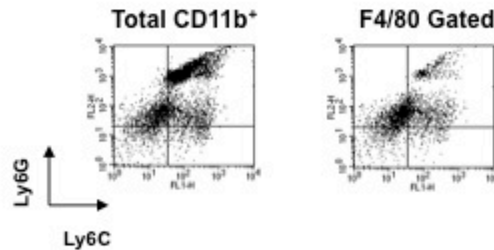
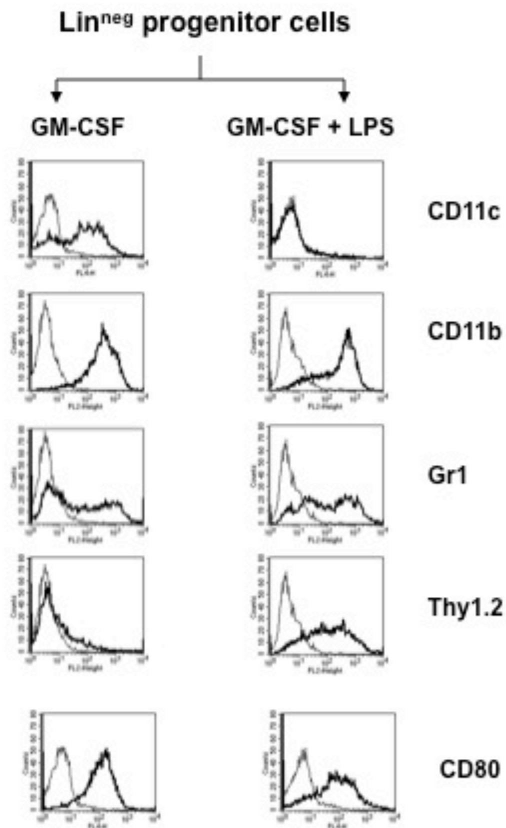
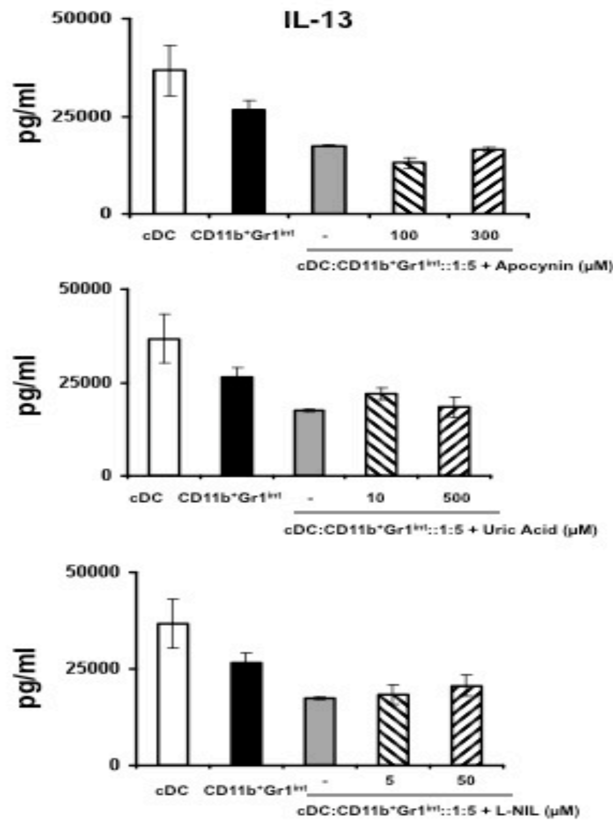
SUPPLEMENTARY MATERIAL



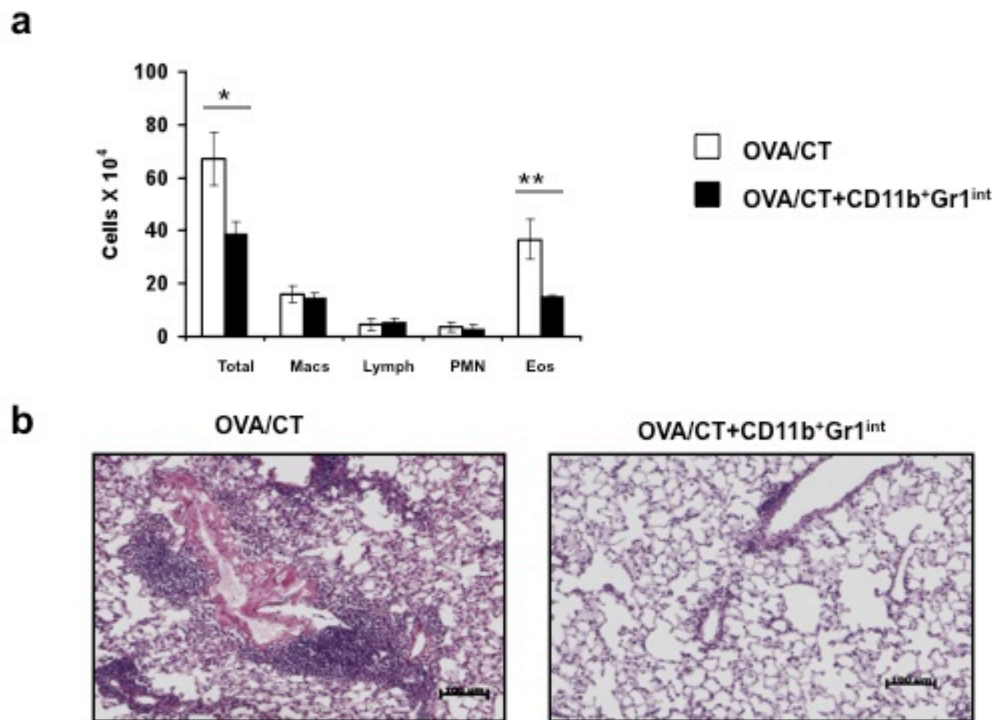
Supplementary Figure 1 LPS administration induces CD11b^{+}Ly6G^{+}Ly6C^{-}F4/80^{+} and CD11b^{+}Ly6G^{+}Ly6C^{+}F4/80^{+} subsets of cells. Cells from the lungs of LPS-treated mice were purified based upon CD11b expression by magnetic bead selection, were stained with Ly6C, Ly6G and F4/80, and were analyzed by flow cytometry. The forward versus side light scatter pattern revealed a distinct non-lymphocytic population of cells where CD11b- and Gr1-expressing cells were concentrated. This population of cells was gated upon for subsequent analyses. The left panel shows the expression profile of Ly6G and Ly6C expression on CD11b^{+} cells. The right panel depicts the expression profile of Ly6G and Ly6C expression on CD11b+ cells gated on F4/80 expression. F4/80 expression was largely confined to CD11b^{+}Ly6G^{int} cells. Among the CD11b^{+} cells gated on F4/80 expression, Ly6G^{int}Ly6C^{-} was the dominant population expanded by LPS treatment with the Ly6G^{int}Ly6C^{lo} population being the minor subset.



Supplementary Figure 2 LPS treatment generates $CD11b^{+}Gr1^{int}$ cells from lin^{neg} progenitor cells. The lin^{neg} population of bone marrow progenitor cells was enriched using a lineage depletion kit and phenotypic analysis was performed by flow cytometry. The lin^{neg} population of bone marrow progenitor cells was cultured with GM-CSF (10 ng/ml) in the absence or presence of LPS (1µg/ml) for 9 days and phenotypic analysis was performed. These results are representative of at least 3 independent experiments.



Supplementary Figure 3 Suppression of Th2 cell response by $CD11b^{+}Gr1^{int}$ cells induced by LPS administration does not involve nitric oxide. Th2 cells were generated *in vitro* using $CD4^{+}$ T cells from DO11.10 mice by incubation under Th2-skewing conditions for 6 days. cDCs and $CD11b^{+}Gr1^{int}$ cells were isolated from the lungs of LPS-treated mice. cDCs and $CD11b^{+}Gr1^{int}$ cells alone (each at 1 X 10^{5} cells/well) or in combination as shown were cultured with Th2 polarized DO11.10 $CD4^{+}$ T cells (1 X 10^{6} cells/well), OVA peptide (5 µg/ml) and various inhibitors at indicated concentrations. Culture supernatants were collected 36 hrs later and analyzed by multiplex cytokine assay for IL-13 levels.



Supplementary Figure 4 Suppression of eosinophilic airway inflammation by CD11b+Gr1^{int} cells isolated from LPS-treated lungs. Mice were antigen-sensitized by three daily consecutive intranasal treatments with OVA plus cholera toxin (CT) followed by 5 d of rest. CD11b^{+}Gr1^{int} cells isolated from the lungs of LPS-treated mice were then adoptively transferred intratracheally (1 x 10^{6} cells/mouse) into mice that had received OVA/CT. Control mice did not receive any cells. Mice were then challenged with aerosolized OVA daily for 7 d. Total and differential cell counts in the BAL fluid.