Fig. S1: Multiple sequence aligment of TrmIs ordered with decreased *ClustalW* score compared to $_{Pab}$ TrmI. This figure was drawn with *ESPript* (1). Putative TrmIs most similar to $_{Pab}$ TrmI and belonging to different taxonomic groups are indicated together with TrmIs with known structure. The consensus sequence motifs are indicated underneath. The secondary structural elements of $_{Pab}$ TrmI are shown above. The cysteine residues forming the intermolecular disulfide bonds between two different monomers and Histidine at position 78, which are present only in the *Pyrococcus* and *Thermoccus* species, are labeled with a star.

Fig. S2: Stereo view of the SAH binding sites in Crystal Forms I and II. A Fobs-Fcalc electron density map omitting SAH calculated at the level of 1.5 standard deviation is superimposed on the structure. **A** In Crystal Form II, the dihedral angle (C4', C5', S, C γ) of SAH is -74°. **B** The folded conformation observed in Crystal form I, molecules C and D. The dihedral angle (C4', C5', S, C γ) of SAH is -152°. **C** The folded conformation observed in Crystal form I, molecules A and B. The dihedral angle (C4', C5', S, C γ) of SAH is -170°.

Fig. S3: A Nucleotide sequence of $_{Pab}tRNA^{Asp}$. **B** $_{Pab}TrmI$ and $_{Tt}TrmI$ modify positions 57 and/or 58 in $_{Pab}tRNA^{Asp}$ but not position 59. Purified enzymes were incubated in the presence of SAM with [α - ^{32}P]ATP or [α - ^{32}P]UTP labeled T7 transcript of $_{Pab}tRNA^{Asp}$ for one hour at 70°C. The resulting [^{32}P]-labeled tRNA was hydrolyzed by nuclease P1 or T2 and the generated nucleosides were analyzed by 2D-TLC on cellulose plates and autoradiography. The nucleosides were quantified by scintillation counting.

Fig. S4: Time course for the dimethylation of *P. abyssi* tRNA^{Asp} by wild-type _{Pab}TrmI (triangles) and the H78Y mutant (circles). An equimolar amount of tRNA and enzyme was incubated in the presence of SAM at 70°C for different incubation times and was then digested with RNase A. The digest was analyzed by MALDI mass spectrometry (Fig. 6). The area of the peak corresponding to the dimethylated fragment m¹Am¹AAUp at m/z 1340.22 was compared to that of the peaks corresponding to the non methylated (m/z 1312.19) and monomethylated (m/z 1326.20) fragments to obtain the percentage of dimethylated tRNA.

1. Gouet, P., Courcelle, E., Stuart, D.I. and Metoz, F. (1999) ESPript: analysis of multiple sequence alignments in PostScript. *Bioinformatics*, **15**, 305-308.

Figure S1

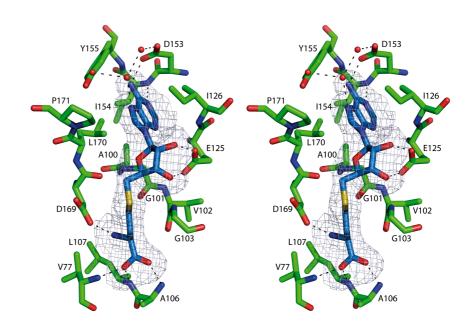
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M.thermautotrophicusstr.	TIEREĨEVRKO.	GTRPRT
D.turgidum	TLLRPWOIEGL	SVRDFH
M.maripaludis	CIVRDIEISEK	GVPPST
A.degensii	VIVRHWOVEGK	CVDDVU
T.maritima	SLFRPYKPVPE	. SVRPIN
S.tokodaii		
	LILREYQIKEN.	
T.acidaminovorans	ILLRYLKTDPR	
S.flavogriseus	SMIRNWHVEGL.	
D.vulgaris	ILIRRWKPVAD	
D.thiodismutans	ILVRRYKPVPE	
M.tuberculosis	TLQRGWNVVGL	
C.Korarchaeum	ILDREYESDER	. R <mark>TRP</mark> AP
T.thermophilus	VGWREWEVRLP	.V <mark>AHP</mark> RF
H.volcanii	TIQRQMDFNDR	. G <mark>SR</mark> PST
H.sapiens	VIVRDWLVCLAKQKNGILAQKVESKINTDVQLDSQEKIGVKGELFQEDDHEESHSDFPYGSFP	YV <mark>AR</mark> PVH
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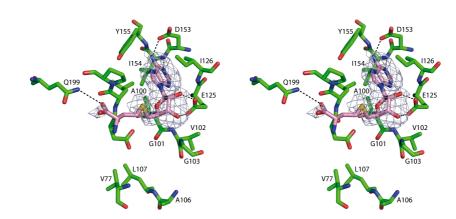
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