A. Pratt, D. A. *J. Org. Chem.* **2008**, *73*, 9326–9333.
- (12) Arukwe, J.; Keilen, G.; Undheim, K. *Acta Chem. Scand.* **1988**, *B42*, 530–536.
- (13) Falck-Pedersen, M. L.; Benneche, T.; Undheim, K. *Acta Chem. Scand.* **1989**, *43*, 251–258.
- (14) Bardhan, S.; Wacharasindhu, S.; Wan, Z.-K.; Mansour, T. S. *Org. Lett.* **2009**, *11*, 2511–2514.