Supplemental Material:

RNA extraction, cDNA synthesis and real-time quantitative polymerase chain reaction (PCR)

Total RNA was extracted with TRIzol reagent (Invitrogen, Grand Island, NY) as per the manufacture's instructions. RNA integrity, quantity, and genomic DNA contamination were assessed using the Agilent Bioanalyzer 2100 and RNA 6000 Labchip kit (Agilent Technologies, Palo Alto, CA). Only those RNA samples with 28S/18S ratios between 1.5 to 2 and no DNA contamination were studied further. RNA samples (2μg) were reverse-transcribed to cDNA using the First Strand cDNA Synthase Kit (MBI Fermentas, Hanover, MD) with random hexamer primer. Real-time quantitative PCR was performed on an iCycler detection system (Bio-Rad, CA). PCR was performed in a 25μl volume using SYBR green Supermix (Bio-Rad, Hercules, CA). Amplification conditions were: 95ºC for 3 min, 50 cycles of 95ºC for 15s, 60ºC for 15s, and 72ºC for 20s. The fold-changes in mRNA expressions for targeted genes were relative to the respective vehicle groups of mice after normalization to 18s rRNA. We selected 18s rRNA for the internal standard in our real-time PCR, based on our preliminary studies showing that there were no significant differences in the 18s rRNA level among the different groups of samples (infected and uninfected). In addition, a search of the literature indicates that 18s rRNA is among the most common endogenous standards currently in use. Primer sequences were designed by using Beacon Designer 5.0 (Premier Biosoft International, Palo Alto, CA), and synthesized by the Biopolymer Laboratory of the University of Maryland.

Primer sequences

The primer sequences are listed on Table I (see supplementary material) or were described previously[14, 16].

Table 1. Primer sequences for real-time quantitative PCR.

Gene	Primer sequences (5' to 3')
F4/80	Forward, AAAGACTGGATTCTGGGAAGTTTGG
	Reverse, CGAGAGTGTTGTGGCAGGTTG
NOS-2	Forward, CGGAGCCTTTAGACCTCAACA
	Reverse, CCCTCGAAGGTGAGCTGAAC
CD206	Forward, TTTGGAATCAAGGGCACAGAG
	Reverse, TGCTCCACAATCCCGAACC
Arginase-I	Forward, CTGGCAGTTGGAAGCATCTCT
	Reverse, GTGAGCATCCACCCAAATGAC
Arginase-II	Forward, GGAACTGGCTGAAGTGGTTAGTAGA
	Reverse, GGGCGTGACCGATAATGGTA
FIZZ1	Forward, CCTCCACTGTAACGAAGACTCTC
	Reverse, GCAAAGCCACAAGCACACC
YM-1	Forward, ATCTATGCCTTTGCTGGAATGC
	Reverse, TGAATGAATATCTGACGGTTCTGAG
IFN-γ	Forward, GCATAGATGTGGAAGAAAAGAGTCTCT
	Reverse, TGGCTCTGCAGGATTTTCATG
IGF-1	Forward, CGTCTTCACACCTCTTCTACC
	Reverse, CGGTCCACACACGAACTG

Immunofluorescent staining

Tissue slides were fixed in cold acetone for 30 min and blocked with 10% normal rat serum in PBS for 1 hr at room temperature. The slides were incubated with anti-F4/80-Alexa647 (1:40, Caltag, Burlingame, CA) and anti-CD206-FITC (MR5D3; Serotek, Raleigh, NC) for 4 hr at room temperature. The slides were then cover-slipped with Vectorshield (Vector Laboratories, Burlingame, CA) and digitally photographed with a Nikon TE 2000-E microscope (Melville, NY) using MetaVue version 6.1 software (Universal Imaging Corporation, Downington, PA).