Supplemental Material

Single Stranded DNA oligonucleotides for construction of IAPP gene:

Wild Type IAPP Forward primer (IAPP-FOR) with NcoI restriction site.

 $$\rm M$$ G 5'- GTGGCTGAGA<u>CCATGG</u>GC AAA TGC AAC ACC GCG ACC TGC GCC ACC CAG CGT CTG GCG AAC TTT CTG GTG CAT AGC AGC AAC AAC

Wild Type IAPP Reverse primer (IAPP-Rev) with BamH I restriction site.

5'- GACGCACCGGATCC ATA GGT GTT GCT GCC CAC GTT GGT GCT GCT CAG AAT CGC GCC AAA GTT GTT GCT GCT ATG CAC CAG

Primers for PCR amplification of full-length IAPP gene:

Forward: 5'-GTGGCTGAGA<u>CCATGG</u>GC AAA Reverse: 5'-GACGCACCGGATCC ATA GGT GTT

Primers for Random Mutagenesis:

Forward Primer: 5'- CTTTAATAAGGAGATATACC**ATG**GGC -3' Reverse Primer: 5'-GCG GAG CCA GCG GAT CC -3'

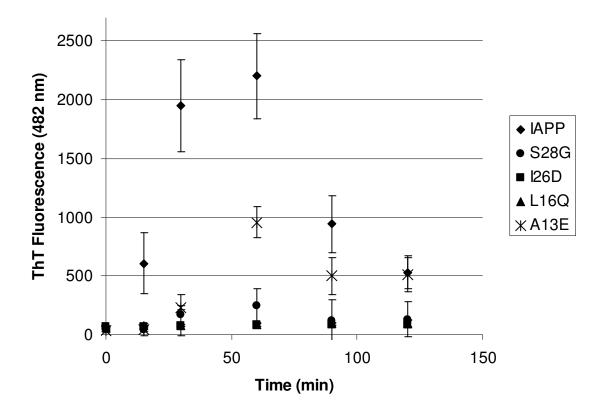


Figure 1S: Comparison of thioflavin T binding time course by selected variants and wild-type human IAPP. One representative time experiment is shown with the error bars representing the standard deviation of all trials ($n \ge 4$). Each aliquot of disaggregated IAPP was thawed and the HFIP removed over a stream of dry nitrogen gas. The resulting solid was dissolved in 20 mM tris buffer pH 7.40 to 106 μ M. The samples were incubated at 37°C with shaking (200 rpm). 17 μ L of each sample was removed at indicated time points and mixed with 663 μ L of 12.0 μ M thioflavin T in 20 mM Tris buffer pH 7.40. The thioflavin T mixture was incubated at room temperature in the dark for 5 minutes before recording the thioflavin T fluorescence spectrum (Ex_{450nm}) using a Hitatchi F-7000 fluorescence spectrophotometer.