Table S1. Oligonucleotide sequences used in this work

siRNA		
ANGPTL4 siRNA	sense	5'-AAAGCTGCAAGATGACCTCAGATGGAGGCTG– 3'
	anti-sense	5'-AAAAGGCTTAAGAAGGGAATCTTCTGGAAGAC-3'
Control siRNA	sense	5'-AAAGCTGTCTTCAAGATTGATATCGAAGACTA-3'
	anti-sense	5'-AAAATAGTCTTCGATATCAAGCTTGAAGACA-3'
Real-time qPCR[a]		
Human ANGPTL4	forward	5'-CTCCCGTTAGCCCCTGAGAG-3'
	reverse	5'-AGGTGCTGCTTCTCCAGGTG-3'
	Taqman probe	5'-(6-FAM)ACCCTGAGGTCCTTCACAGCCTGC(TAMRA)-3'
Mouse ANGPTL4	Forward	5'-GCTTTGCATCCTGGGACGAG-3'
	Reverse	5'-CCCTGACAAGCGTTACCACAG-3'
	Taqman probe	5'-(6-FAM)ACTTGCTGGCTCACGGGCTGCTAC(TAMRA)-3'
L27	Forward	5'- CTGGTGGCTGGAATTGACCGCTA -3'
	Reverse	5'- CAAGGGGATATCCACAGAGTACCTTG -3'
	Taqman probe	5'-(HEX)CTGCCATGGGCAAGAAGAAGATCGCC(BHQ1)-3'

[a] Melting curve analysis was performed to assure that only one PCR product was formed. Primers were designed to generate a PCR amplification product of 100 to 250 bp. Only primer pairs yielding unique amplification products without primer dimer formation were subsequently used for real-time PCR assays.