

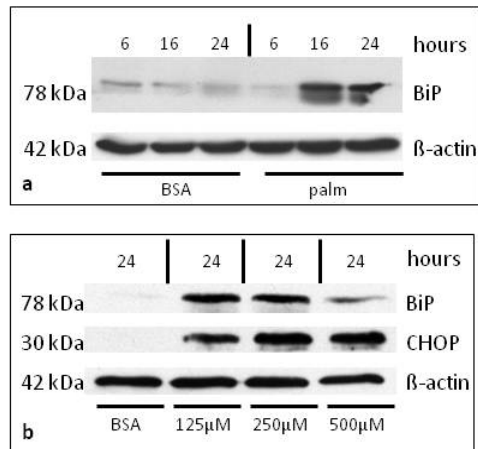
SUPPORTING MATERIAL

MATERIALS AND METHODS

Quantitative real-time PCR of renal biopsies - Human renal biopsy specimens were procured in an international multicenter study, the European Renal cDNA Bank-Kroener-Fresenius Biopsy bank (ERCB-KFB). Biopsies were obtained from patients after informed consent and with approval of the local ethics committees. Following renal biopsy, the tissue was transferred to RNase inhibitor and microdissected into glomerular and tubular fragments. Total RNA was isolated from micro-dissected glomeruli as described previously (1). Reverse transcription and real-time RT-PCR were performed as reported earlier (1). Pre-developed TaqMan reagents were used for human BiP (NM_005347.2), HYOU1 (NM_006389.2), CHOP (NM_004083.4), as well as the reference genes (Applied Biosystems). The expression of BiP, HYOU1 and CHOP was normalized to the mean of three reference genes, GAPDH, 18S rRNA, and synaptopodin. The mRNA expression was analyzed by standard curve quantification (4). For the real time RT-PCR data statistical analysis was performed using Kruskal-Wallis and Mann-Whitney U tests.

SUPPLEMENTAL RESULTS

SFig 1.: Dose and time-dependent induction of the ER-chaperons BiP and CHOP



A. Time-dependent upregulation of BiP protein levels by palmitic acid. β -actin serves as loading control. Representative results of 3 independent experiments. B. Upregulation of BiP and CHOP at 125 μ M, 250 μ M, and 500 μ M palmitic acid. For the control condition (BSA) the BSA concentration was equivalent to cells exposed to 500 μ M palmitic acid complexed to BSA. β -actin serves as loading control. Representative results of 2 independent experiments.

1. Cohen CD, Frach K, Schlondorff D, and Kretzler M. Quantitative gene expression analysis in renal biopsies: a novel protocol for a high-throughput multicenter application. *Kidney Int* 61: 133-140, 2002.
2. Lindenmeyer MT, Rastaldi MP, Ikehata M, Neusser MA, Kretzler M, Cohen CD, and Schlondorff D. Proteinuria and hyperglycemia induce endoplasmic reticulum stress. *J Am Soc Nephrol* 19: 2225-2236, 2008.
3. Liu G, Sun Y, Li Z, Song T, Wang H, Zhang Y, and Ge Z. Apoptosis induced by endoplasmic reticulum stress involved in diabetic kidney disease. *Biochemical and biophysical research communications* 370: 651-656, 2008.
4. Schmid H, Henger A, Cohen CD, Frach K, Grone HJ, Schlondorff D, and Kretzler M. Gene expression profiles of podocyte-associated molecules as diagnostic markers in acquired proteinuric diseases. *J Am Soc Nephrol* 14: 2958-2966, 2003.
5. Vandivier RW, Henson PM, and Douglas IS. Burying the dead: the impact of failed apoptotic cell removal (efferocytosis) on chronic inflammatory lung disease. *Chest* 129: 1673-1682, 2006.
6. Wu J, Zhang R, Torreggiani M, Ting A, Xiong H, Striker GE, Vlassara H, and Zheng F. Induction of diabetes in aged C57B6 mice results in severe nephropathy: an association with oxidative stress, endoplasmic reticulum stress, and inflammation. *Am J Pathol* 176: 2163-2176.